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Synovial mast cells and osteoarthritis: Current understandings and future perspectives

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ABSTRACT

Osteoarthritis (OA) is a prevalent joint disease worldwide that significantly impacts the quality of life of individuals, particularly those in middle-aged and elderly populations. OA was initially considered as non-inflammatory arthritis, but recent studies have identified a substantial number of immune responses in OA, leading to the recognition of inflammation as a key factor in its pathogenesis. An increasing number of studies have found that mast cell (MC) and MC-secreted inflammatory mediators and cytokines are notably increased in the synovial fluid of OA patients, indicating a potential association between MCs and the onset and progression of synovial inflammation. The present review aims to summarize the significance and mechanism of MCs in the pathogenesis of OA. Meanwhile, we also discuss the clinical potential of using MCs as therapeutic target for OA therapy. Modulating the activities of MCs or the mediators of MCs in the synovial fluid inflammatory microenvironment will be promising new options for the treatment of OA.

1. Background

Osteoarthritis (OA) is an increasingly common joint disease worldwide, affecting estimated 240 million individuals [[1](#page-8-0)]. Clinical manifestations of OA typically include joint pain, stiffness, and limited range of motion. The disease progression is usually gradual, but it can eventually lead to joint failure, resulting in persistent pain and disability [\[2\]](#page-8-0). While OA can impact any joint, it is most commonly observed in the knee. The development of OA may be associated with factors such as obesity, age, joint trauma, and biomechanical changes [\[3,4](#page-8-0)]. Interestingly, gender may also contribute to the pathogenesis of OA, as studies have shown a higher prevalence of OA in females than in males [\[5\]](#page-8-0). OA was initially thought to be a non-inflammatory form of arthritis, but as a large number of immune

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responses have been identified in OA, it has been increasingly recognized that inflammation is involved in the pathogenesis of OA [\[6](#page-8-0)–8]. The current treatment strategies for OA include drug therapy (NSAIDs), nondrug therapy (such as education, weight loss, exercise, and physical therapy) $[9,10]$ $[9,10]$ $[9,10]$ $[9,10]$, and surgery. However, surgical treatment can be associated with complications and high costs [\[11](#page-8-0)]. In order to effectively treat the disease, it is crucial to focus on controlling the development of OA.

Mast cells (MCs) are key structural and functional components of the innate and adaptive immune systems and play an important role in responding to inflammation and infection [\[12](#page-8-0)]. Degranulation is the process by which various particles are released from MCs, serving as the primary allergic reaction of MCs. The particles primarily released during this process include histamine, serotonin, proteases, tryptase, lipid mediators (prostaglandins, leukotrienes), as well as cytokines, chemokines and reactive oxygen species [[13\]](#page-8-0). When MCs are not activated properly, they can lead to various diseases, such as allergic diseases and autoimmune diseases [\[14](#page-8-0),[15\]](#page-8-0). MCs in OA were mainly divided into two subgroups according to protease content: the MC_T subgroup expressing tryptase and the MC_{TC} subgroup expressing tryptase, chymotrypsin and carboxypeptidase A3 (Cpa3) [\[16,17](#page-8-0)]. The number of MC_{TC} cells is dominant in normal joint tissues; however, in the tissue of patients with OA and other inflammatory disease, there is a significant increase in the number of cells in the MC_T subgroup $[18,19]$ $[18,19]$. This review aims to enhance the understanding of the biological significance and mechanism of MCs in OA. In addition, we summarized the role of MCs in the synovial fluid microenvironment, offering valuable insights for future OA treatments.

2. MCs and OA

2.1. MCs are involved in the pathogenesis of OA

Multiple studies have provided evidence supporting the involvement of MCs in the pathophysiology of OA. The number of MCs and the and the proportion of degranulation in the synovium of patients with OA were significantly increased, which was characterized by an increase in the release of histamine, Prostaglandin D2 (PGD2) and tryptase [\[20](#page-9-0)–22]. Besides, the number and degranulation status of MCs were positively correlated with the synovitis score and cartilage injury [[23,24\]](#page-9-0). In addition to clinical data, experimental evidence also suggests that MCs have an impact on the progression of OA. Proteomics analysis revealed the accumulation of specific inflammatory proteins in MCs after 9 days of incubation in OA synovial fluid. This accumulation of inflammatory proteins in MCs may further exacerbate the inflammatory process associated with OA [\[20](#page-9-0)]. The injection of MCs exacerbated the development of monosodium iodoacetate-induced OA in mice and stimulated the release of inflammatory cytokines [\[25](#page-9-0)]. Genetic deficiency or pharmacologic inhibition of MCs has been shown to provide protection against the development of OA in mice [\[26,27](#page-9-0)]. These evidences indicate that activated MCs release inflammatory factors through degranulation, thus participating in the pathogenesis of OA.

2.2. Activation pathways of MCs in the pathogenesis of OA

In the microenvironment of osteoarthritis, inflammatory mediators activate MCs. Below we describe the pathways involved in MCs activation.

SCF/C-Kit Receptor Stem cell factor (SCF) is a growth factor expressed by endothelial cells and fibroblasts. It is transmembrane protein synthesized in a soluble or membrane-bound form by enzymatic cleavage of two alternatively spliced mRNAs [[28](#page-9-0)]. SCF exerts its biological functions by binding to membrane tyrosine kinase receptors (C-Kit). The development and survival of MCs are critically dependent on SCF/C-Kit. SCF binds two c-Kit monomers and enables interactions between IgG-like domains 4 and 5 of adjacent c-Kit molecules. The homodimeric state of C-Kit is induced by SCF and subsequently stabilized by IgG 4/5-like domain interactions, allowing for efficient trans-phosphorylation in the lateral membrane region, kinase insert region, kinase domain, and terminal COOH tail. Subsequently, the phosphorylated residues act as docking sites for adjacent signaling molecules, including Src kinase, PI3K, Shc, and phospholipase Cγ (PLCγ). This interaction leads to the activation of the RAS-RAF-MAP kinase (MAPK) cascade, which in turn enhances the activation of transcription factors necessary for various biological functions, results in the promotion of MC proliferation [\[29](#page-9-0),[30\]](#page-9-0). Co-sensing of ATP and IL-33 results in the overactivation of MCs [\[31](#page-9-0)]. A study indicates that the activation of SCF/C-kit is essential for the cytokine response necessary for the co-sensing of ATP and IL-33, thereby facilitating the expansion of pro-inflammatory cytokines and eicosanoid production [[32\]](#page-9-0). This mechanism may be crucial for mast cells in mediating heightened inflammatory responses. Besides, in inflammatory and tumor environment, SCF activates C-Kit receptor to induces MC degranulation, resulting in the expression and release of histamine, proinflammatory cytokines, and chemokines [[33,34\]](#page-9-0). Inhibitor of the SCF/C-Kit pathway, such as RIN3 and β-eudesmol, markedly suppressed SCF-induced MCs migration and reduced MCs infiltration and inflammatory factor content in inflammatory sites [[35,36](#page-9-0)].

IgE/FcεR1 A large number of studies have shown that immunoglobulin E (IgE) and MCs are key factors in the long-term patho-physiological process and tissue remodeling of allergic diseases [\[37,38](#page-9-0)]. According to the structure of FcεR1, the α chain serves as the IgE binding site, and the two γ subunits participate in the initiation and propagation of downstream signals of FceR1. When bound to antigen, FcεR1 forms dimers. Cytoplasmic signaling is activated through the binding of the Src family protein tyrosine kinase Lyn and phosphorylation of the 'immunoreceptor tyrosine activator' (ITAM) located at the terminal of the β and γ subunits. Syk binds to the γ moiety of ITAM and facilitates the phosphorylation of multiple targets [39–[41\]](#page-9-0). During a type I hypersensitivity, the release of IgE and its binding to FcεRI on the surface of tissue-resident MCs initiate the activation of the FcεRI signaling cascade. This process triggers degranulation and the subsequent release of pro-inflammatory cytokines of MCs [[41,42\]](#page-9-0). Early studies have shown that inhibiting the IgE-activated MCs can decrease the release of inflammatory factors and the stimulation of chondrocytes, ultimately leading to improvements in pain and disability among patients with OA [[43,44](#page-9-0)]. Subsequently, a study clarified that, MCs activated by IgE/FcεR1

pathway promote its downstream targets such as MAPK through Syk signaling pathway in OA, resulting in the release of multiple pro-inflammatory factors such as IL-1β, IL-6, CCL2, ADAMTS4 and MMP13, which is not only an important mechanism for the formation of inflammatory environment in osteoarthritis. It also leads to chondrocyte apoptosis, and cartilage breakdown [\[26\]](#page-9-0).

IgG/FCγR Synovial MCs in patients with RA and OA express the IgG receptors FcγRI and FcγRII, but they do not express FcγRIII. When exposed to a specific stimulus, such as IFN- γ , the amount of FC γ R on the MCs surface increases substantially [\[45,46](#page-9-0)]. Fc γ RI mediates the substantial MC degranulation and increase the production of cytokines such as PGD2, tumor necrosis factor (TNF)-α, granulocyte-macrophage colony stimulating factor (GM-CSF), IL-3, and IL-13. Besides, FcγRI activation of MCs through IgG promotes the expression of pro-survival protein A1/Bfl-1 [\[47](#page-9-0),[48\]](#page-9-0). In addition, IgG induces degranulation of MCs by binding to FcγRII, manifested by increased release of histamine and inflammatory substances [\[45\]](#page-9-0). Besides, IgG may be involved in TLR4-FcγR cross-talk, activating the Syk signaling pathway and mediating cytokine production [\[49](#page-9-0)]. Both IgE and IgG activate MAPK, which promotes inflammatory cytokines (IL-1 and TNF-α). Selective inhibition of MAPK can reduce the apoptosis of osteoblasts, thus exerting a positive effect on the treatment of OA [[50\]](#page-9-0).

Toll/TLRs Toll-like receptors (TLRs) are crucial receptors for pathogen recognition and innate immunity. Evidence suggests that TLR1-10 expression in addition to TLR8 has been identified on human mast cells [[51\]](#page-9-0). Peptidoglycan (PGN) from Staphylococcus aureus stimulates mast cells to produce TNF-α, IL-4, IL-5, IL-6, and IL-13 in a TLR2-dependent manner. LPS from E. coli stimulates mast cells to produce TNF-α, il-1β, IL-6, and IL-13 in a TLR4-dependent manner [[52](#page-9-0)]. All TLRs signaling pathways ultimately lead to activation of the transcription factor nuclear factor-κB (NF-κB), which controls the expression of many inflammatory cytokine genes [\[53](#page-9-0)]. Upon activation of TLRs and subsequent activation of NF-kB, various chemokines, including IL-8 and CCL5, as well as cytokines such as IL-1, IL-6, and TNF-α are produced. These molecules play a crucial role in the recruitment of macrophages, granulocytes, and lymphocytes into the synovium of patients with OA to increase local inflammation and cartilage degradation, playing a crucial role in the progression of OA [[54,55\]](#page-9-0). In addition, TLRs has a synergistic effect with FcεRI, and the activation of TLRs sensitizes the MCs to stimulation through FcεRI. This mechanism may further accelerate the inflammatory response [[56,57](#page-9-0)].

NGF/TrkA Receptor Nerve growth factors (NGF) belong to a family of neurotrophin compounds that play an important role in the survival of neurons damaged during development. It is also a potent factor in MC degranulation in vitro and in vivo [\[58](#page-9-0)]. MCs produce TrkA receptors in OA, which are activated by NGF, resulting in the upregulation and release of inflammatory mediators and adverse neuroimmune tissue reactions, resulting in hyperalgesia in patients [[59\]](#page-9-0). NGF promotes the secretion of PGD2 from MCs by activating TrkA receptors to enhance the Ras, phosphoinositide-3 kinase (PI3K). The production of PGD2 may contribute to OA hypersensitivity by activating PGD2 receptor 1 in nociceptors [[60,61](#page-9-0)]. This finding suggests that targeting the TrkA receptor of NGF in MCs could be a promising therapeutic approach for managing OA pain.

In summary, MCs play a role in the inflammatory process of OA through various pathways. By inhibiting MC activation or blocking the pathway between MCs and OA, it is possible to stabilize MCs to some extent, reduce the inflammatory response, and control the development of OA (Fig. 1).

Fig. 1. The Pathways that MCs Activated in OA. MCs may participate in OA activation pathways, including stem cell factor/C-Kit receptor (SCF/ C-Kit), immunoglobulin E/Fcε receptor 1 (IgE/FcεR1), immunoglobulin G/Fcγ receptor (IgG/Fc*γ*R), nerve growth factor/TrkA receptor (NGF/TrkA), and Toll-like cells/receptors (TLRs).

2.3. Mediators of MCs involved in OA

Traditionally, MC media is divided into pre-stored media and self-synthesized media. The latter is due to the rapid production and release of arachidonic acid metabolites and the slow production and secretion of cytokines [\[62](#page-9-0)]. Tryptase is a preformed mediator in MC granules, encoded by TPSB2 and TPSD1 gene. The expression of these genes was found to be upregulated in patients with OA compared to healthy synovium, indicating that MCs have enhanced transcriptional activity in synovium of OA [\[63](#page-9-0)]. Besides, the degranulation of MCs was found to be more pronounced in patients with OA, manifested by significantly increased levels of tryptase, eosinophil cationic protein and histamine [[64\]](#page-9-0). The release of pro-inflammatory mediators by activated MCs can contribute to synovial inflammation and cartilage degeneration (Fig. 2). The available evidence indicates that the activation of MCs and their associated mediators contribute to the development of OA. The following section outlines the established effects of certain mediators on OA (see [Table 1](#page-4-0)).

Tryptase Tryptases are proteases specifically secreted by MCs, serve as markers for MCs [[65\]](#page-9-0). They can be classified into different forms, including α, β, γ and δ. β-Tryptase is the primary isoform found in MCs and has been extensively studied [\[66](#page-10-0)]. Multiple evidences proved that tryptase promotes inflammation response [\[67,68](#page-10-0)]. The extent to which tryptase contributes to the development of inflammatory processes and cartilage damage largely depends on its activation of the protease-activated receptor-2 (PAR-2) receptor. It stimulates the proliferation of synovial fibroblast-like cells (SFC) and triggers the release of pro-inflammatory cytokine IL-8 through PAR-2 [\[69,70](#page-10-0)]. In OA mice lacking PAR-2, cartilage degradation was reduced [\[71](#page-10-0)]. In addition, β-tryptases cleaved Proteoglycan 4 (PRG4), which is related to boundary lubrication and anti-inflammatory. Cleaved PRG4 activates the NF-κB, result in the progression of OA [[72\]](#page-10-0). Tryptases are also matrix metalloproteinase (MMP) convertases. They are biologically active on MMP-1, MMP-2, MMP-3, MMP-9 and MMP-13. Tryptases activation of MMP leads to recombination of the intercellular matrix and degradation of fibrous collagens, proteoglycans and laminins. In OA, they mediate cartilage damage and proteolytic loss of aggrecan proteoglycans in arthritis by activating MMP-3 and MMP-13 [\[73,74](#page-10-0)]. Besides, it has been found that tryptases can induce further degranulation of MCs through a positive feedback mechanism, thus promoting further release of inflammatory mediators [\[75](#page-10-0)]. Tryptases increase microvascular permeability, neutrophilia in vivo, and stimulate MCs to release histamine [[76,77](#page-10-0)].

Histamine The production of histamine in the synovial fluid of OA patients were significantly increased, and was significantly correlated with the number of MCs [\[21](#page-9-0)]. In OA patients, aberrant phenotype of OA chondrocytes express histamine receptors such as H1 and H2 [[78\]](#page-10-0). Histamine can increase the histamine H1 receptor expression in synovial fibroblasts [\[79](#page-10-0)]. Treatment of mice with cetirizine, a histamine H1 receptor antagonist, reduced meniscus severity and OA-related mediators in mice. It was also demonstrated in a cross-sectional study that H1 antihistamines were associated with a reduced prevalence of OA in the knee $[80]$ $[80]$, suggesting that the use of drugs to block histamine activity in MCs could be a therapeutic target for OA. Interestingly, H4 receptors are found to expressed on the surface of MCs, which mediate the release of pro-inflammatory cytokines and chemokines (TGF-β1, TNF-α, TNF-β, PDGF-BB, TIMP-2, M-CSF et al.) [\[81](#page-10-0)].

IL-1β and IL-6 Interleukin-1β (IL-1β) and Interleukin-6 (IL-6) are important pro-inflammatory cytokines in the pathogenesis of OA. They are secreted by MCs and are significantly increased in OA and are associated with the severity of OA [82–[84\]](#page-10-0). In OA cartilage, IL-1β plays a crucial regulatory role in inducing chondrocyte apoptosis [[85\]](#page-10-0). IL-1β promotes the degradation of cartilage by stimulating

Fig. 2. Role of MC Mediators in OA. The activation of MCs leads to the production of cytokines that play a crucial role in the inflammatory process of OA. These cytokines, such as IL-1, IL-6, and TNF-α, are involved in bone erosion and destruction. Additionally, VEGF, IL-6, and IFN-γ contribute to the development of synovitis. Tryptase-induced MMPs promote cartilage degradation, while TGF-β helps inhibit cartilage destruction.

Table 1

Selected MCs mediators and their potential roles in OA.

the expression of matrix MMPs and aggrecanases. It also induces the overexpression of MMP-13 through the p38, JNK, and NF-κB signaling pathways, leading to cartilage destruction $[86,87]$ $[86,87]$. Besides, IL-1 β reduces the production of cartilage-specific macromolecules, such as type II collagen, by regulating the transcription factors Sp1 and Sp3(88). IL-6 can induce increased expression of the aggrecan-degrading enzymes ADAMTS-4 and MMP-3 and the collagen-degrading enzymes MMP-1 and MMP-13 and promote cartilage degradation [\[89](#page-10-0)]. IL-1 and IL-6 also promote osteoclast generation. IL-1β primarily inhibits the synthesis of extracellular matrix (ECM) proteins, promoting the generation and maturation of osteoclasts, and enhancing the inflammatory process of OA. The mechanism of IL-6 promoting osteoclasts is not clear. It may be related to the alterations in the ratio of RANKL to OPG and/or M-CSF expression [\[90](#page-10-0), [91\]](#page-10-0). In addition, IL-1β also regulates NGF expression in synovial fibroblasts, making it a potential target for the treatment of OA pain [\[92](#page-10-0)].

IFN-γ MCs have the ability to regulate the expression of interferon-gamma (IFN- γ) in the immune response. Its mechanism is mainly by stimulating other immune cells to release IFN, such as NK cells and T cells [93–[95\]](#page-10-0). IFN- γ can be detected in the peripheral blood of OA patients [[96\]](#page-10-0). Elevated levels of IFN-γ can activate macrophages and enhance the recruitment and activation of white blood cells, thereby exacerbating the inflammatory response in OA synovitis and promoting the inflammatory process [\[97](#page-10-0),[98\]](#page-10-0). In addition, IFN-γ itself is one of the factors that stimulate the increase of FcεR1 on the surface of MCs [[99\]](#page-10-0).

NGF NGF is significantly elevated in damaged or inflamed tissues and promotes pain sensory conduction in injury-inducing neurons through various mechanisms [\[100\]](#page-10-0). MCs have the ability to synthesize, store, and release NGF upon degranulation, thus playing a crucial role in pain conduction, nerve immunity, and tissue inflammation [\[101,102](#page-10-0)]. In an OA rat model, activated MCs increases NGF in synovial fluid and the sensitivity of NGF is increased by the upregulation of its receptor TrkA, promoting a persistent and intense pain response [\[103\]](#page-10-0). In addition, upregulated NGF in OA stimulates chondrocyte metabolism in the osteoarthritic process [\[104\]](#page-10-0).

TNF-α Tumor necrosis factor–alpha (TNF-α) is pro-inflammatory mediators stored in granules of MCs. Its expression was significantly increased in the synovial tissue of early OA $[105,106]$. TNF- α promotes progression of OA through multiple mechanisms. For example, TNF-α has a detrimental effect on cartilage. TNF-α upregulates the expression of many factors, such as protein-2 (BMP-2), MMP-3 and a disintegrin-like and metalloprotease (reprolysin type) with thrombospondin type 1 motif, 4 (ADAMTS-4), thereby promoting cartilage destruction [107–[110\]](#page-11-0). It can also promote chondrocyte apoptosis, affect bone remodeling, stimulate the proliferation of osteoblasts, and activate osteoclasts [[111,112\]](#page-11-0). In vitro studies have shown that the inhibition of TNF-α has a positive effect on the viability and proliferation of cartilage [\[113](#page-11-0)]. In addition, TNF- α may be key in the pain of OA. Clinical OA patient data showed that the level of TNF- α in synovial fluid was positively correlated with the pain score of knee OA [[114](#page-11-0)].

VEGF As a classic angiogenic cytokine, Vascular endothelial growth factor (VEGF) may be stored and released from the cytoplasmic granules of mast cells [[115](#page-11-0)]. VEGF is significantly increased in the synovial fluid of OA patients and is associated with pain [\[116\]](#page-11-0). Angiogenesis can promote inflammation, chondrocyte hypertrophy and endochondral ossification. VEGF actively promotes angiogenesis and neovascularization and promotes the development of OA inflammation [117–[119\]](#page-11-0). The injection of VEGF into healthy mice resulted in synovial hyperplasia, increased calcification of the articular cartilage, osteosclerosis, and degradation of cartilage [\[120\]](#page-11-0). It also stimulates other inflammatory cells in the microenvironment to release angiogenic mediators, cytokines, and extracellular matrix-degrading proteases, thereby promoting angiogenesis [\[121\]](#page-11-0). In a rat model of OA, knocking down the expression of VEGF helps protect chondrocytes and delays the progression of OA [[122](#page-11-0)].

TGF-β Mast cells also secrete Transforming growth factor-β (TGF-β) [\[123\]](#page-11-0). TGF-β is generally considered to protect against cartilage damage. For instance, IL1 α -induced JAK/STAT signaling is antagonized by TGF-β [\[124\]](#page-11-0). However, recent research shows that TGF-β signaling has conflicting roles of in the development of OA. Synovial joint homeostasis depends on the complex control of TGF-β signal transduction pathways. In osteoarthritic joint, TGF-β activates p38 and Smad1/5/8, result in the increased expression of proteases and proteoglycan degeneration, while the activation of Smad3 protects the cartilage degradation [[125](#page-11-0)]. But another study proved that TGF-β/Smad3 induces the development of OA, indicating that the role of TGF-β relies upon intricate environment of synovium [[126](#page-11-0)].

3. MCs and OA synovial fluid microenvironment

OA synovial fluid microenvironment often changes. For example, inflammatory cytokines, such as IL-17 and IL-22, are increasingly expressed [\[127\]](#page-11-0). Within the microenvironment of synovial fluid, various inflammatory cells contribute to the development of inflammation. We propose that crosstalk between MCs and synovial microenvironment plays an important role in OA. MCs interact with these inflammatory cells, leading to changes in the synovial fluid microenvironment and facilitate the progression of synovial inflammation (Fig. 3).

T Cells The increased expression of many T cell-related cytokines, such as IL-2, IL-6, IL-10, IL-17, IFN-γ, in patients with OA, indicating that T cell activation and differentiation play an important role in the pathogenesis of OA [\[128\]](#page-11-0). During the immune response of OA, immature dendritic cells capture allergens and, upon maturation, migrate to local lymph nodes where they present these antigens to naive T cells. This interaction promotes the differentiation of $CD4^+$ T cells into Th2 cells. Th2 cells subsequently secrete IL-4, which induces isotype switching in B lymphocytes from IgM class antibodies to IgE class antibodies, thereby activating mast cell IgE/FcεRI signaling [[129](#page-11-0)]. In turn, MCs can present native antigens to human T cells, which can directly activate T cells and promote T cell proliferation [\[130,131\]](#page-11-0). For example, research shows that MCs secret IL-1β, TGF-β and IL-6 to induce CD4⁺ T cells to differentiate into Th17 cells [132–[134\]](#page-11-0). Although lower than that in RA patients, the number of Th17 cells in OA patients was significantly higher than that in healthy patients [\[135\]](#page-11-0). The Th17 increase the expression of IL-17, which enhances the inflammatory response in OA pathogenesis [[136](#page-11-0)]. Therefore, the increase in Th17 correlates with the progression of OA [\[133,137](#page-11-0)]. In addition, MC-mediated IL-6 and histamine decrease Treg cells and inhibit their inflammatory suppressive activity [\[138\]](#page-11-0).

Neutrophil There are varying levels of neutrophils present in the synovium of OA [\[139\]](#page-11-0). The role of neutrophils in OA may depends on the level of elastase and TGF-β [[140\]](#page-11-0). MCs are capable of facilitating the release of PGD2 to recruit the neutrophils [[140](#page-11-0)]*.* PGD2 induces lymphocytes to produce IL-8, recruiting more neutrophils into the synovial fluid [\[141\]](#page-11-0). The MC-restricted tryptase mMCP-6 helps recruit neutrophils to the site of bacterial infection and has a key immune role [[142](#page-11-0)]. MCs can also produce many cytokines and growth factors that affect neutrophils, including TNF-α, IL-1β, GM-CSF, and IL-6 [\[143\]](#page-11-0).

Fibroblast MCs may participate in joint destruction by inducing MMPs production to activate fibroblasts. Activation of synovial fibroblasts increases the expression of SCF, which promotes the survival and proliferation of MCs, leading to an increase in MCs in

Fig. 3. The Role of MCs on Other Cells in the OA Synovial Fluid Microenvironment. In the stages of OA, MCs are recruited and activated in the synovial fluid microenvironment. These MCs release various cytokines that have an impact on other cells in the synovial microenvironment, including neutrophils, T cells, macrophages, fibroblasts, osteoclasts, osteoblasts, and chondrocytes.

synovial inflammation [\[144](#page-11-0),[145](#page-11-0)]. This interaction between MCs and synovial fibroblasts forms an important positive feedback loop that plays a crucial role in synovial inflammation in OA. The mediators produced by MCs have profound effects on fibroblasts. For example, TNF-α stimulates fibroblasts in the synovium of obese OA patients to secrete more lactic acid and aerobic glycolysis and produce more IL-6 [[146](#page-11-0)]. TNF-α can also upregulate the expression of NGF in synovial fibroblasts and mediate the pain of OA to a certain extent [[92\]](#page-10-0). MC-derived TNF-α and TGF-β1 also promote fibroblast proliferation [\[147\]](#page-11-0). In vivo studies have shown that the inhibition of TNF-α has a chondroprotective effect, further confirming the detrimental impact of TNF-α on cartilage [[148](#page-11-0)]. In addition, animal studies have also shown that chymase can promote the proliferation of synovial fibroblasts [[149](#page-11-0)].

Macrophage Macrophages play a crucial role in the initiation and development of OA through autocrine and paracrine effects by secreting inflammatory cytokines, growth factors, MMPs, and tissue inhibitors of metalloproteinases (TIMPs) [[150](#page-12-0)]. Macrophages are typically classified into two types: classically activated/inflammatory (M1) type and alternatively activated/immunomodulatory (M2) type. When activated, macrophages can produce a significant amount of proinflammatory cytokines, which can lead to autoimmune diseases and tissue damage [[151,152\]](#page-12-0). For instance, M1 macrophages produce IL-6, TNF-α, IL-1, and IL-12(153), while inducing macrophages polarize into M2 macrophages relieves the osteoarthritis [[154](#page-12-0)]. MCs-produced IL-1β, TNF-α, and IL-6 can activate macrophages, causing them to produce large quantities of proinflammatory cytokines (TNF-α, IL-1, IL-12, IL-18, and IFN-γ), chemokines, and MMPs in the synovitis environment. This process ultimately leads to osteoclast formation, erosion, and progressive joint destruction [\[153,155\]](#page-12-0).

Osteoclast and Osteoblast Osteoclasts have been shown to be important players in the pathogenesis of bone destruction and promote bone resorption [[156](#page-12-0),[157](#page-12-0)]. Osteoblasts play a role in maintaining bone homeostasis. MCs directly act on osteoclasts, osteoclast precursors and osteoblasts by producing histamine and promote osteoclast generation through auto/paracrine signaling mechanisms [\[158,159](#page-12-0)]. The TNF-α, IL-6 and IL-1β produced by MCs can directly stimulate the precursors of osteoclasts, indirectly stimulate the formation and activation of osteoclasts [[90](#page-10-0)[,160,161](#page-12-0)]. In addition, MCs also produce proinflammatory cytokines (IL-1, IL-1β, IL-13, GM-CSF) to induce osteoclast production [\[162,163](#page-12-0)]. IL-1β increases the expression of NF-κB ligand (RANKL) receptor activator through osteoblasts, thereby indirectly inducing osteoclast production and maturation [\[90](#page-10-0)]. MCs can be involved in osteoclast differentiation and activation by producing a signaling system mediated by RANK, a member of the TNF receptor family [\[164\]](#page-12-0). TNF- α directly induces osteoclast differentiation independent of the ODF/RANKL-RANK interaction [[165,166\]](#page-12-0). Additionally, IL-1 α induces osteoblast apoptosis and inhibits osteoblast differentiation by activating the JNK and p38 MAPK pathways [\[167\]](#page-12-0).

Chondrocyte The balance between anabolic and catabolic activities of articular chondrocytes is disrupted during active OA disease [\[168\]](#page-12-0). Besides, as individuals age, the decreased ability of cartilage cells to maintain and repair tissues may also lead to OA [\[169\]](#page-12-0). Chondrocytes have the ability to form an extracellular matrix primarily composed of aggrecan and type II collagen. TNF-α can promote the apoptosis of chondrocytes [\[111\]](#page-11-0). TLRs are increased in OA cartilage lesions, and the ligands of TLR-2 and TLR-4 induce chondrocyte decomposition [\[170\]](#page-12-0). TLRs are also expressed by synovial MCs, and further research is needed to investigate their potential impact on chondrocyte decomposition. The production of TGF-β secreted by MCs, plays a crucial role in regulating chondrocyte metabolism. It stimulates the synthesis of extracellular matrix (ECM) components and helps maintain the balance of chondrocyte activities [[171](#page-12-0)]. In animal studies, cutting off TGF-β and OA signals leads to cartilage degradation and promotes OA development [\[172\]](#page-12-0). However, in OA environment, TGF-β increased MMP-13 and induced chondrocyte damage [\[125\]](#page-11-0).

4. Clinical potential of targeting MCs for OA treatment

The current treatment of OA mainly includes medication for pain relief, patient self-management, exercise and weight loss. Total joint replacement is considered the gold standard of treatment for patients with OA who do not respond to conservative measures or experience a significant decline in their quality of life due to pain [\[173\]](#page-12-0). In addition, the new treatment involves the use of radiofrequency ablation [\[174\]](#page-12-0) and intra-articular injection of platelet-rich plasma (PRP) or hyaluronic acid (HA) to repair cartilage [\[175](#page-12-0)–177]. However, unlike diseases such as cancer, the development of immune cell-based therapies for OA is very limited.

As MCs play a crucial role in inflammatory diseases, such as OA [[178](#page-12-0)], targeting MCs could be an effective approach for OA patients. Disodium cromoglycate, widely described as a "MCs stabilizer," that is, a preparation that blocks the release of MCs mediators after proper cell activation, can be used to inhibit MCs releasing mediators, thereby controlling the development of inflammation [\[179](#page-12-0)–181]. MC-targeted therapy alleviates disease symptoms by blocking MC mediators, inhibiting activation receptors on MCs, neutralizing MC activation signals, silencing MCs and reducing the number of MCs.

Due to the high cost of researching new drugs, stabilizers for MCs are currently being studied more based on the reuse of drugs. Anti-IgE drugs can block the IgE-FcεR1 pathway activated by MCs, thereby reducing MCs numbers and controlling inflammatory allergic reactions [[182](#page-12-0)]. For example, Statins inhibit MC-IgE response by blocking isoprene, thereby reducing the airway inflammation in asthma [[183](#page-12-0)]. The H1 antihistamines associate with a lower prevalence of knee OA implies that it is possible for antihistamines to regulate the impact of MCs on disease management [[184](#page-12-0)]. Monoclonal antibodies and humanized monoclonal antibodies represent a significant advancement in the development of MC stabilizers, offering high specificity and minimal side effects. There are three generations of anti-IgE monoclonal antibodies, which include omalizumab, ligelizumab, kilizumab, and UB-221. Each generation has seen improvements primarily focused on enhancing IgE affinity and reducing IgE production [[185](#page-12-0)]. Besides, some monoclonal antibodies show potential in inhibiting MCs other receptor. For instance, CDX-0159 is an anti-Kit monoclonal antibody that inhibit activation of MCs through suppressing SCF/c-Kit. It has shown safety in animal studies and does not have adverse effects on the blood. In human experiments, the antibody can reduce plasma tryptase levels and inhibit MCs [\[186,187](#page-12-0)]. Several clinical studies have shown that injections of antibodies against NGF in patients with moderate to severe OA can be effective in relieving pain and restoring function [\[188](#page-12-0)–191].

The mechanism of OA control has positive significance for disease treatment. Efforts to identify new pathway inhibitors, such as tyrosine kinase Syk or mediator antagonists, may lead to new successes in this field. Most recently, Mrgprb-2 has been reported as a specific receptor for MC activation. Abnormal activation of PI3K-AKT and MAPK pathways in Mrgprb-2-deficient MCs may serve as a new target for therapy of OA [\[192\]](#page-12-0).

5. Discussion and perspectives

Our review summarizes the crucial role and mechanism of MCs in the inflammation and progression of OA. The activation of MCs during OA leads to the release of pro-inflammatory mediators (histamine, tryptase, IL-1, IL-6, TNF-α, etc.), resulting in synovial inflammation and cartilage degeneration. The synovial microenvironment plays a crucial role in the progression of OA, particularly through the interaction of MCs with other inflammatory cells. Upon MC activation, the release of mediators can have diverse effects on various inflammatory cells, resulting in further intensification of local inflammation.

To date, the role of MCs in OA has been gradually explored. However, there is a lack of research on how MCs specifically affect factors such as gender, age, and obesity, which are important for the progression of OA [[193](#page-13-0),[194](#page-13-0)]. Additionally, although MCs show potential as a target for OA treatment [\[195\]](#page-13-0), many issues may need to be addressed before they can be applied in clinical settings. Currently, most studies on the development and function of MCs are based on murine MCs extracted from mice or from MC-deficient mice [[196](#page-13-0)]. There is a need to explore new methods for obtaining human-derived mast cells to determine which mast cell functions are relevant to humans. Understanding mast cell development and heterogeneity through single-cell sequencing may enhance the application of MCs [[197](#page-13-0)]. Furthermore, MCs not only play a pro-inflammatory role but also exhibit immunomodulatory and tissue homeostasis functions in certain contexts [[198](#page-13-0)]. In the future, it will be essential to better identify the specific subtypes of MCs involved in various stages of OA. Concurrently, advancements in material technologies, including specialized polymers, lipids, and nanozymes, offer the potential to deliver drugs selectively to distinct subtypes of MCs by targeting specific surface receptors. This presents significant opportunities for progress in the development of MC-targeted therapeutic strategies.

CRediT authorship contribution statement

Guanghui Hao: Writing – original draft, Conceptualization. **Shanqian Han:** Writing – original draft, Conceptualization. **Zhangang Xiao:** Writing – review & editing. **Jing Shen:** Writing – review & editing. **Yueshui Zhao:** Writing – review & editing, Writing – original draft, Validation, Supervision, Funding acquisition, Conceptualization. **Qi Hao:** Writing – review & editing, Writing – original draft, Supervision, Investigation, Data curation, Conceptualization.

Ethics approval and consent to participate

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Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Abbreviations

OA Osteoarthritis

- MCs Mast Cells
- RA Rheumatoid Arthritis

PRG4 Proteoglycan 4

References

- [1] [K.D. Allen, L.M. Thoma, Y.M. Golightly, Epidemiology of osteoarthritis, Osteoarthritis Cartilage 30 \(2\) \(2022\) 184](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref1)–195.
- [2] [M. Cross, E. Smith, D. Hoy, S. Nolte, I. Ackerman, M. Fransen, et al., The global burden of hip and knee osteoarthritis: estimates from the global burden of](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref2) [disease 2010 study, Ann. Rheum. Dis. 73 \(7\) \(2014\) 1323](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref2)–1330.
- [3] [L. Lyu, Y. Cai, M. Xiao, J. Liang, G. Zhang, Z. Jing, et al., Causal relationships of general and abdominal adiposity on osteoarthritis: a two-sample mendelian](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref3) [randomization study, J. Clin. Med. 12 \(1\) \(2022\).](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref3)
- [4] [M.F. Rai, L.J. Sandell, Inflammatory mediators: tracing links between obesity and osteoarthritis, Crit. Rev. Eukaryot. Gene Expr. 21 \(2\) \(2011\) 131](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref4)–142.
- [5] [D. Prieto-Alhambra, A. Judge, M.K. Javaid, C. Cooper, A. Diez-Perez, N.K. Arden, Incidence and risk factors for clinically diagnosed knee, hip and hand](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref5) [osteoarthritis: influences of age, gender and osteoarthritis affecting other joints, Ann. Rheum. Dis. 73 \(9\) \(2014\) 1659](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref5)–1664.
- [6] [B.J.E. de Lange-Brokaar, A. Ioan-Facsinay, E. Yusuf, A.W. Visser, H.M. Kroon, S.N. Andersen, et al., Degree of synovitis on MRI by comprehensive whole knee](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref6) semi-quantitative scoring method correlates with histologic and macroscopic features of synovial tissue inflammation in knee osteoarthritis, Osteoarthritis [Cartilage 22 \(10\) \(2014\) 1606](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref6)–1613.
- [7] [F. Berenbaum, Osteoarthritis as an inflammatory disease \(osteoarthritis is not osteoarthrosis!\), Osteoarthritis Cartilage 21 \(1\) \(2013\) 16](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref7)–21.
- [8] [X. Luan, X. Tian, H. Zhang, R. Huang, N. Li, P. Chen, et al., Exercise as a prescription for patients with various diseases, J. Sport Health Sci. 8 \(5\) \(2019\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref8) 422–[441.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref8)
- [9] [S.L. Kolasinski, T. Neogi, M.C. Hochberg, C. Oatis, G. Guyatt, J. Block, et al., 2019 American college of rheumatology/arthritis foundation guideline for the](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref9) [management of osteoarthritis of the hand, hip, and knee, Arthritis Rheumatol. 72 \(2\) \(2020\) 220](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref9)–233.
- [10] [H. Abd-Allah, A.O. Kamel, O.A. Sammour, Injectable long acting chitosan/tripolyphosphate microspheres for the intra-articular delivery of lornoxicam:](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref10) [optimization and in vivo evaluation, Carbohydr. Polym. 149 \(2016\) 263](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref10)–273.
- [11] [N.W. Skelley, S. Namdari, A.M. Chamberlain, J.D. Keener, L.M. Galatz, K. Yamaguchi, Arthroscopic debridement and capsular release for the treatment of](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref11) [shoulder osteoarthritis, Arthroscopy 31 \(3\) \(2015\) 494](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref11)–500.
- [12] D. González-de-Olano, I. Álvarez-Twose, [Mast cells as key players in allergy and inflammation, J Investig. Allergol. Clin. Immunol. 28 \(6\) \(2018\) 365](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref12)-378.
- [13] [S.J. Galli, N. Gaudenzio, M. Tsai, Mast cells in inflammation and disease: recent progress and ongoing concerns, Annu. Rev. Immunol. 38 \(2020\) 49](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref13)–77. [14] [F. Niyonsaba, H. Ushio, M. Hara, H. Yokoi, M. Tominaga, K. Takamori, et al., Antimicrobial peptides human beta-defensins and cathelicidin LL-37 induce the](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref14)
- [secretion of a pruritogenic cytokine IL-31 by human mast cells, J. Immunol. 184 \(7\) \(2010\) 3526](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref14)–3534. [15] [P.A. Nigrovic, K. Shin, Evaluation of synovial mast cell functions in autoimmune arthritis, Methods Mol. Biol. 1220 \(2015\) 423](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref15)–442.
- [16] [S.S. Craig, L.B. Schwartz, Tryptase and chymase, markers of distinct types of human mast cells, Immunol. Res. 8 \(2\) \(1989\) 130](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref16)–148.
- [17] [A.A. Irani, N.M. Schechter, S.S. Craig, G. DeBlois, L.B. Schwartz, Two types of human mast cells that have distinct neutral protease compositions, Proc. Natl.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref17)
- [Acad. Sci. U. S. A. 83 \(12\) \(1986\) 4464](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref17)–4468. [18] [M.G. Buckley, P.J. Gallagher, A.F. Walls, Mast cell subpopulations in the synovial tissue of patients with osteoarthritis: selective increase in numbers of](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref18)
- [tryptase-positive, chymase-negative mast cells, J. Pathol. 186 \(1\) \(1998\) 67](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref18)–74.
- [19] [K. Maaninka, J. Lappalainen, P.T. Kovanen, Human mast cells arise from a common circulating progenitor, J. Allergy Clin. Immunol. 132 \(2\) \(2013\).](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref19)
- [20] P. Kulkarni, A. Harsulkar, A.-G. Märtson, S. Suutre, A. Märtson, S. Koks, Mast cells differentiated in synovial fluid and resident in osteophytes exalt the [inflammatory pathology of osteoarthritis, Int. J. Mol. Sci. 23 \(1\) \(2022\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref20).
- [21] X. Zhao, S. Younis, H. Shi, S. Hu, A. Zia, H.H. Wong, et al., RNA-seq characterization of histamine-releasing mast cells as potential therapeutic target of [osteoarthritis, Clin. Immunol. 244 \(2022\) 109117](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref21).
- [22] [S. Mishima, J.-I. Kashiwakura, S. Toyoshima, T. Sasaki-Sakamoto, Y. Sano, K. Nakanishi, et al., Higher PGD2 production by synovial mast cells from](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref22) [rheumatoid arthritis patients compared with osteoarthritis patients via miR-199a-3p/prostaglandin synthetase 2 axis, Sci. Rep. 11 \(1\) \(2021\) 5738](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref22).
- [23] B.J. de Lange-Brokaar, M. Kloppenburg, S.N. Andersen, A.L. Dorjée, E. Yusuf, L. Herb-van Toorn, et al., Characterization of synovial mast cells in knee [osteoarthritis: association with clinical parameters, Osteoarthritis Cartilage 24 \(4\) \(2016\) 664](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref23)–671.
- [24] L. Farinelli, A. Aquili, M. Mattioli-Belmonte, S. Manzotti, F. D'[Angelo, C. Ciccullo, et al., Synovial mast cells from knee and hip osteoarthritis: histological study](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref24) [and clinical correlations, J Exp Orthop 9 \(1\) \(2022\) 13.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref24)
- [25] [J. Dan, M. Izumi, H. Habuchi, O. Habuchi, S. Takaya, Y. Kasai, et al., A novel mice model of acute flares in osteoarthritis elicited by intra-articular injection of](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref25) [cultured mast cells, J Exp Orthop 8 \(1\) \(2021\) 75.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref25)
- [26] [Q. Wang, C.M. Lepus, H. Raghu, L.L. Reber, M.M. Tsai, H.H. Wong, et al., IgE-mediated mast cell activation promotes inflammation and cartilage destruction in](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref26) [osteoarthritis, Elife 8 \(2019\).](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref26)
- [27] [N. Schubert, J. Dudeck, P. Liu, A. Karutz, S. Speier, M. Maurer, et al., Mast cell promotion of T cell-driven antigen-induced arthritis despite being dispensable](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref27) [for antibody-induced arthritis in which T cells are bypassed, Arthritis Rheumatol. 67 \(4\) \(2015\) 903](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref27)–913.
- [28] P. Besmer, K. Manova, R. Duttlinger, E.J. Huang, A. Packer, C. Gyssler, et al., The kit-ligand (steel factor) and its receptor c-kit/W: pleiotropic roles in [gametogenesis and melanogenesis, Dev. Suppl. \(1993\) 125](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref28)–137.
- [29] [L.R. Teegala, Y. Elshoweikh, R. Gudneppanavar, S. Thodeti, S. Pokhrel, E. Southard, et al., Protein Kinase C](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref29) α and β compensate for each other to promote stem [cell factor-mediated KIT phosphorylation, mast cell viability and proliferation, FASEB \(Fed. Am. Soc. Exp. Biol.\) J. 36 \(5\) \(2022\) e22273](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref29).
- [30] J. Lennartsson, L. Rönnstrand, [Stem cell factor receptor/c-Kit: from basic science to clinical implications, Physiol. Rev. 92 \(4\) \(2012\) 1619](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref30)-1649.
- [31] P.M. Jordan, N. Andreas, M. Groth, P. Wegner, F. Weber, U. Jäger, et al., ATP/IL-33-triggered hyperactivation of mast cells results in an amplified production [of pro-inflammatory cytokines and eicosanoids, Immunology 164 \(3\) \(2021\) 541](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref31)–554.
- [32] [J. Seifert, C. Küchler, S. Drube, ATP/IL-33-Co-Sensing by mast cells \(MCs\) requires activated c-kit to ensure effective cytokine responses, Cells 12 \(23\) \(2023\).](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref32) [33] [Y.J. Choi, J.-S. Yoo, K. Jung, L. Rice, D. Kim, V. Zlojutro, et al., Lung-specific MCEMP1 functions as an adaptor for KIT to promote SCF-mediated mast cell](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref33) [proliferation, Nat. Commun. 14 \(1\) \(2023\) 2045.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref33)
- [34] [T. Annese, R. Tamma, M. Bozza, A. Zito, D. Ribatti, Autocrine/paracrine loop between SCF](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref34)+/c-Kit+ mast cells promotes cutaneous melanoma progression, [Front. Immunol. 13 \(2022\) 794974](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref34).
- [35] S.-Y. Nam, H.-Y. Kim, H.-M. Kim, H.-J. Jeong, В[eta-eudesmol reduces stem cell factor-induced mast cell migration, Int. Immunopharm. 48 \(2017\) 1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref35)–7.
- [36] [C. Janson, N. Kasahara, G.C. Prendergast, J. Colicelli, RIN3 is a negative regulator of mast cell responses to SCF, PLoS One 7 \(11\) \(2012\) e49615.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref36)
- [37] [S.J. Galli, M. Tsai, IgE and mast cells in allergic disease, Nat. Med. 18 \(5\) \(2012\) 693](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref37)–704.
- [38] [G.H. Banafea, S. Bakhashab, H.F. Alshaibi, P. Natesan Pushparaj, M. Rasool, The role of human mast cells in allergy and asthma, Bioengineered 13 \(3\) \(2022\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref38) 7049–[7064](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref38).
- [39] [E. Silveira, A.M.M. Souza, V.M. Mazucato, M.C. Jamur, C. Oliver, Lipid rafts in mast cell biology, J. Lip. 2011 \(2011\) 752906](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref39).
- [40] [U. Blank, J. Rivera, The ins and outs of IgE-dependent mast-cell exocytosis, Trends Immunol. 25 \(5\) \(2004\) 266](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref40)–273.
- [41] [R.P. Siraganian, R.O. de Castro, E.A. Barbu, J. Zhang, Mast cell signaling: the role of protein tyrosine kinase Syk, its activation and screening methods for new](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref41) [pathway participants, FEBS \(Fed. Eur. Biochem. Soc.\) Lett. 584 \(24\) \(2010\) 4933](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref41)–4940.
- [42] P. Dráber, V. Sulimenko, E. Dráberová, Cytoskeleton in mast cell signaling, Front. Immunol. 3 (2012) 130.
- [43] [A. Aquili, L. Farinelli, C. Bottegoni, L. Antonicelli, A. Gigante, The effect of anti-IgE therapy in knee osteoarthritis: a pilot observational study, J. Biol. Regul.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref43) [Homeost. Agents 31 \(4 Suppl. 1\) \(2017\) 1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref43)–5.
- [44] [J.-M. Yoo, J.-H. Yang, Y.S. Kim, H.J. Yang, W.-K. Cho, J.Y. Ma, Inhibitory effects of viscum coloratum extract on IgE/antigen-activated mast cells and mast cell](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref44)[derived inflammatory mediator-activated chondrocytes, Molecules 22 \(1\) \(2016\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref44).
- [45] [H. Lee, J. Kashiwakura, A. Matsuda, Y. Watanabe, T. Sakamoto-Sasaki, K. Matsumoto, et al., Activation of human synovial mast cells from rheumatoid arthritis](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref45) [or osteoarthritis patients in response to aggregated IgG through Fc](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref45)γ receptor I and Fcγ receptor II, Arthritis Rheum. 65 (1) (2013) 109–119.
- [46] [Y. Okayama, A.S. Kirshenbaum, D.D. Metcalfe, Expression of a functional high-affinity IgG receptor, Fc gamma RI, on human mast cells: up-regulation by IFN](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref46)[gamma, J. Immunol. 164 \(8\) \(2000\) 4332](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref46)–4339.
- [47] [M.R. Woolhiser, K. Brockow, D.D. Metcalfe, Activation of human mast cells by aggregated IgG through FcgammaRI: additive effects of C3a, Clin. Immunol. 110](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref47) [\(2\) \(2004\) 172](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref47)–180.
- [48] [M. Karlberg, Z. Xiang, G. Nilsson, Fc gamma RI-mediated activation of human mast cells promotes survival and induction of the pro-survival gene Bfl-1, J. Clin.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref48) [Immunol. 28 \(3\) \(2008\) 250](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref48)–255.
- [49] O. Vargas-Hernández, J.L. Ventura-Gallegos, M.L. Ventura-Ayala, M. Torres, A. Zentella, S. Pedraza-Sánchez, THP-1 cells increase TNF-α production upon LPS + [soluble human IgG co-stimulation supporting evidence for TLR4 and Fc](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref49)γ receptors crosstalk, Cell. Immunol. 355 (2020) 104146.
- [50] [S. Kumar, B.J. Votta, D.J. Rieman, A.M. Badger, M. Gowen, J.C. Lee, IL-1- and TNF-induced bone resorption is mediated by p38 mitogen activated protein](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref50) [kinase, J. Cell. Physiol. 187 \(3\) \(2001\) 294](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref50)–303.
- [51] [H. Sandig, S. Bulfone-Paus, TLR signaling in mast cells: common and unique features, Front. Immunol. 3 \(2012\) 185.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref51)
- [52] [V. Supajatura, H. Ushio, A. Nakao, S. Akira, K. Okumura, C. Ra, et al., Differential responses of mast cell Toll-like receptors 2 and 4 in allergy and innate](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref52) [immunity, J. Clin. Investig. 109 \(10\) \(2002\) 1351](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref52)–1359.
- [53] [T. Kawai, S. Akira, Signaling to NF-kappaB by toll-like receptors, Trends Mol. Med. 13 \(11\) \(2007\) 460](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref53)–469.
- [54] [G. Barreto, M. Manninen, K. K Eklund, Osteoarthritis and toll-like receptors: when innate immunity meets chondrocyte apoptosis, Biology 9 \(4\) \(2020\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref54).
- [55] [J.H. Rosenberg, V. Rai, M.F. Dilisio, D.K. Agrawal, Damage-associated molecular patterns in the pathogenesis of osteoarthritis: potentially novel therapeutic](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref55) [targets, Mol. Cell. Biochem. 434 \(1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref55)–2) (2017) 171–179.
- [56] [H. Qiao, M.V. Andrade, F.A. Lisboa, K. Morgan, M.A. Beaven, FcepsilonR1 and toll-like receptors mediate synergistic signals to markedly augment production](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref56) [of inflammatory cytokines in murine mast cells, Blood 107 \(2\) \(2006\) 610](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref56)–618.
- [57] J. Suurmond, F. Rivellese, A.L. Dorjée, A.M. Bakker, Y.J.P.C. Rombouts, T. Rispens, et al., Toll-like receptor triggering augments activation of human mast cells [by anti-citrullinated protein antibodies, Ann. Rheum. Dis. 74 \(10\) \(2015\) 1915](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref57)–1923.
- [58] [S.K. Kritas, A. Caraffa, P. Antinolfi, A. Saggini, A. Pantalone, M. Rosati, et al., Nerve growth factor interactions with mast cells, Int. J. Immunopathol.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref58) [Pharmacol. 27 \(1\) \(2014\) 15](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref58)–19.
- [59] [S.D. Skaper, Nerve growth factor: a neuroimmune crosstalk mediator for all seasons, Immunology 151 \(1\) \(2017\) 1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref59)–15.
- [60] J. Sousa-Valente, L. Calvo, V. Vacca, R. Simeoli, J.C. Ar´[evalo, M. Malcangio, Role of TrkA signalling and mast cells in the initiation of osteoarthritis pain in the](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref60) [monoiodoacetate model, Osteoarthritis Cartilage 26 \(1\) \(2018\) 84](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref60)–94.
- [61] [J.H. Duan, Y. Wang, D. Duarte, M.R. Vasko, G.D. Nicol, C.M. Hingtgen, Ras signaling pathways mediate NGF-induced enhancement of excitability of small](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref61)[diameter capsaicin-sensitive sensory neurons from wildtype but not Nf1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref61)+/- mice, Neurosci. Lett. 496 (2) (2011) 70–74.
- [62] [J. Hallgren, M.F. Gurish, Granule maturation in mast cells: histamine in control, Eur. J. Immunol. 44 \(1\) \(2014\) 33](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref62)–36.
- [63] [M. Tsuchiya, K. Fukushima, K. Takata, Y. Ohashi, K. Uchiyama, N. Takahira, et al., Increase in TPSB2 and TPSD1 expression in synovium of hip osteoarthritis](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref63) [patients who are overweight, Int. J. Mol. Sci. 24 \(14\) \(2023\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref63).
- [64] [M.G. Buckley, C. Walters, W.M. Wong, M.I. Cawley, S. Ren, L.B. Schwartz, et al., Mast cell activation in arthritis: detection of alpha- and beta-tryptase,](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref64) [histamine and eosinophil cationic protein in synovial fluid, Clin. Sci. \(Lond.\) 93 \(4\) \(1997\) 363](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref64)–370.
- [65] [G.H. Caughey, Mast cell proteases as pharmacological targets, Eur. J. Pharmacol. 778 \(2016\) 44](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref65)–55.
- [66] [J.A. Cairns, Inhibitors of mast cell tryptase beta as therapeutics for the treatment of asthma and inflammatory disorders, Pulm. Pharmacol. Therapeut. 18 \(1\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref66) [\(2005\) 55](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref66)–66.
- [67] [T. Jia, D. Che, Y. Zheng, H. Zhang, Y. Li, T. Zhou, et al., Mast cells initiate type 2 inflammation through tryptase released by MRGPRX2/MRGPRB2 activation in](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref67) [atopic dermatitis, J. Invest. Dermatol. 144 \(1\) \(2024\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref67).
- [68] [S.-W. Oh, C.I. Pae, D.-K. Lee, F. Jones, G.K.S. Chiang, H.-O. Kim, et al., Tryptase inhibition blocks airway inflammation in a mouse asthma model, J. Immunol.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref68) [168 \(4\) \(2002\) 1992](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref68)–2000.
- [69] [S. Nakano, T. Mishiro, S. Takahara, H. Yokoi, D. Hamada, K. Yukata, et al., Distinct expression of mast cell tryptase and protease activated receptor-2 in](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref69) [synovia of rheumatoid arthritis and osteoarthritis, Clin. Rheumatol. 26 \(8\) \(2007\) 1284](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref69)–1292.
- [70] [H.S. Palmer, E.B. Kelso, J.C. Lockhart, C.P. Sommerhoff, R. Plevin, F.G. Goh, et al., Protease-activated receptor 2 mediates the proinflammatory effects of](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref70) [synovial mast cells, Arthritis Rheum. 56 \(11\) \(2007\) 3532](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref70)–3540.
- [71] [C. Huesa, A.C. Ortiz, L. Dunning, L. McGavin, L. Bennett, K. McIntosh, et al., Proteinase-activated receptor 2 modulates OA-related pain, cartilage and bone](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref71) [pathology, Ann. Rheum. Dis. 75 \(11\) \(2016\) 1989](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref71)–1997.
- [72] [N. Das, L.G.N. de Almeida, A. Derakhshani, D. Young, K. Mehdinejadiani, P. Salo, et al., Tryptase](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref72) β regulation of joint lubrication and inflammation via [proteoglycan-4 in osteoarthritis, Nat. Commun. 14 \(1\) \(2023\) 1910.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref72)
- [73] [N.J. Magarinos, K.J. Bryant, A.J. Fosang, R. Adachi, R.L. Stevens, H.P. McNeil, Mast cell-restricted, tetramer-forming tryptases induce aggrecanolysis in](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref73) [articular cartilage by activating matrix metalloproteinase-3 and -13 zymogens, J. Immunol. 191 \(3\) \(2013\) 1404](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref73)–1412.
- [74] [J. Hallgren, G. Pejler, Biology of mast cell tryptase. An inflammatory mediator, FEBS J. 273 \(9\) \(2006\) 1871](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref74)–1895.
- [75] [X. Liu, J. Wang, H. Zhang, M. Zhan, H. Chen, Z. Fang, et al., Induction of mast cell accumulation by tryptase via a protease activated receptor-2 and ICAM-1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref75) [dependent mechanism, Mediat. Inflamm. 2016 \(2016\) 6431574](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref75).
- [76] [S.-H. He, H. Xie, Y.-L. Fu, Inhibition of tryptase release from human colon mast cells by histamine receptor antagonists, Asian Pac. J. Allergy Immunol. 23 \(1\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref76) [\(2005\) 35](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref76)–39.
- [77] [Khedr Mems, A.M. Abdelmotelb, S.L.F. Pender, X. Zhou, A.F. Walls, Neutrophilia, gelatinase release and microvascular leakage induced by human mast cell](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref77) [tryptase in a mouse model: lack of a role of protease-activated receptor 2 \(PAR2\), Clin. Exp. Allergy : J. British Soc. Aller. Clin. Immunol. 48 \(5\) \(2018\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref77) 555–[567](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref77).
- [78] [L.C. Tetlow, D.E. Woolley, Histamine, histamine receptors \(H1 and H2\), and histidine decarboxylase expression by chondrocytes of osteoarthritic cartilage: an](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref78) [immunohistochemical study, Rheumatol. Int. 26 \(2\) \(2005\) 173](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref78)–178.
- [79] [M. Zenmyo, K. Hiraoka, S. Komiya, M. Morimatsu, Y. Sasaguri, Histamine-stimulated production of matrix metalloproteinase 1 by human rheumatoid synovial](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref79) [fibroblasts is mediated by histamine H1-receptors, Virchows Arch. 427 \(4\) \(1995\) 437](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref79)–444.
- [80] [I. Shirinsky, V. Shirinsky, H1-antihistamines are associated with lower prevalence of radiographic knee osteoarthritis: a cross-sectional analysis of the](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref80) [Osteoarthritis Initiative data, Arthritis Res. Ther. 20 \(1\) \(2018\) 116.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref80)
- [81] [E.A. Jemima, A. Prema, E.B. Thangam, Functional characterization of histamine H4 receptor on human mast cells, Mol. Immunol. 62 \(1\) \(2014\) 19](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref81)–28.
- [82] [H.P. McNeil, I. Gotis-Graham, Human mast cell subsets–distinct functions in inflammation? Inflamm. Res. 49 \(1\) \(2000\) 3](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref82)–7.
- [83] M. Laavola, T. Leppänen, M. Hämäläinen, K. Vuolteenaho, T. Moilanen, R. Nieminen, et al., IL-6 in osteoarthritis: effects of pine stilbenoids, Molecules 24 (1) [\(2018\).](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref83)
- [84] [K. Nishida, A. Hasegawa, S. Yamasaki, R. Uchida, W. Ohashi, Y. Kurashima, et al., Mast cells play role in wound healing through the ZnT2/GPR39/IL-6 axis,](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref84) [Sci. Rep. 9 \(1\) \(2019\) 10842.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref84)
- [85] [Z. Gong, Y. Wang, L. Li, X. Li, B. Qiu, Y. Hu, Cardamonin alleviates chondrocytes inflammation and cartilage degradation of osteoarthritis by inhibiting](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref85) [ferroptosis via p53 pathway, Food Chem. Toxicol. : Intern. J. Publis. British Indus. Biolo. Res. Associat. 174 \(2023\) 113644](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref85).
- [86] [S. Ahmed, A. Rahman, A. Hasnain, V.M. Goldberg, T.M. Haqqi, Phenyl N-tert-butylnitrone down-regulates interleukin-1 beta-stimulated matrix](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref86) [metalloproteinase-13 gene expression in human chondrocytes: suppression of c-Jun NH2-terminal kinase, p38-mitogen-activated protein kinase and activating](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref86) [protein-1, J. Pharmacol. Exp. Therapeut. 305 \(3\) \(2003\) 981](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref86)–988.
- [87] [J.A. Mengshol, M.P. Vincenti, C.I. Coon, A. Barchowsky, C.E. Brinckerhoff, Interleukin-1 induction of collagenase 3 \(matrix metalloproteinase 13\) gene](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref87) [expression in chondrocytes requires p38, c-Jun N-terminal kinase, and nuclear factor kappaB: differential regulation of collagenase 1 and collagenase 3,](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref87) [Arthritis Rheum. 43 \(4\) \(2000\) 801](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref87)–811.
- [88] [J.P. Pujol, C. Chadjichristos, F. Legendre, C. Bauge, G. Beauchef, R. Andriamanalijaona, et al., Interleukin-1 and transforming growth factor-beta 1 as crucial](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref88) [factors in osteoarthritic cartilage metabolism, Connect. Tissue Res. 49 \(3\) \(2008\) 293](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref88)–297.
- [89] [F. Legendre, P. Bogdanowicz, K. Boumediene, J.P. Pujol, Role of interleukin 6 \(IL-6\)/IL-6R-induced signal tranducers and activators of transcription and](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref89) [mitogen-activated protein kinase/extracellular, J. Rheumatol. 32 \(7\) \(2005\) 1307](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref89)–1316.
- [90] Y. Cao, I.D. Jansen, S. Sprangers, J. Stap, P.J. Leenen, V. Everts, et al., IL-1β [differently stimulates proliferation and multinucleation of distinct mouse bone](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref90) [marrow osteoclast precursor subsets, J. Leukoc. Biol. 100 \(3\) \(2016\) 513](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref90)–523.
- [91] [G. Gorny, A. Shaw, M.J. Oursler, IL-6, LIF, and TNF-alpha regulation of GM-CSF inhibition of osteoclastogenesis in vitro, Exp. Cell Res. 294 \(1\) \(2004\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref91) 149–[158](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref91).
- [92] [S. Takano, K. Uchida, M. Miyagi, G. Inoue, H. Fujimaki, J. Aikawa, et al., Nerve growth factor regulation by TNF-](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref92)α and IL-1β in synovial macrophages and [fibroblasts in osteoarthritic mice, J Immunol Res 2016 \(2016\) 5706359.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref92)
- [93] [H. Alenius, D. Laouini, A. Woodward, E. Mizoguchi, A.K. Bhan, E. Castigli, et al., Mast cells regulate IFN-gamma expression in the skin and circulating IgE](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref93) [levels in allergen-induced skin inflammation, J. Allergy Clin. Immunol. 109 \(1\) \(2002\) 106](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref93)–113.
- [94] [K. Vosskuhl, T.F. Greten, M.P. Manns, F. Korangy, J. Wedemeyer, Lipopolysaccharide-mediated mast cell activation induces IFN-gamma secretion by NK cells,](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref94) [J. Immunol. 185 \(1\) \(2010\) 119](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref94)–125.
- [95] [D. Elieh Ali Komi, K. Grauwet, Role of mast cells in regulation of T cell responses in experimental and clinical settings, Clin. Rev. Allergy Immunol. 54 \(3\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref95) [\(2018\) 432](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref95)–445.
- [96] V. Drvar, B. Ćurko-Cofek, L. Karleuša, M. Aralica, M. Rogoznica, T. Kehler, et al., Granulysin expression and granulysin-mediated apoptosis in the peripheral [blood of osteoarthritis patients, Biomed. Rep. 16 \(5\) \(2022\) 44](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref96).
- [97] [H. Ishii, H. Tanaka, K. Katoh, H. Nakamura, M. Nagashima, S. Yoshino, Characterization of infiltrating T cells and Th1/Th2-type cytokines in the synovium of](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref97) [patients with osteoarthritis, Osteoarthritis Cartilage 10 \(4\) \(2002\) 277](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref97)–281.
- [98] [Y. Okayama, A.S. Kirshenbaum, D.D. Metcalfe, Expression of a functional high-affinity IgG receptor, Fc gamma RI, on human mast cells: up-regulation by IFN](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref98)[gamma, J. Immunol. 164 \(8\) \(2000\) 4332](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref98)–4339.
- [99] [H. Lee, J-i Kashiwakura, A. Matsuda, Y. Watanabe, T. Sakamoto-Sasaki, K. Matsumoto, et al., Activation of human synovial mast cells from rheumatoid](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref99)
- [arthritis or osteoarthritis patients in response to aggregated IgG through Fc](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref99)γ receptor I and Fcγ receptor II, Arthritis Rheum. 65 (1) (2013) 109–119.
- [100] L. McKelvey, G.D. Shorten, G.W. O'[Keeffe, Nerve growth factor-mediated regulation of pain signalling and proposed new intervention strategies in clinical pain](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref100) [management, J. Neurochem. 124 \(3\) \(2013\) 276](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref100)–289.
- [101] [A. Leon, A. Buriani, R. Dal Toso, M. Fabris, S. Romanello, L. Aloe, et al., Mast cells synthesize, store, and release nerve growth factor, Proc. Natl. Acad. Sci. U. S.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref101) [A. 91 \(9\) \(1994\) 3739](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref101)–3743.
- [102] [S.K. Kritas, A. Caraffa, P. Antinolfi, A. Saggini, A. Pantalone, M. Rosati, et al., Nerve growth factor interactions with mast cells, Int. J. Immunopathol.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref102) [Pharmacol. 27 \(1\) \(2014\) 15](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref102)–19.
- [103] [S. Ashraf, P.I. Mapp, J. Burston, A.J. Bennett, V. Chapman, D.A. Walsh, Augmented pain behavioural responses to intra-articular injection of nerve growth](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref103) [factor in two animal models of osteoarthritis, Ann. Rheum. Dis. 73 \(9\) \(2014\) 1710](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref103)–1718.
- [104] F. Iannone, C. De Bari, F. Dell'[Accio, M. Covelli, V. Patella, Bianco G. Lo, et al., Increased expression of nerve growth factor \(NGF\) and high affinity NGF](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref104) [receptor \(p140 TrkA\) in human osteoarthritic chondrocytes, Rheumatology 41 \(12\) \(2002\) 1413](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref104)–1418.
- [105] [M.J. Benito, D.J. Veale, O. FitzGerald, W.B. van den Berg, B. Bresnihan, Synovial tissue inflammation in early and late osteoarthritis, Ann. Rheum. Dis. 64 \(9\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref105) [\(2005\) 1263](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref105)–1267.
- [106] P. Wojdasiewicz, Ł[.A. Poniatowski, D. Szukiewicz, The role of inflammatory and anti-inflammatory cytokines in the pathogenesis of osteoarthritis, Mediat.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref106) [Inflamm. 2014 \(2014\) 561459](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref106).
- [107] [N. Fukui, Y. Ikeda, T. Ohnuki, A. Hikita, S. Tanaka, S. Yamane, et al., Pro-inflammatory cytokine tumor necrosis factor-alpha induces bone morphogenetic](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref107) [protein-2 in chondrocytes via mRNA stabilization and transcriptional up-regulation, J. Biol. Chem. 281 \(37\) \(2006\) 27229](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref107)–27241.
- [108] [T. Nakase, T. Miyaji, T. Tomita, M. Kaneko, K. Kuriyama, A. Myoui, et al., Localization of bone morphogenetic protein-2 in human osteoarthritic cartilage and](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref108) [osteophyte, Osteoarthritis Cartilage 11 \(4\) \(2003\) 278](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref108)–284.
- [109] [J.C. Fernandes, J. Martel-Pelletier, J.P. Pelletier, The role of cytokines in osteoarthritis pathophysiology, Biorheology 39 \(1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref109)–2) (2002) 237–246.
- [110] [K. Uchida, S. Takano, T. Matsumoto, N. Nagura, G. Inoue, M. Itakura, et al., Transforming growth factor activating kinase 1 regulates extracellular matrix](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref110) [degrading enzymes and pain-related molecule expression following tumor necrosis factor-](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref110)α stimulation of synovial cells: an in vitro study, BMC Muscoskel. [Disord. 18 \(1\) \(2017\) 283](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref110).
- [111] [T. Aizawa, T. Kon, T.A. Einhorn, L.C. Gerstenfeld, Induction of apoptosis in chondrocytes by tumor necrosis factor-alpha, J. Orthop. Res. 19 \(5\) \(2001\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref111) 785–[796.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref111)
- [112] [I. Stanic, A. Facchini, R.M. Borzì, R. Vitellozzi, C. Stefanelli, M.B. Goldring, et al., Polyamine depletion inhibits apoptosis following blocking of survival](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref112) [pathways in human chondrocytes stimulated by tumor necrosis factor-alpha, J. Cell. Physiol. 206 \(1\) \(2006\) 138](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref112)–146.
- [113] [E. Chisari, K.M. Yaghmour, W.S. Khan, The effects of TNF-alpha inhibition on cartilage: a systematic review of preclinical studies, Osteoarthritis Cartilage 28](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref113) [\(5\) \(2020\) 708](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref113)–718.
- [114] [S. Orita, T. Koshi, T. Mitsuka, M. Miyagi, G. Inoue, G. Arai, et al., Associations between proinflammatory cytokines in the synovial fluid and radiographic](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref114) [grading and pain-related scores in 47 consecutive patients with osteoarthritis of the knee, BMC Muscoskel. Disord. 12 \(2011\) 144](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref114).
- [115] Ali Komi D. Elieh, S. Wöhrl, [L. Bielory, Mast cell biology at molecular level: a comprehensive review, Clin. Rev. Allergy Immunol. 58 \(3\) \(2020\) 342](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref115)-365. [116] [S. Takano, K. Uchida, G. Inoue, T. Matsumoto, J. Aikawa, D. Iwase, et al., Vascular endothelial growth factor expression and their action in the synovial](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref116) [membranes of patients with painful knee osteoarthritis, BMC Muscoskel. Disord. 19 \(1\) \(2018\) 204](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref116).
- [117] [L. Haywood, D.F. McWilliams, C.I. Pearson, S.E. Gill, A. Ganesan, D. Wilson, et al., Inflammation and angiogenesis in osteoarthritis, Arthritis Rheum. 48 \(8\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref117) [\(2003\) 2173](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref117)–2177.
- [118] [D.E.A. Komi, F.A. Redegeld, Role of mast cells in shaping the tumor microenvironment, Clin. Rev. Allergy Immunol. 58 \(3\) \(2020\) 313](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref118)–325.
- [119] [C.S. Bonnet, D.A. Walsh, Osteoarthritis, angiogenesis and inflammation, Rheumatology 44 \(1\) \(2005\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref119).
- [120] [A. Ludin, J.J. Sela, A. Schroeder, Y. Samuni, D.W. Nitzan, G. Amir, Injection of vascular endothelial growth factor into knee joints induces osteoarthritis in](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref120) [mice, Osteoarthritis Cartilage 21 \(3\) \(2013\) 491](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref120)–497.
- [121] [D. Ribatti, R. Tamma, T. Annese, Mast cells and angiogenesis in multiple sclerosis, Inflamm. Res. 69 \(11\) \(2020\) 1103](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref121)–1110.
- [122] [X. Zhang, R. Crawford, Y. Xiao, Inhibition of vascular endothelial growth factor with shRNA in chondrocytes ameliorates osteoarthritis, J. Mol. Med. \(Berl.\) 94](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref122) [\(7\) \(2016\) 787](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref122)–798.
- [123] [K. Kyritsi, L. Kennedy, V. Meadows, L. Hargrove, J. Demieville, L. Pham, et al., Mast cells induce ductular reaction mimicking liver injury in mice through mast](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref123) [cell-derived transforming growth factor beta 1 signaling, Hepatology 73 \(6\) \(2021\) 2397](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref123)–2410.
- [124] [G. Biffi, T.E. Oni, B. Spielman, Y. Hao, E. Elyada, Y. Park, et al., IL1-Induced JAK/STAT signaling is antagonized by TGF](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref124)β to shape CAF heterogeneity in [pancreatic ductal adenocarcinoma, Cancer Discov. 9 \(2\) \(2019\) 282](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref124)–301.
- [125] K.N. Bailey, T. Alliston, At the crux of joint crosstalk: TGFβ [signaling in the synovial joint, Curr. Rheumatol. Rep. 24 \(6\) \(2022\) 184](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref125)–197.
- [126] G. Zhai, J. Dor´e, P. Rahman, TGF-β [signal transduction pathways and osteoarthritis, Rheumatol. Int. 35 \(8\) \(2015\) 1283](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref126)–1292.
- [127] [C. Deligne, S. Casulli, A. Pigenet, C. Bougault, L. Campillo-Gimenez, G. Nourissat, et al., Differential expression of interleukin-17 and interleukin-22 in inflamed](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref127) [and non-inflamed synovium from osteoarthritis patients, Osteoarthritis Cartilage 23 \(11\) \(2015\) 1843](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref127)–1852.
- [128] [Y.-S. Li, W. Luo, S.-A. Zhu, G.-H. Lei, T cells in osteoarthritis: alterations and beyond, Front. Immunol. 8 \(2017\) 356](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref128).
- [129] [C.-C. Lee, C.-L. Lin, S.-J. Leu, Y.-L. Lee, Overexpression of Notch ligand Delta-like-1 by dendritic cells enhances their immunoregulatory capacity and exerts](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref129) [antiallergic effects on Th2-mediated allergic asthma in mice, Clin. Immunol. 187 \(2018\) 58](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref129)–67.
- [130] J. Suurmond, A.L. Dorjée, T.W. Huizinga, R.E. Toes, Human mast cells costimulate T cells through a CD28-independent interaction, Eur. J. Immunol. 46 (5) [\(2016\) 1132](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref130)–1141.
- [131] J. Suurmond, J. van Heemst, J. van Heiningen, A.L. Dorjée, M.W. Schilham, F.B. van der Beek, et al., Communication between human mast cells and CD4(+) T [cells through antigen-dependent interactions, Eur. J. Immunol. 43 \(7\) \(2013\) 1758](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref131)–1768.
- [132] [M. Veldhoen, R.J. Hocking, C.J. Atkins, R.M. Locksley, B. Stockinger, TGFbeta in the context of an inflammatory cytokine milieu supports de novo](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref132) [differentiation of IL-17-producing T cells, Immunity 24 \(2\) \(2006\) 179](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref132)–189.
- [133] [H.J. Faust, H. Zhang, J. Han, M.T. Wolf, O.H. Jeon, K. Sadtler, et al., IL-17 and immunologically induced senescence regulate response to injury in](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref133) [osteoarthritis, J. Clin. Invest. 130 \(10\) \(2020\) 5493](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref133)–5507.
- [134] [S. Piconese, G. Gri, C. Tripodo, S. Musio, A. Gorzanelli, B. Frossi, et al., Mast cells counteract regulatory T-cell suppression through interleukin-6 and OX40/](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref134) [OX40L axis toward Th17-cell differentiation, Blood 114 \(13\) \(2009\) 2639](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref134)–2648.
- [135] [H. Yamada, Y. Nakashima, K. Okazaki, T. Mawatari, J. Fukushi, A. Oyamada, et al., Preferential accumulation of activated Th1 cells not only in rheumatoid](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref135) [arthritis but also in osteoarthritis joints, J. Rheumatol. 38 \(8\) \(2011\) 1569](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref135)–1575.
- [136] [J. Xiao, P. Zhang, F.-L. Cai, C.-G. Luo, T. Pu, X.-L. Pan, et al., IL-17 in osteoarthritis: a narrative review, Open Life Sci. 18 \(1\) \(2023\) 20220747](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref136).
- [137] [X. Ye, Q. Lu, A. Yang, J. Rao, W. Xie, C. He, et al., MiR-206 regulates the Th17/Treg ratio during osteoarthritis, Mol. Med. 27 \(1\) \(2021\) 64](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref137).
- [138] [S. Bulfone-Paus, R. Bahri, Mast cells as regulators of T cell responses, Front. Immunol. 6 \(2015\) 394.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref138)
- [139] [B.J.E. de Lange-Brokaar, A. Ioan-Facsinay, G.J.V.M. van Osch, A.M. Zuurmond, J. Schoones, R.E.M. Toes, et al., Synovial inflammation, immune cells and their](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref139) [cytokines in osteoarthritis: a review, Osteoarthritis Cartilage 20 \(12\) \(2012\) 1484](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref139)–1499.
- [140] [M.-F. Hsueh, X. Zhang, S.S. Wellman, M.P. Bolognesi, V.B. Kraus, Synergistic roles of macrophages and neutrophils in osteoarthritis progression, Arthritis](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref140) [Rheumatol. 73 \(1\) \(2021\) 89](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref140)–99.
- [141] [S. Mishima, J.I. Kashiwakura, S. Toyoshima, T. Sasaki-Sakamoto, Y. Sano, K. Nakanishi, et al., Higher PGD\(2\) production by synovial mast cells from](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref141)
- [rheumatoid arthritis patients compared with osteoarthritis patients via miR-199a-3p/prostaglandin synthetase 2 axis, Sci. Rep. 11 \(1\) \(2021\) 5738](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref141). [142] [S.M. Thakurdas, E. Melicoff, L. Sansores-Garcia, D.C. Moreira, Y. Petrova, R.L. Stevens, et al., The mast cell-restricted tryptase mMCP-6 has a critical](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref142) [immunoprotective role in bacterial infections, J. Biol. Chem. 282 \(29\) \(2007\) 20809](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref142)–20815.
- [143] [S.J. Galli, N. Borregaard, T.A. Wynn, Phenotypic and functional plasticity of cells of innate immunity: macrophages, mast cells and neutrophils, Nat. Immunol.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref143) [12 \(11\) \(2011\) 1035](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref143)–1044.
- [144] [T. Kishimoto, W. Ishida, I. Nakajima, O. Taguchi, K. Sugioka, S. Kusaka, et al., Promotion of conjunctival fibroblast-mediated collagen gel contraction by mast](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref144) [cells through up-regulation of matrix metalloproteinase release and activation, Exp. Eye Res. 218 \(2022\) 108980.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref144)
- [145] [M. Leist, C.A. Sünder, S. Drube, C. Zimmermann, A. Geldmacher, M. Metz, et al., Membrane-bound stem cell factor is the major but not only driver of](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref145) [fibroblast-induced murine skin mast cell differentiation, Exp. Dermatol. 26 \(3\) \(2017\) 255](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref145)–262.
- [146] [H. Farah, S.N. Wijesinghe, T. Nicholson, F. Alnajjar, M. Certo, A. Alghamdi, et al., Differential metabotypes in synovial fibroblasts and synovial fluid in hip](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref146) [osteoarthritis patients support inflammatory responses, Int. J. Mol. Sci. 23 \(6\) \(2022\).](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref146)
- [147] [J.C. Kendall, X.H. Li, S.J. Galli, J.R. Gordon, Promotion of mouse fibroblast proliferation by IgE-dependent activation of mouse mast cells: role for mast cell](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref147) [tumor necrosis factor-alpha and transforming growth factor-beta 1, J. Allergy Clin. Immunol. 99 \(1 Pt 1\) \(1997\) 113](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref147)–123.
- [148] [A. Mathiessen, P.G. Conaghan, Synovitis in osteoarthritis: current understanding with therapeutic implications, Arthritis Res. Ther. 19 \(1\) \(2017\) 18](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref148).
- [149] [Y. Chu, J. Wang, X. Zhou, Mast cell chymase in synovial fluid of collagen-induced-arthritis rats regulates gelatinase release and promotes synovial fibroblasts](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref149) [proliferation via FAK/p21 signaling pathway, Biochem. Biophys. Res. Commun. 514 \(1\) \(2019\) 336](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref149)–343.
- [150] [H. Zhang, D. Cai, X. Bai, Macrophages regulate the progression of osteoarthritis, Osteoarthritis Cartilage 28 \(5\) \(2020\) 555](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref150)–561.
- [151] [K. Warmink, P. Vinod, N.M. Korthagen, H. Weinans, J.L. Rios, Macrophage-Driven inflammation in metabolic osteoarthritis: implications for biomarker and](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref151) [therapy development, Int. J. Mol. Sci. 24 \(7\) \(2023\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref151).
- [152] [H. Zhang, C. Lin, C. Zeng, Z. Wang, H. Wang, J. Lu, et al., Synovial macrophage M1 polarisation exacerbates experimental osteoarthritis partially through R](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref152)[spondin-2, Ann. Rheum. Dis. 77 \(10\) \(2018\) 1524](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref152)–1534.
- [153] [N. Wang, H. Liang, K. Zen, Molecular mechanisms that influence the macrophage m1-m2 polarization balance, Front. Immunol. 5 \(2014\) 614](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref153).
- [154] [N. Cao, D. Wang, B. Liu, Y. Wang, W. Han, J. Tian, et al., Silencing of STUB1 relieves osteoarthritis via inducing NRF2-mediated M2 macrophage polarization,](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref154)
- [Mol. Immunol. 164 \(2023\) 112](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref154)–122. [155] [M. Cutolo, R. Campitiello, E. Gotelli, S. Soldano, The role of M1/M2 macrophage polarization in rheumatoid arthritis synovitis, Front. Immunol. 13 \(2022\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref155) [867260.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref155)
- [156] [J. Geurts, A. Patel, M.T. Hirschmann, G.I. Pagenstert, M. Müller-Gerbl, V. Valderrabano, et al., Elevated marrow inflammatory cells and osteoclasts in](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref156) [subchondral osteosclerosis in human knee osteoarthritis, J. Orthop. Res. 34 \(2\) \(2016\) 262](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref156)–269.
- [157] [M.M. McDonald, W.H. Khoo, P.Y. Ng, Y. Xiao, J. Zamerli, P. Thatcher, et al., Osteoclasts recycle via osteomorphs during RANKL-stimulated bone resorption,](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref157) [Cell 184 \(5\) \(2021\).](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref157)
- [158] [V. Fischer, D. Ragipoglu, J. Diedrich, L. Steppe, A. Dudeck, K. Schütze, et al., Mast cells trigger disturbed bone healing in osteoporotic mice, J. Bone Miner. Res.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref158) [: Off. J. Amer. Soc. Bone Miner. Res. 37 \(1\) \(2022\) 137](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref158)–151.
- [159] [M. Biosse-Duplan, B. Baroukh, M. Dy, M.-C. de Vernejoul, J.-L. Saffar, Histamine promotes osteoclastogenesis through the differential expression of histamine](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref159) [receptors on osteoclasts and osteoblasts, Am. J. Pathol. 174 \(4\) \(2009\) 1426](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref159)–1434.
- [160] [K.W. Kim, B.M. Kim, J.Y. Won, H.K. Min, K.A. Lee, S.H. Lee, et al., Regulation of osteoclastogenesis by mast cell in rheumatoid arthritis, Arthritis Res. Ther. 23](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref160) [\(1\) \(2021\) 124.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref160)
- [161] T. Wang, C. He, TNF-А [and IL-6: the link between immune and bone system, Curr. Drug Targets 21 \(3\) \(2020\) 213](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref161)–227.
- [162] [D.S. Amarasekara, H. Yun, S. Kim, N. Lee, H. Kim, J. Rho, Regulation of osteoclast differentiation by cytokine networks, Immune Netw 18 \(1\) \(2018\) e8.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref162) [163] [Y. Otsuka, T. Kondo, H. Aoki, Y. Goto, Y. Kawaguchi, Y. Waguri-Nagaya, et al., IL-1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref163)β promotes osteoclastogenesis by increasing the expression of IGF2 and
- [chemokines in non-osteoclastic cells, J. Pharmacol. Sci. 151 \(1\) \(2023\) 1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref163)–8. [164] [C.W. Ng, B.C.L. Chan, C.H. Ko, I.Y.S. Tam, S.W. Sam, C.B.S. Lau, et al., Human mast cells induce osteoclastogenesis through cell surface RANKL, Inflamm. Res.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref164) 71 (10–[11\) \(2022\) 1261](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref164)–1270.
- [165] [H. Goto, A. Hozumi, M. Osaki, T. Fukushima, K. Sakamoto, A. Yonekura, et al., Primary human bone marrow adipocytes support TNF-](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref165)α-induced osteoclast [differentiation and function through RANKL expression, Cytokine 56 \(3\) \(2011\) 662](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref165)–668.
- [166] Z. Zhao, X. Hou, X. Yin, Y. Li, R. Duan, B.F. Boyce, et al., TNF induction of NF-κ[B RelB enhances RANKL-induced osteoclastogenesis by promoting inflammatory](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref166) [macrophage differentiation but also limits it through suppression of NFATc1 expression, PLoS One 10 \(8\) \(2015\) e0135728](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref166).
- [167] C. Guo, X.G. Yang, F. Wang, X.Y. Ma, IL-1α [induces apoptosis and inhibits the osteoblast differentiation of MC3T3-E1 cells through the JNK and p38 MAPK](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref167) [pathways, Int. J. Mol. Med. 38 \(1\) \(2016\) 319](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref167)–327.
- [168] [K.W. Finnson, Y. Chi, G. Bou-Gharios, A. Leask, A. Philip, TGF-b signaling in cartilage homeostasis and osteoarthritis, Front Biosci \(Schol Ed\) 4 \(2012\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref168) 251–[268.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref168)
- [169] [J.A. Martin, J.A. Buckwalter, Aging, articular cartilage chondrocyte senescence and osteoarthritis, Biogerontology 3 \(5\) \(2002\) 257](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref169)–264.
- [170] [H.A. Kim, M.L. Cho, H.Y. Choi, C.S. Yoon, J.Y. Jhun, H.J. Oh, et al., The catabolic pathway mediated by Toll-like receptors in human osteoarthritic](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref170) [chondrocytes, Arthritis Rheum. 54 \(7\) \(2006\) 2152](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref170)–2163.
- [171] G. Zhen, X. Cao, Targeting TGFβ [signaling in subchondral bone and articular cartilage homeostasis, Trends Pharmacol. Sci. 35 \(5\) \(2014\) 227](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref171)–236.
- [172] [J. Shen, J. Li, B. Wang, H. Jin, M. Wang, Y. Zhang, et al., Deletion of the transforming growth factor](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref172) β receptor type II gene in articular chondrocytes leads to a [progressive osteoarthritis-like phenotype in mice, Arthritis Rheum. 65 \(12\) \(2013\) 3107](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref172)–3119.
- [173] [B. Abramoff, F.E. Caldera, Osteoarthritis: pathology, diagnosis, and treatment options, Med Clin North Am 104 \(2\) \(2020\) 293](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref173)–311.
- [174] [D.T. Goldman, R. Piechowiak, D. Nissman, S. Bagla, A. Isaacson, Current concepts and future directions of minimally invasive treatment for knee pain, Curr.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref174) [Rheumatol. Rep. 20 \(9\) \(2018\) 54.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref174)
- [175] [M. Vetrano, D. Ranieri, M. Nanni, A. Pavan, F. Malisan, M.C. Vulpiani, et al., Hyaluronic Acid \(HA\), Platelet-Rich Plasm and Extracorporeal Shock Wave](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref175) [Therapy \(ESWT\) promote human chondrocyte regeneration in vitro and ESWT-mediated increase of CD44 expression enhances their susceptibility to HA](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref175) [treatment, PLoS One 14 \(6\) \(2019\) e0218740](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref175).
- [176] F. Russo, M. D'Este, G. Vadalà, C. Cattani, R. Papalia, M. Alini, et al., Platelet rich plasma and hyaluronic acid blend for the treatment of osteoarthritis: [rheological and biological evaluation, PLoS One 11 \(6\) \(2016\) e0157048.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref176)
- [177] D. Szwedowski, J. Szczepanek, Ł. Paczesny, J. Zabrzyński, M. Gagat, A. Mobasheri, et al., The effect of platelet-rich plasma on the intra-articular [microenvironment in knee osteoarthritis, Int. J. Mol. Sci. 22 \(11\) \(2021\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref177).
- [178] [A. Loucks, T. Maerz, K. Hankenson, A. Moeser, A. Colbath, The multifaceted role of mast cells in joint inflammation and arthritis, Osteoarthritis Cartilage 31](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref178) [\(5\) \(2023\) 567](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref178)–575.
- [179] [H.Y. Shin, J.S. Kim, N.H. An, R.K. Park, H.M. Kim, Effect of disodium cromoglycate on mast cell-mediated immediate-type allergic reactions, Life Sci. 74 \(23\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref179) [\(2004\) 2877](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref179)–2887.
- [180] [L. Soucek, E.R. Lawlor, D. Soto, K. Shchors, L.B. Swigart, G.I. Evan, Mast cells are required for angiogenesis and macroscopic expansion of Myc-induced](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref180) [pancreatic islet tumors, Nat. Med. 13 \(10\) \(2007\) 1211](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref180)–1218.
- [181] [J. Sun, G.K. Sukhova, M. Yang, P.J. Wolters, L.A. MacFarlane, P. Libby, et al., Mast cells modulate the pathogenesis of elastase-induced abdominal aortic](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref181) [aneurysms in mice, J. Clin. Invest. 117 \(11\) \(2007\) 3359](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref181)–3368.
- [182] [C. Tontini, S. Bulfone-Paus, Novel approaches in the inhibition of IgE-induced mast cell reactivity in food allergy, Front. Immunol. 12 \(2021\) 613461.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref182)
- [183] [E.M. Kolawole, J.J.A. McLeod, V. Ndaw, D. Abebayehu, B.O. Barnstein, T. Faber, et al., Fluvastatin suppresses mast cell and basophil IgE responses: genotype](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref183)[dependent effects, J. Immunol. 196 \(4\) \(2016\) 1461](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref183)–1470.
- [184] [I. Shirinsky, V. Shirinsky, H\(1\)-antihistamines are associated with lower prevalence of radiographic knee osteoarthritis: a cross-sectional analysis of the](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref184) [Osteoarthritis Initiative data, Arthritis Res. Ther. 20 \(1\) \(2018\) 116](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref184).
- [185] [M. Cao, Y. Gao, Mast cell stabilizers: from pathogenic roles to targeting therapies, Front. Immunol. 15 \(2024\) 1418897.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref185)
- [186] [D. Alvarado, M. Maurer, R. Gedrich, S.B. Seibel, M.B. Murphy, L. Crew, et al., Anti-KIT monoclonal antibody CDX-0159 induces profound and durable mast cell](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref186) [suppression in a healthy volunteer study, Allergy 77 \(8\) \(2022\) 2393](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref186)–2403.
- [187] [D. Terhorst-Molawi, T. Hawro, E. Grekowitz, L. Kiefer, K. Merchant, D. Alvarado, et al., Anti-KIT antibody, barzolvolimab, reduces skin mast cells and disease](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref187)
- [activity in chronic inducible urticaria, Allergy 78 \(5\) \(2023\) 1269](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref187)–1279.
[188] N.E. Lane, T.J. Schnitzer, C.A. Birbara, M. Mokhtarani, D.L. Shelton, M.D. Smith, et al., Tanezumab for the treatment of pain from osteoarthri [N. Engl. J. Med. 363 \(16\) \(2010\) 1521](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref188)–1531.
- [189] [H. Nagashima, M. Suzuki, S. Araki, T. Yamabe, C. Muto, Preliminary assessment of the safety and efficacy of tanezumab in Japanese patients with moderate to](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref189) [severe osteoarthritis of the knee: a randomized, double-blind, dose-escalation, placebo-controlled study, Osteoarthritis Cartilage 19 \(12\) \(2011\) 1405](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref189)–1412.
- [190] [A.R. Balanescu, E. Feist, G. Wolfram, I. Davignon, M.D. Smith, M.T. Brown, et al., Efficacy and safety of tanezumab added on to diclofenac sustained release in](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref190) [patients with knee or hip osteoarthritis: a double-blind, placebo-controlled, parallel-group, multicentre phase III randomised clinical trial, Ann. Rheum. Dis. 73](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref190) [\(9\) \(2014\) 1665](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref190)–1672.
- [191] [Z.R. Fan, J.X. Ma, Y. Wang, H.T. Chen, S. Lang, X.L. Ma, Efficacy and safety of tanezumab administered as a fixed dosing regimen in patients with knee or hip](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref191) [osteoarthritis: a meta-analysis of randomized controlled phase III trials, Clin. Rheumatol. 40 \(6\) \(2021\) 2155](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref191)–2165.
- [192] [Z. Ji, J. Li, S. Tao, H. Li, X. Kong, B. Huang, et al., Mrgprb2-mediated mast cell activation exacerbates Modic changes by regulating immune niches, Exp. Mol.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref192) [Med. 56 \(5\) \(2024\) 1178](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref192)–1192.
- [193] [J.A. Martin, J.A. Buckwalter, The role of chondrocyte senescence in the pathogenesis of osteoarthritis and in limiting cartilage repair, J Bone Joint Surg Am](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref193) [\(85-A Suppl 2\) \(2003\) 106](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref193)–110.
- [194] [J.A. Martin, J.A. Buckwalter, Human chondrocyte senescence and osteoarthritis, Biorheology 39 \(1](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref194)–2) (2002) 145–152.
- [195] [S. Bais, R. Kumari, Y. Prashar, N.S. Gill, Review of various molecular targets on mast cells and its relation to obesity: a future perspective, Diabetes Metabol.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref195) [Syndr. 11 \(Suppl 2\) \(2017\) S1001, s7](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref195).
- [196] [Z. Zhang, P.B. Ernst, H. Kiyono, Y. Kurashima, Utilizing mast cells in a positive manner to overcome inflammatory and allergic diseases, Front. Immunol. 13](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref196) [\(2022\) 937120.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref196)
- [197] [G. Cildir, K.H. Yip, H. Pant, V. Tergaonkar, A.F. Lopez, D.J. Tumes, Understanding mast cell heterogeneity at single cell resolution, Trends Immunol. 42 \(6\)](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref197) [\(2021\) 523](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref197)–535.
- [198] [E. Kilinc, I.E. Torun, A. Cetinkaya, F. Tore, Mast cell activation ameliorates pentylenetetrazole-induced seizures in rats: the potential role for serotonin, Eur. J.](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref198) Neurosci. 55 (9–[10\) \(2022\) 2912](http://refhub.elsevier.com/S2405-8440(24)17034-X/sref198)–2924.