

Contribution of hepatitis E virus in acute sporadic hepatitis in north western India

Nidhi Subhash Chandra^{***}, Asha Sharma^{*}, Ramesh Roop Rai^{**} & Bharti Malhotra⁺

^{*}Department of Zoology, University of Rajasthan, ^{**}Department of Gastroenterology, SMS Hospital & ⁺Department of Microbiology, SMS Medical College, Jaipur, India

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Background & objectives: Hepatitis E virus (HEV) causes acute viral hepatitis. Majority of the documented studies on hepatitis E have been focused on the incidence of this disease in northern and south central India. Limited data are available on HEV infection among acute sporadic hepatitis cases in north western India. The present study was undertaken to investigate the contribution of hepatitis E virus infection in sporadic hepatitis cases in Rajasthan and neighbouring States.

Methods: Seven hundred and thirty six patients suspected to have viral hepatitis were screened for the hepatotropic viral markers, hepatitis A, B, C and E by using commercial enzyme immunoassay kits with a high sensitivity and specificity. The acute nature of HEV infection was also confirmed by the detection of HEV RNA by nested RT-PCR.

Results: Hepatitis E was found to be the major cause of acute sporadic viral hepatitis (49.7%) in this region of India. Mixed infections of HEV-HAV (1.2%), HEV-HBV (6.1%), and HEV-HCV (1.7%) were also detected. No viral marker was detected in 32 per cent cases.

Interpretation & conclusion: HEV was found as the major aetiological agent of acute sporadic viral hepatitis in Rajasthan (north western India). It is important to screen primarily for all the common enterically and parenterally transmitted hepatotropic viral markers in acute sporadic viral hepatitis. There is a need to do additional serological and molecular tests to identify the aetiological agent in the cases of acute hepatitis.

Key words Acute hepatitis - HBV - HCV - HEV - hepatitis - PCR - sporadic viral hepatitis

Hepatitis E was first recognised during an epidemic of hepatitis in Kashmir valley in 1978¹. It is an enterically transmitted disease that spreads through faecal contamination of drinking water. It occurs both in the form of epidemics as well as sporadic infection in developing countries²⁻⁵. According to the South East Asia Regional office of the World Health

Organization (WHO), hepatitis E is widespread in developing countries, accounting for up to 90 per cent of all sporadic cases of acute viral hepatitis⁶. Hepatitis E virus affects young to middle aged adults and causes high mortality in pregnant women, 20-30 per cent as compared to 0.2-1 per cent in general population⁷. It has been implicated as an important aetiological agent for

sporadic fulminant hepatic failure (FHF) in developing countries⁸.

Aetiology of acute viral hepatitis (AVH) cannot be differentiated on the basis of mode of presentation; confirmation is done serologically. Hepatitis E virus is an important hepatotropic virus that causes acute viral hepatitis. Most of the studies on prevalence of hepatitis E virus are from epidemic setting and some from sporadic setting. Therefore, the present study was undertaken to investigate the aetiology of AVH sporadic cases in Rajasthan and neighbouring States for hepatitis A virus (HAV), hepatitis B virus (HBV), hepatitis C virus (HCV), and hepatitis E virus (HEV). The acute nature of HEV infection was also confirmed by the detection of HEV RNA by nested RT-PCR.

Material & Methods

Patients: From January 2006 to July 2008, serum samples were collected from 736 individuals (aged 10-70 yr) from consecutive adult acute hepatitis cases aged 14-60 yr and only 4 children (aged 10-14 yr) suspected of having viral hepatitis, attending the OPD and wards of the Department of Gastroenterology, Sawai Man Singh (SMS) Hospital, Jaipur, Rajasthan. SMS hospital is a tertiary care centre where patients come from not only Rajasthan but from neighbouring States of Haryana, Punjab, Uttar Pradesh and Gujarat also. The study was approved by the institutional ethics committee and informed written consent was taken from the patients. Patients with signs and symptoms of jaundice, fever, loss of appetite, abdominal pain, scleral icterus, altered sensorium, encephalopathy and fatigue were included in the study. A detailed history of each patient was recorded, including travel history, blood transfusion, food and water intake from outside sources.

Immunoassays: Blood samples were collected from all patients under aseptic conditions and centrifuged at 1200 g. Serum was separated and stored at -80°C for further analysis. The samples were screened using commercially available Micro ELISA for markers of hepatitis A (IgM anti-HAV, Adaltis, Spain), hepatitis B (HBsAg, Biorad Monalisa HBsAg plus, IgM anti HBc in HBsAg positive cases Abbott laboratories North Chicago, IL) hepatitis C (Innotest HCV ab IV, Belgium) and hepatitis E (EIAgen HEV IgM, Adaltis, Spain). On the basis of serological tests, viral hepatitis was classified as acute hepatitis A (presence of IgM anti-HAV), acute hepatitis B (presence of HBsAg & IgM anti HBc), and hepatitis C may be acute or chronic (presence of anti-HCV) or acute hepatitis E (presence

of IgM anti HEV), acute hepatitis E and B (presence of anti IgM HEV, HBsAg, IgM anti HBc) and HBV carrier (HBsAg positive). The ELISA kit used for IgM anti HEV were coated with recombinant proteins for open reading frame (ORF) 1 and 2 with 98 per cent sensitivity and specificity. ELISA was performed as per manufacturers' protocol.

HEV RNA extraction and detection: HEV RNA detection was done in all 356 IgM anti-HEV positive cases and in 28 patients within first five days of illness but negative for all serological markers. Extracted RNA by Guanidinium thiocyanate (GITC) chloroform phenol method with minor modification⁹ was subjected for cDNA synthesis. cDNA synthesis was carried out using MuLV RT enzyme, reverse primer (20 pmol/ml, Promega) (external anti-sense: 5'- CCG AAT TCA AAG GCA TCC ATG GTG TTT GAG AAT GAC- 3') RNase out (20 U/μl, Invitrogen, UK), 0.1M DTT and 5 μl templates at 42°C for one hour.

After cDNA synthesis PCR amplification was carried out using the specific previously validated primers selected from non-structural ORF-1 region (Gene Bank accession no. M-32400)¹⁰. The primers used were external sense: 5'- CCG GAT CCA CAC ACA TCT GAG CTA CAT TCG TGA GCT- 3', external anti-sense: 5'- CCG AAT TCA AAG GCA TCC ATG GTG TTT GAG AAT GAC- 3', internal sense: 5'- GGA ATT CGA CTC CAC CCA GAA TTA CTT- 3', and internal anti-sense 5'- GGA ATT CAC AGC CGG CGA TCA GGA CAG- 3'. These two sets of primers were designed to produce 343 bp segment of ORF1 region¹⁰. The thermal cycling conditions were initial denaturation 94°C for 5 min followed by 30 cycles of denaturation for 30 sec at 94°C, annealing for 30 sec at 59°C and extension for 30 sec at 72°C, as well as final extension for 7 min at 72°C. The final PCR products of 384 patients (356 IgM antiHEV positive cases and 28 negative for all markers but with <5 day illness) were checked on 2 per cent gel electrophoresis stained with ethidium bromide (0.5 μg/ml) under UV transilluminator. For nested PCR, steps were taken to guard against carry over contamination by working in dedicated and physically separated rooms for sample preparation, master mix, amplification and electrophoresis. Each room had dedicated reagents, consumables, filter tips, micro pipettes, furniture, colour coded footwear and gowns, etc. which were not taken out to any other room. Samples were handled in biosafety cabinet. Positive and negative controls were included; sentinel negative controls were also used

to check for carry over contamination. The first inner PCR product was handled in a separate safety cabinet from the outer final PCR product.

Results

A total of 736 cases of acute sporadic hepatitis were seen over the study period. HEV was the most common cause followed by HBV, HAV and HCV. Of the 736 patients, 356 (48.3 per cent) patients with acute sporadic hepatitis were positive for IgM anti HEV alone (n=285) or in combination with other virus (n=71) by serology. In almost one third of the patients no viral marker could be detected (Table I). Among 356, 176 had acute viral hepatitis (AVH), 142 had fulminant hepatic failure (FHF) and 38 had subacute hepatic failure (SAHF). There was no mortality in 176 patients of AVH, while 16 (11.27%) FHF patients and 3 (7.8%) SAHF patients died. Of the 142 FHF patients, 24 (16.9%) were positive for HBs Ag, four for HAV IgM.

The age of presentation of patients with sporadic acute hepatitis E ranged from 10 to 65 yr (mean \pm SD of 32.43 ± 11.07 yr). Maximum numbers of cases of HEV were seen in the age group 15-40 yr. Among 356 IgM

anti HEV positive acute sporadic hepatitis E cases, 234 (65.73%) were males and 122 (34.26%) were females. Of the 122 consecutive females, 49 (40.16%) were pregnant. The mortality rate in pregnant females was 20.41 per cent (10/49) and in non-pregnant females was 4.10 per cent (3/73). 17 (34.69%) pregnancies ended in abortion or still birth and in 22 (44.89%) females pregnancy continued unaffected by AVH or normal delivery occurred during the disease course.

All 736 patients had abnormal liver function tests (LFT) suggestive of acute hepatitis; the tests were repeated at weekly intervals. The average duration of abnormal LFT was 11 to 15 days after onset of illness.

Serum samples from 356 IgM anti-HEV positive and 28 of 236 (11.86%) patients of acute sporadic hepatitis (within five days of illness and negative for all serological markers) were investigated for presence of HEV-RNA to confirm acute nature of HEV infection. Of the 356 IgM anti-HEV positives, 223 (62.64 %) and 10 out of 28 (35%) were found to be positive for HEV-RNA. Overall, 366 of 736 (49.72%) patients were found to be HEV positive by either ELISA or PCR.

Discussion

Hepatitis E virus is a major hepatotropic virus for acute viral hepatitis. Hepatitis E exists as sporadic hepatitis with periodic resurgence. This infection is responsible for 30-70 per cent cases of acute sporadic hepatitis¹¹ and is the major cause of acute liver failure (ALF)¹². It has been widely reported that HEV primarily affects young adults between 15-40 yr of age in endemic region¹³. Similar findings were observed in the present study.

In the present study, among the 736 serum samples obtained from acute sporadic hepatitis patients with jaundice, 366 (49.7%) were positive for HEV alone or in combination with other hepatitis viruses and responsible for about half of the acute viral hepatitis. HEV positivity ranging from 12.6-78.6 per cent has been reported by others from different parts of India (Table II)¹⁴⁻¹⁹. Positivity depends on sensitivity and specificity of kits used, population studied, type of samples whether sporadic or from outbreaks. Wide variation has been reported from 17-100 per cent in sensitivity of ELISA kits which depends on recombinant HEV antigens or synthetic peptides used in the kit corresponding to HEV epitopes, kits with ORF 2 sequence have shown little variation in geographically diverse strains in comparison to kits targeting ORF3.

Table I. The aetiology of acute sporadic hepatitis (n=736)

Causative agent	N (%)
Based on serological tests	
HEV	285 (38.7)
HAV	23 (3.1)
Total HBV	98 (13.3)
Acute HBV (HBsAg & IgM anti HBc positive)	35 (4.7)
HCV	15 (2)
HEV+HAV	9 (1.2)
Total HEV+HBV	45 (6.1)
{HEV + Acute HBV (HBs Ag & IgM anti HBc positive)	34 (4.61)
HEV+HCV	13 (1.7)
HAV+ HBV (IgM anti HBc negative)	1 (0.13)
HBV+HCV	5 (0.67)
HAV+HCV	1 (0.13)
HAV+HBV+HCV (IgM anti HBc negative)	1 (0.13)
HEV+HBV+HCV (IgM anti HBc negative)	3 (0.4)
HEV+HAV+HBV (IgM anti HBc negative)	1 (0.13)
Negative for all serological markers	236 (32)
[HEV RNA (done in 28/236 patients with < 5 days illness)]	10 (1.3)

Table II. Prevalence of hepatitis E virus in India reported in various studies

Authors	Year	Place	ELISA kits IgM anti HEV	HEV prevalence (%)	Total samples
Kumar <i>et al</i> ¹⁴	2007	Chandigarh	EIAgen Adalitis ORF1&2	~40	843
Chadha <i>et al</i> ¹⁵	1994-1997, 1978-1981	Pune	In house	12.6-42.4 46.4-71.2	276 206
Das <i>et al</i> ¹⁶	2000	Delhi	Genelabs ORF2&3	53.3	75
Jain <i>et al</i> ¹⁷	1999	Delhi	Genelabs ORF 2&3	42	50
Bansal <i>et al</i> ¹⁸	1998	Patna	Western blot (WB) IgG & IgM WB IgM	42 24.5	57
Sarguna <i>et al</i> ¹⁹	2007	Hyderabad	Immunovision ORF2& 3	78.57	546
Present Study	2006-2008	Rajasthan	EIAgen Adalitis ORF1&2	49.7	736

Variation may also occur with manufacturers lot and antibodies to different epitopes may differ in persistence²⁰⁻²². The test may perform differently in endemic and non endemic situation as 33-40 per cent general population has been found to be anti HEV-IgG positive, so difficulty may occur to distinguish between present and past HEV infection^{23,24}. Moreover, in immunocompromised individuals antibodies may persist for 6-10 months. Therefore, anti HEV IgM alone may not be informative for diagnosis of acute sporadic HEV infections at times.

In the present study all anti-HEV IgM positive samples were tested for HEV RNA to confirm acute ongoing infection as it has been reported that PCR may be better indicator for acute HEV infection during the first week of illness than ELISA. In our study, 62.64 per cent anti-HEV IgM positive cases were positive for HEV-RNA also. This could be due to the fact that HEV viraemia is transient, occurs in first two wk of infection and declines after 1 wk of onset of Jaundice²⁵. In patients with less than five days history of illness 80.7 per cent were positive for anti-HEV IgM and 85.3 per cent for HEV RNA as observed by us earlier also²⁶. Clayson *et al*²⁵ observed 93 per cent positivity for HEV RNA and only 79 per cent for IgM anti-HEV while Kumar *et al*¹⁴ reported only 9.4 per cent HEV RNA positivity in AVH cases but 100 per cent positivity in FHF cases in confirmed anti-HEV IgM positive samples. Detection of HEV RNA along with recombinant ORF2 ELISA for anti-HEV IgM could increase positive predictive value for diagnosis of acute HEV infection specially in endemic areas¹⁴. However, IgM antibody detection is a better choice for most diagnostic laboratories with serology facilities; HEV RNA testing may also be done in better equipped laboratories within one week

of onset of symptoms when other serological tests are negative.

In endemic areas, infection with HEV has been seen in association with other hepatotropic viruses (HAV, HBV and HCV) as also observed in the present study^{14,27-29}. Dual infection with HAV, HBV and HCV in acute HEV patients was observed in 1.2, 6.1 and 1.7 per cent cases, respectively, without any sequel. All the HAV and HEV dual infections were possibly co-infections as these have a common route of transmission and all infected patients were less than 14 yr of age. Malathi *et al*²⁷ observed dual infection of HEV and HAV in 13.4 per cent patients of acute hepatitis in children and Kumar *et al*²⁸ observed 4.4 per cent patients of acute hepatitis in adults.

In our study, dual acute infection of HEV and HBV was reported in 4.61 per cent and acute HEV on chronic HBV infection was seen in 1.5 per cent of acute hepatitis cases. It could be due to two reasons; one, the population corresponds to meso endemic zone of HBV; second reason could be attributed to the reactivation of latent HBV due to clinical HEV. However, there is little data available to determine such type of infection. In a study from Chennai²⁹, dual infection of HEV and HBV was observed in 5.4 per cent of adults patients of acute hepatitis. Anti-HEV IgM seropositivity rate ranging from 3.2 to 31 per cent in HBsAg carriers has been reported in different studies^{14,30} and hepatitis E has also been suspected to cause super-infection in carriers of hepatitis B³¹.

The HCV - HEV super-infection was observed in 1.7 per cent cases, all of whom had a history of prior exposure to procedures relating to parenteral route (7 had blood transfusion 4 had surgery and 2 were

drug abusers). Studies from Delhi and Chandigarh^{8,28} reported dual infection of HEV - HCV in 11 and 7.4 per cent cases, respectively. HCV positivity varies in different geographical areas. Low positivity of HCV has been reported from Rajasthan earlier; 0.28 per cent by Sood *et al*³² from a new private hospital and 1.693 per cent by Sharma *et al*³³ from a tertiary care hospital. Mehta *et al*³⁴ from Jodhpur reported 0.27 per cent HEV positivity in blood donors. Since our study was from known AVH cases, 5.1 per cent HCV positivity and 1.7 per cent HEV-HCV positivity are justified as per local trends of HCV infection.

In our study 32 per cent patients were found to be negative for all common hepatotropic viral markers by serological tests, further testing for HEV RNA in 28 samples negative for all serological test in patients with less than five days of illness helped in diagnosing HEV infection in additional patients. However, additional serological and molecular tests should be planned for identifying the aetiological agents in serologically negative cases for HAV, HBV, HCV and HEV to determine whether other viruses like hepatitis G (HGV), Sen Virus, TT virus, *etc.* exist in this part of India or variant of any other known pathogen.

References

1. Khuroo MS. Study of an epidemic of non-A, non-B hepatitis. Possibility of another human hepatitis virus distinct from post-transfusion non-A, non-B type. *Am J Med* 1980; 68 : 818-24.
2. Bradley DW. Hepatitis E: epidemiology, aetiology and molecular biology. *Rev Med Virol* 1992; 2 : 19-28.
3. Labrique AB, Thomas DL, Stoszek SK, Nelson KE. Hepatitis E: an emerging infectious disease. *Epidemiol Rev* 1999; 21 : 162-79.
4. Worm HC, Van der poel WH, Brandstatter G. Hepatitis E: an overview. *Microbes Infect* 2002; 4 : 657-66.
5. Arankalle VA, Chobe LP, Jha J, Chadha MS, Banerjee K, Favorov MO, *et al.* Aetiology of acute sporadic non-A, non-B viral hepatitis in India. *J Med Virol* 1993; 40 : 121-5.
6. Aggarwal R, Naik S. Epidemiology of hepatitis E: Current status. *J Gastroenterol Hepatol* 2009; 24 : 1484-93.
7. Chandra V, Taneja S, Kalia M, Jameel S. Molecular biology and pathogenesis of hepatitis E virus. *J Biosci* 2008; 33 : 451-64.
8. Nanda SK, Yalcinkaya K, Panigrahi AK, Acharya SK, Jameel S, Panda SK. Etiological role of hepatitis E virus in sporadic fulminant hepatitis. *J Med Virol* 1994; 42 : 133-7.
9. Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal Biochem* 1987; 162 : 156-9.
10. Jameel S, Durgapal H, Habibullah CM, Khuroo MS, Panda SK. Enteric non-A, non-B hepatitis: epidemics, animal transmission, and hepatitis E virus detection by the polymerase chain reaction. *J Med Virol* 1992; 37 : 263-70.
11. Panda SK, Thakral D, Rehman S. Hepatitis E virus. *Rev Med Virol* 2007; 17 : 151-80.
12. Acharya SK, Panda SK, Saxena A, Gupta SD. Acute hepatic failure in India: a perspective from the East. *J Gastroenterol Hepatol* 2000; 15 : 473-9.
13. Arankalle VA, Tsarev SA, Chadha MS, Alling DW, Emerson SU, Banerjee K, *et al.* Age-specific prevalence of antibodies to hepatitis A and E viruses in Pune, India, 1982 and 1992. *J Infect Dis* 1995; 171 : 447-50.
14. Kumar S, Ratho RK, Chawla YK, Chakraborti A. The incidence of sporadic viral hepatitis in North India: a preliminary study. *Hepatobiliary Pancreat Dis Int* 2007; 6 : 596-9.
15. Chadha MS, Walimbe AM, Chobe LP, Arankalle VA. Comparison of aetiology of sporadic acute and fulminant viral hepatitis in hospitalized patients in Pune, India during 1978-81 and 1994-97. *Indian J Gastroenterol* 2003; 22 : 11-5.
16. Das K, Agarwal A, Andrew R, Frösner GG, Kar P. Role of hepatitis E and other hepatotropic virus in aetiology of sporadic acute viral hepatitis: a hospital based study from urban Delhi. *Eur J Epidemiol* 2000; 16 : 937-40.
17. Jain A, Kar P, Madan K, Das UP, Budhiraja S, Gopalkrishna V, *et al.* Hepatitis C virus infection in sporadic fulminant viral hepatitis in North India: cause or co-factor. *Eur J Gastroenterol Hepatol* 1999; 11 : 1231-7.
18. Bansal J, He J, Yarbough PO, Sen S, Constantine NT, Sen D. Hepatitis E virus infection in eastern India. *Am J Trop Med Hyg* 1998; 59 : 258-60.
19. Sarguna P, Rao A, Sudha Ramana KN. Outbreak of acute viral hepatitis due to hepatitis E virus in Hyderabad. *Indian J Med Microbiol* 2007; 25 : 378-82.
20. Mast EE, Alter MJ, Holland PV, Purcell RH. Evaluation of assays for antibody to hepatitis E virus by a serum panel. Hepatitis E Virus Antibody Serum Panel Evaluation Group. *Hepatology* 1998; 27 : 857-61.
21. Ticehurst JR. Hepatitis E virus. In: Murray PR, Baron EJ, Pfaller MA, Tenover FC, Tenover RH, editors. *Manual of clinical microbiology*. 7th ed. Washington, DC: American Society for Microbiology; 1999.
22. Longer CF, Denny SL, Caudill JD, Miele TA, Asher LV, Myint KS, *et al.* Experimental hepatitis E: pathogenesis in cynomolgus macaques (*Macaca fascicularis*). *J Infect Dis* 1993; 168 : 602-9.
23. Lin CC, Wu JC, Chang TT, Chang WY, Yu ML, Tam AW, *et al.* Diagnostic value of immunoglobulin G (IgG) and IgM anti-hepatitis E virus (HEV) tests based on HEV RNA in an area where hepatitis E is not endemic. *J Clin Microbiol* 2000; 38 : 3915-8.
24. Favorov MO, Fields HA, Purdy MA, Yashina TL, Aleksandrov AG, Alter MJ, *et al.* Serologic identification of hepatitis E virus infections in epidemic and endemic settings. *J Med Virol* 1992; 36 : 246-50.
25. Clayson ET, Myint KS, Snitbhan R, Vaughn DW, Innis BL, Chan L, *et al.* Viremia, fecal shedding, and IgM and IgG

- responses in patients with hepatitis E. *J Infect Dis* 1995; 172 : 927-33.
26. Chandra NS, Sharma A, Malhotra B, Rai RR. Dynamics of HEV viremia, fecal shedding and its relationship with transaminases and antibody response in patients with sporadic acute hepatitis E. *Viol J* 2010; 7 : 213-9.
 27. Malathi S, Mohanavalli B, Menon T, Srilatha P, Sankaranarayanan VS, Raju BB, *et al.* Clinical and viral marker pattern of acute sporadic hepatitis in children in Madras, South India. *J Trop Pediatr* 1998; 44 : 275-8.
 28. Kumar S, Ratho RK, Chawla YK, Chakraborti A. Virological investigation of a hepatitis E epidemic in North India. *Singapore Med J* 2006; 47 : 769-73.
 29. Mohanavalli B, Menon T, Mohan KV, Ramathilagam B, Dinakaran N, Malathi S, *et al.* Serological profile of hepatitis A-E viruses in sporadic acute viral hepatitis in adults in Madras. *Indian J Gastroenterol* 1996; 15 : A116.
 30. Khuroo MS, Rustgi VK, Dawson GJ, Mushahwar IK, Yattoo GN, Kamili S, *et al.* Spectrum of hepatitis E virus infection in India. *J Med Virol* 1994; 43 : 281-6.
 31. Khuroo MS, Deurmeyer W, Zargar SA, Ahanger MA, Shah MA. Acute sporadic non-A, non-B hepatitis in India. *Am J Epidemiol* 1983; 118 : 360-4.
 32. Sood S, Malvankar S. Seroprevalence of hepatitis B surface antigen, antibodies to the hepatitis C virus, and human immunodeficiency virus in a hospital-based population in Jaipur, Rajasthan. *Indian J Commun Med* 2010; 35 : 165-9.
 33. Sharma R, Sinha P, Bachiwal R, Rishi S. Seroprevalence of anti-hepatitis C virus antibody in a hospital-based population of Jaipur, Rajasthan. *Indian J Commun Med* 2007; 32 : 158-59.
 34. Mehta NM, Purohit A, Haag A, Mathur A, Joshi KC, Joshi R, *et al.* Co-infection rate of HIV and hepatitis C Virus (HCV) among blood donors in Rajasthan, India. *Int Conf AIDS*, 2002; 14 : abstract no. TuPeA4401.

Reprint requests: Dr Bharti Malhotra, Associate Professor of Microbiology & Nodal Officer Advanced Research & TB Lab, SMS Medical College, Jaipur 302 004, India
e-mail: drbhartimalhotra@gmail.com