



# Complete Genome Sequences of the *Carlavirus Sweet potato chlorotic fleck virus* from East Timor and Australia

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We present here the first complete genome sequences of *Sweet potato chlorotic fleck virus* (SPCFV) from sweet potato in Australia and East Timor, and we compare these with four complete SPCFV genomes from Korea and one from Uganda. The Australian, East Timorese, Korean, and Ugandan genomes differed considerably from each other.

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To examine possible connectivity between viruses infecting important crops in Australia and South-East Asia, we studied sweet potato viruses from East Timor and Australia. *Sweet potato chlorotic fleck virus* (SPCFV) is a single-stranded RNA virus belonging to the genus *Carlavirus*, family *Betaflexiviridae* (1). It has not been reported from East Timor, but it was found once in Australia, from where a single SPCFV coat protein sequence (Aus-Can) is available under accession number EF990647 (2). Currently, only four complete SPCFV genomes from Korea and one from Uganda are available in GenBank (1, 3). Isolate AusCan and an SPCFV isolate from East Timor were sequenced and their complete genomes were obtained.

Fifteen East Timorese sweet potato samples blotted onto Fast Technology for Analysis of nucleic acids (FTA) cards (4) were sent to Australia. A plant infected with isolate AusCan was planted, and a scion from it graft-inoculated to indicator plant *Ipomoea setosa*. Total RNA was extracted from the FTA cards and an I. setosa leaf sample with virus symptoms using the ZR Plant RNA Mini-Prep kit (Zymo Research) and treated with RNase-free DNase (Invitrogen) measured using Qubit (Invitrogen). RNA integrity was confirmed using RNA screen Tape (TapeStation 2200, Agilent Technologies). Libraries were prepared from total RNA using a TruSeqstranded Total RNA sample preparation kit with Ribozero-Plant (catalogue number RS-122-2401, Illumina). Final size and concentration of each library was verified using Qubit and D1000 ScreenTape (TapeStation 2200, Agilent Technologies). Sequencing was by HiSeq 2500 using a TruSeq SBS kit V4 (Illumina) with 151 cycles of paired-end reads. The reads were assembled and the genomes were annotated using CLC Genomics Workbench version 6.5 (CLC bio) and Geneious version 8.1.7 (Biomatters) (5).

Only one FTA card sample (Tm37) collected in May 2015 from the Dili district in East Timor contained SPCFV. It yielded 14,721,488 reads and, after trimming, 14,546,337 remained. *De novo* assembly generated 1,307 contigs and 21,003 reads mapped to the contig of interest with coverage of 397×. Sample AusCan yielded 6,260,728 reads and, after trimming, 5,405,928 remained. *De novo* assembly generated 832 contigs with 333,024 reads mapped to the contig of interest, giving coverage of  $6,302 \times$ . Both Tm37 and AusCan sequences had the six intact open reading frames (ORFs) typical of carlaviruses (6). Pairwise nucleotide identity between Tm37 and AusCan was 72%. The closest match to Tm37 by BLAST was 87% to KP715159 from Korea, and to AusCan it was 87% to KP115606, also from Korea. Although the Tm37 and AusCan genomes only had 72.4% nucleotide identities, these are within the *Carlavirus* spp. demarcation of <72% identity (7). This high SPCFV genome sequence divergence provides no evidence of connectivity between East Timor and Australia. The same applies to connectivity between either of them and the Korean and Ugandan genomes, which differed by at least 13%.

Nucleotide sequence accession numbers. The sequences were deposited in GenBank under accession numbers KU720565 (Tm37) and KU707475 (AusCan).

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# REFERENCES

- 1. Aritua V, Barg E, Adipala E, Vetten HJ. 2007. Sequence analysis of the entire RNA genome of a sweet potato chlorotic fleck virus isolate reveals that it belongs to a distinct carlavirus species. Arch Virol 152:813–818. http://dx.doi.org/10.1007/s00705-006-0891-z.
- 2. Jones RAC, Dwyer GI. 2007. Detection of *Sweet potato chlorotic fleck virus* and *Sweet potato feathery mottle virus*—strain O in Australia. Australas Plant Pathol 36:591–594. http://dx.doi.org/10.1071/AP07069.
- Kwak H-R, Kim J, Kim M-K, Seo J-K, Jung M-N, Kim J-S, Lee S, Choi H-S. 2015. Molecular characterization of five potyviruses infecting Korean sweet potatoes based on analyses of complete genome sequences. Plant Pathol J 31:388–401. http://dx.doi.org/10.5423/PPJ.OA.04.2015.0072.
- 4. Ndunguru J, Taylor NJ, Yadav J, Aly H, Legg JP, Aveling T, Thompson G, Fauquet CM. 2005. Application of FTA technology for sampling, recov-

ery and molecular characterization of viral pathogens and virus-derived transgenes from plant tissues. Virol J 2:45. http://dx.doi.org/10.1186/1743 -422X-2-45.

- Kehoe MA, Coutts BA, Buirchell BJ, Jones RAC. 2014. Plant virology and next generation sequencing: experiences with a *Potyvirus*. PLoS One 9:e104580. http://dx.doi.org/10.1371/journal.pone.0104580.
- Deng X-G, Peng X-J, Zhu F, Chen Y-J, Zhu T, Qin S-B, Xi D-H, Lin H-H. 2015. A critical domain of *Sweet potato chlorotic fleck virus* nucleotidebinding protein (NaBp) for RNA silencing suppression, nuclear localization and viral pathogenesis. Mol Plant Pathol 16:365–375. http:// dx.doi.org/10.1111/mpp.12186.
- 7. King AMQ, Adams MJ, Lefkowitz EJ. 2011. Virus taxonomy: classification and nomenclature of viruses: ninth report of the international committee on taxonomy of viruses. Elsevier Academic Press, San Diego, CA.