RESEARCH ARTICLE

Open Access

The lungs were on fire: a pilot study of ¹⁸F-FDG PET/CT in idiopathic-inflammatory-myopathy-related interstitial lung disease



Junyu Liang^{1†}, Heng Cao^{1†}, Yinuo Liu², Bingjue Ye³, Yiduo Sun¹, Yini Ke¹, Ye He¹, Bei Xu¹ and Jin Lin^{1*}

Abstract

Background: Interstitial lung disease (ILD) and its rapid progression (RP) are the main contributors to unfavourable outcomes of patients with idiopathic inflammatory myopathy (IIM). This study aimed to identify the clinical value of PET/CT scans in IIM-ILD patients and to construct a predictive model for RP-ILD.

Methods: Adult IIM-ILD patients who were hospitalized at four divisions of the First Affiliated Hospital, Zhejiang University School of Medicine (FAHZJU), from 1 January 2017 to 31 December 2020 were reviewed. PET/CT scans and other characteristics of patients who met the inclusion and exclusion criteria were collected and analysed.

Results: A total of 61 IIM-ILD patients were enrolled in this study. Twenty-one patients (34.4%) developed RP-ILD, and 24 patients (39.3%) died during follow-up. After false discovery rate (FDR) correction, the percent-predicted diffusing capacity of the lung for carbon monoxide (DLCO%, P = 0.014), bilateral lung mean standard uptake value (SUVmean, P = 0.014) and abnormal mediastinal lymph node (P = 0.045) were significantly different between the RP-ILD and non-RP-ILD groups. The subsequent univariate and multivariate logistic regression analyses verified our findings. A "DLM" model was established by including the above three values to predict RP-ILD with a cut-off value of \geq 2 and an area under the curve (AUC) of 0.905. Higher bilateral lung SUVmean (P = 0.019) and spleen SUVmean (P = 0.011) were observed in IIM-ILD patients who died within 3 months, and a moderate correlation was recognized between the two values.

Conclusions: Elevated bilateral lung SUVmean, abnormal mediastinal lymph nodes and decreased DLCO% were significantly associated with RP-ILD in IIM-ILD patients. The "DLM" model was valuable in predicting RP-ILD and requires further validation.

Keywords: Idiopathic inflammatory myopathy, Interstitial lung disease, PET/CT scan, Dermatomyositis, Polymyositis

Introduction

Interstitial lung disease (ILD) is a frequent complication in patients with idiopathic inflammatory myopathy, showing a predominantly non-specific interstitial pneumonia (NSIP) pattern in histopathological findings [1–

3]. A large proportion of idiopathic-inflammatory-myopathy-related ILD (IIM-ILD) cases are rapidly progressive (RP) and refractory to conventional immuno-suppressive therapy, making the clinical management of IIM-ILD challenging [4]. In recent studies, the survival of IIM patients with ILD, RP-ILD in particular, is far from satisfactory. Specifically, the mortality rate of IIM-ILD patients varied from 19.0 to 27.0% during follow-up, and a 3-month mortality rate of over 30.0% was identified for RP-ILD [5–7]. Therefore, it is vital to search for

Zhejiang Province, People's Republic of China Full list of author information is available at the end of the article



© The Author(s). 2021 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

^{*} Correspondence: linjinzju@zju.edu.cn

[†]Junyu Liang and Heng Cao contributed equally to this work. ¹Department of Rheumatology, The First Affiliated Hospital, Zhejiang University School of Medicine, #79 Qingchun Road, Hangzhou 310003,

efficient biomarkers or tools for predicting the development and progression of RP-ILD in order to avoid unfavourable outcomes.

Activation and infiltration of immune cells and cytokine release participate in the development of idiopathic pulmonary fibrosis (IPF), connective tissue diseaserelated ILD (CTD-ILD) and coronavirus disease 2019 (COVID-19) [8-11]. Since elevated ¹⁸F-fluorodeoxyglucose (FDG) uptake on PET/CT scans can indicate focal immune activation, a series of studies have focused on the clinical value of PET/CT scans in these diseases. In systemic sclerosis-related ILD (SSc-ILD), pulmonary FDG uptake was significantly increased and correlated with the severity of ILD [12, 13]. In addition, elevated pulmonary FDG uptake also predicted the progression of SSc-ILD [13]. Dr Morita reported an IIM-ILD patient who presented with high pulmonary FDG uptake and subsequently died of RP-ILD [14]. Subsequently, two small-sample cohort studies found that FDG uptake (maximum standard uptake value, SUVmax) in the lung was correlated with ILD severity and might predict the occurrence of RP-ILD [15, 16]. However, the sample sizes were too small, and the included organs were only the lungs and muscles. Meanwhile, the correlation between abnormal FDG uptake and the survival of IIM-ILD patients remains unclear. A systemic evaluation of FDG uptake in IIM-ILD patients is necessary to acquire a broader view of the clinical value of PET/CT scans in IIM-ILD.

A systemic evaluation of FDG uptake in a larger cohort of IIM-ILD patients was thus implemented to clarify the predictive value of FDG uptake in multiple organs for RP-ILD as well as its ability to predict unfavourable outcomes. The mean value of the region of interest (ROI) instead of the maximum value at a single pixel was calculated to avoid interference and to acquire a more representative FDG uptake of the targeted organ. In addition, we constructed a model to predict RP-ILD and unfavourable outcomes in IIM-ILD patients by incorporating statistically significant values on PET/CT scans, lung function testing, and other variables.

Patients and methods

Patients

A retrospective cohort study was conducted at the inpatient department of Qingchun, Chengzhan, Zhijiang and Yuhang divisions of the First Affiliated Hospital, Zhejiang University School of Medicine (FAHZJU). After acquiring approval from the Institutional Review Board (IRB) of FAHZJU (Reference Number: 2021-194) and written informed consent from all of the patients involved, in accordance with the Declaration of Helsinki, a case search was retrospectively conducted of the inpatient electronic medical record (EMR) system for patients with discharge diagnoses of dermatomyositis

(DM), polymyositis (PM) or amyopathic dermatomyositis (ADM), from 1 January 2017 to 31 December 2020. The inclusion criteria for this study were (1) age over 18 years old; (2) a definite/probable diagnosis of DM, PM or ADM applying the 2017 ACR/EULAR classification criteria, as confirmed by two experienced rheumatologists (Heng Cao and Bei Xu) [17]; (3) ILD on highresolution computed tomography (HRCT) within the first week of admission, including the usual interstitial pneumonia (UIP) pattern and non-UIP patterns (NSIP, cryptogenic organizing pneumonia, the coexistence of more than one CT pattern), as confirmed by an experienced radiologist and a respiratory specialist (Yinuo Liu and Bingjue Ye); and (4) a PET/CT scan performed during hospitalization. The exclusion criteria were (1) clarified overlap syndromes with other connective tissue diseases (CTDs); (2) myopathy related to thyroid dysfunction, strenuous exercise, inherited metabolic disorders, drug-induced myositis (lamivudine, statins, Chinese herbal medicine, etc.; (3) hospitalization for reasons unrelated to myositis and its complications, such as fracture, pregnancy, acquired immunodeficiency syndrome, cataract, etc.; (4) newly identified or unresolved malignancies; and (5) loss to follow-up without death from any cause within 3 months after hospitalization.

Data collection

Data from all of the enrolled patients were retrospectively collected by referring to the EMR system of FAHZJU. Clinical records, including demographic information, the course of the disease, the duration of diagnostic delay, disease activity assessment, complications, radiological/laboratory results, lung function testing, and immunosuppressive medications, were acquired and analysed. Survival data were acquired from the follow-up data. Specifically, IIM patients were followed from hospitalization until the end of the follow-up period or death, whichever happened first. For patients who died during hospitalization, their dates of death were clearly documented in the EMR system. For patients who were discharged, a routine return visit was arranged 2 weeks after discharge. In addition to the regular inpatient or outpatient visits, a brief telephone interview was performed 3 months after discharge and then every year. The end of follow-up could be due to death from any cause, loss to follow-up or closure of follow-up for the purpose of this study (31 March 2021).

Baseline disease activity assessment, laboratory tests, lung HRCT scans and lung function testing were carried out within the first week of hospitalization. On-admission IIM disease activity was routinely assessed using the Myositis Disease Activity Assessment Visual Analogue Scale (MYOACT) [18]. ILD and its rapid progression were confirmed by experienced respiratory

specialist and radiologist (Bingjue Ye and Yinuo Liu) using lung HRCTs. Cases with definite or probable UIP patterns were identified based on their classic HRCT manifestation: the presence of basal-dominant reticular opacities and a predominantly basal and subpleural distribution of honeycomb lesions, with multiple equalsized cystic lesions of two to ten mm diameter with a thick wall [19].

A subgroup of RP-ILD patients was considered who presented with progressive dyspnoea and progressive hypoxemia, with acute worsening of interstitial changes chest radiographs within 1 month hospitalization or the onset of respiratory symptoms [20-22]. The included IIM-ILD patients were divided into an RP-ILD group and a non-RP-ILD group (control group). Since pulmonary infection is easily confused with interstitial lung disease, the identification of bacterial, fungal, or tuberculosis infection requires a careful and comprehensive investigation based on the essential microbiological findings in sputum or blood, the clinical manifestations, and the radiographic and laboratory abnormalities. In addition, the diagnosis of Epstein-Barr virus (EBV) and cytomegalovirus (CMV) infection relied on the detection of serum antibodies and DNA.

All of the included patients received potent immuno-suppressive medications: (1) systemic prednisolone (PSL) or methylprednisolone (mPSL) with a maximum dosage ≥ 1 mg/kg/day (calculated for prednisolone) and (2) combined therapy of PSL/mPSL, disease-modifying anti-rheumatic drugs (DMARDs) or Janus kinase (JAK) inhibitors, with or without intravenous immunoglobulin (IVIG). The DMARDs used for these patients included mycophenolate, tacrolimus, cyclosporine, methotrexate, thalidomide, hydroxychloroquine and cyclophosphamide. JAK inhibitors mainly referred to tofacitinib and baricitinib.

The profiles of 12 myositis-specific antibodies (MSAs, anti-MDA5, anti-TIF1γ, anti-Jo-1, anti-OJ, anti-EJ, anti-PL-12, anti-PL-7, anti-Mi-2α, anti-Mi-2β, anti-NXP2, anti-SRP and anti-SAE1) and four myositis-associated antibodies (MAAs, anti-Ku, anti-PM-Scl75, anti-PM-Scl100 and anti-Ro-52) were assessed by an immuno-blotting assay utilizing the EUROLINE Autoimmune Inflammatory Myopathies 16 Ag (IgG) commercial line blot assay (Euroimmun, Lübeck, Germany) encompassing a membrane strip with the 16 autoantigens following the manufacturer's instructions. Serum samples were routinely acquired from suspected IIM patients within the first week of hospitalization and subjected to testing.

Whole-body CT and PET, which were performed with a combined PET/CT scanner (Biograph, Sensation 16, Siemens systems), covered a region ranging from the meatus of the ear to the mid-thigh. Patients fasted overnight or for at least 6 h prior to PET/CT detection.

Blood glucose levels were confirmed to be within normal limits before the injection of 4.0 MBq/kg [18F]FDG. Patients rested for 30 min to minimize non-specific FDG uptake by muscles. Imaging acquisition was systematically implemented at 60 min post-injection. SUV (standard uptake value) was calculated by the following formula: SUV (g/ml) = regional radioactivity concentration (Bq/ml)/[injected dose (Bq)/body weight (g)]. The ROI (20 mm diameter) was manually placed by a single trained radiologist (Yinuo, Liu) in the region with the highest FDG uptake in the liver, spleen, and bone marrow (thoracic, T10-12, lumbar, L2-L4) [23], oesophagus, stomach, small intestine, colon/rectum, bilateral lung, bilateral cerebellum, bilateral proximal muscles (namely, trapezius, deltoid, biceps, iliopsoas, gluteus medius, gluteus maximus and quadriceps) [24], excluding the regions prominently influenced by FDG uptake in other anatomical structures. To avoid noise and acquire the value representing a certain volume of the targeted organs, SUV was calculated as the mean value of ROI (SUVmean) instead of the maximum value at a single pixel [25]. For bilaterally distributed organs, the SUVmean was documented as the maximum SUVmean value of the symmetrical sides. Abnormal hilar or mediastinal lymph nodes were defined as those with swelling and elevated FDG uptake. The radiologist was blinded to the complications and outcome of the included patients when evaluating the SUVmean value of each and every targeted organ.

Statistical analysis

Statistical analysis was performed using SPSS 22.0 (Chicago, IL, USA), GraphPad Prism 8.0 and R 3.6.1. An independent sample t test was utilized to compare normally distributed continuous variables between the RP-ILD and control groups. The Mann-Whitney *U* test was used to compare skewed continuous variables or ordinal categorical variables. The chi-square test and Fisher's exact test were applied to compare unordered categorical variables. Logistic regression analyses were used to further verify clinical factors significantly associated with RP-LD. Survival in different groups was assessed by the Kaplan-Meier method with the log-rank test. Cox proportional hazards regression analyses were subsequently adopted to identify the effect of clinical factors on the time to death from any cause. P values of the comparisons and univariate analyses were adjusted by false discovery rate (FDR) correction, utilizing the p.adjust function in R.3.6.1, to acquire adjusted p values and to minimize type I error. Explanatory factors with P < 0.05 in the univariate analyses were entered into the subsequent multivariate analyses. The correlation between two continuous variables was quantified with Pearson linear analysis. Receiver operating characteristic (ROC)

curve analysis was performed to evaluate the predictive value of the continuous variables. All tests were two-sided, and P < 0.05 was deemed statistically significant.

Results

From 1 January 2017 to 31 December 2020, 274 adult IIM-ILD patients were admitted to the Qingchun, Chengzhan, Zhijiang and Yuhang divisions of FAHZJU. Among them, 61 patients who satisfied the inclusion/exclusion criteria were included in the study (Additional file 1), encompassing 40 with DM, nine with PM and 12 with ADM. Their mean age was 56.72 ± 11.28 years old and 25 were men (41.0%). Twenty-four patients (39.3%) died during follow-up, and the median follow-up time was 11.90 (4.00, 23.80) months. Among the included patients, 21 patients (34.4%) developed RP-ILD (Fig. 1). The other 40 patients without RP-ILD constituted the control group. All of the RP-ILD events were identified after taking PET/CT scans. IIM patients with RP-ILD were found to suffer from shorter survival (P = 0.005, Fig. 2A), with 9 RP-ILD patients (42.9%) dying within 3 months after hospitalization.

An unadjusted comparison between IIM-ILD patients with or without RP-ILD showed that patients who later developed RP-ILD had more complications of pulmonary bacterial infection (P = 0.036) and gastrointestinal haemorrhage (P = 0.044), higher MYOACT score (P =0.006), a higher bilateral lung SUVmean (P < 0.001, Fig. 2B), more abnormal mediastinal (P = 0.002) and hilar (P= 0.018) lymph nodes, lower total lung capacity (TLC, P = 0.049), lower percent-predicted diffusing capacity of the lung for carbon monoxide (DLCO%, P < 0.001, Fig. 2C) and positivity of anti-MDA5 antibody (P = 0.016). However, after FDR correction, only DLCO% (P = 0.014), bilateral lung SUVmean (P = 0.014) and abnormal mediastinal lymph nodes (P = 0.045) remained significant (Table 1). The subsequent univariate and multivariate logistic regression analyses also revealed the statistical significance of DLCO% (P = 0.003), bilateral lung SUVmean (P = 0.001) and abnormal mediastinal lymph nodes (P = 0.013) in association with RP-ILD (Additional files 2 and 3). No significant correlation was identified among the three clinical factors (P = 0.491 and r = -0.090 for bilateral lung SUVmean and DLCO%, P = 0.243 for bilateral lung SUV mean and abnormal mediastinal lymph node, P = 0.077 for DLCO% and abnormal mediastinal lymph node, Additional file 4).

Utilizing ROC curve analysis, the optimal cut-off value of the bilateral lung SUVmean for RP-ILD was > 0.454, with a sensitivity of 95.2% and a specificity of 62.5%. The area under the curve (AUC) was 0.805 (Fig. 2D). Meanwhile, the optimal cut-off value of DLCO% for RP-ILD was < 49.0%, with a sensitivity of 87.5%, a specificity of 66.7% and an AUC of 0.802 (Fig. 2E). Moreover,

pulmonary bacterial infection (P = 0.007) and pulmonary fungal infection (P = 0.034) were associated with elevated bilateral lung SUVmean. A moderate correlation was also recognized between the MYOACT score (P = 0.004, r = 0.360) and the bilateral lung SUVmean. Nevertheless, no correlation was identified between the bilateral lung SUVmean and the time gap after the onset of respiratory symptoms (P = 0.395, r = -0.111) (Additional file 5).

To develop a scoring system for the prediction of RP-ILD, we rounded up the cut-off values of bilateral SUVmean and DLCO% to > 0.450 and < 50.0%, respectively. Each was allotted one point. The identification of abnormal mediastinal lymph nodes was also allotted one point. Each included patient was then assigned a cumulative score (maximum score of three, minimum score of zero). The scoring system was named "DLM" using the initials of DLCO%, lung and mediastinum. Then, the predictive value of the DLM model was evaluated in 61 patients. None of the 15 patients with the minimum possible score of zero developed RP-ILD. In contrast, all nine patients with a maximum possible score of three developed RP-ILD (Table 2, Fig. 3A). In the ROC curve analysis (Fig. 3B), we found a cut-off value of ≥ 2 , AUC of 0.905, sensitivity of 85.7%, specificity of 82.5%, positive predictive value (PPV) of 72.0%, negative predictive value (NPV) of 91.7% and accuracy of 83.6%. Furthermore, the predictive value of the DLM model was verified in univariate and multivariate logistic regression analyses for RP-ILD in IIM-ILD patients. After incorporating the DLM score into the statistical model, the DLM score was significantly correlated with RP-ILD with P < 0.001, and all of the other factors were excluded from the model (Additional file 6).

To explore the predictive value of PET/CT scans for survival, a univariate Cox proportional hazards regression analysis identified pulmonary bacterial infection (P < 0.001), RP-ILD (P = 0.008), MYOACT score (P <0.001), DLCO% (P = 0.016), bilateral lung SUVmean (P= 0.007), spleen SUVmean (P = 0.008) and the use of steroid+IVIG (P = 0.015) as significantly correlated with survival during follow-up (Additional file 7). The subsequent multivariate Cox proportional hazards regression analysis identified pulmonary bacterial infection (P =0.013) and MYOACT score (P < 0.001) as factors significantly related to the survival of IIM-ILD patients (Additional file 8). Furthermore, IIM-ILD patients who died within 3 months were found to have a higher bilateral lung SUVmean (P = 0.019, Fig. 4A) and a higher spleen SUVmean (P = 0.011, Fig. 4B). In addition, a moderate correlation was recognized between spleen SUVmean and bilateral lung SUVmean (P = 0.006, r = 0.346, Fig. 4C). Nevertheless, in the RP-ILD subgroups, bilateral lung SUVmean (P = 0.598, Additional file 9) and spleen

(2021) 23:198 Page 5 of 12

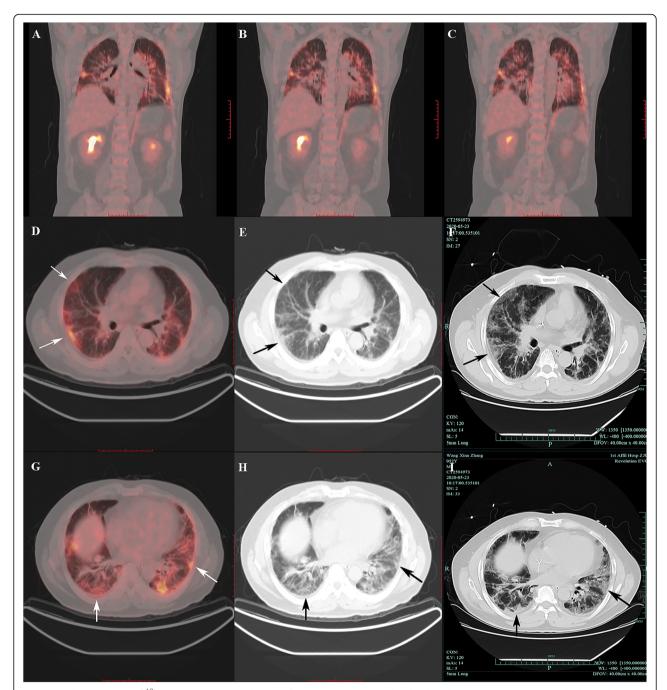


Fig. 1 Visual examination of ¹⁸F-FDG-PET/CT scan and HRCT of one IIM-ILD patients who developed RP-ILD. **A** to **C**. Prominently elevated FDG uptake in bilateral lungs of one IIM-ILD patient who later developed RP-ILD. **D** to **F** and **G** to **I**. Prominently elevated FDG uptake in bilateral lungs of one IIM-ILD patient indicated future RP-ILD in one week (where the arrow pointed). FDG: Fluorodeoxyglucose; HRCT: High resolution CT; IIM-ILD: Idiopathic-inflammatory-myopathy-related interstitial lung disease; RP-ILD: Rapidly progressive interstitial lung disease

SUVmean (P = 0.161, Additional file 9) were not recognized to be significantly different between patients who died within 3 months or survived beyond this threshold.

Discussion

Liang et al. Arthritis Research & Therapy

Apart from its uses in oncologic imaging, ¹⁸F-FDG PET/CT has also been found to reflect immune cell activation

in the spleen, malignant tissues and lungs [25, 26]. In a bleomycin-induced pulmonary fibrosis mouse model, the early stage of FDG uptake increase was probably related to the early recruitment and activation of leukocytes, while the later and persistent elevation of FDG uptake might be associated with aerobic glycolysis of myofibroblasts [12, 27, 28]. Furthermore, elevated

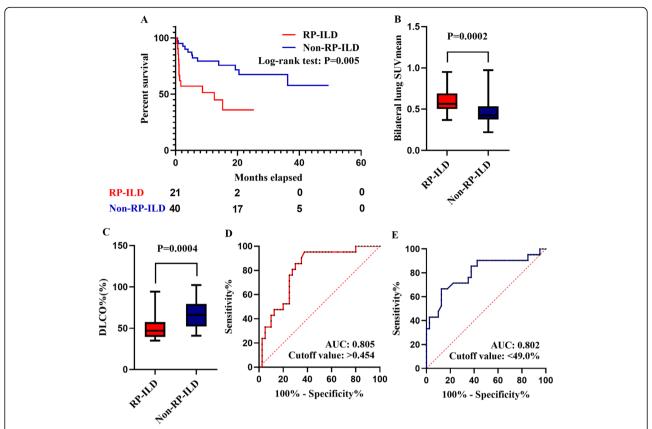


Fig. 2 Evaluation of IIM-ILD patients with or without RP-ILD. A. Survival of IIM-ILD patients with or without RP-ILD. B. Comparison of bilateral lung SUVmean in RP-ILD and non-RP-ILD groups. C. Comparison of DLCO% in RP-ILD and non-RP-ILD groups. D. ROC curve of bilateral lung SUVmean predicting RP-ILD. E. ROC curve of DLCO% predicting RP-ILD. IIM-ILD: Idiopathic-inflammatory-myopathy-related interstitial lung disease; RP-ILD: Rapidly progressive interstitial lung disease; SUVmean: mean standard uptake value; DLCO%: Percent-predicted diffusing capacity of the lung for carbon monoxide; ROC: Receiver operating characteristic; AUC: Area under the curve

pulmonary FDG uptake was also found to be associated with macrophage and neutrophil activation in acute or chronic lung diseases [29, 30]. Elevated ¹⁸F-FDG uptake could therefore serve as a marker for focal and global inflammatory activation and elevated metabolism in bilateral lungs. In addition, two antifibrotic drugs, namely, nintedanib and pirfenidone, were found to significantly decrease pulmonary FDG uptake in bleomycin-treated mice, indicating the value of PET/CT scans in evaluating the response to antifibrotic medications [31, 32].

However, the role of PET/CT scans requires further validation in IPF and CTD-ILD patients. To the best of our knowledge, this is the largest cohort to investigate the clinical value of pulmonary FDG uptake in IIM-ILD patients and the first study to systemically explore FDG uptake in extra-pulmonary organs (liver, spleen, digestive tract, etc.) in IIM-ILD patients.

After initially confirming the role of FDG uptake in reflecting focal immune and metabolic activation, we used a retrospective cohort to identify the clinical value of PET/CT scans for RP-ILD in IIM-ILD patients. Both increased pulmonary FDG uptake and abnormal

mediastinal lymph nodes were significantly correlated with RP-ILD in IIM-ILD patients. Compared with conventional HRCT, PET/CT scans seemed to be more intuitive for predicting RP-ILD. In IPF patients, PET/CT scans have been widely studied and found to be valuable for the evaluation of disease activity and therapy efficacy as well as the prediction of disease progression and survival [31, 33, 34]. In patients with lung cancer, elevated pulmonary FDG uptake is associated with acute exacerbation of ILD after chemotherapy or surgery [35–37]. In Ssc-ILD patients, pulmonary FDG uptake is correlated with ILD severity [13]. Together with abnormal mediastinal/hilar lymph nodes, it is associated with the progression of ILD within 2 years [13]. The predictive value of pulmonary FDG uptake for ILD progression crossed the boundary of IPF, tumour-associated ILD and CTD-ILD and needs to be verified with a broader ILD spectra and larger cohorts. In addition, in our study, elevated pulmonary FDG uptake was associated with pulmonary bacterial/fungal infection and elevated IIM disease activity. These findings reflect the focal immune abnormality of both IIM and an infectious background, consistent

Table 1 Comparisons of multiple factors between RP-ILD and non-RP-ILD groups

Factors	RP-ILD (21)	Non-RP-ILD (40)	P value	P-adjusted
Age(y)	54.86 ± 11.72	57.70 ± 11.06	0.354	0.824
Sex (male/female)	8/13	17/23	0.740	1.000
Course of disease(m)	2.00 (1.00, 2.75)	2.25 (1.00, 5.00)	0.253	0.748
Duration of diagnosis delay(m)	1.50 (0.50, 2.00)	1.00 (1.00, 3.00)	0.274	0.756
Clinical manifestations or complications				
Pulmonary bacterial infection	7 (33.3%)	4 (10.0%)	0.036	0.347
Pulmonary fungal infection	5 (23.8%)	4 (10.0%)	0.253	0.748
Tuberculosis infection	0 (0.0%)	0 (0.0%)	NA	NA
EBV infection	6 (28.6%)	5 (12.5%)	0.164	0.605
CMV infection	2 (9.5%)	0 (0.0%)	0.115	0.559
Carcinoma	1 (4.8%)	6 (15.0%)	0.405	0.824
Gastrointestinal haemorrhage	4 (19.0%)	1 (2.5%)	0.044	0.347
UIP pattern	3 (14.3%)	4 (10.0%)	0.683	0.962
Pneumomediastinum	1 (4.8%)	1 (2.5%)	1.000	1.000
Laboratory finding				
Ferritin (ng/ml)	612.50 (302.05, 2110.25)	664.05 (314.40, 2061.23)	0.495	0.863
ESR (mm/h)	16.00 (6.50, 32.00)	17.50 (9.50, 42.00)	0.466	0.857
CRP (mg/L)	5.40 (3.10, 19.80)	6.35 (3.00, 36.73)	0.660	0.962
ALT(U/L)	37.00 (22.50, 99.50)	91.50 (33.75, 212.50)	0.051	0.347
AST(U/L)	45.00 (33.00, 97.00)	54.50 (34.00, 235.50)	0.224	0.725
LDH(U/L)	329.00 (275.50, 478.50)	323.00 (247.75, 461.50)	0.897	1.000
CK(U/L)	84.00 (50.50, 255.00)	86.00 (44.25, 415.25)	0.838	1.000
Disease activity				
MYOACT score	13.00 (9.00, 15.50)	9.00 (7.00, 10.75)	0.006	0.102
Lung function testing				
FVC% (%)	65.33 ± 20.08	71.86 ± 15.61	0.165	0.605
FEV1% (%)	64.85 ± 19.40	74.42 ± 18.25	0.062	0.383
FEV1/FVC	0.81 (0.75, 0.85)	0.81 (0.77,0.85)	0.849	1.000
TLC(L)	3.43 ± 1.01	3.98 ± 1.02	0.049	0.347
DLCO% (%)	50.29 ± 15.58	66.92 ± 16.67	< 0.001	0.014
¹⁸ F-FDG PET/CT scan findings				
Time gap after RS onset*(d)	20.00 (12.50, 25.00)	16.50 (13.00, 24.75)	0.767	1.000
Bilateral lung SUVmean	0.60 ± 0.15	0.45 ± 0.14	< 0.001	0.014
Abnormal mediastinal lymph node	14 (66.7%)	10 (25.0%)	0.002	0.045
Abnormal hilar lymph node	11 (52.4%)	9 (22.5%)	0.018	0.204
Liver SUVmean	1.83 ± 0.34	1.73 ± 0.47	0.412	0.824
Spleen SUVmean	2.02 ± 0.50	1.82 ± 0.46	0.126	0.571
Bone marrow SUVmean	1.72 ± 0.50	1.62 ± 0.44	0.402	0.824
Cardiac SUVmean	1.38 (1.00, 2.03)	1.60 (1.12, 2.39)	0.379	0.824
Oesophagus SUVmean	1.51 (1.19, 1.83)	1.34 (1.07, 1.74)	0.213	0.724
Stomach SUVmean	0.69 (0.49, 1.00)	0.71 (0.57, 0.83)	0.927	1.000
Small intestine SUVmean	1.05 ± 0.33	1.03 ± 0.32	0.878	1.000
Colon and rectum SUVmean	1.07 (0.80, 1.32)	1.02 (0.83, 1.20)	0.693	0.962
Bilateral cerebellum SUVmean	5.17 ± 1.26	5.24 ± 1.61		· · · ·

Table 1 Comparisons of multiple factors between RP-ILD and non-RP-ILD groups (Continued)

Factors	RP-ILD (21)	Non-RP-ILD (40)	P value	P-adjusted
Bilateral trapezius SUVmean	0.82 (0.70, 0.96)	0.79 (0.68, 1.00)	0.585	0.944
Bilateral deltoid SUVmean	0.71 (0.58, 1.27)	0.78 (0.61, 1.09)	0.590	0.944
Bilateral biceps SUVmean	0.75 (0.67, 1.01)	0.77 (0.60, 1.10)	0.879	1.000
Bilateral iliopsoas SUVmean	1.10 ± 0.36	1.12 ± 0.36	0.850	1.000
Bilateral gluteus maximus SUVmean	0.85 ± 0.27	0.79 ± 0.32	0.462	0.857
Bilateral gluteus medius SUVmean	0.92 ± 0.29	0.98 ± 0.29	0.473	0.857
Bilateral quadriceps SUVmean	0.78 (0.62, 0.96)	0.82 (0.69, 1.05)	0.370	0.824
Myositis-specific antibodies and myositis-a	associated antibodies			
Anti-MDA5	13 (61.9%)	12 (30.0%)	0.016	0.204
Anti-PL-7	4 (19.0%)	2 (5.0%)	0.169	0.605
Anti-PL-12	1 (4.8%)	1 (2.5%)	1.000	1.000
Anti-EJ	0 (0.0%)	1 (2.5%)	1.000	1.000
Anti-OJ	0 (0.0%)	1 (2.5%)	1.000	1.000
Anti-Jo-1	2 (9.5%)	3 (7.5%)	1.000	1.000
Anti-TIF1γ	1 (4.8%)	3 (7.5%)	1.000	1.000
Anti-Mi-2α	0 (0.0%)	2 (5.0%)	0.541	0.920
Anti-Mi-2β	0 (0.0%)	4 (10.0%)	0.289	0.756
Anti-SAE1	0 (0.0%)	5 (12.5%)	0.154	0.605
Anti-NXP2	1 (4.8%)	6 (15.0%)	0.405	0.824
Anti-SRP	1 (4.8%)	2 (5.0%)	1.000	1.000
Anti-Ku	1 (4.8%)	1 (2.5%)	1.000	1.000
Anti-PM-Scl75	1 (4.8%)	2 (5.0%)	1.000	1.000
Anti-PM-Scl100	0 (0.0%)	0 (0.0%)	NA	NA
Anti-Ro-52	14 (66.7%)	17 (42.5%)	0.073	0.414
Therapies				
Steroid monotherapy	6 (28.6%)	14 (35.0%)	0.611	0.944
Steroid + DMARDs	7 (33.3%)	19 (47.5%)	0.288	0.756
Steroid+IVIG	2 (9.5%)	2 (5.0%)	0.602	0.944
Steroid+DMARDs+IVIG	3 (14.3%)	4 (10.0%)	0.683	0.962
Steroid + JAK inhibitor	3 (14.3%)	1 (2.5%)	0.113	0.559
IIM subtypes				
DM	13 (61.9%)	27 (67.5%)	0.662	0.962
PM	2 (9.5%)	7 (17.5%)	0.479	0.857
ADM	6 (28.6%)	6 (15.0%)	0.309	0.778

RP-ILD, rapidly progressive interstitial lung disease; *P*-adjusted, adjusted *P* value after false discovery rate correction; y, years; m, months; NA, not available; EBV, Epstein-Barr virus; CMV, cytomegalovirus; UIP pattern, usual interstitial pneumonia pattern; ESR, erythrocyte sedimentation rate; CRP, C-reactive protein; ALT, alanine transaminase; AST, aspartate transaminase; LDH, lactate dehydrogenase; CK, creatine kinase; MYOACT, Myositis Disease Activity Assessment Visual Analogue Scales; FVC%, percent-predicted forced vital capacity; FEV1%, percent-predicted forced expiratory volume in 1 s; FEV1/FVC, ratio of FEV1 over FVC; TLC, total lung capacity; DLCO%, percent-predicted diffusing capacity of the lung for carbon monoxide; RS, respiratory symptoms; d, days; FDG, fluorodeoxyglucose; SUVmean, mean standard uptake value; DMARDs, disease-modifying anti-rheumatic drugs; IVIG, intravenous immunoglobulin; JAK, Janus kinase; IIM, idiopathic inflammatory myopathy; DM, dermatomyositis; PM, polymyositis; ADM, amyopathic dermatomyositis

with previous reports that CTD-related immune alterations and infectious triggers played important roles in the progressive phenotype of CTD-ILD [38, 39].

Before FDR correction, pulmonary and splenic FDG uptake was found to be correlated with the survival of IIM-ILD patients. The outcome-predicting value of

^{*}Time gap after RS onset referred to the time gap between onset of respiratory symptoms (evident feelings of chest distress and shortness of breath) and PET/CT scan

Table 2 Distribution of RP-ILD and non-RP-ILD in IIM-ILD patients with different DLM score

DLM score	Total patients (N)	RP-ILD % (N)	Non-RP-ILD % (N)
0	15	0.0% (0)	100.0% (15)
1	21	14.3% (3)	85.7% (18)
2	16	56.3% (9)	43.7% (7)
3	9	100.0% (9)	0.0% (0)

RP-ILD, rapidly progressive interstitial lung disease; IIM-ILD, idiopathic-inflammatory-myopathy-related interstitial lung disease; N, number

pulmonary FDG uptake has been previously proposed for IPF [40]. Fraioli and his colleagues proposed that including pulmonary FDG uptake in the conventional GAP (gender, age and physiology) model would significantly increase the model's accuracy for outcome prediction [41]. Through Pearson linear analysis, we also found a moderate correlation between the spleen SUVmean and bilateral lung SUVmean. As the largest lymphoid organ containing specific subsets of myeloid cells and lymphocytes, the spleen is involved in CTD development [42]. We hypothesized that there might be a link between the spleen and lung during the development of CTD-ILD. In traditional Chinese medicine, the spleen is the organ that produces and replenishes Qi, which tonifies and nourishes the lung [43]. It will be interesting and meaningful to identify cross-talk between the spleen and lung from the perspective of modern medicine. However, only higher disease activity and complications of pulmonary infection remained statistically significant after FDR correction and multivariate analysis, which was consistent with the findings of previous studies [44, 45].

RP-ILD is a severe and fatal complication of IIM patients, and it is thus necessary to search for early prediction tools to enable the application of timely and potent therapeutic regimens for patients at high risk of RP-ILD. Previous studies indicated that anti-MDA5 antibody,

anti-Ro52 antibody, DLCO%, serum ferritin, etc., were correlated with the development of RP-ILD [46, 47]. After FDR correction and multivariate logistic regression analysis, bilateral lung SUVmean, abnormal mediastinal lymph node and DLCO% were significantly different between patients with RP-ILD and the control group. The three clinical factors were tested independently and were then incorporated to form the DLM model. The DLM model had satisfying AUC, sensitivity, specificity, PPV, NPV and accuracy. However, due to the scarcity of IIM-ILD patients receiving PET/CT scans, we could not construct another cohort of IIM-ILD patients to validate the DLM score.

There were multiple limitations of this study. In this small-sample study, selection bias occurred since patients with fever, signs of cytopenia, higher disease activity and dyspnoea tended to receive PET/CT scans to exclude malignancy or a relapse of malignancy. The proportion of anti-MDA5 antibody-positive patients was thus higher than that reported previously, which might impede the identification of the role of anti-MDA5 antibody in RP-ILD and survival. Due to its retrospective nature, the small sample size and the high heterogeneity of the disease profile, we could not study additional clinical factors, such as Kreb Von Den Lungen-6 and the HRCT score. The lack of timely follow-up lung function testing in multiple patients made it impossible to incorporate deterioration of lung function into the criteria of RP-ILD. The scarcity of IIM patients receiving PET/CT scans made it impossible to verify the predictive value of the DLM model in a validation cohort. Healthy people or IIM patients without ILD seldom receive PET/CT scans, making it impossible to construct a healthy control group or select IIM patients without ILD as controls. To make the prediction model easier to construct, the bilateral lung SUVmean was calculated at one site with the highest FDG uptake instead of multiple sites.

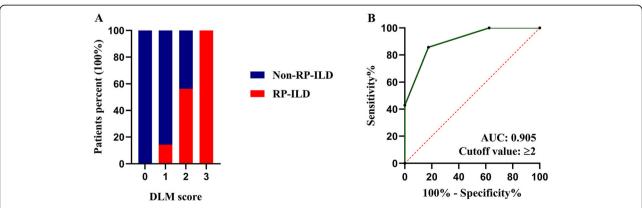


Fig. 3 Evaluation of DLM model in predicting RP-ILD in IIM-ILD patients. A. Distribution of RP-ILD and non-RP-ILD patients in each DLM score group. B. ROC curve of DLM model predicting RP-ILD. RP-ILD: Rapidly progressive interstitial lung disease; IIM-ILD: Idiopathic-inflammatory-myopathy-related interstitial lung disease; ROC: Receiver operating characteristic; AUC: Area under the curve

Liang et al. Arthritis Research & Therapy (2021) 23:198 Page 10 of 12

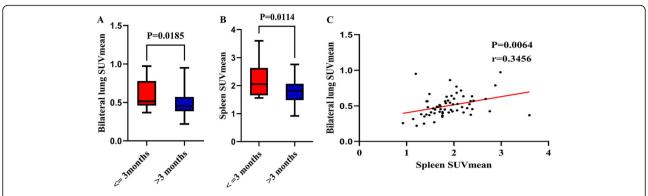


Fig. 4 Evaluation of abnormal FDG uptake in IIM-ILD patients with different survival. **A.** Comparison of bilateral lung SUVmean in IIM-ILD patients who died within three months or survived beyond this threshold. **B.** Comparison of spleen SUVmean in IIM-ILD patients who died within three months or survived beyond three months. **C.** Correlation between bilateral lung SUVmean and spleen SUVmean in IIM-ILD patients. FDG: Fluorodeoxyglucose; IIM-ILD: Idiopathic-inflammatory-myopathy-related interstitial lung disease; SUVmean: mean standard uptake value

Despite all of these limitations, we were able to identify the clinical value of PET/CT scans in IIM-ILD and constructed a scoring system for the prediction of RP-ILD, and our findings suggest future research approaches for IIM-ILD.

Conclusions

Elevated pulmonary FDG uptake, abnormal mediastinal lymph nodes and decreased DLCO% were significantly correlated with the development of RP-ILD in IIM-ILD patients. A DLM model was constructed and initially proved efficient in predicting RP-ILD in these patients. Higher disease activity and complications of bacterial infection were significantly related to unfavourable outcomes. In addition, IIM-ILD patients with higher pulmonary and splenic FDG uptake (which were correlated) showed a tendency to suffer from unfavourable outcomes, indicating a systemic hyperinflammatory status and a possible linkage between the two organs.

Abbreviations

ILD: Interstitial lung disease; NSIP: Non-specific interstitial pneumonia; IIM-ILD: Idiopathic-inflammatory-myopathy-related ILD; RP: Rapidly progressive; IPF: Idiopathic pulmonary fibrosis; CTD-ILD: Connective tissue disease-related ILD; COVID-19: Coronavirus disease 2019; FDG: Fluorodeoxyglucose; SSc-ILD: Systemic sclerosis-related ILD; SUV: Standard uptake value; ROI: Region of interest; FAHZJU: The First Affiliated Hospital, Zhejiang University School of Medicine; IRB: Institutional Review Board; EMR: Electronic medical record; ICD-10: International Classification of Diseases, tenth version; DM: Dermatomyositis; PM: Polymyositis; ADM: Amyopathic dermatomyositis; HRCT: High-resolution computed tomography; UIP: Usual interstitial pneumonia; CTD: Connective tissue disease; MYOACT: Myositis Disease Activity Assessment Visual Analogue Scales; PSL: Prednisolone; mPSL: Methylprednisolone; DMARDs: Disease modifying anti-rheumatic drugs; JAK: Janus kinase; IVIG: Intravenous immunoglobulin; FDR: False discovery rate; ROC: Receiver operating characteristic; TLC: Total lung capacity; DLCO%: Percent-predicted diffusing capacity of the lung for carbon monoxide; AUC: Area under the curve; PPV: Positive predictive value; NPV: Negative predictive value; GAP: Gender, age and physiology

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s13075-021-02578-9.

Additional file 1. Enrollment and groupings of IIM-ILD patients

Additional file 2. Univariate logistic regression analyses of RP-ILD in IIM-ILD patients

Additional file 3. Multivariate logistic regression analysis of RP-ILD in IIM-ILD patients

Additional file 4. Correlation of DLCO%, bilateral lung SUVmean and AML

Additional file 5. Correlation of bilateral lung SUVmean, infection, MYOACT score and time gap after RS onset

Additional file 6. Multivariate logistic regression analysis of RP-ILD after inclusion of DLM score

Additional file 7. Univariate Cox proportional hazards regression analyses of survival in IIM-ILD patients

Additional file 8. Multivariate Cox proportional hazards regression analysis of survival in IIM-ILD patients

Additional file 9. Comparisons of bilateral lung SUVmean and spleen SUVmean in RP-ILD patients with different survival

Additional file 10. Dataset supporting the conclusions of this article

Acknowledgements

Not applicable.

Authors' contributions

Study design: JLiang, HC and JLin. Data collection: JLiang, YS, YK and YH. Verification of IIM diagnosis: HC and BX. Reevaluation of FDG uptake in multiple organs: YL. Identification of ILD and RP-ILD: YL and BY. Statistical analysis: JLiang. Writing: JLiang and HC. Proof reading: JLiang, HC and JLin. All authors read and approved the final manuscript.

Funding

This study was supported by the grants from National Natural Science Foundation of China (81701602).

Availability of data and materials

The dataset supporting the conclusions of this article is presented as Additional file 10 (seen in additional files of this article).

Declarations

Ethics approval and consent to participate

The study was approved (2021-194) by the IRB of FAHZJU. All participants signed written informed consent at admission.

Consent for publication

Consents for publication from the patients included were also acquired prior to submission

Competing interests

The authors declare that they have no competing interests.

Author details

¹Department of Rheumatology, The First Affiliated Hospital, Zhejiang University School of Medicine, #79 Qingchun Road, Hangzhou 310003, Zhejiang Province, People's Republic of China. ²PET Center, The First Affiliated Hospital, Zhejiang University School of Medicine, #79 Qingchun Road, Hangzhou 310003, Zhejiang Province, People's Republic of China. ³Department of Respiratory Diseases, The First Affiliated Hospital, Zhejiang University School of Medicine, #79 Qingchun Road, Hangzhou 310003, Zhejiang Province, People's Republic of China.

Received: 22 April 2021 Accepted: 9 July 2021 Published online: 23 July 2021

References

- Tsuji H, Nakashima R, Hosono Y, Imura Y, Yagita M, Yoshifuji H, et al. Multicenter prospective study of the efficacy and safety of combined immunosuppressive therapy with high-dose glucocorticoid, tacrolimus, and cyclophosphamide in interstitial lung diseases accompanied by antimelanoma differentiation-associated gene 5-positive dermatomyositis. Arthritis Rheumatol. 2020;72(3):488–98. https://doi.org/10.1002/art.41105.
- Vojinovic T, Cavazzana I, Ceruti P, Fredi M, Modina D, Berlendis M, et al. Predictive features and clinical presentation of interstitial lung disease in inflammatory myositis. Clin Rev Allergy Immunol. 2021;60(1):87–94. https://doi.org/10.1007/s12016-020-08814-5.
- Sugiyama Y, Yoshimi R, Tamura M, Takeno M, Kunishita Y, Kishimoto D, et al. The predictive prognostic factors for polymyositis/dermatomyositisassociated interstitial lung disease. Arthritis Res Ther. 2018;20(1):7. https://doi. org/10.1186/s13075-017-1506-7.
- Jablonski R, Bhorade S, Strek ME, Dematte J. Recognition and management of myositis-associated rapidly progressive interstitial lung disease. *Chest*. 2020;158(1):252–63. https://doi.org/10.1016/j.chest.2020.01.033.
- Gono T, Masui K, Nishina N, Kawaguchi Y, Kawakami A, Ikeda K, et al. Risk prediction modeling based on a combination of initial serum biomarker levels in polymyositis/dermatomyositis-associated interstitial lung disease. Arthritis Rheumatol. 2021;73(4):677–86. https://doi.org/10.1002/art.41566.
- Li S, Sun Y, Shao C, Huang H, Wang Q, Xu K, et al. Prognosis of adult idiopathic inflammatory myopathy-associated interstitial lung disease: a retrospective study of 679 adult cases. *Rheumatology (Oxford)*. 2021;60(3): 1195–204. https://doi.org/10.1093/rheumatology/keaa372.
- Barba T, Fort R, Cottin V, Provencher S, Durieu I, Jardel S, et al. Treatment of idiopathic inflammatory myositis associated interstitial lung disease: a systematic review and meta-analysis. *Autoimmun Rev.* 2019;18(2):113–22. https://doi.org/10.1016/j.autrev.2018.07.013.
- Heukels P, Moor CC, von der Thusen JH, Wijsenbeek MS, Kool M. Inflammation and immunity in IPF pathogenesis and treatment. Respir Med. 2019;147:79–91. https://doi.org/10.1016/j.rmed.2018.12.015.
- Lescoat A, Lelong M, Jeljeli M, Piquet-Pellorce C, Morzadec C, Ballerie A, et al. Combined anti-fibrotic and anti-inflammatory properties of JAKinhibitors on macrophages in vitro and in vivo: perspectives for scleroderma-associated interstitial lung disease. *Biochem Pharmacol*. 2020; 178:114103. https://doi.org/10.1016/j.bcp.2020.114103.
- Enomoto Y, Suzuki Y, Hozumi H, Mori K, Kono M, Karayama M, et al. Clinical significance of soluble CD163 in polymyositis-related or dermatomyositisrelated interstitial lung disease. *Arthritis Res Ther.* 2017;19(1):9. https://doi. org/10.1186/s13075-016-1214-8.
- Rendeiro AF, Ravichandran H, Bram Y, Chandar V, Kim J, Meydan C, et al. The spatial landscape of lung pathology during COVID-19 progression. Nature. 2021;593(7860):564–9. https://doi.org/10.1038/s41586-021-03475-6.

- Peelen DM, Zwezerijnen B, Nossent EJ, Meijboom LJ, Hoekstra OS, Van der Laken CJ, et al. The quantitative assessment of interstitial lung disease with positron emission tomography scanning in systemic sclerosis patients. *Rheumatology (Oxford)*. 2020;59(6):1407–15. https://doi.org/10.1093/rheumatology/kez483.
- Ledoult E, Morelle M, Soussan M, Mekinian A, Behal H, Sobanski V, et al. (18)F-FDG positron emission tomography scanning in systemic sclerosisassociated interstitial lung disease: a pilot study. *Arthritis Res Ther.* 2021;23(1): 76. https://doi.org/10.1186/s13075-021-02460-8.
- Morita Y, Kuwagata S, Kato N, Tsujimura Y, Mizutani H, Suehiro M, et al. 18F-FDG PET/CT useful for the early detection of rapidly progressive fatal interstitial lung disease in dermatomyositis. *Intern Med.* 2012;51(12):1613–8. https://doi.org/10.2169/internalmedicine.51.6813.
- Li Y, Zhou Y, Wang Q. Multiple values of (18)F-FDG PET/CT in idiopathic inflammatory myopathy. Clin Rheumatol. 2017;36(10):2297–305. https://doi. org/10.1007/s10067-017-3794-3.
- Motegi SI, Fujiwara C, Sekiguchi A, Hara K, Yamaguchi K, Maeno T, et al. Clinical value of (18) F-fluorodeoxyglucose positron emission tomography/ computed tomography for interstitial lung disease and myositis in patients with dermatomyositis. *J Dermatol.* 2019;46(3):213–8. https://doi.org/1 0.1111/1346-8138.14758.
- Lundberg IE, Tjarnlund A, Bottai M, Werth VP, Pilkington C, Visser M, et al. 2017 European League Against Rheumatism/American College of Rheumatology classification criteria for adult and juvenile idiopathic inflammatory myopathies and their major subgroups. *Ann Rheum Dis*. 2017; 76(12):1955–64. https://doi.org/10.1136/annrheumdis-2017-211468.
- Liang J, Xu D, Sun C, Chen W, Cao H, Lin J. Hemophagocytic lymphohistiocytosis: prevalence, risk factors, outcome, and outcome-related factors in adult idiopathic inflammatory myopathies. *J Rheumatol.* 2020; 47(10):1532–40. https://doi.org/10.3899/jrheum.190542.
- Izuka S, Yamashita H, Iba A, Takahashi Y, Kaneko H. Acute exacerbation of rheumatoid arthritis-associated interstitial lung disease: clinical features and prognosis. *Rheumatology (Oxford)*. 2020;60(5):2348–54. https://doi.org/10.1 093/rheumatology/keaa608.
- Abe Y, Matsushita M, Tada K, Yamaji K, Takasaki Y, Tamura N. Clinical characteristics and change in the antibody titres of patients with anti-MDA5 antibody-positive inflammatory myositis. *Rheumatology (Oxford)*. 2017;56(9): 1492–7. https://doi.org/10.1093/rheumatology/kex188.
- Horiike Y, Suzuki Y, Fujisawa T, Yasui H, Karayama M, Hozumi H, et al. Successful classification of macrophage-mannose receptor CD206 in severity of anti-MDA5 antibody positive dermatomyositis associated ILD. Rheumatology (Oxford). 2019;58(12):2143–52. https://doi.org/10.1093/rheumatology/kez185.
- 22. Hozumi H, Fujisawa T, Nakashima R, Johkoh T, Sumikawa H, Murakami A, et al. Comprehensive assessment of myositis-specific autoantibodies in polymyositis/dermatomyositis-associated interstitial lung disease. *Respir Med.* 2016;121:91–9. https://doi.org/10.1016/j.rmed.2016.10.019.
- Kim J, Yoo SW, Kang SR, Bom HS, Song HC, Min JJ. Clinical implication of F-18 FDG PET/CT in patients with secondary hemophagocytic lymphohistiocytosis. *Ann Hematol.* 2014;93(4):661–7. https://doi.org/10.1007/s00277-013-1906-y.
- Tanaka S, Ikeda K, Uchiyama K, Iwamoto T, Sanayama Y, Okubo A, et al. [18F]FDG uptake in proximal muscles assessed by PET/CT reflects both global and local muscular inflammation and provides useful information in the management of patients with polymyositis/dermatomyositis. *Rheumatology* (Oxford). 2013;52(7):1271–8. https://doi.org/10.1093/rheumatology/ket112.
- Bronte V, Pittet MJ. The spleen in local and systemic regulation of immunity. *Immunity*. 2013;39(5):806–18. https://doi.org/10.1016/j.immuni.2013.10.010.
- Tomita M, Suzuki M, Kono Y, Nakajima K, Matsuda T, Kuge Y, et al. Influence on [(18)F]FDG uptake by cancer cells after anti-PD-1 therapy in an enforcedimmune activated mouse tumor. *EJNMMI Res.* 2020;10(1):24. https://doi. org/10.1186/s13550-020-0608-4.
- Bondue B, Sherer F, Van Simaeys G, Doumont G, Egrise D, Yakoub Y, et al. PET/CT with 18F-FDG- and 18F-FBEM-labeled leukocytes for metabolic activity and leukocyte recruitment monitoring in a mouse model of pulmonary fibrosis. J Nucl Med. 2015;56(1):127–32. https://doi.org/10.2967/ jnumed.114.147421.
- Xie N, Tan Z, Banerjee S, Cui H, Ge J, Liu RM, et al. Glycolytic reprogramming in myofibroblast differentiation and lung fibrosis. Am J Respir Crit Care Med. 2015;192(12):1462–74. https://doi.org/10.1164/rccm.201 504-0780OC.

- Savelli G, Bonacina M, Rizzo A, Zaniboni A. Activated macrophages are the main inflammatory cell in COVID-19 interstitial pneumonia infiltrates. Is it possible to show their metabolic activity and thus the grade of inflammatory burden with (18)F-Fluorocholine PET/CT? Med Hypotheses. 2020;144:109885. https://doi.org/10.1016/j.mehy.2020.109885.
- Chen DL, Schiebler ML, Goo JM, van Beek EJR. PET imaging approaches for inflammatory lung diseases: current concepts and future directions. Eur J Radiol. 2017;86:371–6. https://doi.org/10.1016/j.ejrad.2016.09.014.
- Tanguy J, Goirand F, Bouchard A, Frenay J, Moreau M, Mothes C, et al.
 [(18)F]FMISO PET/CT imaging of hypoxia as a non-invasive biomarker of
 disease progression and therapy efficacy in a preclinical model of
 pulmonary fibrosis: comparison with the [(18)F]FDG PET/CT approach. Eur J
 Nucl Med Mol Imaging. 2021. https://doi.org/10.1007/s00259-021-05209-2.
- Bondue B, Castiaux A, Van Simaeys G, Mathey C, Sherer F, Egrise D, et al. Absence of early metabolic response assessed by 18F-FDG PET/CT after initiation of antifibrotic drugs in IPF patients. *Respir Res.* 2019;20(1):10. https://doi.org/10.1186/s12931-019-0974-5.
- Justet A, Laurent-Bellue A, Thabut G, Dieudonne A, Debray MP, Borie R, et al. [(18)F]FDG PET/CT predicts progression-free survival in patients with idiopathic pulmonary fibrosis. *Respir Res.* 2017;18(1):74. https://doi.org/10.11 86/s12931-017-0556-3.
- 34. Lee EY, Wong CS, Fung SL, Yan PK, Ho JC. SUV as an adjunct in evaluating disease activity in idiopathic pulmonary fibrosis a pilot study. *Nucl Med Commun.* 2014;35(6):631–7. https://doi.org/10.1097/MNM. 0000000000000083.
- Akaike K, Saruwatari K, Oda S, Shiraishi S, Takahashi H, Hamada S, et al. Predictive value of (18)F-FDG PET/CT for acute exacerbation of interstitial lung disease in patients with lung cancer and interstitial lung disease treated with chemotherapy. *Int J Clin Oncol*. 2020;25(4):681–90. https://doi. org/10.1007/s10147-019-01584-x.
- Fukunaga K, Nagatani Y, Nakagawa H, Nitta-Seko A, Nagata T, Nishizono M, et al. Increased (18)F-FDG accumulation in less-affected lung area in patients with non-small cell lung cancer and postoperative acute exacerbation of interstitial lung disease. Eur J Radiol. 2021;135:109477. https://doi.org/10.1016/j.ejrad.2020.109477.
- Yamamichi T, Shimada Y, Masuno R, Ohira T, Abe S, Yoshimura A, et al. Association between F-18 fluorodeoxyglucose uptake of noncancerous lung area and acute exacerbation of interstitial pneumonia in patients with lung cancer after resection. *J Thorac Cardiovasc Surg.* 2020;159(3):1111–8 e1112. https://doi.org/10.1016/j.jtcvs.2019.07.100.
- Spagnolo P, Distler O, Ryerson CJ, Tzouvelekis A, Lee JS, Bonella F, et al. Mechanisms of progressive fibrosis in connective tissue disease (CTD)associated interstitial lung diseases (ILDs). Ann Rheum Dis. 2021;80(2):143–50. https://doi.org/10.1136/annrheumdis-2020-217230.
- Ricci A, Pagliuca A, Vermi M, Pizzirusso D, Innammorato M, Sglavo R, Scarso F, Salemi S, Lagana B, Di Rosa R et al: The role of lung colonization in connective tissue disease-associated interstitial lung disease. *Microorganisms* 2021, 9(5). doi: https://doi.org/10.3390/microorganisms9050932, The Role of Lung Colonization in Connective Tissue Disease-Associated Interstitial Lung Disease, 9, 5
- Umeda Y, Demura Y, Morikawa M, Anzai M, Kadowaki M, Ameshima S, et al. Prognostic value of dual-time-point 18F-FDG PET for idiopathic pulmonary fibrosis. J Nucl Med. 2015;56(12):1869–75. https://doi.org/10.2967/jnumed.11 5.163360.
- Fraioli F, Lyasheva M, Porter JC, Bomanji J, Shortman RI, Endozo R, et al. Synergistic application of pulmonary (18)F-FDG PET/HRCT and computer-based CT analysis with conventional severity measures to refine current risk stratification in idiopathic pulmonary fibrosis (IPF). Eur J Nucl Med Mol Imaging. 2019;46(10):2023–31. https://doi.org/10.1007/s00259-019-04386-5.
- Khoo JK, Barnes H, Key S, Glaspole IN, Ostor AJ. Pulmonary adverse events of small molecule JAK inhibitors in autoimmune disease: systematic review and meta-analysis. *Rheumatology (Oxford)*. 2020;59(9):2217–25. https://doi. org/10.1093/rheumatology/keaa117.
- Luan F, Ji Y, Peng L, Liu Q, Cao H, Yang Y, et al. Extraction, purification, structural characteristics and biological properties of the polysaccharides from Codonopsis pilosula: a review. *Carbohydr Polym.* 2021;261:117863. https://doi.org/10.1016/j.carbpol.2021.117863.
- Murray SG, Schmajuk G, Trupin L, Lawson E, Cascino M, Barton J, et al. A population-based study of infection-related hospital mortality in patients with dermatomyositis/polymyositis. *Arthritis Care Res (Hoboken)*. 2015;67(5): 673–80. https://doi.org/10.1002/acr.22501.

- Liang J, Sun C, Xu L, Xu G, Cao H, Lin J. Community-acquired pneumonia and hospital-acquired pneumonia in adult patients with idiopathic inflammatory myopathy: outcome and antibiotic therapy. *Rheumatol Ther*. 2021;8(1):255–72. https://doi.org/10.1007/s40744-020-00268-7.
- Li Y, Gao X, Li Y, Jia X, Zhang X, Xu Y, et al. Predictors and mortality of rapidly progressive interstitial lung disease in patients with idiopathic inflammatory myopathy: a series of 474 patients. Front Med (Lausanne). 2020;7:363. https://doi.org/10.3389/fmed.2020.00363.
- Chen F, Zuo Y, Li S, Shi J, Wang G, Shu X. Clinical characteristics of dermatomyositis patients with isolated anti-Ro-52 antibody associated rapid progressive interstitial lung disease: data from the largest single Chinese center. Respir Med. 2019;155:127–32. https://doi.org/10.1016/j.rmed.2019.07.020.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

