

HHS Public Access

Author manuscript

Obesity (Silver Spring). Author manuscript; available in PMC 2013 June 01.

Published in final edited form as:

Obesity (Silver Spring). 2012 December; 20(12): 2431–2437. doi:10.1038/oby.2012.162.

Copy number variations associated with obesity related traits in African Americans: a joint analysis between GENOA and HyperGEN

Wei Zhao¹, Nathan E. Wineinger^{2,3}, Hemant K. Tiwari², Thomas H. Mosley⁴, Ulrich Broeckel⁵, Donna K. Arnett⁶, Sharon L. R. Kardia¹, Edmond K. Kabagambe⁶, and Yan V. Sun^{1,7}

¹Department of Epidemiology, School of Public Health, University of Michigan, Ann Arbor, MI

²Department of Biostatistics, University of Alabama at Birmingham, Birmingham, AL

³Scripps Translational Science Institute, Scripps Health, San Diego, CA

⁴Department of Medicine, University of Mississippi Medical Center, Jackson, MS

⁵Department of Pediatrics and Medicine & Human and Molecular Genetics Center, Medical College of Wisconsin, Milwaukee, WI

⁶Department of Epidemiology, University of Alabama at Birmingham, Birmingham, AL

⁷Department of Epidemiology, Rollins School of Public Health, Emory University, Atlanta, GA

Abstract

Obesity is a highly heritable trait and a growing public health problem. African Americans are a genetically diverse, yet understudied population with a high prevalence of obesity (body mass index (BMI) greater than 30 kg/m²). Recent studies based upon single nucleotide polymorphisms (SNPs) have identified genetic markers associated with obesity. However, a large proportion of the heritability of obesity remains unexplained. Copy number variation (CNV) has been cited as a possible source of missing heritability in common diseases such as obesity. We conducted a CNV genome-wide association study of BMI in two African American cohorts from GENOA and HyperGEN. We performed independent and identical association analyses in each study, then combined the results in a meta-analysis. We identified three CNVs associated with BMI, obesity, and other obesity-related traits after adjusting for multiple testing. These CNVs overlap the *PARK2*, *GYPA* and *SGCZ* genes. Our results suggest that CNV may play a role in the etiology of obesity in African Americans.

Users may view, print, copy, and download text and data-mine the content in such documents, for the purposes of academic research, subject always to the full Conditions of use:http://www.nature.com/authors/editorial_policies/license.html#terms

Corresponding author: Yan V. Sun, Ph.D., Emory University, Rollins School of Public Health, Department of Epidemiology, 1518 Clifton Road NE #3049, Atlanta, GA 30322, Phone: (404) 727-9090, Fax: (404) 727-8737, yvsun@emory.edu.

Keywords

Obesity; CNVs; Meta-analysis; BMI; African Americans

INTRODUCTION

Obesity is a major public health problem, particularly among African Americans (AAs). In 2008, the prevalence of obesity among AAs in the Unites States was 37.3% in adult men and 49.6% in adult women, whereas the U.S. national prevalence was 32.2% and 35.5% in adult men and women, respectively (1). Obesity increases the likelihood of developing a variety of chronic diseases including diabetes, hypertension, high cholesterol, stroke, heart disease, cancers, and arthritis (2), leading to excess mortality and morbidity (3) and exerting a severe economic burden on the healthcare system (4). Environmental factors such as excessive energy intake and sedentary lifestyle are known to contribute to obesity. However, genetic factors have been found to strongly modulate an individual's susceptibility to obesity. Twin and family studies have estimated the heritability of body mass index (BMI) to range from 40–70% (5, 6). However, genome-wide association studies (GWAS) based upon single nucleotide polymorphisms (SNPs) have only accounted for a modest proportion of the total genetic variation – a common finding known as missing heritability (7, 8). Structural variants such as copy number variations (CNVs) have been suggested as a potential source of this missing heritability (7, 8).

CNVs have been reported to be associated with human disease, disorders (9, 10, 11, 12) and quantitative gene expression levels (13, 14) that, in some cases, are likely to have functional effects (14, 15). A recent study demonstrated that the Caucasian patients with onset of obesity before 10 years of age had significantly more large, rare deletions than controls (16). This finding was replicated in obese Caucasians regardless of the age of onset (17). However, obese AAs were found to possess less burden of rare CNVs, measured by the total span of CNVs per individual and average length of CNVs, compared to AA controls. (18). A genome-wide association study (GWAS) of obesity in children found eight rare CNVs that were exclusive to obese Caucasian and African American children (19). A study on BMI in Chinese identified a copy number variable region which overlaps a gene regulating energy homeostasis (20). This finding was replicated in Caucasians (21). These studies suggest that CNVs may play an important role in the genetic architecture of obesity. A large scale survey of CNVs is needed to further understand the genetics of obesity in AAs.

We conducted a CNV genome-wide association study on BMI and other obesity-related traits in two established African American cohorts: the Genetic Epidemiology Network of Arteriopathy (GENOA) and Hypertension Genetic Epidemiology Network (HyperGEN) from the National Heart, Lung and Blood Institute (NHLBI) Family Blood Pressure Program. Participants in both studies were genotyped on the Affymetrix 6.0 array – a platform which includes probes specifically designed for CNV analysis. We found three CNVs that were significantly associated with BMI and obesity-related traits after correcting for multiple testing. We also examined three previously reported CNVs linked to obesity (21, 22, 23), but found no evidence of association.

METHODS

Sample

The GENOA study is a community-based study of hypertensive sibships that aims to identify genes influencing blood pressure (24). AA subjects were enrolled in the field center in Jackson, MS. Sibships were recruited who met the eligibility requirement: at least two adults were clinically diagnosed with essential hypertension before age 60. Other siblings were invited to participate regardless of affection status. Data was collected in two phases: between 1995 and 2000, 1,854 subjects were examined; then between 2000 and 2004, 1,482 of these participants were re-examined. The GENOA study was approved by the Institutional Review Boards (IRB) of the participating institutions, and each participant provided written informed consent.

The HyperGEN study is a family based cross-sectional study designed to identify genetic contributors to hypertension (25). AA men and women were enrolled from centers in Birmingham, AL and Forsyth County, NC. In the first recruitment phase, sibships were recruited who met eligibility requirements: probands with onset of hypertension by age 60 and one or more hypertensive siblings who were willing to participate in the study (1995–2000). In the second phase, the offspring of the hypertensive siblings were recruited (2000–2003). At present, 1,224 AA subjects have been enrolled in HyperGEN and have provided the necessary phenotypic and genetic data. The HyperGEN study was approved by the IRB of the participating institutions, and each participant provided written informed consent.

Copy Number Variation Analysis

Study participants in GENOA (N=1,355) and HyperGEN (N=1,224) were genotyped using the Affymetrix Genome-wide Human SNP Array 6.0 platform. Genetic samples were excluded if they had an overall SNP call rate less than 95% or sex mismatch between genotypic and phenotypic measurement based upon quality control implementations. HyperGEN samples who had a disproportionately large number of copy number polymorphisms (greater than 25%) or were genotyped in small batches (16 or less subjects per batch) were also excluded. Following these quality control procedures, 1,263 AAs from GENOA and 1,026 AAs from HyperGEN remained for the current study (92 and 198 were excluded from GENOA and HyperGEN respectively). The Human Genome 18 reference build was used for probe localization.

The GENOA study used the Affymetrix Genotyping Console 3.0.1 to generate a reference genome by using all 1,263 raw intensity files (.CEL files) from the Human SNP Array 6.0 platform (Affymetrix Genotyping Console 3.0.1 User Manual). The boundaries of the common CNV segments were determined using the predefined CNV regions (26). Genotype calls for these CNVs were determined using the Canary algorithm (26). Any CNV call whose confidence score was less than 0.8 was excluded from the analysis (lower scores indicate more uncertainty), resulting in a 90% call rate. In total, we identified 1,130 CNV regions. Among these, 24 regions were monomorphic (normal type for all subjects) and were excluded from the analyses.

The HyperGEN study also used the Human SNP Array 6.0 platform to generate raw intensity files. CNVs were called using the Canary application within Birdsuite software, Version 1.5.5 (27). Samples were analyzed by batch to eliminate batch effects (28). CNV genotype calls with confidence values greater than 0.1 were removed from the analysis (higher scores indicate more uncertainty), resulting in a 92% call rate. In total, we identified 1,285 CNV regions. Among these, 84 regions were monomorphic and were excluded from the analyses. The chromosomal boundaries and the copy number state in each study were exported and used in the association analyses of BMI and related traits.

The Genotyping Console and the Birdsuite software have different implementations of the same Canary algorithm. Therefore the confidence scores from the two software packages have different ranges for high-quality CNV call. The confidence scores were used to exclude CNV calls with poor confidence. We chose the threshold to have about 10% overall missingness in both analysis datasets.

Statistical Analysis

We used a linear mixed effect model to test for CNV-BMI associations in each identified CNV region in each study population. This model included CNV, age, age-squared and sex as fixed effect covariates and family as a random effect to adjust for familial correlations within the families. CNV was treated as an additive trait, using the integer copy number values obtained from Canary. Principal component (PC) analysis was used to adjust for population stratification (29). The first 10 PCs based on 762,766 autosomal SNPs (MAF greater than 0.01 and call rate greater than 95%) from GENOA samples were calculated and incorporated into the mixed model as fixed effect covariates. Similarly, the first 10 PCs based on 823,728 autosomal SNPs from HyperGEN samples were included as covariates in the HyperGEN analysis. In extracting PCs, SNPs were removed from the calculation if they exhibited Mendelian inheritance errors, missingness greater than 1%, MAF less than 1%, or failure of Hardy-Weinberg equilibrium in founders (HWE p-value less than 0.001).

Associated CNVs were examined with other obesity-related traits: weight, hip circumference, and waist circumference. Identical linear mixed models were fit to perform the analyses on these continuous traits. We also stratified the sample into obese (BMI greater than or equal to 30 kg/m²) and non-obese (BMI less than 30 kg/m²). The relationship between CNV and obesity status was examined using generalized estimating equations.

An inverse variance based meta-analysis was carried out using METAL software (30) to combine the results from GENOA and HyperGEN. This approach calculated a weighted sum effect size, where the effect size for each study was weighted by the estimated standard errors. False discovery rate (FDR) was calculated to adjust for multiple testing and a FDR q-value of 0.1 was set as the threshold for significance.

All statistical analyses were performed with R statistical environment version 2.9.0 from R Project (http://www.r-project.org/) and METAL (30).

RESULTS

Basic descriptive statistics of the sample from each study population are summarized in Table 1. The GENOA and HyperGEN studies have similar sample sizes and phenotypic measurements on the traits we examined.

CNVs identified in this study were classified into three categories: deletion (copy number is less than 2), duplication (copy number is more than 2) and mixture type (observed copy numbers are less and more than 2). Among the 1,106 CNVs called in GENOA, 186 CNVs were mixed type, 750 CNVs were deletion type and 170 were duplication type. Similarly, among the 1,201 CNVs called in HyperGen, 221 CNVs were mixed type, 779 CNVs were deletion type and 201 were duplication type. There were 782 CNVs whose call types are consistent between these two studies, which suggest high quality CNV calls. Among these CNVs, 566 were deletion type, 95 duplication type and 121 mixed type. In this study, we focused on previously reported CNV regions with relatively common frequency in the HapMap sample. We did not observe any large CNVs (length > 500 kilobases) with frequency lower than 1% in both GENOA and HyperGEN AA populations.

There were a total of 1,037 CNVs that were shared between GENOA and HyperGEN. Among the 1,037 CNVs that were included in the meta-analysis, 55 CNV regions had pvalue less than 0.05 in the meta-analysis. After adjusting for multiple testing, three CNVs had q-value less than 0.1: CNP11162, CNP10809, and CNP11421 (Table 2). CNP11162 is located on chromosome 6, 162,416,281 to 162,423,724 bp with 1.7% of deletion; CNP10809 is located on chromosome 4, 145,220,925 to 145,232,498 bp with 4.2% of deletion and 1.7% of duplication; and CNP11421 is located on chromosome 8, 14,553,275 to 14,559,579 bp with 1.2% of deletion and 0.04% of duplication. The percentage of deletion (0 and 1 copies) and duplication (3 and 4 copies) were the percentage of individuals who carried loss or gain among all the individuals from both GENOA and HyperGEN. All the three CNVs included deletions. Although CNP10809 was mixture type, a greater number of deletions than duplications were observed in this region (82 individuals had loss and 34 individuals had gain). CNP11421 in HyperGEN was mixture type as well with only one individual who had gain. All of the three CNV regions show negative association with BMI, suggesting deletions in these regions may increase the risk of obesity. Figure 1 shows how BMI changes over different copies of CNVs in each study population.

Association analyses of these three CNVs with obesity, weight, hip circumference, and waist circumference were also conducted. Results are included in Table 3. As expected, these CNVs were found to be associated with most of the other obesity-related traits. The number of each type of CNVs in obese and non-obese groups is shown in Table 4.

We also examined the three previous reported CNV regions associated with obesity: CNP59 located on chromosome 1, 72,528,701 to 72,535,958 bp; CNP2150 located on chromosome 16, 19,853,151 to 19,874,863 bp, and CNP1732 located on chromosome 11, 55,130,608 to 55,209,585 (21, 22, 23). In our study, we found no evidence of association with these three CNVs: the p-value for CNP59 was 0.367 in GENOA and 0.255 in HyperGEN; the p-value for CNP2150 was 0.420 in GENOA and 0.594 in HyperGEN; and the p-value for CNP1732

was 0.406 in GENOA and 0.005 in HyperGEN. Previous reports suggest low copy number in CNP1732 is correlated with obesity in children (21). However, although we found CNP1732 to be significantly associated with BMI in HyperGEN, the association was in the opposite direction: lower copy number is associated with lower BMI. A meta-analysis examining these CNVs did not show evidence of an association with BMI (p-value = 0.150, 0.336, and 0.346 for CNV59, CNV2150, and CNV1732, respectively). Furthermore, we looked at the number of each CNV type in the obese group and the non-obese group. In GENOA, the number of deletions for CNP59, CNP2150 and CNP1732 were 31, 140, and 150 in the obese group compared to 38, 137 and 131 in the non-obese group. In HyperGEN, there were 97, 92, and 113 deletions in the obese group compared to 82, 71, and 107 in the non-obese group, respectively. There was no evidence of an enrichment of CNV in either group.

DISCUSSION

In this study we performed genome wide association analyses between CNVs and BMI in two AA populations. A meta-analysis suggested CNP11162, CNP10809 and CNP11421 were significantly associated with BMI. We performed additional association analyses of those CNVs with other obesity-related traits including weight, waist circumference, hip circumference, and dichotomized obesity, and found consistently significant associations. These variants overlap with genes *PARK2* (Parkinson protein 2, E3 ubiquitin protein ligase), *GYPA* (Glycophorin A) and *SGCZ* (Sarcoglycan, Zeta), respectively.

PARK2 encodes for parkin, a ubiquitin ligase. Recessive mutations in the PARK2 gene have been found in familial Parkinson's disease (31), which was characterized by dopamine degeneration in substantial nigral pathway. Animal studies confirmed that knockout of this gene in mice disrupted dopaminergic transmission in striatal area (32, 33). Dopamine is known to play an important role in modulating reward sensitivity, conditioning and high level cognitive control, which are all involved in the regulation of food intake (34) and therefore have a potential role in obesity. In one study, palatable food was shown to release dopamine in the dorsal striatum that is highly correlated with the level of pleasure subjects reported from eating the food (35). There is evidence from human imaging studies suggesting that obese individuals might have impairments in the dopaminergic pathway (36, 37). Thus, it is possible that a deletion in part of PARK2 gene leads to deficit in dopamine transmission that is involved in homeostatic regulation of food intake, and results in excessive food intake and obesity. Most recently, PARK2 knockout mice were found to resist body weight gain when they were exposed to a high fat diet during the age of 12 to 18 weeks, which suggests that this gene is involved in the regulation of fat intake (38). More interestingly, a recent study found an intronic SNP in the PARK2 gene associated with levels of several serum amino acids that are directly involved in metabolic pathway (39). The putative functional role of PARK2 on serum metabolites may assist further understanding of the relationship between the deletion in PARK2 gene and obesity-related traits.

GYPA is a gene that bears the antigenic determinants for the MN and Ss blood groups (40), and SGCZ encodes a protein that is part of sarcoglycan complex which bridges the inner

cyroskeleton and the extra-cellular matrix (41). Neither of these two genes has been reported to be associated with any obesity-related traits.

In a recent large scale GWAS, two CNV regions on chromosome 1 and 16 were reported to be associated with BMI in Caucasians (22, 23). Another recent family-based GWAS reported one CNV region on chromosome 11 that was associated with early onset of extreme obesity (21). We identified CNVs overlapping with these reported regions (CNP59 overlaps with the chromosome 1 region, CNP2150 overlaps with the chromosome 16 region and CNP1732 overlaps with the chromosome 11 region) and specifically examined whether these CNVs were associated with BMI in our study cohorts. However, we were unable to replicate these associations with BMI or other obesity-related traits. The frequencies of the three reported CNVs on chromosome 1, 16 and 11 (all of them are deletion polymorphisms) were 62%, 13%, and 28% in the reported studies of Caucasians (21, 22, 23) whereas the frequencies of the overlapping CNVs (CNP59, CNP2150 and CNP1732) were 12%, 21%, and 22% in our study cohorts. Also, the large sample sizes (32,387 and 249,796 participants) or the family design of the reported studies increased the power to detect small effect sizes. Our combined study of 2,289 AAs may have limited power to detect effects of that size. For example, if we assume the frequency of the non-normal variants is 0.3 and effect size is 0.17, the power of our study is only 0.14 at an alpha level of 0.05.

We identified three CNVs that were associated with BMI and obesity in AA populations. While our results should be interpreted within the context of obesity in hypertensive families, we have highlighted a potential causal pathway in one of these genes (*PARK2*) that may lead to dysfunction in brain rewarding and cognitive control regulating food intake, and thus result in excessive or compulsive food intake and obesity. As CNVs in our reported genes have not been previously found to be associated with BMI in studies based on Caucasian populations, our findings reinforce the need to stratify or account for population differences in genetic studies, particularly when considering traits like obesity that exhibit differing patterns among populations.

ACKNOWLEDGEMENTS

This study was supported by National Institute of Health grant HL100245, HL087660, HL055673, and HL079888, as well as the University of Alabama at Birmingham's Alumni Associations' Marie and Emmett Carmichael Fund for Graduate Students in Biosciences. The opinions expressed herein are those of the authors and not necessarily those of the NIH or any organization with which the authors are affiliated.

REFERENCES

- 1. Flegal KM, Carroll MD, Ogden CL, et al. Prevalence and trends in obesity among US adults 1999–2008. JAMA. 2010; 303:235–241. [PubMed: 20071471]
- 2. Malnick SD, Knobler H. The medical complications of obesity. QJM. 2006; 99:565–579. [PubMed: 16916862]
- 3. Flegal KM, Graubard BI, Williamson DF, et al. Cause-specific excess deaths associated with underweight, overweight, and obesity. JAMA. 2007; 298:2028–2037. [PubMed: 17986696]
- 4. Finkelstein EA, Trogdon JG, Brown DS, et al. The lifetime medical cost burden of overweight and obesity: implications for obesity prevention. Obesity (Silver Spring). 2008; 16:1843–1848. [PubMed: 18535543]

5. Maes HH, Neale MC, Eaves LJ. Genetic and environmental factors in relative body weight and human adiposity. Behav Genet. 1997; 27:325–351. [PubMed: 9519560]

- Atwood LD, Heard-Costa NL, Cupples LA, et al. Genomewide linkage analysis of body mass index across 28 years of the Framingham Heart Study. Am J Hum Genet. 2002; 71:1044–1050. [PubMed: 12355400]
- 7. Manolio TA, Collins FS, Cox NJ, et al. Finding the missing heritability of complex diseases. Nature. 2009; 461:747–753. [PubMed: 19812666]
- 8. Eichler EE, Flint J, Gibson G, et al. Missing heritability and strategies for finding the underlying causes of complex disease. Nat Rev Genet. 2010; 11:446–450. [PubMed: 20479774]
- 9. Pinto D, Pagnamenta AT, Klei L, et al. Functional impact of global rare copy number variation in autism spectrum disorders. Nature. 2010; 466:368–372. [PubMed: 20531469]
- Xu B, Woodroffe A, Rodriguez-Murillo L, et al. Elucidating the genetic architecture of familial schizophrenia using rare copy number variant and linkage scans. Proc Natl Acad Sci U S A. 2009; 106:16746–16751. [PubMed: 19805367]
- 11. Niederer HA, Willcocks LC, Rayner TF, et al. Copy number, linkage disequilibrium and disease association in the FCGR locus. Hum Mol Genet. 2010; 19:3282–3294. [PubMed: 20508037]
- 12. Grayson BL, Smith ME, Thomas JW, et al. Genome-wide analysis of copy number variation in type 1 diabetes. PLoS One. 2010; 5:e15393. [PubMed: 21085585]
- 13. Stranger BE, Forrest MS, Dunning M, et al. Relative impact of nucleotide and copy number variation on gene expression phenotypes. Science. 2007; 315:848–853. [PubMed: 17289997]
- 14. Sun YV, Peyser PA, Kardia SL. A common copy number variation on chromosome 6 association with the gene expression level of endothelin 1 in transformed B lymphocytes from three racial groups. Circ Cardiovasc Genet. 2009; 2:483–488. [PubMed: 20031624]
- Sun YV, Kardia SL. Identification of epistatic effects using a protein-protein interaction database. Hum Mol Genet. 2010; 19:4345–4352. [PubMed: 20736252]
- 16. Bochukova EG, Huang N, Keogh J, et al. Large, rare chromosomal deletions associated with severe early-onset obesity. Nature. 2010; 463:666–670. [PubMed: 19966786]
- 17. Wang K, Li WD, Glessner JT, et al. Large copy-number variations are enriched in cases with moderate to extreme obesity. Diabetes. 2010; 59:2690–2694. [PubMed: 20622171]
- Kang SJ, Chiang CW, Palmer CD, et al. Genome-wide association of anthropometric traits in African- and African-derived populations. Hum Mol Genet. 2010; 19:2725–2738. [PubMed: 20400458]
- 19. Glessner JT, Bradfield JP, Wang K, et al. A genome-wide study reveals copy number variants exclusive to childhood obesity cases. Am J Hum Genet. 2010; 87:661–666. [PubMed: 20950786]
- 20. Sha BY, Yang TL, Zhao LJ, et al. Genome-wide association study suggested copy number variation may be associated with body mass index in the Chinese population. J Hum Genet. 2009; 54:199–202. [PubMed: 19229253]
- 21. Jarick I, Vogel CI, Scherag S, et al. Novel common copy number variation for early onset extreme obesity on chromosome 11q11 identified by a genome-wide analysis. Hum Mol Genet. 2010
- 22. Willer CJ, Speliotes EK, Loos RJ, et al. Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. Nat Genet. 2009; 41:25–34. [PubMed: 19079261]
- 23. Speliotes EK, Willer CJ, Berndt SI, et al. Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. Nat Genet. 2010; 42:937–948. [PubMed: 20935630]
- 24. Daniels PR, Kardia SL, Hanis CL, et al. Familial aggregation of hypertension treatment and control in the Genetic Epidemiology Network of Arteriopathy (GENOA) study. Am J Med. 2004; 116:676–681. [PubMed: 15121494]
- 25. Williams RR, Rao DC, Ellison RC, et al. NHLBI family blood pressure program: methodology and recruitment in the HyperGEN network. Hypertension genetic epidemiology network. Ann Epidemiol. 2000; 10:389–400. [PubMed: 10964005]
- 26. Korn JM, Kuruvilla FG, McCarroll SA, et al. Integrated genotype calling and association analysis of SNPs, common copy number polymorphisms and rare CNVs. Nat Genet. 2008; 40:1253–1260. [PubMed: 18776909]

27. Korn JM, Kuruvilla FG, McCarroll SA, et al. Integrated genotype calling and association analysis of SNPs, common copy number polymorphisms and rare CNVs. Nat Genet. 2008; 40:1253–1260. [PubMed: 18776909]

- 28. Leek JT, Scharpf RB, Bravo HC, et al. Tackling the widespread and critical impact of batch effects in high-throughput data. Nat Rev Genet. 2010; 11:733–739. [PubMed: 20838408]
- 29. Price AL, Patterson NJ, Plenge RM, et al. Principal components analysis corrects for stratification in genome-wide association studies. Nat Genet. 2006; 38:904–909. [PubMed: 16862161]
- 30. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. Bioinformatics. 2010; 26:2190–2191. [PubMed: 20616382]
- 31. Kitada T, Asakawa S, Hattori N, et al. Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. Nature. 1998; 392:605–608. [PubMed: 9560156]
- 32. Itier JM, Ibanez P, Mena MA, et al. Parkin gene inactivation alters behaviour and dopamine neurotransmission in the mouse. Hum Mol Genet. 2003; 12:2277–2291. [PubMed: 12915482]
- 33. Oyama G, Yoshimi K, Natori S, et al. Impaired in vivo dopamine release in parkin knockout mice. Brain Res. 2010; 1352:214–222. [PubMed: 20620130]
- Volkow ND, Wang GJ, Baler RD. Reward, dopamine and the control of food intake: implications for obesity. Trends Cogn Sci. 2010
- 35. Small DM, Jones-Gotman M, Dagher A. Feeding-induced dopamine release in dorsal striatum correlates with meal pleasantness ratings in healthy human volunteers. Neuroimage. 2003; 19:1709–1715. [PubMed: 12948725]
- 36. Stoeckel LE, Weller RE, Cook EW 3rd, et al. Widespread reward-system activation in obese women in response to pictures of high-calorie foods. Neuroimage. 2008; 41:636–647. [PubMed: 18413289]
- 37. Stice E, Spoor S, Bohon C, et al. Relation of reward from food intake and anticipated food intake to obesity: a functional magnetic resonance imaging study. J Abnorm Psychol. 2008; 117:924–935. [PubMed: 19025237]
- 38. Kim KY, Stevens MV, Akter MH, et al. Parkin is a lipid-responsive regulator of fat uptake in mice and mutant human cells. J Clin Invest. 2011; 121:3701–3712. [PubMed: 21865652]
- 39. Gieger C, Geistlinger L, Altmaier E, et al. Genetics meets metabolomics: a genome-wide association study of metabolite profiles in human serum. PLoS Genet. 2008; 4:e1000282. [PubMed: 19043545]
- 40. Palacajornsuk P. Review: molecular basis of MNS blood group variants. Immunohematology. 2006; 22:171–182. [PubMed: 17430076]
- 41. Wheeler MT, Zarnegar S, McNally EM. Zeta-sarcoglycan, a novel component of the sarcoglycan complex, is reduced in muscular dystrophy. Hum Mol Genet. 2002; 11:2147–2154. [PubMed: 12189167]

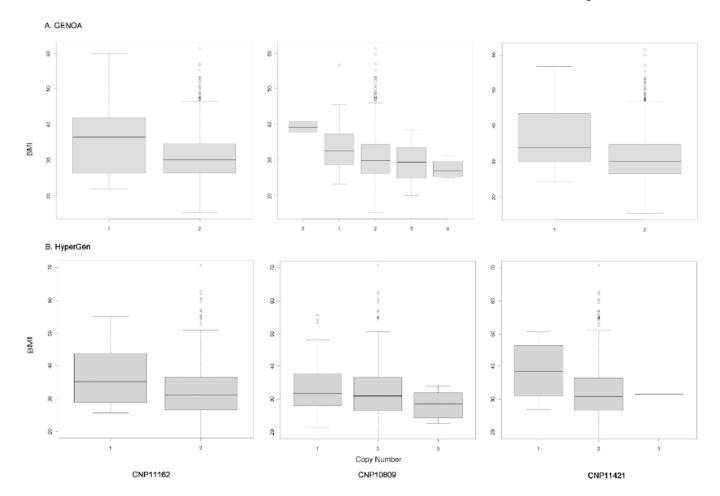


Figure 1.

The relationship of BMI and the three significant CNVs (CNP11162, CNP10809 and CNP11421). A: Box plots of BMI with different copies of CNVs in GENOA; B: Box plots of BMI with different copies of CNVs in HyperGEN

Table 1

Basic descriptive statistics of the samples

		GENOA			HyperGEN	
Trait	Total (N=1,263)	Male (N=398)	Fotal (N=1,263) Male (N=398) Female (N=865) Total (N=1,026) Male (N=337) Female (N=689)	Total (N=1,026)	Male (N=337)	Female (N=689)
Age (years), Mean (SD)	58.0 (10.07)	59.3 (9.62)	57.4 (10.22)	44.3 (13.43)	44.3 (13.43) 43.1 (13.63)	45.0 (13.29)
Weight (kg), Mean (SD)	88.3 (18.21)	90.3 (17.06)	87.4 (18.66)	90.9 (23.55)	92.3 (21.21)	90.4 (24.61)
Height (cm), Mean (SD)	168.8 (8.94)	177.9 (6.47)	164.5 (6.39)	167.6 (8.90)	167.6 (8.90) 176.4 (6.81)	163.3 (6.26)
BMI, kg/m², Mean (SD)	31.1 (6.49)	28.5 (4.95)	32.3 (6.76)	32.4 (8.12)	29.6 (6.26)	33.8 (8.56)
Waist Circumference (cm), Mean (SD) 103.4 (16.25) 100.3 (12.10) 104.8 (17.67)	103.4 (16.25)	100.3 (12.10)	104.8 (17.67)	102.6 (18.71) 99.7 (16.57)	99.7 (16.57)	104.0 (19.51)
Hip Circumference (cm), Mean (SD)	113.2 (14.11)	105.6 (10.38)	113.2 (14.11) 105.6 (10.38) 116.7 (14.25)	114.4 (16.35)	107.9 (12.76)	117.7 (16.95)
Obese (BMI 30 kg/m^2)	640 (50.7%)	128 (32.2%)	512 (59.2%)	570 (55.7%)	136 (40.5%)	434 (63.1%)

Table 2

CNVs associated with BMI

	l e	2	2	
FDR	q-valı	0.052	0.05	0.070
Meta Analysis	Beta SE p-value p-value q-value	0.0001	0.0001	0.0002
	p-value	0.0081	0.1712	0.0026
	SE	2.24	1.32	2.05
HyperGEN	Beta	-5.93	-1.75	-6.12
Ħ	Beta SE p-value N Type	Deletion	Mixture ^a	Mixture ^a
	Z	1,021	886	1,023
	p-value	0.0036	0.0003	0.0229
	SE	1.39	0.63	1.89
GENOA	Beta	-4.05	-2.31	-4.31
J	Type	Deletion	Mixture ^a	Deletion
	Z	1,263	1,008	1,258
	End position	.81 162,423,724 1,263 Deletion -4.05 1.39 0.0036 1,021 Deletion -5.93 2.24 0.0081 0.0001 0.052	$25 145,232,498 1,008 \text{Mixture}^{a} -2.31 0.63 0.0003 938 \text{Mixture}^{a} -1.75 1.32 0.1712 0.0001 0.052$	14,559,579
	CNV Chr Start position End position N Type	162,416,281	145,220,925	14,553,275 14,559,579 1,258 Deletion -4.31 1.89 0.0229 1,023 Mixture -6.12 2.05 0.0026 0.0002 0.070
	Chr	9	4	~
	CNV	CNP11162 6 162,416,2	CNP10809	CNP11421

a observed copy numbers can be less or more than 2 (i.e., some individuals had either deletion or amplification creating mixture of individuals with deletion, amplification, or normal)

Author Manuscript

Author Manuscript

Table 3

Association of BMI-associated CNVs and other obesity related traits

021 Deletion -18.49 6.56 0.0048 0.0009 938 Mixturea -3.75 3.88 0.3197 0.00034 .023 Mixturea -21.02 6.04 0.0005 0.0001 .021 Deletion -11.23 4.47 0.0120 0.0004 .023 Mixturea -13.62 4.07 0.0007 0.0005 .021 Deletion -11.98 5.20 0.0202 0.0017 .023 Mixturea -5.71 3.05 0.0568 0.0008 .023 Mixturea -11.61 4.70 0.0125 0.0024 .021 Deletion -0.91 0.77 0.2291 0.06203 .023 Mixturea -0.26 0.45 1.000 0.0006 .023 Mixturea -1.23 0.80 0.0898 0.0454	GENOA N Type Beta SE p-value	GENOA Beta SE	SE		p-value		z	Type	HyperGEN Beta	SE	p-value	Meta Analysis p-value
-18.49 6.56 0.0048 -3.75 3.88 0.3197 -21.02 6.04 0.0005 -11.23 4.47 0.0120 -3.14 2.64 0.2234 -13.62 4.07 0.0007 -11.98 5.20 0.0202 -5.71 3.05 0.0568 -11.61 4.70 0.0125 -0.91 0.77 0.2291 -0.26 0.45 1.000 -1.23 0.80 0.0898	Weight (kg)											
xurrea -5.08 1.82 0.0055 938 Mixturea -3.75 3.88 0.3197 letion -11.42 5.48 0.0375 1,023 Mixturea -21.02 6.04 0.0005 runca -2.31 0.63 0.0003 938 Mixturea -3.14 2.64 0.0120 nh -4.31 1.89 0.0229 1,023 Mixturea -13.62 4.07 0.0007 nh -8.00 3.60 0.0265 1,021 Deletion -11.98 5.20 0.0202 letion -8.00 3.60 0.0265 1,021 Deletion -11.98 5.0 0.0568 letion -8.80 4.92 0.0723 1,023 Mixturea -5.71 3.05 0.0568 letion -0.66 0.45 0.0723 1,023 Mixturea -5.71 4.70 0.0125 xumea -0.66 0.45 0.1409 1,021 Deletion -0.91 0.77 0.2291	.62	1,263	Deletion	-8.71	4.02	0.0305	1,021	Deletion	-18.49	6.56	0.0048	0.0000
letion -11.42 5.48 0.0375 1,023 Mixturea -21.02 6.04 0.0005 letion -4.05 1.39 0.0036 1,021 Deletion -11.23 4.47 0.0120 nuxturea -2.31 0.63 0.0003 938 Mixturea -3.14 2.64 0.0120 letion -4.31 1.89 0.0229 1,023 Mixturea -3.16 4.07 0.0007 nuxturea -8.00 3.60 0.0265 1,021 Deletion -11.98 5.20 0.0268 xturea -4.60 1.64 0.0052 938 Mixturea -5.71 3.05 0.0568 letion -8.85 4.92 0.0723 1,023 Mixturea -11.61 4.70 0.0125 xturea -0.80 0.23 0.004 938 Mixturea -0.91 0.77 0.2291 xturea -0.80 0.23 0.004 938 Mixturea -0.26 0.45 <	809	1,008	$Mixture^a$	-5.08	1.82	0.0055	938	Mixture ^a	-3.75	3.88	0.3197	0.0034
letion -4.05 1.39 0.0036 1,021 Deletion -11.23 4.47 0.0120 xturea -2.31 0.63 0.0003 938 Mixturea -3.14 2.64 0.0234 n) Aixturea -3.14 2.64 0.234 letion -4.31 1.89 0.0229 1,023 Mixturea -13.62 4.07 0.0007 letion -8.00 3.60 0.0265 1,021 Deletion -11.98 5.20 0.0268 letion -8.00 3.60 0.0062 938 Mixturea -5.71 3.05 0.0568 letion -0.66 0.45 0.0723 1,023 Mixturea -0.91 0.77 0.2291 xturea -0.80 0.23 0.0004 938 Mixturea -0.26 0.45 1.000 xture -0.80 0.62 0.1804 1,023 Mixturea -0.26 0.45 1.000	421	1,258	Deletion	-11.42	5.48	0.0375	1,023	Mixture ^a	-21.02	6.04	0.0005	0.0001
tetion 4.05 1.39 0.0036 1,021 Deletion -11.23 4.47 0.0120 xturrea -2.31 0.63 0.0003 938 Mixturea -3.14 2.64 0.0234 letion -4.31 1.89 0.0229 1,023 Mixturea -13.62 4.07 0.0007 sturrea -8.00 3.60 0.0265 1,021 Deletion -11.98 5.20 0.0202 sturrea -4.60 1.64 0.0052 938 Mixturea -5.71 3.05 0.0568 letion -8.85 4.92 0.0723 1,023 Mixturea -11.61 4.70 0.0125 xturrea -0.66 0.45 0.1409 1,021 Deletion -0.91 0.77 0.2291 xturrea -0.80 0.23 0.0004 938 Mixturea -0.26 0.45 1.000 xturrea -0.80 0.62 0.1804 1.023 Mixturea -0.23 0.80	rcumf	erence (cm)									
xumea -2.31 0.63 0.0003 938 Mixturea -3.14 2.64 0.2234 n A A A A A A A A A A A A A A A A B		1,263	Deletion	-4.05	1.39	0.0036	1,021	Deletion	-11.23	4.47	0.0120	0.0004
nh -4.31 1.89 0.0229 1,023 Mixturea -13.62 4.07 0.0007 nh sturea -8.00 3.60 0.0265 1,021 Deletion -11.98 5.20 0.0202 xturea -4.60 1.64 0.0052 938 Mixturea -5.71 3.05 0.0268 letion -8.85 4.92 0.0723 1,023 Mixturea -11.61 4.70 0.0125 letion -0.66 0.45 0.1409 1,021 Deletion -0.91 0.77 0.2291 xturea -0.80 0.23 0.0004 938 Mixturea -0.26 0.45 1.000 letion -0.83 0.62 0.1804 1,023 Mixturea -0.26 0.45 1.000		1,008	Mixture ^a	-2.31	0.63	0.0003	938	Mixture ^a	-3.14	2.64	0.2234	0.0001
n) n a n a	421	1,258	Deletion	-4.31	1.89	0.0229	1,023	Mixturea	-13.62	4.07	0.0007	0.0005
letion -8.00 3.60 0.0265 $1,021$ Deletion -11.98 5.20 0.0202 xturrea -4.60 1.64 0.0052 938 Mixturea -5.71 3.05 0.0568 letion -8.85 4.92 0.0723 $1,023$ Mixturea -11.61 4.70 0.0125 letion -0.66 0.45 0.1409 $1,021$ Deletion -0.91 0.77 0.2291 xturrea -0.80 0.23 0.0004 938 Mixturea -0.26 0.45 1.000 letion -0.83 0.62 0.1804 $1,023$ Mixturea -0.26 0.45 1.000	Circun	nferenc	e (cm)									
xture a -4.60 1.64 0.0052 938 Mixture a -5.71 3.05 0.0568 letion -8.85 4.92 0.0723 1,023 Mixture a -11.61 4.70 0.0125 letion -0.66 0.45 0.1409 1,021 Deletion -0.91 0.77 0.2291 xture a -0.80 0.23 0.0004 938 Mixture a -0.26 0.45 1.000 letion -0.83 0.62 0.1804 1,023 Mixture a -1.23 0.80 0.0898	162	1,263	Deletion	-8.00	3.60	0.0265	1,021	Deletion	-11.98	5.20	0.0202	0.0017
letion -8.85 4.92 0.0723 1,023 Mixturea -11.61 4.70 0.0125 letion -0.66 0.45 0.1409 1,021 Deletion -0.91 0.77 0.2291 xturea -0.80 0.23 0.0004 938 Mixturea -0.26 0.45 1.000 letion -0.83 0.62 0.1804 1,023 Mixturea -1.23 0.80 0.0898	608	1,008	Mixture ^a	-4.60	1.64	0.0052	938	Mixture ^a	-5.71	3.05	0.0568	0.0008
letion -0.66 0.45 0.1409 $1,021$ Deletion -0.91 0.77 0.2291 xture a -0.80 0.23 0.0004 938 Mixture a -0.26 0.45 1.000 letion -0.83 0.62 0.1804 $1,023$ Mixture a -1.23 0.80 0.0898	421	1,258	Deletion	-8.85	4.92	0.0723	1,023	Mixture ^a	-11.61	4.70	0.0125	0.0024
1,263 Deletion -0.66 0.45 0.1409 1,021 Deletion -0.91 0.77 0.2291 1,008 Mixturea -0.80 0.23 0.0004 938 Mixturea -0.26 0.45 1.000 1,258 Deletion -0.83 0.62 0.1804 1,023 Mixturea -1.23 0.80 0.0898	y (BMI)	1 30 kg	/m ²)									
1,008 Mixture ^a -0.80 0.23 0.0004 938 Mixture ^a -0.26 0.45 1.000 1,258 Deletion -0.83 0.62 0.1804 1,023 Mixture ^a -1.23 0.80 0.0898		1,263	Deletion	-0.66	0.45	0.1409	1,021	Deletion	-0.91	0.77	0.2291	0.06203
1,258 Deletion -0.83 0.62 0.1804 1,023 Mixturea -1.23 0.80 0.0898		1,008	Mixture ^a	-0.80	0.23	0.0004	938	Mixture ^a	-0.26	0.45	1.000	0.0006
	421	1,258	Deletion	-0.83	0.62	0.1804	1,023	Mixture ^a	-1.23	0.80	0.0898	0.0454

a observed copy numbers can be less or more than 2 (i.e., some individuals had either deletion or amplification creating mixture of individuals with deletion, amplification, or normal)

Zhao et al.

Table 4

The Frequency Table of CNVs in Obese Group vs Non-obese Group

			GENOA	P(HyperGEN	EN		TOTAL	Т
CNV	Copy Number	Z	Opese	Non-Obese	Z	Obese	Non-Obese	N	Opese	Non-Obese
CNP11162	1	1,263	15	L	1,021	11	9	2,284	26	13
	2		625	919		557	447		1182	1063
CNP10809	0	1,008	2	0	886	0	0	1,946	2	0
	1		32	15		17	13		52	28
	2		450	8/4		501	401		951	628
	3		01	14		2	4		12	18
	4		1	3		0	0		1	3
CNP11421	1	1,258	8	3	1,023	14	3	2,281	22	9
	2		630	617		554	451		1,084	1,068
	3		0	0		1	0		1	0

Page 14