Original Article

Lead exposure-induced changes in hematology and biomarkers of hepatic injury: protective role of TrévoTM supplement

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Abstract

Lead exposure has been linked to health challenges involving multiple organ failure. More than fifty percent of lead present in the human body is accumulated in the liver causing hepatic injury. A major mechanism of lead toxicity is oxidative stress. Trévo[™] is a nutritional supplement with numerous bioactive natural products with detoxifying and antioxidant properties. This study was designed to investigate the hepatoprotective effects of Trévo[™] dietary supplements against lead-hepatotoxicity in male Wistar rats. Thirty-five healthy animals were divided into five groups of seven each as follows: Group I=control; II=15 mg/kg of lead acetate (PbA); III= 2 mL/kg of Trévo™ + PbA; IV= 5 mL/kg of Trévo™ + PbA;V=5 mL/kg of Trévo™. Animals were orally treated with Trévo™ for two days before co-administration with PbA intraperitoneally for 12 consecutive days. Animals were sacrificed 24 h after the last administration and blood were collected via cardiac puncture and processed for hematological parameters and assessment of alanine aminotransferase (ALT), aspartate aminotransferase (AST), and albumin (ALB). The liver was excised and processed for markers of oxidative stress and histopathological examination. Intraperitoneal administration of 15 mg/kg of PbA caused a significant increase in serum concentration of AST, ALT, while the concentration of ALB was significantly decreased (P<0.001). PbA caused a significant reduction in packed cell volume, hemoglobin while the total white blood cell count, neutrophils, lymphocytes, monocytes, eosinophils, and basophils were increased. Oxidative stress was significantly pronounced in the liver of rats exposed to PbA as observed in the high concentration of malonedialdehyde, decreased concentration of glutathione, the activity of catalase, superoxide dismutase, and glutathione-S-transferase. Pretreatment with TrévoTM was able to significantly prevent the anemic, oxidative damage, and hepatic injury initiated by PbA. Histological examination also corroborated the biochemical results. In conclusion, the study reveals that Trévo™ is effective in attenuating PbA-induced hepatotoxicity in male Wistar rats.

Keywords: Trévo™, lead, hepatotoxicity, oxidative stress, hematology

Introduction

The liver is one of the largest and most important organs of the human body; it is the main organ for the detoxification and metabolism of drugs. Other functions of the liver include nutritional homeostasis, cholesterol, and glucose metabolism, and synthesis of clotting factors [1,2]. These functions have exposed it to the toxic effect of drugs and chemicals as a result of ingestion or other means of exposure to the chemicals. These toxic agents can be deleterious to the role of the liver as a detoxifying organ [3]. This chemical works by attacking functional macromolecules such as lipid, protein, and nucleic acids, either through the generation of free radicals, depletion of antioxidant molecules, inflammation, and apoptosis [4-6]. Some distortion in liver tissue includes damage to the membrane of the hepatocyte and organelles leading to swelling, damage, and necrosis. Health challenges and death from liver disorders are on the rise globally [7]. Some of the common liver diseases include biliary disease, hepatitis B and C, and alcoholic liver disease. Lead is one of the most toxic heavy metals that humans are often exposed to, making it a metal of public health concern. Lead exposure has been reported to be toxic to most organs, such as the liver, kidney, heart, brain, testes, and hematopoietic tissues [8-13]. Exposure to lead can be through various routes- orally through ingesting food and water contaminated with lead, inhalation of polluted air, from dust, burning fuel, and fossil. Due to the colorless and odorless nature of lead, it persists for a longer time in the environment. It can only be detected at a very high concentration, a stage where it becomes harmful to the environment and

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living organisms including humans. Sources of lead poison include contaminated food and water, lead-containing paint and gasoline, industrial emission [2,14]. Mudipalli [15] reported that the liver is the main storage organ of lead accumulation. More than 33% of accumulated lead in the human body is found in the liver, followed by the kidney. The animal experiment involving lead hepatotoxicity has shown that lead exposure altered enzymes and molecules involved in the metabolism of xenobiotics, metabolism of cholesterol, and liver hyperplasia. The toxicity of lead has been reported to cause liver injury, osteoporosis, neurological disorders, and cardiovascular diseases [3]. The accepted mechanism of lead toxicity is oxidative stress [2-4,16,17]. This is often achieved by generating free radicals and depleting the antioxidant systems. Lead has been reported to replace the cations in enzymes and proteins resulting in loss of activities and functions respectively [18]. Some of the macromolecules oxidized by lead include lipids (measured as malonedialdehyde (MDA), reduced glutathione (GSH), and antioxidants such as superoxide dismutase (SOD) and catalase (SOD)). The role of natural products in combatting the toxic effects of heavy metals and other poisonous chemicals is on the rise. Since the major mechanism of lead toxicity is oxidative stress, natural products rich in antioxidants can be a good antidote against lead poison and can be used along with common lead chelators. Several compounds from natural products with confirmed antioxidant activities have been used as a hepatoprotective agent against lead position [2,3,7,16,17].

Trévo™ is a phytonutrient supplement that is manufactured in the USA under the trade name TRÉVO (TRÉVO LLCTM) by United Int'l Lab LLC, TX 75244, USA. Trèvo formula contains potential antioxidants according to the manufacturer. It contains more than 174 ingredients extracted from various plant sources. The constituents involve many ingredients from phytonutrients of common garden fruits and vegetables, herbs, and coral calcium complex containing vitamins (A, C, and E), amino acids, trace minerals, essential fatty acids, digestive enzymes, and coenzymes, apart from primary essential antioxidants. Trévo™ is a mixture of various plant-rich extracts such as maqui berry, amalaki fruit, schizandra fruit, borojo fruit, goji fruit, noni, mangosteen, acai berry, gac fruit, camu camu, pomegranate, star fruit, and acerola cherry, antioxidant compounds such as beta-carotene, lycopene, ascorbic acid, α -tocopherol, and retinoic acid, essential fatty acids such as Omega-3 and Omega-6 from borage seed oil and flaxseed oil. All these substances have been reported to boost the immune system, cardiovascular functions, reproductive and nervous systems. In addition, these substances also have the potential to repair and regenerate new cells. According to the manufacturer, daily intake of Trévo™ provides the body with a high concentration of antioxidants that can prevent the generation of free radicals, thus countering or slowing down the effect of toxic chemicals and poison. Recent investigations show that Trévo™ was protective against acetaminophen, cyanide-induced liver damage [19,20], in addition to its neuroprotective and cardioprotective activities [21, 22]. This study investigated the effectiveness of Trévo[™] to protect against lead-induced biochemical and histological changes in the liver of male Wistar rats.

Materials and Methods

Chemicals and reagents

Reduced glutathione, Trévo[™] was a product of Trèvo[™] LLC, Oklahoma City, USA. Other chemicals were of analytical grade.

Animal care and handling

Thirty-five male Wistar rats weighing 170±10g were purchased from the Central Animal House, University of Benin, Edo State, Nigeria were used for this experiment. The animals were housed in well-ventilated cages and provided water and food *ad libitum*.

Experimental design

Male Wistar rats were randomly divided into 5 groups of 7 rats each as follows:

Group 1: administered vehicle (distilled water)

Group 2: administered 15 mg/kg of lead acetate (PbA) intraperitoneally for 12 consecutive days [4].

Group 3: orally administered 2 mL/kg of Trévo[™] for 2 days before co-administration with PbA for 12 consecutive days. Group 4: orally administered 5 mL/kg of Trévo[™] for 2 days before co-administration with PbA for 12 consecutive days. Group 5: orally administered 5 mL/kg of Trévo[™] for 14 consecutive days.

Processing of the liver

24 h after last administration, animals were sacrificed via cervical dislocation, and the liver excised, rinsed, and homogenized in a phosphate buffer saline (0.1M, pH 7.4) to obtain a 10% w/v homogenate. The homogenate was centrifuged at 15000 rpm for 10 min with the temperature set at 4 $^{\circ}$ C to obtain a clear supernatant that was used for biochemical assays.

Biomarkers of liver function

Serum activity of alanine aminotransferase, aspartate aminotransferase, and concentration of albumin was

determined in the serum following the instruction from the kit manual.

Biochemical assay

Estimation of oxidants in the hepatic tissues

Lipid peroxidation was determined by measuring the formation of thiobarbituric acid reactive substances according to the method of Varshney and Kale [23]. The malondialdehyde level was calculated using a molar extinction coefficient of 1.56×105 M -1 cm -1.

Estimation of antioxidants in the hepatic tissues

GSH content was estimated according to the method of Jollow et al. [24]. The reaction is based on the fact that the thiol group of GSH reacts with 5, 5'-dithiobis-(2-nitrobenzoic acid) to form thionitrobenzoic acid, which was monitored at 412 nm. Catalase (CAT) activity was determined as described by [25]. Superoxide dismutase (SOD) activity was evaluated by the method of Misra and Fridovich [26] by monitoring the absorbance of a mixture containing the brain supernatant, carbonate buffer (0.05 M, pH 10.2), and adrenalin (0.3 mM) for 150 s at 480 nm.

GST assay

Glutathione transferase (GST) activity was determined by measuring the conjugation of GSH with 1-chloro-2, 4dinitrobenzene at 340 nm as described by Habig et al. [27].

Histopathological evaluation

Liver tissues were taken from the eviscerated rats and fixed in 10% formalin for 24 h, and then processed to obtain paraffin blocks. Sections of 4-6 µm thickness were cut using a microtome and stained with hematoxylin and eosin (H and E) stain by using the method of Stevens and Wilson [28].

Statistical analysis

All grouped data were statistically performed with Prism (GraphPad Prism, 6.01) software. Differences among groups were evaluated by one-way analysis of variance followed by Duncan's multiple range tests. All values were expressed as the mean±standard deviation of seven animals per group.

Results

Effect of lead acetate (PbA) and pretreatment with Trévo™ (2 and 5 mL/kg) on hematological parameters in male Wistar rats.

Table 1 shows that PbA caused a significant decrease in PCV and a mild decrease in Hb as compared to the control (P<0.05), while the level of WBC, Neu, lymph, mono, eosin, and baso significantly increased when compared with the control (P<0.05). Pretreatment with TrévoTM (2-and 5 mL/kg) was able to prevent the anemic and inflammation induced by PbA as observed in the significant increase in PCV and Hb level as compared to the untreated group (P<0.05). In addition, the level of other blood parameters (Neu, lymph, mono, eosin, and baso) was significantly decreased as compared to the untreated group (P<0.05). Administration of the rats with 5 mL/kg of TrévoTM had no significant effect on hematology parameters when compared to the control (P<0.05)

Group	PCV (HB	TOTAL WBC count	NEUT (LYM (MONO (x1	EOSINO (x1	BASO (x1
	%)	(g/dL)	(g/dL)	%)	%)	0ºL)	0ºL)	0ºL)
Control	44.7±5.	14.9±1.	5.4±0.10	49.7±2.6	35.5±3.	5.9±0.69	2.3±0.15	0.2±0.01
	77	54		3	98			
PbA	29.7±3.	10.2±1.	9.0±1.05*	55.3±5.9	47.2±3.	7.4±0.51*	3.7±0.03*	0.6±0.04*
	07*	67*		6*	19*			
2 mL/kg Trévo™	42.3±4.	14.1±1.	5.9±0.79 [#]	50.0±4.5	39.3±4.	7.0±0.89	3.0±0.08#	0.3±0.03#
+PbA	46#	49#		4#	04#			
5 mL/kg Trévo™	43.7±3.	14.6±1.	8.2±0.52#	51.4±4.3	40.9±4.	6.7±0.89#	3.0±0.14#	0.0±0.00#
+PbA	89#	16#		0#	50#			
5 mL/kg Trévo™	42.3±3.	14.1±1.	5.5±0.62	48.0±5.0	30.0±3.	5.7±0.60	2.0±0.02	0.0±0.00
	72	26		6	68			

Table 1. The effect of Trévo[™] on blood hematology of rats exposed to PbA.

Data are mean±SD (n=7). Statistically significant differences:

*P<0.05= control group vs. PbA; #P<0.05= PbA vs. treatment

PCV=Packed Cell Volume; Hb= Hemoglobin; WBC= White Blood Cells; NEUT= neutrophil; LYM= lymphocyte; MONO= monocyte; EOSINO=eosinophil; BASO= basophil; PbA= lead acetate

Effect of lead acetate (PbA) and pretreatment with Trévo[™] (2 and 5 mL/kg) in serum markers of hepatotoxicity.

Table 2 shows that exposure of rats to lead acetate caused a significant increase in AST, ALT, and ALB levels as compared to the control group (P<0.001). Pretreatment of the rat with 2- and 5 mL/kg of TrévoTM reduced the hepatotoxic effect of PbA as observed in the significant decrease in the serum level of AST, ALT, and ALB when compared to PbA (P<0.05). Treatment with 5 mL/kg of TrévoTM had no significant effect on serum markers of hepatotoxicity when compared to the control (P>0.05).

Table 2. Serum ALT and AST activities and tota	albumin levels in control and in	n all experimental groups.
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Groups	AST (IU)	ALT (IU)	ALB (mg/dL)
Control	13.6±1.51	8.5±0.97	7.0±0.33
PbA	19.5±2.07***	14.5±1.56***	3.6±0.26***
2 mL/kg Trévo™ +PbA	18.0±1.10	8.1±0.52###	5.6±0.62###
5 mL/kg Trévo™ +PbA	17.7±1.10	11.5±1.14###	6.0±0.59###
5mL/kg Trévo™	13.4±1.21	8.0±0.53	7.1±0.30

Data are mean±SD (n=7). Statistically significant differences:

***P<0.001= control group vs. PbA; ###P<0.001 PbA vs. treatment

ALT = alanine transaminase; AST= aspartate transaminase; ALB= albumin; PbA= lead acetate

Effect of PbA and TrévoTM on markers of oxidative stress

Figure 1 and 2 shows the effect of PbA and pretreatment with Trévo™ on the concentration of malonedialdehyde (MDA) and reduced glutathione (GSH) respectively.

PbA caused an insignificant increase (P<0.05) in the level of MDA and a significant decrease (P<0.05) in the level of GSH when compared to the control. Pretreatment with TrévoTM (2- and 5 mL/kg) was able to reduce the level of MDA, though not significantly different when compared to the untreated group (P>0.05). For GSH, pretreatment of the rats with 2 mL/kg of TrévoTM had no significant effect on GSH concentration, when compared to the untreated group (P>0.05), while pretreatment with 5 mL/kg of TrévoTM caused a significant increase in the concentration of GSH when compared to the untreated group (P<0.05). Administration of 5 mL/kg of TrévoTM only to the rats had no significant effect on GSH concentration when compared to the control group (P>0.05).

Figures 3 and 4 show the effect of PbA and pretreatment with Trévo™ on catalase (CAT) and superoxide

dismutase (SOD) activities respectively. PbA caused a significant decrease in CAT and SOD activities as compared to the control (P<0.001 and 0.05 respectively). Pretreatment with 2- and 5 mL/kg of Trévo[™] was able to prevent the inhibitory effect of PbA on CAT and SOD as observed in the significant increase activity of CAT and SOD as compared to the untreated group (P<0.05 and 0.001 respectively). The result also showed that a high dose of Trévo[™] was more effective in increasing the activity of CAT and SOD than a low dose. Administration of rats with 5 mL/kg of Trévo[™] had no significant effect on the activities of CAT and SOD as compared to the control (P>0.05).



Figure 1. The concentration of lipid peroxides product - malondialdehyde (MDA), in the liver tissue of male rats after 2 day-pretreatment with $Trévo^{TM}$ and 12-day co-administration with lead acetate (15 mg/kg) via intraperitoneal administration. Data are shown as mean±standard deviation (SD) for 7 animals.



Figure 2. The concentration of non-enzymatic antioxidant- reduced glutathione (GSH) in the liver tissue of male rats after 2 day-pretreatment with Trévo[™] and 12-day co-administration with lead acetate (15 mg/kg) via intraperitoneal administration. Data are shown as mean±standard deviation (SD) for 7 animals. Statistically significant differences: ***P<0.001= control vs. PbA; [#]P<0.05= PbA vs. Trévo[™] (5 mL/kg) + PbA



Figure 3. The catalase activity in the liver tissue of male rats after 7 day-pretreatment with Trévo[™] and 12-day concomitant exposure to lead acetate (15 mg/kg) via intraperitoneal administration. Data are shown as mean±standard deviation (SD) for 7 animals. Statistically significant differences: ***P<0.001= control vs. PbA; ^{##}P<0.001= PbA vs. treatment groups



Figure 4. The superoxide dismutase (SOD) activity in the kidney tissue of male rats after 7 day-pretreatment with TrévoTM and 12-day concomitant exposure to lead acetate (15 mg/Kg) via intraperitoneal administration. Data are shown as mean±standard deviation (SD) for 7 animals. Statistically significant differences: ***P<0.001= control vs. PbA; $^{*}P<0.05=$ PbA vs. TrévoTM (2 mL/kg) + PbA; $^{**}P<0.01=$ PbA vs. TrévoTM (5 mL/kg) + PbA



Figure 5. The glutathione-S-transferase (GST) activity in the liver tissue of male rats after 7 day-pretreatment with TrévoTM and 12-day concomitant exposure to lead acetate (15 mg/kg) via intraperitoneal administration. Data are shown as mean±standard deviation (SD) for 7 animals. Statistically significant differences: ***P<0.001= control vs. PbA; ***P<0.01= PbA vs. TrévoTM (2 mL/kg) + PbA; ***P<0.001= PbA vs. TrévoTM (5 mL/kg) + PbA

Effect of pretreatment with Trévo[™] and lead acetate (PbA) on glutathione-S-transferase (GST) activity.

PbA caused a significant decrease in the activity of GST as compared to control (P<0.001). However, pretreatment with 2-and 5 mL/kg of TrévoTM significantly increase the activity of GST as compared to the PbA group (P<0.05). An interesting observation is a significant decrease in the GST activity in animals administered only TrévoTM when compared to the control (P<0.05).

Histology

Liver from PbA administered rats was examined for histopathological changes. As shown in Figure 6 rats administered PbA only shows diffuse hepatic vacuolation with periportal hepatic necrosis and cellular infiltration, a situation that was ameliorated in the group pretreated with TrévoTM 2-and 5 mL/kg) before the co-administration with PbA with very mild portal necrosis.



Figure 6. Photomicrograph of rat liver sections stained with hematoxylin and eosin (X100) showing alterations in the liver after treatment with trévo and exposure to PbA. The blue arrow indicates congestion. The green arrow indicates restoration and normal architecture of hepatic cords. The red arrows indicate normal hepatic artery and portal vein.

Discussion

The hepatotoxicity of PbA has been linked to its ability to form a complex with some proteins abundant in the liver. In addition, the liver is one of the major organs involved in Pb excretion. We hypothesized that Trévo™ a supplement drink that contains numerous phytochemicals with metal chelating properties can prevent lead hepatotoxicity. Our results show that Trévo™ at the two doses based on the method of [19] was able to prevent PbA-induced hepatotoxicity. The anemic effect of PbA as observed in the low percentage of PCV and hemoglobin was reversed by treatment with Trévo™, which was able to increase the percentage of PCV and Hb volume. Treatment with Trévo™ also reduced the total WBC and other blood parameters such as neutrophil, lymphocyte, monocyte, eosinophil, and basophil. A decrease in Hb content by PbA has been linked to the ability of PbA to inhibit erythropoiesis or its binding to RBC, which often results in low Hb content and short life for RBC respectively. Another anemic effect of PbA can also be due to its displacement of Fe from Hb or inhibiting the activity of aminolevulinic acid dehydratase (ALAD), all culminating in the reduction of Hb and lifespan of RBCs. The anemic effect of toxic heavy metals has been reported by various investigators and all supported our findings [28-32]. PbA administration for 10 days caused a significant increase in serum levels of AST ad ALT. The increased level of these enzymes might be due to leakage from the tissue as a result of damage caused by PbA. This observation is similar to the report of [2,3]. They reported that elevated serum levels of ALT and AST can be linked to damage of the membrane due to oxidative stress induced by PbA. This increased the permeability of the enzymes from the hepatocytes into blood circulation. In addition, PbA exposure also caused a significant decrease in serum albumin. Albumin plays an important role in blood detoxification. The depletion of albumin can further enhance the toxicity of PbA. One of the ways by which PbA caused albumin reduction is through forming a complex with the protein, thereby making it lose its biological function [33,34]. Pretreatment with Trévo™ before concurrent administration of PbA caused a significant reduction in the serum level of AST ad ALT and increase concentration of albumin. The hepatoprotective effect of Trévo™ against cyanide ad acetaminophen-induced liver injury has been reported [21,22]. Trévo ameliorated the hepatotoxicity of PbA by preventing damage to the hepatic membrane and leakage of the enzyme into the blood. In addition, it is possible that Trévo™ chelate PbA, thus preventing it from binding to a functional molecule such as albumin. Our result also supported the oxidative stress-induced mechanism of PbA as reported by other researchers [2-4,16,17]. PbA exposure caused a mild increase in MDA level as well as a significant decrease in GSH concentration, CAT, SOD, and GST activities in liver tissue. The mechanism of PbA-induced oxidative stress involves, might involve increased production of ROS as a result of damage to enzymes involved in mitochondria respiration. The ROS generated react with biomolecules increasing peroxide products such as lipid peroxide ad protein carbonyl, further stressing the antioxidant system, as observed in the low level of CAT, SOD, and GST. The histopathological examination of the liver tissue in rats administered PbA corroborates the findings of the biochemical assays. Morphological changes observed include congestion of the central venules, hepatic necrosis, and abscess in the liver parenchyma decreased portal triads and necrotized hepatocytes. Pretreatment with Trévo™ prevents the morphological changes observed. Some of the phytochemicals present in Trévo[™] have been reported to improve liver functions and the purification of the organ. Pomegranate, Amalaki, and Phyllanthus emblica [36,37]. These are compounds with established antioxidant and hepatoprotective properties. Therefore, the ability of Trévo™ to prevent enzyme leakages and the oxidative effect of PbA might not be unconnected with the phytochemicals present in the product.

Conclusions

In conclusion, our results show that Trévo[™] was hepatoprotective against lead toxicity and the activity might be linked to the presence of numerous antioxidants phytochemicals present in Trévo[™].

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Conflict of interest

The authors declare no competing interest.

CRediT author statement

OBI: Conceptualization, Methodology, Investigation, Data Curation, Writing- Original draft preparation, EFA: Writing-Review & Editing, Formal analysis; TTO: Conceptualization, Data curation; BC: Resources

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References

- [1] Bischoff K, Mukai M, Ramaiah SK. Liver toxicity. In Veterinary Toxicology; Academic Press Elsevier: Amsterdam, The Netherlands 2018. 239–257.
- [2] Al-Megrin WA, Alkhuriji AF, Yousef AOS, Metwally DM, Habotta OA, Kassab RB, et al. Antagonistic efficacy of luteolin against lead acetate exposure-associated with hepatotoxicity is mediated via antioxidant, anti-inflammatory, and antiapoptotic activities. Antioxidants 2019;9(1):10. <u>https://doi.org/10.3390/antiox9010010</u>
- [3] Alhusaini A, Fadda L, Hasan IH, Zakaria E, Alenazi AM, Mahmoud AM. Curcumin ameliorates lead-induced hepatotoxicity by suppressing oxidative stress and inflammation, and modulating Akt/GSK-3 β Signaling Pathway. Biomolecules 2019;9(11):703. https://doi.org/10.3390/biom9110703
- [4] Abdelhamid FM, Mahgoub HA, & Ateya AI. Ameliorative effect of curcumin against lead acetate-induced hematobiochemical alterations, hepatotoxicity, and testicular oxidative damage in rats. Environmental Science and Pollution Research 2020;27(10):10950-10965. <u>https://doi.org/10.1007/s11356-020-07718-3</u>
- [5] Hsu PC, Guo YL. Antioxidant nutrients and lead toxicity. Toxicology 2002;180(1):33-44. <u>https://doi.org/10.1016/s0300-483x(02)00380-3</u>
- [6] Sabath E, Robles-Osorio ML. Medio ambiente y riñón: nefrotoxicidad por metales pesados. Nefrología (Madrid) 2012;32(3):279-286. <u>https://doi.org/10.3265/Nefrologia.pre2012.Jan.10928</u>
- [7] Al-Dbass AM, Al- Daihan SK, Bhat RS. Agaricus blazei Murill as an efficient hepatoprotective and antioxidant agent against CCl4-induced liver injury in rats. Saudi Journal of Biological Sciences 2012;19(3):303-309. <u>https://doi.org/10.1016/j.sjbs.2012.03.004</u>
- [8] Assi MA, Hezmee MNM, Abd Wahid Haron MYM, Sabri MAR. The detrimental effects of lead on human and animal health. Vet. World 2016;9(6):660. <u>https://doi.org/10.14202/vetworld.2016.660-671</u>
- [9] Basgen JM, Sobin C. Early chronic low-level lead exposure produces glomerular hypertrophy in young C57BL/6J mice. Toxicol. Lett. 2014;225(1):48-56. <u>https://doi.org/10.1016/j.toxlet.2013.11.031</u>
- [10] Mujaibel LM, Kilarkaje N. Mitogen-activated protein kinase signaling and its association with oxidative stress and apoptosis in lead-exposed hepatocytes. Environ. Toxicol. 2015;30(5):513-529. <u>https://doi.org/10.1002/tox.21928</u>
- [11] Liu J, Lewis G. Environmental toxicity and poor cognitive outcomes in children and adults. J. Environ. Health 2014;76(6):130.
- [12] Pant N, Kumar G, Upadhyay AD, Patel DK, Gupta YK, Chaturvedi PK. Reproductive toxicity of lead, cadmium, and phthalate exposure in men. Environ. Sci. Pollut. Res. 2014;21(18):11066-11074. <u>https://doi.org/10.1007/s11356-014-2986-5</u>
- [13] Abdel Moneim AE. Flaxseed oil as a neuroprotective agent on lead acetate-induced monoamineric alterations and neurotoxicity in rats. Biol Trace Elem Res 2012;148(3):363-370. <u>https://doi.org/10.1007/s12011-012-9370-4</u>
- [14] Wu X, Cobbina SJ, Mao G, Xu H, Zhang Z, Yang L. A review of toxicity and mechanisms of individual and mixtures of heavy metals in the environment. Environ Sci Pollut Res 2016;23(9):8244-8259. <u>https://doi.org/10.1007/s11356-016-6333-x</u>
- [15] Mudipalli A. Lead hepatotoxicity & potential health effects. Indian J Med Res 2007;126(6):518.
- [16] Adli DEH, Brahmi M, Kahloula K, Arabi W, Bouzouira B, Talatizi M, et al. The therapeutic effect of Cinnamomum cassia essential oil against hepatotoxicity induced by co-exposure to lead and manganese in developing Wistar rats, Journal of Drug Delivery and Therapeutics 2020;10(5):49-55.
- [17] Chen C, Lin B, Qi S, He J, Zheng H. Protective effects of salidroside on lead acetate-induced oxidative stress and hepatotoxicity in Sprague-Dawley Rats. Biological Trace Element Research 2019;191(2):426-434 <u>https://doi.org/10.1007/s12011-019-1635-8</u>
- [18] Flora SJ, Flora G, Saxena G. Environmental occurrence, health effects and management of lead poisoning. In Lead Chemistry, Analytical Aspects, Environmental Impacts and Health Effects; Cascas, S.B., Sordo, J., Eds.;Elsevier Publication: Amsterdam, The Netherlands 2006;158-228. <u>https://doi.org/10.1016/B978-044452945-9/50004-X</u>

- [19] Ilesanmi OB, Ikpesu T. Neuromodulatory activity of trèvo on cyanide-induced neurotoxicity viz neurochemical, antioxidants, cytochrome C oxidase and p53. Advances in Traditional Medicine 2021;21(2):297-304. <u>https://doi.org/10.1007/s13596-020-00450-w</u>
- [20] Ilesanmi OB, Atanu FO, Odewale TT, Adeogun E, Chikere BN, Alaneme CU, et al. Effect of a phytonutrient-rich product and administration time on cyanide-induced cardiotoxicity. Trop J Nat Prod Res 2020;4(7):304-309.
- [21] Akinmoladun AC, Oguntunde KO, Owolabi LO, Ilesanmi OB, Ogundele JO, Olaleye MT, et al. Reversal of acetaminophen-generated oxidative stress and concomitant hepatotoxicity by a phytopharmaceutical product. Food Science and Human Wellness 2017;6(1):20-27. <u>https://doi.org/10.1016/j.fshw.2016.11.001</u>
- [22] Babatunde IO, Abigail A. Ameliorative effect of a multi-nutrient-rich product against cyanide induced hepatorenotoxicity. Drug Discovery 2021;15(35):34-42.
- [23] Varshney R, Kale RK. Effects of calmodulin antagonists on radiation-induced lipid peroxidation in microsomes. Int J Radiat Biol 1990;58(5):733–743. <u>https://doi.org/10.1080/09553009014552121</u>
- [24] Jollow DJ, Mitchell JR, Potter WZ, Davis DC, Gillette JR, Brodie BB. Acetaminophen-induced hepatic necrosis. II. Role of covalent binding in vivo. J Pharmacol Exp Ther 1973;187(1):195-202.
- [25] Aebi H. Methods of Enzymatic Analysis 2nd ed. Academic Press; 1974;2:673-684. <u>https://doi.org/10.1016/B978-0-12-091302-2.X5001-4</u>
- [26] Misra HP, Fridovich I. The univalent reduction of oxygen by reduced flavins and quinones. J Biol Chem 1972;247(1):188-192. <u>https://doi.org/10.1016/S0021-9258(19)45773-6</u>
- [27] Habig WH, Pabst MJ, Jakoby WB. Glutathione S-transferases: the first enzymatic step in mercapturic acid formation. J Biol Chem 1974;249(22):7130-7139. <u>https://doi.org/10.1016/S0021-9258(19)42083-8</u>
- [28] Dewanjee S, Sahu R, Karmakar S, Gangopadhyay M. Toxic effects of lead exposure in Wistar rats: involvement of oxidative stress and the beneficial role of edible jute (Corchorus olitorius) leaves. Food Chem Toxico 2013;55:78–91. <u>https://doi.org/10.1016/j.fct.2012.12.040</u>
- [29] Yuan G, Dai S, Yin Z, Lu H, Jia R, Xu J, et al. Toxicological assessment of combined lead and cadmium: acute and subchronic toxicity study in rats. Food Chem Toxicol 2014; 65:260-268. <u>https://doi.org/10.1016/j.fct.2013.12.041</u>
- [30] Abdel-Moneim AM, El-Toweissy MY, Ali AM, Awad Allah AAM, Darwish HS, Sadek IA. Curcumin ameliorates Lead (Pb²⁺)-induced hemato-biochemical alterations and renal oxidative damage in a rat model. Biological trace element research 2015;168(1):206-220. <u>https://doi.org/10.1007/s12011-015-0360-1</u>
- [31] El-Boshy ME, Refaat B, Qasem AH, Khan A, Ghaith M, Almasmoum H, et al. The remedial effect of Thymus vulgaris extract against lead toxicity-induced oxidative stress, hepatorenal damage, immunosuppression, and hematological disorders in a rats. Environmental Science and Pollution Research 2019;26(22):22736-22746. https://doi.org/10.1007/s11356-019-05562-8
- [32] Offor SJ, Mbagwu HO, Orisakwe OE. Lead induced hepato-renal damage in male albino rats and effects of activated charcoal. Front Pharmacol 2017;8:107. <u>https://doi.org/10.3389/fphar.2017.00107</u>
- [33] Mahaldar K, Hossain A, Islam F, Islam S, Islam MA, Shahriar M, et al. Antioxidant and hepatoprotective activity of Piperretro fractum against Paracetamol-induced hepatotoxicity in Sprague-Dawley rat. Natural Product Research 2020; 34(22):3219-3225. <u>https://doi.org/10.1080/14786419.2018.1550768</u>
- [34] Ahmed RA. Hepatoprotective and antiapoptotic role of aged black garlic against hepatotoxicity induced by cyclophosphamide. The Journal of Basic and Applied Zoology 2018;79(1):1-8. <u>https://doi.org/10.1186/s41936-018-0017-7</u>
- [35] Huang CZ, Tung YT, Hsia SM, Wu CH, Yen GC. The hepatoprotective effect of Phyllanthus emblica L. fruit on high fat dietinduced non-alcoholic fatty liver disease (NAFLD) in SD rats. Food Funct. 2017;8(2):842-850. <u>https://doi.org/10.1039/C6FO01585A</u>
- [36] Kumar V, Kshemada K, Ajith KG, Binil RS, Deora N, Sanjay G, et al. Amalaki rasayana, a traditional Indian drug enhances cardiac mitochondrial and contractile functions and improves cardiac function in rats with hypertrophy. Sci Rep 2017;7(1):1-17. <u>https://doi.org/10.1038/s41598-017-09225-x</u>

[37] Husain H, Latief U, Ahmad R. Pomegranate action in curbing the incidence of liver injury triggered by Diethylnitrosamine by declining oxidative stress via Nrf2 and NFκB regulation. Sci Rep 2018;8(1):1-17. https://doi.org/10.1038/s41598-018-26611-1