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Review article

# Unveiling the multifaceted pathogenesis and therapeutic drugs of Alzheimer's disease: A comprehensive review

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# ABSTRACT

Alzheimer's disease (AD) is a severe neurodegenerative disorder characterized by the accumulation of β-amyloid (Aβ) plaques and tau phosphorylation-induced neurofibrillary tangles. This review comprehensively summarizes AD pathogenesis and related factors, drawing on a wealth of authoritative reports and research findings. Specifically, we delve into the intricate mechanisms underlying AD pathology, including Aβ deposition, tau protein phosphorylation, cholinergic dysfunction, neuroinflammation, mitochondrial oxidative stress, ferroptosis, imbalance in the gut microbiota, and microRNA dysregulation. We also explored the effects of these factors on the brain, including synaptic damage and cognitive impairment. Moreover, our review highlights the associations between the pathogenesis of AD and inflammatory cytokines in the peripheral blood and cerebrospinal fluid, dysbiosis of the gut microbiota, and changes in microRNA expression. Overall, we provided a systematic and illustrative overview of the pathogenesis and therapeutic drugs for AD, offering help in the prevention and treatment of this condition.

# **1. Introduction**

With global economic and medical advancements, population aging is becoming increasingly evident, making dementia in older adults a significant challenge. Among the various types of dementia, Alzheimer's disease (AD) is the most prevalent. The hallmark features in patients with AD are β-amyloid (Aβ) deposition, neurofibrillary tangles (NFTs), and the loss of neurons and synapses in the brain. The 2018 World Health Organization (WHO) report on AD highlights the global crisis posed by dementia, estimating 50 million patients worldwide in 2018. Projections indicate that this number will continue to rise, reaching 82 million by 2030, imposing a significant healthcare burden on a global scale [\[1\]](#page-10-0). Consequently, an increasing number of researchers are investigating the etiological mechanisms of AD [\[2\]](#page-10-0). AD is named after Alois Alzheimer, the neurologist who first discoverd it [[3](#page-10-0)]. His discovery paved the way for further research into the pathogenesis of AD, with the initial identification of Aβ plaques as a contributing factor [[4](#page-11-0)]. Subsequently,

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<span id="page-1-0"></span>Iqbal et al. isolated NFTs from the brains of patients with AD [[5](#page-11-0)]. As research has progressed, an increasing number of hypotheses regarding AD have been proposed. Extensive research has revealed that the pathogenesis of AD is associated with Aβ deposition [\[6\]](#page-11-0), tau protein phosphorylation [[7](#page-11-0)], cholinergic dysfunction [[8](#page-11-0)], neuroinflammation [\[9\]](#page-11-0), mitochondrial oxidative stress [\[10](#page-11-0)], ferroptosis [\[11](#page-11-0)], an imbalance in the gut microbiota [[12\]](#page-11-0), and alterations in microRNA (miRNA) expression [[6](#page-11-0)]. These factors can interact with and exacerbate each other, further accelerating AD progression. This review aims to organize the multiple mechanisms and medications for AD to provide insights into its prevention and treatment.

# **2. Molecular mechanisms of AD**

# *2.1. Aβ*

There are two hallmark pathological features in the periphery of hippocampal neurons in patients with AD, one of which is Aβ deposition, primarily formed by smaller fragments of amyloid precursor protein (APP). APP generates Aβ peptides of different lengths after successively undergoing β-secretase and γ-secretase [\[13](#page-11-0)]. Aβ1-40 and Aβ1-42 are particularly neurotoxic and are frequently associated with AD [\[14](#page-11-0)]. The in-depth understanding of the process of secretase-induced Aβ formation and the mechanism of Aβ damage to neurons provides more theoretical basis for the future development of Aβ-targeted drugs for the treatment of AD.

### *2.1.1. Role of secretory enzymes on Aβ formation*

β-secretase 1 (BACE1), an aspartyl protease predominantly found in brain neurons, is known for cleaving APP to produce Aβ [\[13](#page-11-0), [15\]](#page-11-0). Scientists have studied BACE1 in various aspects, including the genes that regulate BACE1 and its post-translational modifications [\[16](#page-11-0)]. Bao et al. found that both the activity and stability of BACE1 were enhanced following SUMOylation at residue K501 [\[17](#page-11-0)]. The stability-enhanced BACE1 is not easily degraded by the lysosome, which can promote APP shearing, leading to Aβ accumulation [[17\]](#page-11-0). BACE2, which shares 64 % amino acid similarity with BACE1, is predominantly expressed in peripheral tissues [\[18,19](#page-11-0)]. However, its level is increased in the brains of patients with AD and has been regarded as a marker that is highly correlated with AD. Moreover, the



**Fig. 1.** Tau phosphorylated sites and related kinases. (A) The adult full-length tau protein has 85 potential phosphorylation sites, including 45 serine, 35 threonine, and 5 tyrosine residues. The phosphorylation sites shown in the figure are associated with the development of AD. (B) Aβ oligomers bind to the receptor, which then upregulates E2F1. This, in turn, induces the expression of PAX6 and c-Myb. PAX6 and c-Myb can directly regulate the transcription of GSK-3β, thereby promoting tau phosphorylation. CDK5 also promotes tau phosphorylation, and CDK5 is regulated by p35, p39, and p25. In addition, tau interacts with Nup98, leading to defects in nucleocytoplasmic transport.

amount and activity of BACE2 are related to BACE1 levels in the brain [\[20](#page-11-0)]. Thus, BACE2 was initially assumed to have a similar function to BACE1 in cleaving APP at the β-site to produce Aβ. However, later research questioned this idea and proposed that BACE2 may play a protective role for neurons [\[21](#page-11-0),[22\]](#page-11-0). Evidence indicates that elevated levels of BACE2 do not exacerbate the deposition of Aβ40 and Aβ42 but rather reduce them [\[23](#page-11-0)]. However, Wang Z et al. has found that under specific conditions, BACE2 is able to cleave APP at the β-site [\[2\]](#page-10-0). Hence, scientists hold different views on the role of BACE2 in Aβ formation and AD processes. We are currently unable to clearly determine whether BACE2 promotes or inhibits the production of Aβ, possibly due to varying experimental conditions. Therefore, its specific mechanism of action remains to be studied.

#### *2.1.2. Aβ is aggressive to nerve cells*

Research suggests that Aβ impairs neuronal cell function and reduces cell viability through multiple pathways.

Shankar et al. revealed that Aβ oligomers impaired learning and memory abilities in rats [[24\]](#page-11-0). Evidence indicates that Aβ oligomers significantly increased long-term depression, and the density of dendritic spines was decreased around the rodent hippocampus [[24\]](#page-11-0). Clinical trials revealed that removing amyloid plaques does not improve symptoms of AD, whereas inhibiting Aβ oligomers resulted in better clinical outcomes, so Aβ oligomers are considered the more neurotoxic forms [[25\]](#page-11-0). Moreover, Aβ oligomers are duplicated in astrocytes over time and induce neural damage [[26\]](#page-11-0). Teng et al. discovered that the number of the C-terminal fragment of the synaptic adhesion protein N-cadherin amplifies the synaptotoxic effects of Aβ [\[27](#page-11-0)]. Furthermore, a delay in synaptic vesicle endocytosis promotes the binding of Aβ oligomers to synaptic vesicle membranes and facilitates their internalization, resulting in synaptic impairment. Aβ not only affects hippocampal neurons but also causes injury to microglia. Microglia are mononuclear macrophages of the central nervous system (CNS) that relate to the growth and development of neuronal cells [[24\]](#page-11-0). Baron has found that the microglia surrounding the Aβ acquired the specifically activated phenotype, accompanied by a light increase in cytokines [[25\]](#page-11-0). Additionally, Liu Q et al. discovered that Aβ aggregates up-regulate the expression and function of α7 nicotinic acetylcholine receptor (nAChR), leading to decreased cell viability [\[23](#page-11-0)].

#### *2.2. Tau phosphorylation*

Tau-phosphorylated proteins, commonly found in the mammalian brain, are primarily located in axons, and regulate axonal transport [\[28](#page-11-0)]. The C-terminal domain of tau comprises four repeating regions: R1, R2, R3, and R4 [\[29](#page-11-0)]. Phosphorylated tau (p-tau) is a key factor in the formation of NFTs. We examined both the phosphorylation sites of tau and the effects of p-tau on neuronal structure and function to understand the role of phosphorylated tau in AD pathogenesis.

#### *2.2.1. Tau phosphorylated sites involved in AD*

Tau protein has 85 sites potentially linked to phosphorylation [\[30](#page-11-0)]. At these sites, approximately 40 residues are phosphorylated, which are considered high-risk factors for AD [[31\]](#page-11-0).

Aberrant phosphorylation of Ser289 and Ser293 significantly destabilizes microtubules and promotes tau accumulation by inducing structural changes in the monomeric R2 peptides [\(Fig. 1A](#page-1-0)) [\[30](#page-11-0)]. In the monomer, Ser289 phosphorylation enhances ordered-disordered structural transitions and intramolecular interactions, resulting in a more compact phosphorylated R2. In contrast, in dimers, phosphorylation of Ser289 promotes β-fold formation, which can lead to the oligomerization of R2 peptides. Oligomerization of tau during aggregation is the most neurotoxic form, inducing neuroinflammatory factors, binding to astrocytes and microglia, and triggering apoptosis [\[32](#page-11-0)].

Furthermore, a lot of tau phosphorylation sites involved in AD, including Ser199, Ser214, and Ser231, are reported [\(Fig. 1A](#page-1-0)) [\[33](#page-11-0)–36]. However, our current understanding of these sites is limited by their association with tau phosphorylation, and investigating their upstream and downstream mechanisms and interactions poses challenges for future research.

#### *2.2.2. Kinase promoting tau phosphorylation*

Phosphorylation of tau relies on the action of phosphorylases, and an in-depth examination of the function of phosphorylases contributes to a further understanding of the pathogenesis of AD and provides a crucial theoretical basis for targeting p-tau for the treatment of AD.

Glycogen synthase kinase-3β (GSK-3β) is an important phosphorylating enzyme that affects axonal transport function and rapidly inhibits mitochondrial and neurotrophic factor receptor TrkA movement [[37,38\]](#page-11-0). Moreover, Singh T et al. revealed that tau proteins need to be preprocessed by CDK-5 before GSK-3β [\[39](#page-11-0)]. Additionally, Aβ can enhance tau phosphorylation by mediating GSK-3β and CDK-5, particularly GSK-3β [\(Fig. 1](#page-1-0)B). Recent findings by Yalun Zhang suggest that the upstream pathway leading to GSK-3β-induced tau phosphorylation is regulated by multiple factors [[40\]](#page-11-0). Aβ upregulates E2F1, which in turn induces the expression of PAX6 and c-Myb. Both PAX6 and c-Myb are direct targets of E2F1 ([Fig. 1](#page-1-0)B). PAX6 directly regulates GSK-3β transcription and induces tau phosphorylation at Ser356, Ser396, and Ser404 [[40\]](#page-11-0).

#### *2.2.3. P-tau promotes neuronal nuclear envelope damage*

While the sites and kinases associated with tau phosphorylation are known, the mechanism through which p-tau damages neurons and induces AD remains unclear. The nuclear envelope (NE) is vital for maintaining a stable nuclear environment. The nuclear pore complex, located on the NE, facilitates the exchange of materials and information between the cytoplasm and nucleus [\(Fig. 1](#page-1-0)B). Lisa Diez et al. suggests that the accumulation of p-tau near the neuronal NE impairs the transport of substances within and outside the nucleus [[41\]](#page-12-0). Further research has demonstrated that tau interacts with phenylalanine-glycine (FG)-rich nucleoporins 98 (Nup98), forming aggregates in the nuclear pores and obstructing material exchange [\[41\]](#page-12-0). Impaired transport of substances within the nucleus may contribute to cellular dysfunction and death, which are closely associated with AD [\[42](#page-12-0)].

# **3. Systemic factors in AD**

# *3.1. Cholinergic dysfunction*

Acetylcholine (ACh), the first neurotransmitter discovered in humans, is crucial for neuronal function. The cholinergic hypothesis suggests that cognitive impairment causes the destruction of cholinergic neurons. Cholinergic neurons participate in various physiological functions of the brain, including attention and memory [\[43](#page-12-0)]. Three key factors affect the action of ACh: (1) its synthesis; (2) binding of ACh to its receptor; and (3) degradation of ACh by acetylcholinesterase (AChE).

# *3.1.1. ACh synthesis disorder*

In cholinergic neuronal terminals, ACh is formed from choline and acetyl coenzyme A (acetyl-CoA) by the enzyme choline acetyltransferase (ChAT) (Fig. 2) and plays an important role in transporting information between reportors [\[44\]](#page-12-0). ACh is a product of the pyruvate produced during the tricarboxylic acid (TCA) cycle. However, the deposition of Aβ significantly reduces the efficiency of pyruvate participation in the TCA cycle [[45\]](#page-12-0), leading to decreased production of ACh and ATP. Excess glutamate causes postsynaptic  $Ca<sup>2+</sup>$ , activating pyruvate dehydrogenase kinase, which inhibits pyruvate dehydrogenase complex and compromises mitochondrial



**Fig. 2.** ACh plays an important role in the nervous system and influences memory and recognition. **(1)** Synthesis of ACh. Acetyl-CoA and choline are the raw materials for the synthesis of ACh, and most of the ACh comes from the TCA cycle. However, Aβ reduces pyruvate utilization, thus leading to a reduction in ACh production during the TCA cycle.  $Ca^{2+}$  overload can activate pyruvate dehydrogenase kinase (PDHK), which inhibits pyruvate dehydrogenase complex (PDHC) and ultimately affects the TCA cycle and production of ACh. **(2)** Receptors for ACh. ACh is transported out of the axonal region by VAChT and then combined with muscarinic or nicotinic receptors. This process plays an important role in information transfer. However, Aβ binds α7nAChR, thereby reducing ACh binding to α7nAChR. AF2671 (M1 mAChR) increased α-secretase and decreased Aβ synthesis. Meanwhile, AF2671 decreased the level of tau by inhibiting the activation of GSK-3β. (Image created using Biorender.com.)

activity [\[46,47](#page-12-0)]. According to the cholinergic hypothesis, cognitive decline in patients with AD is the result of a combination of ACh and ATP deficiencies.

# *3.1.2. Impaired ACh receptor function and impaired transport*

ACh activates two types of receptors: muscarinic and nicotinic. Among the five known isoforms of muscarinic ACh receptors (mAChRs), M1 mAChR has the highest distribution in the CNS and is involved in numerous brain activities [\[48](#page-12-0),[49\]](#page-12-0). Stimulation of AF267B (an M1 mAChR agonist) increases α-secretase activity and decreases Aβ synthesis [[50,51](#page-12-0)]. Additionally, stimulation of AF267B inhibits GSK-3β activity, resulting in reduced tau levels. These studies suggest that the activation state of M1 mAChR affects brain cognitive functions by regulating Aβ and tau. The neuronal nAChR consists of α and β subunits, which combine to form a pentameric receptor complex [\[52](#page-12-0)]. In the CNS, most nicotinic receptors are expressed on the membranes of presynaptic neurons, and their main role is to regulate the release of neurotransmitters [[43,53](#page-12-0)–55] such as glutamate, GABA, dopamine, 5-hydroxytryptamine, norepinephrine, and ACh [56–[61\]](#page-12-0). Moreover, Aβ demonstrates a strong affinity for α7nAChRs [\[62](#page-12-0)], which results in compromised ACh transport. In addition, Aβ interacts with cholinergic components in various ways, including impairing cholinergic function, inhibiting the activity of ChAT and ACh, and increasing AChE activity [[63\]](#page-12-0). Aβ and p-tau can not only directly damage neurons but also indirectly impair neuronal function by disrupting the cholinergic system.

# *3.2. Neuroinflammation*

Neuroinflammation is the inflammation of the CNS, which is regulated by the involvement of neuroglia, including microglia and astrocytes [\[9\]](#page-11-0). The activation of microglia and astrocytes contributes to neuronal growth, development, and repair; however, prolonged activation induces neuroinflammation, impairs the ability of glial cells to clear amyloids, and exacerbates neurodegenerative diseases.

#### *3.2.1. Microglia-related neuroinflammation*

Microglia, the brain's innate immune cells [[64\]](#page-12-0) and about 10 % of the cells in the nervous system [\[65](#page-12-0)]. Microglia are divided into resting (M0) and activated (M1 and M2) states and play different roles in the brain. M0 microglia are highly branched, constantly moving through synaptic stretches, and act as the CNS [[66\]](#page-12-0). After being activated by pathogens and Aβ, M0 microglia differentiate through the regulation of damage-associated molecular patterns or pathogen-associated molecular patterns [[67,68](#page-12-0)]. M1 microglia, activated by Toll-like receptors or interferon-gamma, lead to neuronal damage and impaired phagocytosis through the production of large amounts of NO, reactive oxygen species (ROS), interleukin (IL)-1β, IL-6, IL-18, and TNF-α [\[69,70](#page-12-0)]. In contrast, M2 microglia is activated by IL-4 or IL-13, which exerts reparative effects and phagocytosis by releasing anti-inflammatory factors, including transforming growth factor-β, IL-4, IL-10, and IL-13 [[69,70](#page-12-0)]. However, the M1/M2 phenotypic nomenclature is not widely recognized, mainly because it crudely divides microglia into two types.

Recent studies have revealed that the microglial activation phenotype extends far beyond the M1/M2 phenotype. Analysis of all immune cells (CD45<sup>+</sup>) derived from the brains of 5XFAD mice using massive parallel single-cell RNA-seq revealed two clusters of microglia expressing unique genes, including apolipoprotein E (ApoE), lipoprotein lipase, and Cystatin F, and defined microglia as disease-associated microglia (DAM) [\[71](#page-12-0)]. DAM aggregates near Aβ and exhibits phagocytic activity [[71](#page-12-0)]. To date, a variety of microglia surface receptors have been found to interact with Aβ, including scavenger receptors, TLR, CD36, RAGE, TREM 2, and late glycosylation end product receptors [[72,73\]](#page-12-0). In addition to classical HLA-DR upregulation, recently identified microglia activation markers include F4/80, CD68, CD45, and ionized calcium-binding adapter molecule 1 [[66,74\]](#page-12-0).

#### *3.2.2. Astrocytes-related neuroinflammation*

Astrocytes are involved in the regulation of blood-brain barrier (BBB) stability, secretion of neurotrophic factors, and modulation of synaptic function and plasticity [[66,75](#page-12-0)], and play an important role in the maintenance of homeostasis in the CNS. Astrocytes stimulated by pathogens or Aβ can develop two phenotypes with completely opposite activation states, the neurotoxic A1 phenotype (A1 astrocytes) and the neuroprotective A2 phenotype (A2 astrocytes) [\[76](#page-12-0)–78]. Activated microglia induce A1 astrocytes by IL-1α, TNF, and C1q, resulting in the loss of the ability of A1 astrocytes to promote neuronal survival, growth, synaptogenesis, and phagocytosis [\[79](#page-13-0)]. In the brain of patients with AD, A1 astrocytes highly express glial cell acidic protein, S100 calcium-binding protein B (S100B), and complement C3 (C3) [\[78](#page-12-0)]. S100B is a cytokine that activates cyclooxygenase-2 in microglia by upregulating nitric oxide synthase, ultimately leading to neuronal death [[64](#page-12-0)]. Astrocytes extensively release inflammatory cytokines, including IL-1, IL-6, and TNF-α, when activated by pathogens or Aβ, which can have beneficial or harmful consequences [[80](#page-13-0)]. Additionally, since astrocytes express BACE and presenilin-1, cytokine stimulation may induce  $\mathbb{A}\beta$  production in astrocytes [\[64](#page-12-0)].

In AD, astrocytes that specifically express ApoE are thought to have a role in degrading Aβ [\[81](#page-13-0)]. A variety of receptors exist on the surface of astrocytes that can bind to Aβ, including the receptor for advanced glycosylation end products, lipoprotein receptor-associated proteins, membrane-associated proteoglycans, and scavenger receptor-like receptors [\[82,83](#page-13-0)]. Therefore, we know that the phagocytosis of glial cells is very important for the protection of the CNS.

#### *3.3. Mitochondrial oxidative stress*

Mitochondria produce ATP through oxidative phosphorylation (OXPHOS), which plays a key role in maintaining endogenous neuroprotective and repair mechanisms [\[84](#page-13-0)]. However, with age, mitochondria accumulate oxidative damage, leading to <span id="page-5-0"></span>neurodegeneration, impaired synaptic plasticity, and associated cognitive deficits [[84\]](#page-13-0). An in-depth investigation of the relationship between mitochondrial functional impairment and neurons can help further understand AD pathogenesis and provide a more comprehensive theoretical basis for the future development of AD therapeutic drugs.

The severe imbalance between ROS and reactive nitrogen species production and oxidative stress induces antioxidant defenses, which are significantly increased in the brains of patients with AD and occur earlier than Aβ accumulation [[85\]](#page-13-0). The cell membrane contains more unsaturated fatty acids, whereas the catalase content of neurons is low; therefore, the brain is more prone to oxidative damage [\[86,87](#page-13-0)]. Recently, studies have shown that Aβ can insert into cellular membranes, leading to the generation of ROS and the occurrence of lipid peroxidation in the membranes [\[10](#page-11-0)[,88](#page-13-0)]. Even Aβ aggregation at the mitochondrial membrane disrupts mitochondrial transport, leading to a decrease in OXPHOS enzyme activity and a decrease in the transmembrane electrochemical gradient, which impairs mitochondrial function and increases ROS levels [[89,90\]](#page-13-0). In tau-knockout mice, ROS are significantly reduced, mitochondrial fusion is increased, mitochondrial permeability transition pore and cyclophilin D is inhibited, and ATP production is increased, suggesting that tau overexpression can cause aberrant mitochondrial fusion and oxidative stress [\[91,92](#page-13-0)]. Tau inhibits the binding of dynamin-related protein 1 to mitochondria by binding to the actin cytoskeleton, resulting in mitochondrial elongation and increased fusion [[86\]](#page-13-0). Mitochondria provide energy to support membrane ion exchange and synaptic transmission, and maintain Ca<sup>2+</sup> homeostasis [\[86,90](#page-13-0)]. When mitochondria take up excess  $Ca^{2+}$ , it impairs mitochondrial function, leading to elevated ROS levels, inducing Aβ deposition and tau phosphorylation, further damaging neurons [\[93](#page-13-0)].



**Fig. 3.** Gut microbes activate C/EBPβ/AEP signaling, elevating pro-inflammatory enzymes and resulting in increasing levels of APOE4, APP, Aβ, and Tau. These metabolites can be released from the intestinal epithelium into the peripheral bloodstream. exacerbating the inflammatory response, including neuroinflammation. Bile acids may increase BBB permeability and allow intestinal metabolites to enter the CNS, leading to inflammation and AD. (Image created using Figdraw).

#### *3.4. Ferroptosis*

Iron is a trace element in the body that is involved in the regulation of many physiological activities. Fe $^{3+}$  and Fe $^{2+}$  are storage and transport forms, respectively, in the CNS. However, under conditions of iron overload or excessive free iron, the equilibrium of the antioxidant system is disrupted [\[94,95](#page-13-0)], leading to oxidative stress and neuronal cell death [\[96,97](#page-13-0)].

Normally, ferroportin 1 acts as a remote regulator of intracellular iron homeostasis and transports excess ferritin outside the cells [\[98](#page-13-0)]. However, nuclear receptor coactivator 4 degrades ferritin to free iron, and excessive intracellular iron accumulation can lead to elevated ROS levels and oxidative stress, promoting cellular ferroptosis [[98,99](#page-13-0)], leading to neuronal cell damage. Iron transporter protein 1 (Fpn-1) is the only known non-heme iron transporter protein in mammals [\[100\]](#page-13-0). Its primary function is to regulate systemic iron homeostasis by binding to transferrin and transporting iron to iron-demanding tissues. Bao observed morphological and molecular features indicative of ferroptosis in Fpnfl/fl/NEXcre and AD mice [\[100\]](#page-13-0). Fpn deficiency in the hippocampus is accompanied by AD-like phenomena, including brain atrophy and memory deficits.

NADPH oxidase 4 is a major source of ROS and induces ferroptosis by impairing mitochondrial function in astrocytes [\[101\]](#page-13-0). Philip et al. found that white matter-degenerating microglia are enriched in the iron-binding protein light chain ferritin, which accumulates lipid droplets and undergoes peroxidative damage  $[102]$ . GPX4 is indispensable for the reduction of  $H_2O_2$  and is the only enzyme capable of reducing phospholipid hydroperoxides. The loss of GPX4 activity is an important cause of lipid peroxide formation and accelerates ROS production, ultimately inducing ferroptosis in cells [\[103,104\]](#page-13-0). Recent research suggests that ferroptosis is not independently involved in the pathological response to AD; it also forms a complex network of linkages with p-tau to participate in AD. This can occur via Cys-Cys binding or hyperphosphorylation of tau proteins via the kinase pathway and ferroptosis [[105](#page-13-0)].

The concept of ferroptosis was first introduced 10 years ago by Dr. Brent R. Stockwell and is related to the development of diverse diseases, including AD. However, ferroptosis remains a relatively new area of research, and its study in the context of AD has a large scope.

#### *3.5. Imbalance of intestinal flora*

The human gut contains 10–100 trillion commensal microbial cells [\[106\]](#page-13-0). Gut microorganisms secrete neurotransmitters, neu-romodulators, and other amino acid-derived metabolites [[107](#page-13-0),[108\]](#page-13-0). Indeed, both the microbes and the synthetics released have the potential to cause an inflammatory response or accelerate amyloid formation, leading to impaired memory and cognitive functioning. Therefore, maintaining a balance between intestinal microorganisms is important.

#### *3.5.1. Gut microbial metabolites and inflammation*

Several genetic changes in gut microbes are associated with CCAAT-enhancer-binding protein (C/EBPβ)/asparagine endopeptidase (AEP) signaling, which increases levels of pro-inflammatory enzymes associated with polyunsaturated fatty acid metabolism [\(Fig. 3](#page-5-0)) [\[12](#page-11-0),[109](#page-13-0)]. In addition, the activation of C/EBPβ signaling would further elevate AEP, resulting in increasing levels of APOE4, APP, Aβ, and tau [\[109\]](#page-13-0), eventually inducing inflammation and cognitive impairment. The gut flora and the gut form a microbial gut axis that communicates bidirectionally through cytokines, hormones, and neural signals ([Fig. 3\)](#page-5-0) [[110](#page-13-0)]. Furthermore, the diversity of gut microorganisms is strongly linked to the oral flora [[111,112\]](#page-13-0). Periodontitis greatly increases the number and variety of oral pathogens, such as lipopolysaccharides (LPS), flagellins, peptides, as well as pro-inflammatory molecules. If these molecules overactivate systemic inflammatory responses, they may lead to neuroinflammation and AD [\[113\]](#page-13-0).

#### *3.5.2. Gut-microbe-brain axis*

Intercommunication between the gut flora and brain was first introduced by William James and Carl Lange in the 1880s [\[114\]](#page-13-0). Evidence indicates that the microbiota continuously produces LPS and amyloids in healthy individuals. When these substances accumulate to a certain level in the body, they can be harmful, particularly when the permeability of the gut-blood-brain barrier undergoes significant changes in adult individuals  $[110,115]$  $[110,115]$  $[110,115]$  $[110,115]$ . When tight junctions between gut cells are compromised, permeation is enhanced and these tight junctions are unable to prevent LPS from entering the circulation, exacerbating the inflammatory response [\(Fig. 3\)](#page-5-0) [\[116](#page-13-0)]. Intraperitoneal administration of LPS promotes the production of inflammatory factors and Αβ in the mice hippocampus. Additionally, mice develop cognitive dysfunction [[117](#page-13-0)]. LPS activates microglia surface receptors through myeloid differentiation factor 88 and nuclear factor-kappa beta (NF-κB)-dependent signaling pathways, thereby promoting cytokine [\(Fig. 3](#page-5-0)) and chemokine production [[118,119\]](#page-14-0). Extracellular LPS has also been reported to trigger microglial NOD-like receptor protein 3 inflammasome activation, increasing the level of ROS and IL-1β [[120\]](#page-14-0), which may be related to AD. In conclusion, the intestinal secretion of LPS induces an inflammatory response by binding to multiple neuronal cell receptors and promoting AD.

Gut microbes also produce bile acids (BAs) [[121](#page-14-0)]. Increased bacterially produced BAs may increase BBB permeability and allow peripheral cholesterol to reach the CNS by disrupting tight junctions. This refers to the allowance of BAs, or cholesterol, from the periphery for entry into the CNS [\[122\]](#page-14-0). Cellular cholesterol in the brain can directly bind to APP, contributing to the insertion of APP into the lipid raft phospholipid monolayers that form Aβ and ultimately promoting Aβ production [\[123\]](#page-14-0). Microbial amyloid and Aβ42 are also recognized by the TLR2/TLR1 receptor, which activates the production of inflammatory factors. Microglia play an important role in preventing Aβ damage to neurons in healthy humans. Normally, activated microglia surround Aβ and prevent its diffusion, thus reducing the binding of Aβ to nearby neurons and mitigating neuronal damage [[124](#page-14-0)]. Moreover, maintenance of intestinal microbial homeostasis is a prerequisite for the maturation and function of microglia, thereby protecting neurons from Aβ damage [\[125\]](#page-14-0).

In summary, compounds produced and secreted by bacteria induce a systemic inflammatory response, increase BBB permeability,

<span id="page-7-0"></span>and ultimately contribute to the development of neurodegenerative diseases, including AD [\[126,127](#page-14-0)].

#### *3.6. miRNA dysregulation*

miRNAs are usually dysregulated in the brains of patients with AD; they regulate neuronal growth, synapse formation, and plasticity.

## *3.6.1. miRNA dysregulation leads to Aβ deposition*

Research has shown that miR-409-5p [[128](#page-14-0)] and miR-30b [[129](#page-14-0)] are upregulated in the brains of patients with AD. These miRNAs are thought to be involved in the process of AD by regulating Aβ formation. The overexpression of miR-409-5p had deleterious effects on neurite growth, reduced neuronal survival, and accelerated Aβ accumulation [\[128\]](#page-14-0). Song et al. provided the first evidence that miR-30b strongly upregulates ephrin type-B receptor 2 (ephB2), sirtuin1 (sirt1), and glutamate receptor subunit 2 (GluA2) to protect synapse integrity [[129](#page-14-0)]. Their further research found that Aβ42 and cytokines promote miR-30b overexpression through NF-κB and then impair synaptic integrity by downregulating ephB2, sirt1, and GluA2, ultimately resulting in AD. In addition, Long J.M. et al. identified a novel miRNA region, miR-346, which up-regulates APP by targeting the 5′-untranslated region (UTR) of APP mRNA [\[130\]](#page-14-0).

Additionally, miRNAs regulate Aβ levels by controlling the expression of BACE1 and APP. Wang et al. discovered that specific cortical regions implicated in AD pathology showed decreased neuronal miR-107 expression and increased BACE1 levels [[131](#page-14-0)]. The 3′-UTR of BACE1 mRNA is targeted by miR-107, promoting the production of Aβ and contributing to the development of AD. In addition to BACE1, miRNAs can influence Aβ production by modulating α-secretase. α-Secretase ADAM10 inhibits Aβ formation, while miR-144 promotes AD progression by suppressing ADAM10 production through its 3′-UTR [\[132\]](#page-14-0).

#### *3.6.2. miRNA dysregulation leads to tau phosphorylation*

Tau is a critical marker of AD and, several miRNAs regulate tau phosphorylation. In vivo, the expression of miR-125b contributes to tau phosphorylation, and Banzhaf-Strathmann found that miR-125b inhibits the expression of DUSP6, PPP1CA, and Bcl-W [\[133\]](#page-14-0). Downregulation of phosphatases and Bcl-W is accompanied by tau phosphorylation. Signals involved in tau phosphorylation, including p35, CDK5, and p44/42-MAPK, were upregulated [\[133\]](#page-14-0). Additionally, the deletion of miR-132 increases tau phosphorylation by activating ERK1/2 and BACE1 and upregulating inositol 1,4,5-trisphosphate 3-kinase B [[134](#page-14-0)]. Downregulation of miR-34a

#### **Table 1**

miRNAs associated with AD**.**



**Abbreviation:** Ple, pleckstrin; Sdcbp2, syndecan Binding Protein 2; UTR, untranslated region; EphB2, ephrin type-B receptor 2; Sirt1, sirtuin1; GluA2, glutamate ionotropic receptor AMPA type subunit 2; APP, amyloid-β peptide precursor protein; BACE1, β-site APP cleaving enzyme 1; DUSP6, dualspecific phosphatase 6; PPP1CA, protein phosphatase 1 catalytic subunit alpha isoform; Bcl-W, Bcl-2-like protein 2; 15-LOX, 15-lipoxygenase; mRNAs, VDR messenger RNAs; ITPKB, inositol 1,4,5-trisphosphate 3-kinase B; CDK5, Cyclin-dependent Kinase 5; mdh2, mitochondrial tricarboxylic acid cycle gene; Sqstm1, Sequestosome 1; Optn, Optineurin; PTPN1, tyrosine-protein phosphatase non-receptor type 1; HEY2, split (Hes)-related with YRPW motif protein 2.

<span id="page-8-0"></span>[\[135\]](#page-14-0) and miR-106b [[136](#page-14-0)] is also involved in tau phosphorylation, resulting in cognitive impairment.

In addition to Aβ and tau phosphorylation, miRNAs can also regulate various AD processes, such as mitochondrial function, enzyme, and microbial activities. The relevant information is presented in [Table 1.](#page-7-0)

#### **4. Drugs for the treatment of Alzheimer's disease**

# *4.1. FDA-approved drugs in the past*

Over the past 20 years, only five drugs have been approved by the FDA for the treatment of AD, the most common of which are AChE inhibitors, including tacrine, donepezil, rivastigmine, galantamine, and an N-methyl-D-aspartic acid (NMDA) receptor antagonist, memantine. Tacrine was the first clinically approved cholinesterase inhibitor but was withdrawn because of its high hepatotoxicity [[146](#page-14-0)]. Donepezil demonstrated equal efficacy and a better safety profile than other AChE inhibitors [[147](#page-14-0)]. Memantine was the first drug approved by the FDA to treat moderate-to-severe AD in 2003 [\[148,149\]](#page-14-0). These drugs have a significant limitation in that they can only alleviate symptoms without preventing or slowing disease progression, and both classes of drugs are associated with serious side effects. As a result, the current phase of drug development has shifted to targeting the pathogenesis of AD, with the largest number of drugs targeting Aβ and Tau for the treatment of AD. In recent years, AD drugs approved by the Food and Drug Administration (FDA) include aducanumab, lecanemab, and donanemab. The details are presented in Table 2.

# *4.2. FDA-approved drugs in recent years*

# *4.2.1. Aducanumab*

Aducanumab, a monoclonal antibody to the Aβ protein, was approved by the FDA in 2021 [[150](#page-14-0)]. However, the aducanumab research process has not been completed. Aducanumab's two global Phase III clinical trials, code-named ENGAGE and EMERGE,

**Table 2** 

FDA-approved drugs in AD.				
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**Abbreviation:** THA, Tacrine; AchE, acetylcholinesterase; NMDA, N-Methyl-D-aspartic acid; CDR-SB score, Clinical Dementia Rating–Sum of Boxes score; MMSE, Mini-Mental State Examination; ADAS-Cog 13, Alzheimer's Disease Assessment Scale–Cognitive Subscale–13 items; ADCS-ADL-MCI, Alzheimer's Disease Cooperative Study-Activities of Daily Living Scale for Mild Cognitive Impairment; ARIA, amyloid-associated imaging abnormalities; ARIA-E, Amyloid-related imaging abnormalities due to edema/sulcal effusion; ARIA-H, Amyloid-related imaging abnormalities due to haemosiderin deposition; ADAS-cog14 score, 14-item cognitive subscale of the Alzheimer's Disease Assessment Scale; ADCOMS, Alzheimer's Disease Composite Score; iADRS, integrated Alzheimer Disease Rating Scale.

<span id="page-9-0"></span>showed different results. However, after analyzing a larger dataset, medical data statisticians found that aducanumab had a significant effect compared with the placebo. Clinical Dementia Rating-Sum of Boxes (CDR-SB) is the primary endpoint score, with higher scores indicating greater impairment. High-dose aducanumab (10 mg/kg target dose) reduced CDR-SB scores by 22 % at week 78 compared to the placebo group. Three other pre-specified secondary endpoints were as follows: an 18 % decrease in Minimum Mental State Examination, a 27 % decrease in Alzheimer's Disease Assessment Scale–Cognitive Subscale13, and a 40 % decrease in The Alzheimer's Disease Cooperative Study - Activities of Daily Living Scale for use in Mild Cognitive Impairment in the high-dose group relative to the placebo group. The details are presented in [Table 2.](#page-8-0)

# *4.2.2. Lecanemab*

Lecanemab, a humanized IgG1 monoclonal antibody that binds with high affinity to Aβ soluble protofibrils, is being tested in patients with early AD, which reverses the pathological progression of AD and delays the clinical course of the disease by removing toxic Aβ proteins from the brain. This study [[154\]](#page-14-0) showed a 27 % slowing of cognitive and memory function decline in patients after 18 months of treatment with lecanemab compared to placebo. Lecanemab became the first Aβ-targeted drug in history to be fully approved by the FDA for AD treatment. The details are presented in [Table 2](#page-8-0).

# *4.2.3. Donanemab*

Donanemab is an antibody drug that targets N3pG (modified β amyloid plaques), a subtype of amyloid [\[156\]](#page-15-0). Donanemab binds to and promotes the clearance of amyloid plaques. The results showed that donanemab significantly alleviated cognitive decline in patients with AD, with a more pronounced effect in patients with low-to-moderate tau levels. Patients with low-to-moderate tau levels had a 35 % decrease in the Integrated Alzheimer's Disease (AD) Rating Scale (iADRS) (p *<* 0.0001) and a 36 % decrease in CDR-SB scores (p *<* 0.0001) compared with 22 % and 29 % for all patients, respectively [[155\]](#page-14-0). The details are presented in [Table 2.](#page-8-0)

# *4.3. Common side effects of drugs*

Amyloid-related imaging abnormalities (ARIA) are prevalent side effects of current Aβ antibody drugs, which can manifest as brain edema or sulcal effusion (ARIA-E) or as hemosiderin deposits in the brain parenchyma (ARIA-H microhemorrhage) or on the pial surface (ARIA-H superficial siderosis) [\[157\]](#page-15-0). The most common side effects of aducanumab are ARIA-E, headache, cerebral microhaemorrhage (ARIA-H microhaemorrhage), nasopharyngitis, falls, localized superficial scurfing (ARIA-H superficial scurfing), and dizziness [[150](#page-14-0)]. In terms of safety, the incidence of cerebral edema and cerebral hemorrhage caused by lecanemab was relatively low compared to other comparable anti-Aβ drugs, at 12.6 % and 17.3 %, respectively [[154](#page-14-0)]. In addition, a higher incidence of ARIA was found in patients who were homozygous for the ApoE ε4 allele after receiving lecanemab, and lecanemab should therefore be used with caution [[154](#page-14-0)]. Of the patients treated with donanemab, 24 % developed ARIA-E and 31 % developed ARIA-H, most of which were mild to moderate. The details are presented in [Table 2.](#page-8-0)

# *4.4. Future directions for drug development*

Based on the existing FDA-approved drugs, most of the drugs are AChE inhibitors and Aβ amyloid-targeting drugs. Future investigations may focus on targeting the upstream molecules of Aβ, such as α-secretase, BACE1, BACE2, and γ-secretase to inhibit Aβ deposition, including lanabecestat and umibecestat. However, lanabecestat has been associated with adverse events during clinical trials [[158](#page-15-0)], forcing the termination of the trials. Neuregulin 1 plays an important role in normal human psychiatric behaviors, and

**Table 3**  Selected AD drugs in clinical trial phase 3. The information comes from [[160,161\]](#page-15-0).

No.	Name of the Drug	Mechanism of Action	Clinical Trial
1	Gantenerumab	Anti-amyloid monoclonal antibody	NCT01760005
$\overline{2}$	Remternetug	Anti-amyloid monoclonal antibody	NCT05463731
3	Solanezumab	Anti-amyloid monoclonal antibody	NCT01760005
4	Lanabecestat	BACE1 reversible inhibition	NCT0224573.
			NCT02783573
5	Umibecestat	BACE1 reversible inhibition	NCT03131453
6	Verubecestat	BACE1 reversible inhibition	NCT02910739
7	Elenbecestat	<b>BACE1</b> reversible inhibition	NCT02956486
8	Atabecestat	BACE1 reversible inhibition	NCT03587376,
			NCT02569398
9	Semagacestat	γ-secretase inhibitor	NCT01035138
10	Tarenflurbil (MPC-	$\gamma$ -secretase inhibitor	NCT00322036
	7869)		
11	E2814	Anti-tau monoclonal antibody	NCT01760005,
			NCT05269394
12	Fosgonimeton	Activates signaling via the HGF/MET receptor system; promotes survival of neurons, enhances	NCT04488419,
		hippocampal synaptic plasticity.	NCT04886063
13	Levetiracetam	Modulator of the SV2A to reduce aberrant neuronal hyperactivity	NCT05986721

**Abbreviation:** HGF, hepatocyte growth factor; SV2A, synaptic vesicle protein.

<span id="page-10-0"></span>seizure protein 6 contributes to the maintenance of dendritic strength and prolonged sustained tension, both of which are substrates for BACE1; thus, the use of BACE1 inhibitors leads to adverse psychiatric events [[158](#page-15-0)]. Although, BACE inhibitors have shown good performance in reducing Aβ deposition, they are inadequate in terms of safety. Other BACE inhibitors may also cause adverse reactions for similar reasons. Additional information is presented in [Table 3](#page-9-0). In addition, the pathogenesis of AD is complex and varied, including p-tau, neuroinflammation, ferroptosis, imbalance of the intestinal flora, miRNA dysregulation, and many other mechanisms. In future, we plan to develop more targeted and universal drugs based on these pathogenic mechanisms of AD. Additional information is presented in [Table 3.](#page-9-0)

In addition to the pathogenesis of AD, some scientists are now proposing that neural stem cell (NSC) transplantation could emerge as a novel therapy for neurodegenerative diseases [[159](#page-15-0)]. The primary objective of NSC replacement was to restore degenerating neurons, thereby delaying neuronal function and cognitive decline. Furthermore, recent reports suggest that NSCs also exhibit the ability to promote neurotrophin secretion [[159](#page-15-0)]. The pathogenesis of AD is complex, and AD drugs cannot be studied in a single direction. The development of AD drugs should not only improve the cognitive dysfunction of patients with AD and slow down the disease process, but also, increase the safety of the drugs as much as possible. Therefore, based on an in-depth study of the different mechanisms, discovering the interactions between different mechanisms is very important for the development of new AD drugs.

# **5. Conclusion**

AD involves various triggers, including family genetics, old age, and lifestyle habits. The prevalence of AD is increasing every year as the standard of living improves and the population ages significantly. In response to this global health problem, the drugs approved by the FDA are mainly divided into three categories, namely AChE inhibitors, NMDA receptor antagonists, and Aβ protein monoclonal antibodies [\[146,150\]](#page-14-0). To date, no drug can completely cure AD, which may be related to the complex and intertwined pathogenic mechanisms of AD. Consequently, many drugs that target pathogenic mechanisms, such as p-tau, neuroinflammation, mitochondrial oxidative stress, ferroptosis, the intestinal environment, and miRNA, are present [[160\]](#page-15-0). Most drugs are terminated during clinical trials for a variety of reasons, with common reasons such as drug ineffectiveness and serious side effects. Therefore, the development of new therapeutics for AD remains challenging. However, owing to the complex pathological mechanisms of AD, a single target may have little effect. Perhaps we should consider a combination of drugs for multi-targeted therapy. For example, the combination of donepezil and memantine is associated with greater improvements in cognitive and daily activities and neuropsychiatric symptoms than monotherapy [[162](#page-15-0)]. In the future, multi-target AD therapy may be a powerful means of treating AD.

#### **CRediT authorship contribution statement**

**Liting Peng:** Writing – original draft, Visualization. **Zhiming Zhang:** Writing – review & editing, Writing – original draft. **Qi Li:**  Writing – review & editing, Visualization. **Zhenjiang Song:** Visualization. **Canqun Yan:** Writing – review & editing. **Hongyan Ling:**  Writing – review & editing, Conceptualization.

# **Data availability**

Data availability is not applicable to this article as no new data were created or analyzed in this study.

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# **Declaration of competing interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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# **References**

- [1] [World Alzheimer Report 2018 The state of the art of dementia research: New frontiers \(2018\)](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref1).
- [2] Z. Wang, Q. Xu, F. Cai, X. Liu, Y. Wu, W. Song, BACE2, a conditional beta-secretase, contributes to Alzheimer's disease pathogenesis, JCI Insight 4 (2019), [https://doi.org/10.1172/jci.insight.123431.](https://doi.org/10.1172/jci.insight.123431)
- [3] D.J. Selkoe, Cell biology of protein misfolding: the examples of Alzheimer's and Parkinson's diseases, Nat. Cell Biol. 6 (2004) 1054–1061, [https://doi.org/](https://doi.org/10.1038/ncb1104-1054) [10.1038/ncb1104-1054](https://doi.org/10.1038/ncb1104-1054).
- <span id="page-11-0"></span>[4] G. Blessed, B.E. Tomlinson, M. Roth, The association between quantitative measures of dementia and of senile change in the cerebral grey matter of elderly subjects, Br. J. Psychiatry 114 (1968) 797–811, <https://doi.org/10.1192/bjp.114.512.797>.
- [5] K. Iqbal, H.M. Wisniewski, M.L. Shelanski, S. Brostoff, B.H. Liwnicz, R.D. Terry, Protein changes in senile dementia, Brain Res. 77 (1974) 337–343, [https://doi.](https://doi.org/10.1016/0006-8993(74)90798-7) [org/10.1016/0006-8993\(74\)90798-7](https://doi.org/10.1016/0006-8993(74)90798-7).
- [6] N. Ma, C. Tie, B. Yu, W. Zhang, J. Wan, Identifying lncRNA-miRNA-mRNA networks to investigate Alzheimer's disease pathogenesis and therapy strategy, Aging (Albany NY) 12 (2020) 2897–2920, [https://doi.org/10.18632/aging.102785.](https://doi.org/10.18632/aging.102785)
- [7] R. Ossenkoppele, R. van der Kant, O. Hansson, Tau biomarkers in Alzheimer's disease: towards implementation in clinical practice and trials, Lancet Neurol. 21 (2022) 726–734, [https://doi.org/10.1016/S1474-4422\(22\)00168-5.](https://doi.org/10.1016/S1474-4422(22)00168-5)
- [8] M. El-Hussieny, M.A. Abd-El-Maksoud, F.M. Soliman, M.A. Fouad, M.K. El-Ashrey, Dual-target ligand discovery for Alzheimer's disease: triphenylphosphoranylidene derivatives as inhibitors of acetylcholinesterase and beta-amyloid aggregation, J Enzyme Inhib Med Chem 38 (2023) 2166040, <https://doi.org/10.1080/14756366.2023.2166040>.
- [9] D. Singh, Astrocytic and microglial cells as the modulators of neuroinflammation in Alzheimer's disease, J. Neuroinflammation 19 (2022) 206, [https://doi.](https://doi.org/10.1186/s12974-022-02565-0) [org/10.1186/s12974-022-02565-0.](https://doi.org/10.1186/s12974-022-02565-0)
- [10] D.A. Butterfield, Amyloid beta-peptide (1-42)-induced oxidative stress and neurotoxicity: implications for neurodegeneration in Alzheimer's disease brain. A review, Free Radic. Res. 36 (2002) 1307-1313, [https://doi.org/10.1080/1071576021000049890.](https://doi.org/10.1080/1071576021000049890)
- [11] F. Wang, J. Wang, Y. Shen, H. Li, W.D. Rausch, X. Huang, Iron dyshomeostasis and ferroptosis: a new Alzheimer's disease hypothesis? Front. Aging Neurosci. 14 (2022) 830569 <https://doi.org/10.3389/fnagi.2022.830569>.
- [12] C. Chen, J. Liao, Y. Xia, X. Liu, R. Jones, J. Haran, B. McCormick, T.R. Sampson, A. Alam, K. Ye, Gut microbiota regulate Alzheimer's disease pathologies and cognitive disorders via PUFA-associated neuroinflammation, Gut 71 (2022) 2233–2252, <https://doi.org/10.1136/gutjnl-2021-326269>.
- [13] S. Patel, A.V. Bansoad, R. Singh, G.L. Khatik, BACE1: a key regulator in Alzheimer's disease progression and current development of its inhibitors, Curr. Neuropharmacol. 20 (2022) 1174–1193, <https://doi.org/10.2174/1570159X19666211201094031>.
- [14] J.L. Dage, A. Eloyan, M. Thangarajah, D.B. Hammers, A.M. Fagan, J.D. Gray, S.E. Schindler, C. Snoddy, K.N.H. Nudelman, K.M. Faber, et al., Cerebrospinal fluid biomarkers in the longitudinal early-onset Alzheimer's disease study, Alzheimers Dement (2023), <https://doi.org/10.1002/alz.13399>.
- [15] H.A. Taylor, K.J. Simmons, E.M. Clavane, C.J. Trevelyan, J.M. Brown, L. Przemylska, N.T. Watt, L.C. Matthews, P.J. Meakin, PTPRD and dcc are novel BACE1 substrates differentially expressed in Alzheimer's disease: a data mining and bioinformatics study, Int. J. Mol. Sci. 23 (2022), https://doi.org/10.3390, ijms23094568
- [16] H. Hampel, R. Vassar, B. De Strooper, J. Hardy, M. Willem, N. Singh, J. Zhou, R. Yan, E. Vanmechelen, A. De Vos, et al., The beta-Secretase BACE1 in Alzheimer's Disease, Biol Psychiatry 89 (2021) 745–756, <https://doi.org/10.1016/j.biopsych.2020.02.001>.
- [17] J. Bao, M. Qin, Y.A.R. Mahaman, B. Zhang, F. Huang, K. Zeng, Y. Xia, D. Ke, Q. Wang, R. Liu, et al., BACE1 SUMOylation increases its stability and escalates the protease activity in Alzheimer's disease, Proc Natl Acad Sci U S A 115 (2018) 3954–3959, <https://doi.org/10.1073/pnas.1800498115>.
- [18] N.M. Moussa-Pacha, S.M. Abdin, H.A. Omar, H. Alniss, T.H. Al-Tel, BACE1 inhibitors: current status and future directions in treating Alzheimer's disease, Med. Res. Rev. 40 (2020) 339–384, [https://doi.org/10.1002/med.21622.](https://doi.org/10.1002/med.21622)
- [19] S.V. Ovsepian, J. Horacek, V.B. O'Leary, C. Hoschl, The ups and downs of BACE1: walking a fine line between neurocognitive and other psychiatric symptoms of Alzheimer's disease, Neuroscientist 27 (2021) 222–234, [https://doi.org/10.1177/1073858420940943.](https://doi.org/10.1177/1073858420940943)
- [20] C.J. Holler, R.L. Webb, A.L. Laux, T.L. Beckett, D.M. Niedowicz, R.R. Ahmed, Y. Liu, C.R. Simmons, A.L. Dowling, A. Spinelli, et al., BACE2 expression increases in human neurodegenerative disease, Am. J. Pathol. 180 (2012) 337–350, [https://doi.org/10.1016/j.ajpath.2011.09.034.](https://doi.org/10.1016/j.ajpath.2011.09.034)
- [21] [Y.Q. Sha, Cellular smears in the diagnosis of oral pemphigus\], Zhonghua Kou Qiang Ke Za Zhi 21 \(1986\) 34](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref21)–35.
- [22] J. Saez-Valero, R. Perez-Gonzalez, BACE2 beyond beta-processing of APP, its neuroprotective role in cerebrovascular endothelium, J. Neurochem. 166 (2023) 887–890, <https://doi.org/10.1111/jnc.15940>.
- [23] B.D. Bennett, S. Babu-Khan, R. Loeloff, J.C. Louis, E. Curran, M. Citron, R. Vassar, Expression analysis of BACE2 in brain and peripheral tissues, J. Biol. Chem. 275 (2000) 20647–20651, [https://doi.org/10.1074/jbc.M002688200.](https://doi.org/10.1074/jbc.M002688200)
- [24] G.M. Shankar, S. Li, T.H. Mehta, A. Garcia-Munoz, N.E. Shepardson, I. Smith, F.M. Brett, M.A. Farrell, M.J. Rowan, C.A. Lemere, et al., Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory, Nat Med 14 (2008) 837-842, <https://doi.org/10.1038/nm1782>.
- [25] M. Tolar, J. Hey, A. Power, S. Abushakra, Neurotoxic soluble amyloid oligomers drive Alzheimer's pathogenesis and represent a clinically validated target for slowing disease progression, Int. J. Mol. Sci. 22 (2021), [https://doi.org/10.3390/ijms22126355.](https://doi.org/10.3390/ijms22126355)
- [26] W. Wang, T.T. Hou, L.F. Jia, Q.Q. Wu, M.N. Quan, J.P. Jia, Toxic amyloid-beta oligomers induced self-replication in astrocytes triggering neuronal injury, EBioMedicine 42 (2019) 174–187, <https://doi.org/10.1016/j.ebiom.2019.03.049>.
- [27] Z. Teng, G.I. Kartalou, S. Dagar, P.C. Fraering, V. Lessmann, K. Gottmann, A delay in vesicle endocytosis by a C-terminal fragment of N-cadherin enhances Abeta synaptotoxicity, Cell Death Discov 9 (2023) 444, [https://doi.org/10.1038/s41420-023-01739-w.](https://doi.org/10.1038/s41420-023-01739-w)
- [28] R. Dixit, J.L. Ross, Y.E. Goldman, E.L. Holzbaur, Differential regulation of dynein and kinesin motor proteins by tau, Science 319 (2008) 1086–1089, [https://](https://doi.org/10.1126/science.1152993) oi.org/10.1126/science.1152993
- [29] V.V. Bhandare, B.V. Kumbhar, A. Kunwar, Differential binding affinity of tau repeat region R2 with neuronal-specific beta-tubulin isotypes, Sci. Rep. 9 (2019) 10795,<https://doi.org/10.1038/s41598-019-47249-7>.
- [30] V.H. Man, X. He, J. Gao, J. Wang, Phosphorylation of tau R2 repeat destabilizes its binding to microtubules: a molecular dynamics simulation study, ACS Chem. Neurosci. 14 (2023) 458–467, [https://doi.org/10.1021/acschemneuro.2c00611.](https://doi.org/10.1021/acschemneuro.2c00611)
- [31] Y. Xia, S. Prokop, B.I. Giasson, "Don't Phos over Tau": recent developments in clinical biomarkers and therapies targeting tau phosphorylation in Alzheimer's disease and other tauopathies, Mol. Neurodegener. 16 (2021) 37, [https://doi.org/10.1186/s13024-021-00460-5.](https://doi.org/10.1186/s13024-021-00460-5)
- [32] H. Zhang, W. Wei, M. Zhao, L. Ma, X. Jiang, H. Pei, Y. Cao, H. Li, Interaction between abeta and tau in the pathogenesis of Alzheimer's disease, Int. J. Biol. Sci. 17 (2021) 2181–2192, <https://doi.org/10.7150/ijbs.57078>.
- [33] S. Mondragon-Rodriguez, G. Perry, J. Luna-Munoz, M.C. Acevedo-Aquino, S. Williams, Phosphorylation of tau protein at sites Ser(396-404) is one of the earliest events in Alzheimer's disease and Down syndrome, Neuropathol. Appl. Neurobiol. 40 (2014) 121-135,<https://doi.org/10.1111/nan.12084>
- [34] D. Kyriakou, E. Bletsa, V. Moussis, Y. Deligiannakis, G. Malandrinos, Coordination properties of Cu(II) ions towards a phosphorylated fragment from the R1 domain of the tau protein and the effect of Ser phosphorylation on Cu(II) binding affinity, Dalton Trans. 52 (2022) 58–69, [https://doi.org/10.1039/](https://doi.org/10.1039/d2dt02838g) [d2dt02838g.](https://doi.org/10.1039/d2dt02838g)
- [35] A. Sengupta, J. Kabat, M. Novak, Q. Wu, I. Grundke-Iqbal, K. Iqbal, Phosphorylation of tau at both Thr 231 and Ser 262 is required for maximal inhibition of its binding to microtubules, Arch. Biochem. Biophys. 357 (1998) 299–309, [https://doi.org/10.1006/abbi.1998.0813.](https://doi.org/10.1006/abbi.1998.0813)
- [36] K. Stefanoska, M. Gajwani, A.R.P. Tan, H.I. Ahel, P.R. Asih, A. Volkerling, A. Poljak, A. Ittner, Alzheimer's disease: ablating single master site abolishes tau hyperphosphorylation, Sci. Adv. 8 (2022) eabl8809, <https://doi.org/10.1126/sciadv.abl8809>.
- [37] H. Decker, K.Y. Lo, S.M. Unger, S.T. Ferreira, M.A. Silverman, Amyloid-beta peptide oligomers disrupt axonal transport through an NMDA receptor-dependent mechanism that is mediated by glycogen synthase kinase 3beta in primary cultured hippocampal neurons, J. Neurosci. 30 (2010) 9166-9171, [https://doi.org/](https://doi.org/10.1523/JNEUROSCI.1074-10.2010) [10.1523/JNEUROSCI.1074-10.2010.](https://doi.org/10.1523/JNEUROSCI.1074-10.2010)
- [38] Y. Rui, P. Tiwari, Z. Xie, J.Q. Zheng, Acute impairment of mitochondrial trafficking by beta-amyloid peptides in hippocampal neurons, J. Neurosci. 26 (2006) 10480–10487, [https://doi.org/10.1523/JNEUROSCI.3231-06.2006.](https://doi.org/10.1523/JNEUROSCI.3231-06.2006)
- [39] T.J. Singh, I. Grundke-Iqbal, W.Q. Wu, V. Chauhan, M. Novak, E. Kontzekova, K. Iqbal, Protein kinase C and calcium/calmodulin-dependent protein kinase II phosphorylate three-repeat and four-repeat tau isoforms at different rates, Mol. Cell. Biochem. 168 (1997) 141–148, [https://doi.org/10.1023/a:](https://doi.org/10.1023/a:1006807105059)  006807105059
- [40] Y. Zhang, Y. Zhang, Y. Aman, C.T. Ng, W.H. Chau, Z. Zhang, M. Yue, C. Bohm, Y. Jia, S. Li, et al., Amyloid-beta toxicity modulates tau phosphorylation through the PAX6 signalling pathway, Brain 144 (2021) 2759–2770, [https://doi.org/10.1093/brain/awab134.](https://doi.org/10.1093/brain/awab134)
- <span id="page-12-0"></span>[41] L. Diez, L.E. Kapinos, J. Hochmair, S. Huebschmann, A. Dominguez-Baquero, A. Vogt, M. Rankovic, M. Zweckstetter, R.Y.H. Lim, S. Wegmann, Phosphorylation but not oligomerization drives the accumulation of tau with nucleoporin Nup98, Int. J. Mol. Sci. 23 (2022), [https://doi.org/10.3390/](https://doi.org/10.3390/ijms23073495)
- [ijms23073495.](https://doi.org/10.3390/ijms23073495) [42] N. Nag, T. Tripathi, Tau-FG-nucleoporin98 interaction and impaired nucleocytoplasmic transport in Alzheimer's disease, Brief Funct Genomics 22 (2023) 161–167, [https://doi.org/10.1093/bfgp/elac022.](https://doi.org/10.1093/bfgp/elac022)
- [43] T.H. Ferreira-Vieira, I.M. Guimaraes, F.R. Silva, F.M. Ribeiro, Alzheimer's disease: targeting the cholinergic system, Curr. Neuropharmacol. 14 (2016) 101–115, <https://doi.org/10.2174/1570159x13666150716165726>.
- [44] E. Akyuz, A.K. Polat, E. Eroglu, I. Kullu, E. Angelopoulou, Y.N. Paudel, Revisiting the role of neurotransmitters in epilepsy: an updated review, Life Sci. 265 (2021) 118826, [https://doi.org/10.1016/j.lfs.2020.118826.](https://doi.org/10.1016/j.lfs.2020.118826)
- [45] H. Bielarczyk, A. Jankowska-Kulawy, C. Hofling, A. Ronowska, S. Gul-Hinc, S. Rossner, R. Schliebs, T. Pawelczyk, A. Szutowicz, AbetaPP-Transgenic 2576 mice mimic cell type-specific aspects of acetyl-CoA-linked metabolic deficits in Alzheimer's disease, J Alzheimers Dis 48 (2015) 1083-1094, [https://doi.org/](https://doi.org/10.3233/JAD-150327) [10.3233/JAD-150327](https://doi.org/10.3233/JAD-150327).
- [46] L. Guo, J. Tian, H. Du, Mitochondrial dysfunction and synaptic transmission failure in Alzheimer's disease, J Alzheimers Dis 57 (2017) 1071–1086, [https://doi.](https://doi.org/10.3233/JAD-160702) [org/10.3233/JAD-160702.](https://doi.org/10.3233/JAD-160702)
- [47] K.Y. Wong, J. Roy, M.L. Fung, B.C. Heng, C. Zhang, L.W. Lim, Relationships between mitochondrial dysfunction and neurotransmission failure in Alzheimer's disease, Aging Dis 11 (2020) 1291–1316, <https://doi.org/10.14336/AD.2019.1125>.
- [48] S. Jiang, Y. Li, C. Zhang, Y. Zhao, G. Bu, H. Xu, Y.W. Zhang, M1 muscarinic acetylcholine receptor in Alzheimer's disease, Neurosci. Bull. 30 (2014) 295–307, [https://doi.org/10.1007/s12264-013-1406-z.](https://doi.org/10.1007/s12264-013-1406-z)
- [49] A.A. Davis, J.J. Fritz, J. Wess, J.J. Lah, A.I. Levey, Deletion of M1 muscarinic acetylcholine receptors increases amyloid pathology in vitro and in vivo, J. Neurosci. 30 (2010) 4190-4196, <https://doi.org/10.1523/JNEUROSCI.6393-09.2010>.
- [50] R. Haring, D. Gurwitz, J. Barg, R. Pinkas-Kramarski, E. Heldman, Z. Pittel, A. Wengier, H. Meshulam, D. Marciano, Y. Karton, et al., Amyloid precursor protein secretion via muscarinic receptors: reduced desensitization using the M1-selective agonist AF102B, Biochem. Biophys. Res. Commun. 203 (1994) 652–658, <https://doi.org/10.1006/bbrc.1994.2232>.
- [51] A. Caccamo, S. Oddo, L.M. Billings, K.N. Green, H. Martinez-Coria, A. Fisher, F.M. LaFerla, M1 receptors play a central role in modulating AD-like pathology in transgenic mice, Neuron 49 (2006) 671–682,<https://doi.org/10.1016/j.neuron.2006.01.020>.
- [52] C. Lena, J.P. Changeux, Allosteric nicotinic receptors, human pathologies, J. Physiol. Paris 92 (1998) 63–74, [https://doi.org/10.1016/S0928-4257\(98\)80140-](https://doi.org/10.1016/S0928-4257(98)80140-X)
- [X.](https://doi.org/10.1016/S0928-4257(98)80140-X) [53] J. Whyte, R. Harrison, G.G. Lunt, S. Wonnacott, Subcellular fractionation and distribution of cholinergic binding sites in fetal human brain, Neurochem. Res. 11 (1986) 1011–1023, [https://doi.org/10.1007/BF00965590.](https://doi.org/10.1007/BF00965590)
- [54] K.A. Radcliffe, J.A. Dani, Nicotinic stimulation produces multiple forms of increased glutamatergic synaptic transmission, J. Neurosci. 18 (1998) 7075–7083, [https://doi.org/10.1523/JNEUROSCI.18-18-07075.1998.](https://doi.org/10.1523/JNEUROSCI.18-18-07075.1998)
- [55] R. Gray, A.S. Rajan, K.A. Radcliffe, M. Yakehiro, J.A. Dani, Hippocampal synaptic transmission enhanced by low concentrations of nicotine, Nature 383 (1996) 713–716, <https://doi.org/10.1038/383713a0>.
- [56] K.A. Radcliffe, J.L. Fisher, R. Gray, J.A. Dani, Nicotinic modulation of glutamate and GABA synaptic transmission of hippocampal neurons, Ann. N. Y. Acad. Sci. 868 (1999) 591-610,<https://doi.org/10.1111/j.1749-6632.1999.tb11332.x>.
- [57] D.S. McGehee, M.J. Heath, S. Gelber, P. Devay, L.W. Role, Nicotine enhancement of fast excitatory synaptic transmission in CNS by presynaptic receptors, Science 269 (1995) 1692–1696, [https://doi.org/10.1126/science.7569895.](https://doi.org/10.1126/science.7569895)
- [58] G.I. Wilkie, P. Hutson, J.P. Sullivan, S. Wonnacott, Pharmacological characterization of a nicotinic autoreceptor in rat hippocampal synaptosomes, Neurochem. Res. 21 (1996) 1141–1148, [https://doi.org/10.1007/BF02532425.](https://doi.org/10.1007/BF02532425)
- [59] P.B. Clarke, M. Reuben, Release of [3H]-noradrenaline from rat hippocampal synaptosomes by nicotine: mediation by different nicotinic receptor subtypes from striatal [3H]-dopamine release, Br. J. Pharmacol. 117 (1996) 595–606,<https://doi.org/10.1111/j.1476-5381.1996.tb15232.x>.
- [60] M. Nisell, G.G. Nomikos, T.H. Svensson, Systemic nicotine-induced dopamine release in the rat nucleus accumbens is regulated by nicotinic receptors in the ventral tegmental area, Synapse 16 (1994) 36-44, <https://doi.org/10.1002/syn.890160105>.
- [61] [X. Yang, H.E. Criswell, G.R. Breese, Nicotine-induced inhibition in medial septum involves activation of presynaptic nicotinic cholinergic receptors on gamma](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref61)[aminobutyric acid-containing neurons, J Pharmacol Exp Ther 276 \(1996\) 482](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref61)–489.
- [62] E.Y.L. Liu, Y. Xia, X. Kong, M.S.S. Guo, A.X.D. Yu, B.Z.Y. Zheng, S. Mak, M.L. Xu, K.W.K. Tsim, Interacting with alpha 7 nAChR is a new mechanism for AChE to enhance the inflammatory response in macrophages, Acta Pharm. Sin. B 10 (2020) 1926-1942, <https://doi.org/10.1016/j.apsb.2020.05.005>.
- [63] Z.R. Chen, J.B. Huang, S.L. Yang, F.F. Hong, Role of cholinergic signaling in Alzheimer's disease, Molecules 27 (2022), [https://doi.org/10.3390/](https://doi.org/10.3390/molecules27061816) [molecules27061816](https://doi.org/10.3390/molecules27061816).
- [64] M.A. Rather, A. Khan, S. Alshahrani, H. Rashid, M. Qadri, S. Rashid, R.M. Alsaffar, M.A. Kamal, M.U. Rehman, Inflammation and Alzheimer's disease: mechanisms and therapeutic implications by natural products, Mediators Inflamm 2021 (2021) 9982954, [https://doi.org/10.1155/2021/9982954.](https://doi.org/10.1155/2021/9982954)
- [65] L.J. Lawson, V.H. Perry, P. Dri, S. Gordon, Heterogeneity in the distribution and morphology of microglia in the normal adult mouse brain, Neuroscience 39 (1990) 151–170, [https://doi.org/10.1016/0306-4522\(90\)90229-w.](https://doi.org/10.1016/0306-4522(90)90229-w)
- [66] S. Hashioka, Z. Wu, A. Klegeris, Glia-driven neuroinflammation and systemic inflammation in Alzheimer's disease, Curr. Neuropharmacol. 19 (2021) 908–924, <https://doi.org/10.2174/1570159X18666201111104509>.
- [67] M.T. Heneka, M.J. Carson, J. El Khoury, G.E. Landreth, F. Brosseron, D.L. Feinstein, A.H. Jacobs, T. Wyss-Coray, J. Vitorica, R.M. Ransohoff, et al., Neuroinflammation in Alzheimer's disease, Lancet Neurol. 14 (2015) 388–405, [https://doi.org/10.1016/S1474-4422\(15\)70016-5.](https://doi.org/10.1016/S1474-4422(15)70016-5)
- [68] H. Kettenmann, U.K. Hanisch, M. Noda, A. Verkhratsky, Physiology of microglia, Physiol. Rev. 91 (2011) 461–553, [https://doi.org/10.1152/](https://doi.org/10.1152/physrev.00011.2010)  [physrev.00011.2010.](https://doi.org/10.1152/physrev.00011.2010)
- [69] W.Y. Wang, M.S. Tan, J.T. Yu, L. Tan, Role of pro-inflammatory cytokines released from microglia in Alzheimer's disease, Ann. Transl. Med. 3 (2015) 136, <https://doi.org/10.3978/j.issn.2305-5839.2015.03.49>.
- [70] C. Wang, S. Zong, X. Cui, X. Wang, S. Wu, L. Wang, Y. Liu, Z. Lu, The effects of microglia-associated neuroinflammation on Alzheimer's disease, Front. Immunol. 14 (2023) 1117172, [https://doi.org/10.3389/fimmu.2023.1117172.](https://doi.org/10.3389/fimmu.2023.1117172)
- [71] H. Keren-Shaul, A. Spinrad, A. Weiner, O. Matcovitch-Natan, R. Dvir-Szternfeld, T.K. Ulland, E. David, K. Baruch, D. Lara-Astaiso, B. Toth, et al., A unique microglia type associated with restricting development of Alzheimer's disease, Cell 169 (2017) 1276–1290 e1217, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.cell.2017.05.018) [cell.2017.05.018](https://doi.org/10.1016/j.cell.2017.05.018).
- [72] S. Merighi, M. Nigro, A. Travagli, S. Gessi, Microglia and Alzheimer's disease, Int. J. Mol. Sci. 23 (2022), <https://doi.org/10.3390/ijms232112990>.
- [73] Y. Yu, R.D. Ye, Microglial Abeta receptors in Alzheimer's disease, Cell. Mol. Neurobiol. 35 (2015) 71–83, [https://doi.org/10.1007/s10571-014-0101-6.](https://doi.org/10.1007/s10571-014-0101-6)
- [74] T.L. Tay, J.C. Savage, C.W. Hui, K. Bisht, M.E. Tremblay, Microglia across the lifespan: from origin to function in brain development, plasticity and cognition, J Physiol. 595 (2017) 1929–1945, <https://doi.org/10.1113/JP272134>.
- [75] R. Daneman, L. Zhou, A.A. Kebede, B.A. Barres, Pericytes are required for blood-brain barrier integrity during embryogenesis, Nature 468 (2010) 562–566, [https://doi.org/10.1038/nature09513.](https://doi.org/10.1038/nature09513)
- [76] J.T. Hinkle, V.L. Dawson, T.M. Dawson, The A1 astrocyte paradigm: new avenues for pharmacological intervention in neurodegeneration, Mov. Disord. 34 (2019) 959–969, [https://doi.org/10.1002/mds.27718.](https://doi.org/10.1002/mds.27718)
- [77] M. Neal, J. Luo, D.S. Harischandra, R. Gordon, S. Sarkar, H. Jin, V. Anantharam, L. Desaubry, A. Kanthasamy, A. Kanthasamy, Prokineticin-2 promotes chemotaxis and alternative A2 reactivity of astrocytes, Glia 66 (2018) 2137–2157, <https://doi.org/10.1002/glia.23467>.
- [78] J. Kim, I.D. Yoo, J. Lim, J.S. Moon, Pathological phenotypes of astrocytes in Alzheimer's disease, Exp. Mol. Med. 56 (2024) 95–99, [https://doi.org/10.1038/](https://doi.org/10.1038/s12276-023-01148-0) [s12276-023-01148-0](https://doi.org/10.1038/s12276-023-01148-0).
- <span id="page-13-0"></span>[79] S.A. Liddelow, K.A. Guttenplan, L.E. Clarke, F.C. Bennett, C.J. Bohlen, L. Schirmer, M.L. Bennett, A.E. Munch, W.S. Chung, T.C. Peterson, et al., Neurotoxic reactive astrocytes are induced by activated microglia, Nature 541 (2017) 481–487, [https://doi.org/10.1038/nature21029.](https://doi.org/10.1038/nature21029)
- [80] M. Fakhoury, Microglia and astrocytes in Alzheimer's disease: implications for therapy, Curr. Neuropharmacol. 16 (2018) 508–518, [https://doi.org/10.2174/](https://doi.org/10.2174/1570159X15666170720095240) [1570159X15666170720095240.](https://doi.org/10.2174/1570159X15666170720095240)
- [81] M. Koistinaho, S. Lin, X. Wu, M. Esterman, D. Koger, J. Hanson, R. Higgs, F. Liu, S. Malkani, K.R. Bales, et al., Apolipoprotein E promotes astrocyte colocalization and degradation of deposited amyloid-beta peptides, Nat Med 10 (2004) 719-726, https://doi.org/10.1038/nm105
- [82] T. Wyss-Coray, J. Rogers, Inflammation in Alzheimer disease-a brief review of the basic science and clinical literature, Cold Spring Harb Perspect Med. 2 (2012) a006346, [https://doi.org/10.1101/cshperspect.a006346.](https://doi.org/10.1101/cshperspect.a006346)
- [83] M. Ries, M. Sastre, Mechanisms of abeta clearance and degradation by glial cells, Front. Aging Neurosci. 8 (2016) 160, [https://doi.org/10.3389/](https://doi.org/10.3389/fnagi.2016.00160)  [fnagi.2016.00160.](https://doi.org/10.3389/fnagi.2016.00160)
- [84] C. Sharma, S. Kim, Y. Nam, U.J. Jung, S.R. Kim, Mitochondrial dysfunction as a driver of cognitive impairment in Alzheimer's disease, Int. J. Mol. Sci. 22 (2021), [https://doi.org/10.3390/ijms22094850.](https://doi.org/10.3390/ijms22094850)
- [85] D.A. Butterfield, B. Halliwell, Oxidative stress, dysfunctional glucose metabolism and Alzheimer disease, Nat. Rev. Neurosci. 20 (2019) 148-160, [https://doi.](https://doi.org/10.1038/s41583-019-0132-6) [org/10.1038/s41583-019-0132-6.](https://doi.org/10.1038/s41583-019-0132-6)
- [86] A. Misrani, S. Tabassum, L. Yang, Mitochondrial dysfunction and oxidative stress in Alzheimer's disease, Front. Aging Neurosci. 13 (2021) 617588, [https://doi.](https://doi.org/10.3389/fnagi.2021.617588) [org/10.3389/fnagi.2021.617588](https://doi.org/10.3389/fnagi.2021.617588).
- [87] X. Ren, L. Zou, X. Zhang, V. Branco, J. Wang, C. Carvalho, A. Holmgren, J. Lu, Redox signaling mediated by thioredoxin and glutathione systems in the central nervous system, Antioxid Redox Signal 27 (2017) 989–1010, [https://doi.org/10.1089/ars.2016.6925.](https://doi.org/10.1089/ars.2016.6925)
- [88] [M. Levine, K. Harding, Spatial regulation of homeo box gene expression in Drosophila, Oxf Surv Eukaryot Genes. 4 \(1987\) 116](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref88)–142.
- [89] P.H. Reddy, M.F. Beal, Amyloid beta, mitochondrial dysfunction and synaptic damage: implications for cognitive decline in aging and Alzheimer's disease, Trends Mol. Med. 14 (2008) 45–53, [https://doi.org/10.1016/j.molmed.2007.12.002.](https://doi.org/10.1016/j.molmed.2007.12.002)
- [90] M.C. Jurcau, F.L. Andronie-Cioara, A. Jurcau, F. Marcu, D.M. Tit, N. Pascalau, D.C. Nistor-Cseppento, The link between oxidative stress, mitochondrial dysfunction and neuroinflammation in the pathophysiology of Alzheimer's disease: therapeutic implications and future perspectives, Antioxidants 11 (2022), [https://doi.org/10.3390/antiox11112167.](https://doi.org/10.3390/antiox11112167)
- [91] X.C. Li, Y. Hu, Z.H. Wang, Y. Luo, Y. Zhang, X.P. Liu, Q. Feng, Q. Wang, K. Ye, G.P. Liu, et al., Human wild-type full-length tau accumulation disrupts mitochondrial dynamics and the functions via increasing mitofusins, Sci. Rep. 6 (2016) 24756, <https://doi.org/10.1038/srep24756>.
- [92] R. Kandimalla, M. Manczak, X. Yin, R. Wang, P.H. Reddy, Hippocampal phosphorylated tau induced cognitive decline, dendritic spine loss and mitochondrial abnormalities in a mouse model of Alzheimer's disease, Hum. Mol. Genet. 27 (2018) 30–40, <https://doi.org/10.1093/hmg/ddx381>.
- [93] P. Jadiya, D.W. Kolmetzky, D. Tomar, A. Di Meco, A.A. Lombardi, J.P. Lambert, T.S. Luongo, M.H. Ludtmann, D. Pratico, J.W. Elrod, Impaired mitochondrial calcium efflux contributes to disease progression in models of Alzheimer's disease, Nat. Commun. 10 (2019) 3885, [https://doi.org/10.1038/s41467-019-](https://doi.org/10.1038/s41467-019-11813-6)  [11813-6](https://doi.org/10.1038/s41467-019-11813-6).
- [94] B.R. Stockwell, J.P. Friedmann Angeli, H. Bayir, A.I. Bush, M. Conrad, S.J. Dixon, S. Fulda, S. Gascon, S.K. Hatzios, V.E. Kagan, et al., Ferroptosis: a regulated cell death nexus linking metabolism, Redox Biology, and Disease, Cell 171 (2017) 273–285,<https://doi.org/10.1016/j.cell.2017.09.021>.
- [95] H. Mao, Y. Zhao, H. Li, L. Lei, Ferroptosis as an emerging target in inflammatory diseases, Prog. Biophys. Mol. Biol. 155 (2020) 20–28, [https://doi.org/](https://doi.org/10.1016/j.pbiomolbio.2020.04.001) [10.1016/j.pbiomolbio.2020.04.001](https://doi.org/10.1016/j.pbiomolbio.2020.04.001).
- [96] R.J. Bridges, N.R. Natale, S.A. Patel, System xc(-) cystine/glutamate antiporter: an update on molecular pharmacology and roles within the CNS, Br. J. Pharmacol. 165 (2012) 20–34, <https://doi.org/10.1111/j.1476-5381.2011.01480.x>.
- [97] S. Ayton, Y. Wang, I. Diouf, J.A. Schneider, J. Brockman, M.C. Morris, A.I. Bush, Brain iron is associated with accelerated cognitive decline in people with Alzheimer pathology, Mol Psychiatry 25 (2020) 2932–2941, [https://doi.org/10.1038/s41380-019-0375-7.](https://doi.org/10.1038/s41380-019-0375-7)
- [98] G. Gao, J. Li, Y. Zhang, Y.Z. Chang, Cellular iron metabolism and regulation, Adv. Exp. Med. Biol. 1173 (2019) 21–32, [https://doi.org/10.1007/978-981-13-](https://doi.org/10.1007/978-981-13-9589-5_2) [9589-5\\_2](https://doi.org/10.1007/978-981-13-9589-5_2).
- [99] R.J. Ward, F.A. Zucca, J.H. Duyn, R.R. Crichton, L. Zecca, The role of iron in brain ageing and neurodegenerative disorders, Lancet Neurol. 13 (2014) 1045–1060, [https://doi.org/10.1016/S1474-4422\(14\)70117-6.](https://doi.org/10.1016/S1474-4422(14)70117-6)
- [100] W.D. Bao, P. Pang, X.T. Zhou, F. Hu, W. Xiong, K. Chen, J. Wang, F. Wang, D. Xie, Y.Z. Hu, et al., Loss of ferroportin induces memory impairment by promoting ferroptosis in Alzheimer's disease, Cell Death Differ. 28 (2021) 1548–1562, <https://doi.org/10.1038/s41418-020-00685-9>.
- [101] M.W. Park, H.W. Cha, J. Kim, J.H. Kim, H. Yang, S. Yoon, N. Boonpraman, S.S. Yi, I.D. Yoo, J.S. Moon, NOX4 promotes ferroptosis of astrocytes by oxidative stress-induced lipid peroxidation via the impairment of mitochondrial metabolism in Alzheimer's diseases, Redox Biol. 41 (2021) 101947, https://doi.org/ [10.1016/j.redox.2021.101947.](https://doi.org/10.1016/j.redox.2021.101947)
- [102] P.A. Adeniyi, X. Gong, E. MacGregor, K. Degener-O'Brien, E. McClendon, M. Garcia, O. Romero, J. Russell, T. Srivastava, J. Miller, et al., Ferroptosis of microglia in aging human white matter injury, Ann. Neurol. 94 (2023) 1048–1066,<https://doi.org/10.1002/ana.26770>.
- [103] [L.P. Singh, T. Yumnamcha, T.S. Devi, Mitophagy, ferritinophagy and ferroptosis in retinal pigment epithelial cells under high glucose conditions: implications](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref103) [for diabetic retinopathy and age-related retinal diseases, JOJ Ophthalmol 8 \(2021\) 77](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref103)–85.
- [104] J.A. Thompson, C.C. White, D.P. Cox, J.Y. Chan, T.J. Kavanagh, N. Fausto, C.C. Franklin, Distinct Nrf1/2-independent mechanisms mediate as 3+-induced glutamate-cysteine ligase subunit gene expression in murine hepatocytes, Free Radic. Biol. Med. 46 (2009) 1614–1625, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.freeradbiomed.2009.03.016) [freeradbiomed.2009.03.016](https://doi.org/10.1016/j.freeradbiomed.2009.03.016).
- [105] Y. Soeda, M. Yoshikawa, O.F. Almeida, A. Sumioka, S. Maeda, H. Osada, Y. Kondoh, A. Saito, T. Miyasaka, T. Kimura, et al., Toxic tau oligomer formation blocked by capping of cysteine residues with 1,2-dihydroxybenzene groups, Nat. Commun. 6 (2015) 10216, [https://doi.org/10.1038/ncomms10216.](https://doi.org/10.1038/ncomms10216) [106] L.K. Ursell, J.L. Metcalf, L.W. Parfrey, R. Knight, Defining the human microbiome, Nutr. Rev. 70 (Suppl 1) (2012) S38–S44, [https://doi.org/10.1111/j.1753-](https://doi.org/10.1111/j.1753-4887.2012.00493.x)
- [4887.2012.00493.x](https://doi.org/10.1111/j.1753-4887.2012.00493.x).
- [107] F. Angelucci, K. Cechova, J. Amlerova, J. Hort, Antibiotics, gut microbiota, and Alzheimer's disease, J. Neuroinflammation 16 (2019) 108, [https://doi.org/](https://doi.org/10.1186/s12974-019-1494-4) [10.1186/s12974-019-1494-4.](https://doi.org/10.1186/s12974-019-1494-4)
- [108] T.G. Dinan, J.F. Cryan, The microbiome-gut-brain Axis in health and disease, Gastroenterol Clin North Am 46 (2017) 77–89, [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.gtc.2016.09.007) [gtc.2016.09.007](https://doi.org/10.1016/j.gtc.2016.09.007).
- [109] J. Xiong, Z. Zhang, K. Ye, C/EBPbeta/AEP signaling drives Alzheimer's disease pathogenesis, Neurosci. Bull. 39 (2023) 1173–1185, [https://doi.org/10.1007/](https://doi.org/10.1007/s12264-023-01025-w) [s12264-023-01025-w](https://doi.org/10.1007/s12264-023-01025-w).
- [110] [Y. Zhao, W.J. Lukiw, Microbiome-generated amyloid and potential impact on amyloidogenesis in Alzheimer](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref110)'s disease (AD), J. Nat. Sci. 1 (2015).
- [111] M. Iwauchi, A. Horigome, K. Ishikawa, A. Mikuni, M. Nakano, J.Z. Xiao, T. Odamaki, S. Hironaka, Relationship between oral and gut microbiota in elderly people, Immun Inflamm Dis 7 (2019) 229–236, <https://doi.org/10.1002/iid3.266>.
- [112] M.M.H. Abdelbary, M. Hatting, A. Bott, A. Dahlhausen, D. Keller, C. Trautwein, G. Conrads, The oral-gut axis: salivary and fecal microbiome dysbiosis in patients with inflammatory bowel disease, Front. Cell. Infect. Microbiol. 12 (2022) 1010853, [https://doi.org/10.3389/fcimb.2022.1010853.](https://doi.org/10.3389/fcimb.2022.1010853)
- [113] M. Sochocka, K. Donskow-Lysoniewska, B.S. Diniz, D. Kurpas, E. Brzozowska, J. Leszek, The gut microbiome alterations and inflammation-driven pathogenesis of Alzheimer's disease-a critical review, Mol. Neurobiol. 56 (2019) 1841–1851, <https://doi.org/10.1007/s12035-018-1188-4>.
- [114] R.J. Mulders, K.C.G. de Git, E. Schele, S.L. Dickson, Y. Sanz, R.A.H. Adan, Microbiota in obesity: interactions with enteroendocrine, immune and central nervous systems, Obes. Rev. 19 (2018) 435–451, [https://doi.org/10.1111/obr.12661.](https://doi.org/10.1111/obr.12661)
- [115] J.M. Hill, W.J. Lukiw, Microbial-generated amyloids and Alzheimer's disease (AD), Front. Aging Neurosci. 7 (2015) 9, [https://doi.org/10.3389/](https://doi.org/10.3389/fnagi.2015.00009) [fnagi.2015.00009](https://doi.org/10.3389/fnagi.2015.00009).
- [116] X. Hu, T. Wang, F. Jin, Alzheimer's disease and gut microbiota, Sci. China Life Sci. 59 (2016) 1006–1023, <https://doi.org/10.1007/s11427-016-5083-9>.
- [117] M.S. Kahn, D. Kranjac, C.A. Alonzo, J.H. Haase, R.O. Cedillos, K.A. McLinden, G.W. Boehm, M.J. Chumley, Prolonged elevation in hippocampal Abeta and cognitive deficits following repeated endotoxin exposure in the mouse, Behav. Brain Res. 229 (2012) 176–184, <https://doi.org/10.1016/j.bbr.2012.01.010>.
- <span id="page-14-0"></span>[118] W.J. Lukiw, Bacteroides fragilis lipopolysaccharide and inflammatory signaling in Alzheimer's disease, Front. Microbiol. 7 (2016) 1544, [https://doi.org/](https://doi.org/10.3389/fmicb.2016.01544) [10.3389/fmicb.2016.01544](https://doi.org/10.3389/fmicb.2016.01544).
- [119] C.Y. Liu, X. Wang, C. Liu, H.L. Zhang, Pharmacological targeting of microglial activation: new therapeutic approach, Front. Cell. Neurosci. 13 (2019) 514, [https://doi.org/10.3389/fncel.2019.00514.](https://doi.org/10.3389/fncel.2019.00514)
- [120] B.I. Arioz, B. Tastan, E. Tarakcioglu, K.U. Tufekci, M. Olcum, N. Ersoy, A. Bagriyanik, K. Genc, S. Genc, Melatonin attenuates LPS-induced acute depressive-like behaviors and microglial NLRP3 inflammasome activation through the SIRT1/nrf2 pathway, Front. Immunol. 10 (2019) 1511, [https://doi.org/10.3389/](https://doi.org/10.3389/fimmu.2019.01511) [fimmu.2019.01511](https://doi.org/10.3389/fimmu.2019.01511).
- [121] J.M. Calderon, D. Erazo, T.J. Kieran, N.L. Gottdenker, C. Leon, J. Cordovez, F. Guhl, T.C. Glenn, C. Gonzalez, How microclimatic variables and blood meal sources influence Rhodnius prolixus abundance and Trypanosoma cruzi infection in Attalea butyracea and Elaeis guineensis palms? Acta Trop. 212 (2020) 105674 <https://doi.org/10.1016/j.actatropica.2020.105674>.
- [122] T. Li, J.Y. Chiang, Bile acids as metabolic regulators, Curr. Opin. Gastroenterol. 31 (2015) 159–165, <https://doi.org/10.1097/MOG.0000000000000156>.
- [123] P. Gamba, G. Testa, B. Sottero, S. Gargiulo, G. Poli, G. Leonarduzzi, The link between altered cholesterol metabolism and Alzheimer's disease, Ann. N. Y. Acad. Sci. 1259 (2012) 54–64, [https://doi.org/10.1111/j.1749-6632.2012.06513.x.](https://doi.org/10.1111/j.1749-6632.2012.06513.x)
- [124] C. Condello, P. Yuan, A. Schain, J. Grutzendler, Microglia constitute a barrier that prevents neurotoxic protofibrillar Abeta42 hotspots around plaques, Nat. Commun. 6 (2015) 6176, <https://doi.org/10.1038/ncomms7176>.
- [125] D. Erny, A.L. Hrabe de Angelis, D. Jaitin, P. Wieghofer, O. Staszewski, E. David, H. Keren-Shaul, T. Mahlakoiv, K. Jakobshagen, T. Buch, et al., Host microbiota constantly control maturation and function of microglia in the CNS, Nat. Neurosci. 18 (2015) 965–977, <https://doi.org/10.1038/nn.4030>.
- [126] Z. Zhao, J. Ning, X.Q. Bao, M. Shang, J. Ma, G. Li, D. Zhang, Fecal microbiota transplantation protects rotenone-induced Parkinson's disease mice via suppressing inflammation mediated by the lipopolysaccharide-TLR4 signaling pathway through the microbiota-gut-brain axis, Microbiome 9 (2021) 226, [https://doi.org/10.1186/s40168-021-01107-9.](https://doi.org/10.1186/s40168-021-01107-9)
- [127] E.M.M. Quigley, Microbiota-brain-gut Axis and neurodegenerative diseases, Curr. Neurol. Neurosci. Rep. 17 (2017) 94, [https://doi.org/10.1007/s11910-017-](https://doi.org/10.1007/s11910-017-0802-6) [0802-6](https://doi.org/10.1007/s11910-017-0802-6).
- [128] J. Guo, Y. Cai, X. Ye, N. Ma, Y. Wang, B. Yu, J. Wan, MiR-409-5p as a regulator of neurite growth is down regulated in APP/PS1 murine model of Alzheimer's disease, Front. Neurosci. 13 (2019) 1264, <https://doi.org/10.3389/fnins.2019.01264>.
- [129] Y. Song, M. Hu, J. Zhang, Z.Q. Teng, C. Chen, A novel mechanism of synaptic and cognitive impairments mediated via microRNA-30b in Alzheimer's disease, EBioMedicine 39 (2019) 409–421, [https://doi.org/10.1016/j.ebiom.2018.11.059.](https://doi.org/10.1016/j.ebiom.2018.11.059)
- [130] J.M. Long, B. Maloney, J.T. Rogers, D.K. Lahiri, Novel upregulation of amyloid-beta precursor protein (APP) by microRNA-346 via targeting of APP mRNA 5' untranslated region: implications in Alzheimer's disease, Mol Psychiatry 24 (2019) 345–363, [https://doi.org/10.1038/s41380-018-0266-3.](https://doi.org/10.1038/s41380-018-0266-3)
- [131] P.T. Nelson, W.X. Wang, MiR-107 is reduced in Alzheimer's disease brain neocortex: validation study, J Alzheimers Dis 21 (2010) 75-79, [https://doi.org/](https://doi.org/10.3233/JAD-2010-091603) [10.3233/JAD-2010-091603](https://doi.org/10.3233/JAD-2010-091603).
- [132] C. Cheng, W. Li, Z. Zhang, S. Yoshimura, Q. Hao, C. Zhang, Z. Wang, MicroRNA-144 is regulated by activator protein-1 (AP-1) and decreases expression of Alzheimer disease-related a disintegrin and metalloprotease 10 (ADAM10), J. Biol. Chem. 288 (2013) 13748–13761, [https://doi.org/10.1074/jbc.](https://doi.org/10.1074/jbc.M112.381392)  [M112.381392.](https://doi.org/10.1074/jbc.M112.381392)
- [133] J. Banzhaf-Strathmann, E. Benito, S. May, T. Arzberger, S. Tahirovic, H. Kretzschmar, A. Fischer, D. Edbauer, MicroRNA-125b induces tau
- hyperphosphorylation and cognitive deficits in Alzheimer's disease, EMBO J. 33 (2014) 1667-1680, https://doi.org/10.15252/embj.201387
- [134] E. Salta, A. Sierksma, E. Vanden Eynden, B. De Strooper, miR-132 loss de-represses ITPKB and aggravates amyloid and TAU pathology in Alzheimer's brain, EMBO Mol. Med. 8 (2016) 1005–1018, [https://doi.org/10.15252/emmm.201606520.](https://doi.org/10.15252/emmm.201606520)
- [135] J.R. Dickson, C. Kruse, D.R. Montagna, B. Finsen, M.S. Wolfe, Alternative polyadenylation and miR-34 family members regulate tau expression, J. Neurochem. 127 (2013) 739–749, [https://doi.org/10.1111/jnc.12437.](https://doi.org/10.1111/jnc.12437)
- [136] L.H. Sun, T. Ban, C.D. Liu, Q.X. Chen, X. Wang, M.L. Yan, X.L. Hu, X.L. Su, Y.N. Bao, L.L. Sun, et al., Activation of Cdk5/p25 and tau phosphorylation following chronic brain hypoperfusion in rats involves microRNA-195 down-regulation, J. Neurochem. 134 (2015) 1139–1151, [https://doi.org/10.1111/jnc.13212.](https://doi.org/10.1111/jnc.13212)
- [137] Y. Zhao, S. Bhattacharjee, B.M. Jones, J. Hill, P. Dua, W.J. Lukiw, Regulation of neurotropic signaling by the inducible, NF-kB-sensitive miRNA-125b in Alzheimer's disease (AD) and in primary human neuronal-glial (HNG) cells, Mol. Neurobiol. 50 (2014) 97–106,<https://doi.org/10.1007/s12035-013-8595-3>.
- [138] Q. Shi, G.E. Gibson, Up-regulation of the mitochondrial malate dehydrogenase by oxidative stress is mediated by miR-743a, J. Neurochem. 118 (2011) 440–448, [https://doi.org/10.1111/j.1471-4159.2011.07333.x.](https://doi.org/10.1111/j.1471-4159.2011.07333.x)
- [139] C. Liang, T. Zou, M. Zhang, W. Fan, T. Zhang, Y. Jiang, Y. Cai, F. Chen, X. Chen, Y. Sun, et al., MicroRNA-146a switches microglial phenotypes to resist the pathological processes and cognitive degradation of Alzheimer's disease, Theranostics 11 (2021) 4103–4121, <https://doi.org/10.7150/thno.53418>.
- [140] M.L. Chen, C.G. Hong, T. Yue, H.M. Li, R. Duan, W.B. Hu, J. Cao, Z.X. Wang, C.Y. Chen, X.K. Hu, et al., Inhibition of miR-331-3p and miR-9-5p ameliorates Alzheimer's disease by enhancing autophagy, Theranostics 11 (2021) 2395–2409, <https://doi.org/10.7150/thno.47408>.
- [141] D. Chen, G. Lan, R. Li, Y. Mei, X. Shui, X. Gu, L. Wang, T. Zhang, C.L. Gan, Y. Xia, et al., Melatonin ameliorates tau-related pathology via the miR-504-3p and CDK5 axis in Alzheimer's disease, Transl. Neurodegener. 11 (2022) 27, <https://doi.org/10.1186/s40035-022-00302-4>.
- [142] X. Wang, D. Liu, H.Z. Huang, Z.H. Wang, T.Y. Hou, X. Yang, P. Pang, N. Wei, Y.F. Zhou, M.J. Dupras, et al., A novel MicroRNA-124/PTPN1 signal pathway mediates synaptic and memory deficits in Alzheimer's disease, Biol Psychiatry 83 (2018) 395–405, [https://doi.org/10.1016/j.biopsych.2017.07.023.](https://doi.org/10.1016/j.biopsych.2017.07.023)
- [143] P. Xia, J. Chen, Y. Liu, X. Cui, C. Wang, S. Zong, L. Wang, Z. Lu, MicroRNA-22-3p ameliorates Alzheimer's disease by targeting SOX9 through the NF-kappaB signaling pathway in the hippocampus, J. Neuroinflammation 19 (2022) 180, [https://doi.org/10.1186/s12974-022-02548-1.](https://doi.org/10.1186/s12974-022-02548-1)
- [144] A. Aschrafi, A.D. Schwechter, M.G. Mameza, O. Natera-Naranjo, A.E. Gioio, B.B. Kaplan, MicroRNA-338 regulates local cytochrome c oxidase IV mRNA levels and oxidative phosphorylation in the axons of sympathetic neurons, J. Neurosci. 28 (2008) 12581-12590, https://doi.org/10.1523/JNEUROSCI.3338-[08.2008](https://doi.org/10.1523/JNEUROSCI.3338-08.2008).
- [145] F.Z. Chen, Y. Zhao, H.Z. Chen, MicroRNA-98 reduces amyloid beta-protein production and improves oxidative stress and mitochondrial dysfunction through the Notch signaling pathway via HEY2 in Alzheimer's disease mice, Int. J. Mol. Med. 43 (2019) 91–102,<https://doi.org/10.3892/ijmm.2018.3957>.
- [146] A. Bubley, A. Erofeev, P. Gorelkin, E. Beloglazkina, A. Majouga, O. Krasnovskaya, Tacrine-based hybrids: past, present, and future, Int. J. Mol. Sci. 24 (2023), <https://doi.org/10.3390/ijms24021717>.
- [147] N. Zhang, M.L. Gordon, Clinical efficacy and safety of donepezil in the treatment of Alzheimer's disease in Chinese patients, Clin. Interv. Aging 13 (2018) 1963–1970, [https://doi.org/10.2147/CIA.S159920.](https://doi.org/10.2147/CIA.S159920)
- [148] D. Lo, G.T. Grossberg, Use of memantine for the treatment of dementia, Expert Rev. Neurother. 11 (2011) 1359–1370, <https://doi.org/10.1586/ern.11.132>. [149] T. Pardo-Moreno, A. Gonzalez-Acedo, A. Rivas-Dominguez, V. Garcia-Morales, F.J. Garcia-Cozar, J.J. Ramos-Rodriguez, L. Melguizo-Rodriguez, Therapeutic
- approach to Alzheimer's disease: current treatments and new perspectives, Pharmaceutics 14 (2022), [https://doi.org/10.3390/pharmaceutics14061117.](https://doi.org/10.3390/pharmaceutics14061117)
- [150] J. Sevigny, P. Chiao, T. Bussiere, P.H. Weinreb, L. Williams, M. Maier, R. Dunstan, S. Salloway, T. Chen, Y. Ling, et al., The antibody aducanumab reduces Abeta plaques in Alzheimer's disease, Nature 537 (2016) 50–56, [https://doi.org/10.1038/nature19323.](https://doi.org/10.1038/nature19323)
- [151] G. Marucci, M. Buccioni, D.D. Ben, C. Lambertucci, R. Volpini, F. Amenta, Efficacy of acetylcholinesterase inhibitors in Alzheimer's disease, Neuropharmacology 190 (2021) 108352, [https://doi.org/10.1016/j.neuropharm.2020.108352.](https://doi.org/10.1016/j.neuropharm.2020.108352)
- [152] [R.A. Hansen, G. Gartlehner, A.P. Webb, L.C. Morgan, C.G. Moore, D.E. Jonas, Efficacy and safety of donepezil, galantamine, and rivastigmine for the treatment](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref152) of Alzheimer'[s disease: a systematic review and meta-analysis, Clin. Interv. Aging 3 \(2008\) 211](http://refhub.elsevier.com/S2405-8440(24)15248-6/sref152)–225.
- [153] S. Budd Haeberlein, P.S. Aisen, F. Barkhof, S. Chalkias, T. Chen, S. Cohen, G. Dent, O. Hansson, K. Harrison, C. von Hehn, et al., Two randomized phase 3 studies of aducanumab in early Alzheimer's disease, J Prev Alzheimers Dis 9 (2022) 197–210, <https://doi.org/10.14283/jpad.2022.30>.
- [154] C.H. van Dyck, C.J. Swanson, P. Aisen, R.J. Bateman, C. Chen, M. Gee, M. Kanekiyo, D. Li, L. Reyderman, S. Cohen, et al., Lecanemab in early Alzheimer's disease, N. Engl. J. Med. 388 (2023) 9–21,<https://doi.org/10.1056/NEJMoa2212948>.
- [155] J.R. Sims, J.A. Zimmer, C.D. Evans, M. Lu, P. Ardayfio, J. Sparks, A.M. Wessels, S. Shcherbinin, H. Wang, E.S. Monkul Nery, et al., Donanemab in early symptomatic alzheimer disease: the TRAILBLAZER-ALZ 2 randomized clinical trial, JAMA 330 (2023) 512–527, <https://doi.org/10.1001/jama.2023.13239>.
- <span id="page-15-0"></span>[156] S. Shcherbinin, C.D. Evans, M. Lu, S.W. Andersen, M.J. Pontecorvo, B.A. Willis, I. Gueorguieva, P.M. Hauck, D.A. Brooks, M.A. Mintun, et al., Association of amyloid reduction after donanemab treatment with tau pathology and clinical outcomes: the TRAILBLAZER-ALZ randomized clinical trial, JAMA Neurol. 79 (2022) 1015–1024,<https://doi.org/10.1001/jamaneurol.2022.2793>.
- [157] R.A. Sperling, C.R. Jack Jr., S.E. Black, M.P. Frosch, S.M. Greenberg, B.T. Hyman, P. Scheltens, M.C. Carrillo, W. Thies, M.M. Bednar, et al., Amyloid-related imaging abnormalities in amyloid-modifying therapeutic trials: recommendations from the Alzheimer's Association Research Roundtable Workgroup, Alzheimers Dement 7 (2011) 367–385, [https://doi.org/10.1016/j.jalz.2011.05.2351.](https://doi.org/10.1016/j.jalz.2011.05.2351)
- [158] Error in the biomarker subsection, JAMA Neurol. 77 (2020) 1179, [https://doi.org/10.1001/jamaneurol.2020.2416.](https://doi.org/10.1001/jamaneurol.2020.2416)
- [159] R. De Gioia, F. Biella, G. Citterio, F. Rizzo, E. Abati, M. Nizzardo, N. Bresolin, G.P. Comi, S. Corti, Neural stem cell transplantation for neurodegenerative diseases, Int. J. Mol. Sci. 21 (2020), <https://doi.org/10.3390/ijms21093103>.
- [160] J. Cummings, Y. Zhou, G. Lee, K. Zhong, J. Fonseca, F. Cheng, Alzheimer's disease drug development pipeline: 2024, Alzheimers Dement (N Y) 10 (2024) e12465, [https://doi.org/10.1002/trc2.12465.](https://doi.org/10.1002/trc2.12465)
- [161] Food and drug administration US. [https://www.clinicaltrials.gov/,](https://www.clinicaltrials.gov/) 2024. (Accessed 10 July 2024).
- [162] J. Guo, Z. Wang, R. Liu, Y. Huang, N. Zhang, R. Zhang, Donepezil Memantine, Or combination therapy-what is the best therapy for Alzheimer's disease? A network meta-analysis, Brain Behav 10 (2020) e01831, <https://doi.org/10.1002/brb3.1831>.