IMMUNOLOGY: RESEARCH ARTICLE

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Common bone marrow signature in COVID-19-associated multisystem inflammatory syndrome in children: A first-wave small case series experience

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Abstract

The hyper-inflammatory response, also known as multisystem inflammatory syndrome in children (MIS-C), represents a major concern in children with SARS-CoV-2 infection. We report bone marrow features of three patients with MIS-C who were diagnosed during the first wave of the SARS-CoV-2 pandemic. A bone marrow evaluation was performed at onset of the inflammatory condition in order to exclude secondary hemophagocytic lymphohistiocytosis (sHLH). The bone marrows of the patients presented common features: the erythroid and megakaryocytic lineages were prominently affected and hemophagocytosis was moderately increased, differently than observed in sHLH. Megakaryocytopoiesis was increased, representing a peculiar feature of MIS-C differing from sHLH. SARS-CoV-2 RT-PCR and viral panel were studied in bone marrow aspiration samples. MIS-C is a rare complication of SARS-CoV-2 infections in children. An immuno-dysregulation considering both innate and adaptive immunity together with vascular inflammation and endothelial dysfunction play a major role. Our observations, although limited due to the small sample size, suggest that there are unique features in the bone marrow of patients with MIS-C that are likely secondary to immuno-dysregulation, and there are notable differences in bone marrow features compared to those reported in sHLH.

KEYWORDS bone marrow, MIS-C, SARS-CoV-2

Abbreviations: BM, bone marrow; CXCL10, C-X-C motif chemokine ligand 10; CXCL9, C-X-C motif chemokine ligand 9; GvHD, graft-versus-host disease; IL, interleukin; MIS-C, multisystem inflammatory syndrome in children; MPO, myeloperoxidase; OPBG, Bambino Gesù Children's Hospital; sHLH, secondary hemophagocytic lymphohistiocytosis; TFNα, tumor necrosis factor α.

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1 INTRODUCTION

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Children are susceptible to SARS-CoV-2 infection, but are less likely to be symptomatic or present severe respiratory symptoms with less than 1% of critically ill patients^{1–5}; however, a hyperinflammatory response, also known as multisystem inflammatory syndrome in children (MIS-C), represents a major concern in pediatric population.^{6–13} MIS-C occurs in children 4–6 weeks after mild or asymptomatic SARS-CoV-2 infection due to a delayed immune response. The case definition for MIS-C is broad, including the presence of fever with laboratory evidence of inflammation and of severe multisystem involvement: cardiac, renal, respiratory, hematologic, gastrointestinal, dermatologic, or neurologic.^{5–13}

During the first SARS-CoV-2 wave, specifically at the first observations of hyperinflammatory syndrome, we performed a bone marrow (BM) evaluation in order to detect secondary hemophagocytic lymphohistiocytosis (sHLH). We report the BM features with plasma cytokine profiles of three children who met the criteria for the diagnosis of MIS-C. We compared BM with patients who had sHLH.

2 METHODS

We focused on three patients admitted at Bambino Gesù Children's Hospital (OPBG) with a hyperinflammatory syndrome consistent with MIS-C¹³ in which a BM evaluation was performed at diagnosis. The hyperinflammatory status of all three patients was diagnosed by the end of April 2020; at that time, MIS-C had not yet been described as a pathological entity uniquely associated with the SARS-CoV-2 infection, and a BM evaluation was performed to exclude/confirm an sHLH. The plasma from the three patients was collected before any anti-inflammatory therapy. Clinical characteristics are summarized in Table 1 and Table S1. All patients were alive 24 months after MIS-C diagnosis. A viral, bacterial, and fungal infection workup was performed on blood, BM, nasopharyngeal swab, and urine and stool samples. All three patients presented a positive RT-PCR nasopharyngeal swab for SARS-CoV-2 within the last 4 weeks prior to MIS-C diagnosis.

Cytokine profile data were obtained at diagnosis: interleukin (IL)-1 β), IL-6, IL-8, tumor necrosis factor α (TFN α), C-X-C motif chemokine ligand 9 (CXCL9), C-X-C motif chemokine ligand 10 (CXCL10), and IL-18 were considered for this report. Cytokine levels were measured using an automated ELISA assay (ELLA microfluidic analyzer, Protein Simple) according to the manufacturer's instructions.

2.1 Bone marrow morphological study

BM biopsies were fixed in acetic zinc formalin, decalcified in ethylenediaminetetraacetic acid (EDTA), and paraffin embedded. Hematoxylin and eosin, periodic acid-Schiff, May–Grunwald–Giemsa, reticulin, and Pearls iron stains were performed. The immunohistochemical panel included glycophorin, CD71, myeloperoxidase (MPO), CD61, CD10, TdT, CD20, CD79, CD3, CD4, CD8, CD68, CD163, CD138, and CD117. As controls, BM biopsies of five patients with sHLH diagnosed in the last 3 years at OPBG were compared to BM biopsies of MIS-C patients. The patients with sHLH had an HLH of unknown etiology, and no infection or underline disease had been identified. The Ethics Committee (2159_OPBG_2020) approved this retrospective study. All investigations were conducted in accordance with principles expressed in the Declaration of Helsinki.

3 | RESULTS

A BM evaluation was performed in three pediatric patients with MIS-C. Co-infection were excluded. A viral panel including RT-PCR for *Cytomegalovirus*, Epstein–Barr virus, parvovirus B19, herpes simplex virus 1 and 2, human herpes virus 6, and adenovirus were negative on peripheral and BM blood. Blood, urine, and sputum cultures were negative, as was the viral panel on nasopharyngeal swab. The level of plasma cytokines increased in all patients (Table 1).

Pathology findings of BM biopsies are summarized in Table 2. Cellularity was normal in two patients and decreased in one. Erythroid precursors were reduced in all BM biopsies, with isolated early erythroblasts that rarely formed small aggregates. Myeloid population (stained with MPO) was relatively increased with left shift and focal disarray. Megakaryocytes (identified with a CD61 staining) were increased in number, with cell size variable from small to large along with the presence of rare micro-megakaryocytes. The nuclei appeared hypo- or hyper-lobated and hyper-chromatic. A mild lymphocytic infiltrate was seen with a prevalent T phenotype. CD68 and CD163 highlighted mild histiocytic hyperplasia, with a small number of histiocytes containing vacuoles, granules, and ingested blood elements (Figure 1). The plasma cell and mast cell counts were normal. No vascular alterations were found (Figure 2). Reticulin stain showed a mildly increased reticulin network.

4 DISCUSSION

In the first wave of the COVID-19 pandemic, the pediatric population had been considered less affected by COVID-19.3-5 By the end of April 2020, clinicians and scientists were warned over a rising number of children presenting with a multisystem inflammatory condition needing intensive care, in connection to the pandemic; the presenting signs and symptoms overlapped with another inflammatory disease such as Kawasaki disease and macrophage activation syndrome (MAS)/sHLH.5-14 The acronym MIS-C is used to identify a clinical condition characterized by an inflammatory status with frequent gastrointestinal manifestations and circulatory failure, including myocardial injury. Despite a severe disease with often emerging life-threatening symptoms, the mortality rate is low at 1.9%.^{4–14} The pathogenesis is not yet fully understood, but it seems to be due to a delayed immunological phenomenon associated with proinflammatory triggers following either symptomatic or asymptomatic SARS-CoV-2 infection related to age-specific immune response mechanisms.¹⁵⁻¹⁸

TABLE 1 Laboratory findings at multisystem inflammatory syndrome in children (MIS-C) onset

	Patient 1	Patient 2	Patient 3
Lymphocyte count per mm ³ , N (%) (5.5–15) (40%–57%)	600 (10%)	810 (7.9%)	1430 (11%)
Hemoglobin, g/dl (10.5-15)	10.7	11.3	10.4
Platelets per mm ³ (150–450)	116,000	215,000	186,000
C-reactive protein, mg/dl (<0.5)	19	22	28
Procalcitonin, ng/ml (<0.5)	7.9	2.28	1.2
Erythrocyte sedimentation rate, mm/h (<20)	57	53	74
ThsTnT (<14 pg/ml)	86.5	93	173
NT-proBNP (<230 pg/ml)	812	2950	190
D-dimer (>0.5 mg/L)	5.42	3.18	>20
Ferritin (<300 µg/ml)	772	23.345	419
Cholesterol, mg/dl (<170)	76	85	151
Triglyceride, mg/dl(<0.5)	86	196	124
Alanine aminotransferase (<41 U/L)	19	90	21
Aspartate aminotransferase (<40 U/L)	25	97	27
Lactate dehydrogenase (120-300 U/L)	228	569	255
Uric acid (3.4–7 mg/dl)	3	3.2	4.9
Serum albumin (3.8–5.4 g/dl)	3.3	2.6	4.0
Fibrinogen (162-401 mg/dl)	564	400	643
Sodium (136–145 mEq/L)	135	137	137
IL-1b, pg/ml (HD <0.04)	1.3	6.2	2.5
IL-6, pg/ml (HD 1.2; 0.91–1.89)	76.4	90.7	50.3
IL-8, pg/ml (HD 1.55–1.6)	40.9	408.1	209.7
TNF-a, pg/ml (HD 7.47; 5.4–9.4)	27.3	20.9	20.9
CXCL9*, pg/ml (NV 429.5-801.5)	2760	1267	515
CXCL10*, pg/ml (NV 111.0-379.3)	2034	560	244
IL-18*, pg/ml (NV <300 pg/ml)	<2048	246,492	<2048
Perforin	Normal	Normal	Normal
Cytotoxicity assay	Reduced	Reduced	Reduced
Degranulation assays	Reduced	Reduced	Reduced

Note: N, absolute count. For ThsTnT, NT-proBNP, D-dimer, and ferritin are added at the upper local limit; for IL1b, IL 6, IL8, and TNF-a are added median value and range in healthy donors (HD).

There is scarce information reporting on BM evaluation in COVID-19 patients that showed hemophagocytosis; the COVID-19-related hyperinflammatory status is considered as a form of sHLH.¹⁹⁻²⁹ Recently, Lacinel et al. reported the presence of hemophagocytosis on BM aspirates from three children with MIS-C.³⁰

The three reported children presented an inflammatory condition consistent with MIS-C after the SARS-CoV-2 infection. Plasma cytokine levels were increased in all patients, as observed in recent MIS-C immunological studies.¹⁵⁻¹⁸ Surprisingly, the BM presented common features in all three patients; the erythroid and megakaryocytic lineages were prominently affected and hemophagocytosis was evident. Hemophagocytosis was previously described in adults on BM, lymph nodes, and spleen (mostly in autopsy findings) and can be found in critically ill patients with no other secondary HLH features, most likely as a result of increased BM turnover.^{19-26,31,32} It is noteworthy that in the presented MIS-C cases, only a mild hemophagocytosis was observed with almost normal cellularity, while the erythroid and megakaryocytic lineages were found to be prominently affected with almost normal peripheral blood findings. The erythroid hypoplasia with lack of erythroid maturation resembles the primary cytopathic effect of the parvovirus B19 infection on BM,³³ while the mild peripheral anemia could be explained by a compensatory extension of the half-life of red blood cells, as discussed by Patel et al.³⁴

The megakaryocytopoiesis alterations seem to be a peculiar feature of MIS-C differing from sHLH; the BM of sHLS do not show any significant alterations of megakaryocytopoiesis (see Table S2). The hyperplasia of megakaryocytes in addition to the presence of polynuclear cells and immature forms with asynchrony nucleus/cytoplasm could be due to an overstimulation by thrombopoietin as a compensatory mechanism of peripheral autoimmune thrombocytopenia, as

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TABLE 2 Morphologic bone marrow findings in three multisystem inflammatory syndrome in children (MIS-C) patients

	Patient 1	Patient 2	Patient 3	
Cellularity	Decreased	Normal	Normal	
Reticulin fibrosis	Focally increased	Focally increased	Focally increased	
Collagen fibrosis	Not increased	Not increased	Not increased	
Abnormal iron storage	No	No	No	
Erythropoiesis	Decreased	Decreased	Decreased	
	Small island	Small island	Small island	
	Frequent single immature cells	Frequent single immature cells	Frequent single immature cells	
Myelopoiesis	Left shift	Left shift	Left shift	
	Mild disarray	Mild disarray	Mild disarray	
Megakaryocytopoiesis	Increased	Increased	Increased	
	Pleomorphic	Pleomorphic	Pleomorphic	
	Megakaryocytes: small hypolobated and large hypolobated	Megakaryocytes: small hypolobated and large hypolobated	Megakaryocytes: small hypolobated and large hypolobated	
	Micromegakaryocytes	Micromegakaryocytes	Micromegakaryocytes	
	Hyperchromatic nuclei	Hyperchromatic nuclei	Hyperchromatic nuclei	
Lymphocytes	Mild increased	Mild increased	Mild increased	
Plasma cells	Not increased	Not increased	Not increased	
Hematogones	Not increased	Not increased	Not increased	
Histiocytes	Moderately increased	Moderately increased	Moderately increased	
Mast cells	Not increased	Not increased	Not increased	
Vascularity	Not increased	Not increased	Not increased	

CD163 Expression

CD71 Expression



FIGURE 1 The differences in CD163 and CD71 expression pattern on bone marrow in multisystem inflammatory syndrome in children (MIS-C) (A and B) and secondary hemophagocytic lymphohistiocytosis (sHLH) (C and D)

reported in adults.³⁵ Nevertheless, a prominent thrombocytopenia was not observed in our patients. Moreover, the nuclear hyperchromasia could be a sign of virus colonization and virus-induced apoptosis, as seen in HIV infection.³⁶ However, as underlined before, we did



FIGURE 2 The erythroid and megakaryocytic lineages are more affected in multisystem inflammatory syndrome in children (MIS-C) (arrows, A and B) than in secondary hemophagocytic lymphohistiocytosis (sHLH) (C and D), while the hemophagocytosis was less evident in MIS-C (A and B) than in sHLH (arrows, C and D)

not find SARS-CoV-2 RNA or any other virus on BM, and co-infection were excluded. The BM alterations seem to be a consequence of an inflammatory microenvironment as suggested by the plasma cytokine profile.

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As reported, the inflammatory response in MIS-C presents differences from the cytokine storm of severe acute COVID-19, sharing several features with Kawasaki disease, while autoantibodies could be involved in the pathogenesis.¹⁶ Recently, particularly marked elevations of IL-1, IL-10, and TFN α were observed in patients affected by MIS-C compared to patients with severe COVID-19 infection, as observed in our series.^{37,38}

More likely, the aberrant activation of the immune network caused by the cell-extrinsic microenvironmental factors driven by IL-6 and IL-8 plays a fundamental role, leading to a final common pathway of organ damage that involves BM as observed in other conditions such as graftversus-host disease (GvHD). Acute GvHD is a major life-threatening complication after allogeneic hematopoietic cell transplantation, with classical target organs including the intestines, liver, and skin and nonclassical such as BM; since the early 1990s, the cytokine dysregulation in GVDH has been well known, which reflects an immune network and underlying complex mechanism.^{39–43}

MIS-C is a rare complication of SARS-CoV-2 infection in children; great efforts were made to distinguish this entity from other inflammatory states of childhood and clarify its pathogenesis. An immuno-dysregulation, considering both innate and adaptive immunity together with vascular inflammation and endothelial dysfunction, plays a major role with a distinct, but not specific, cytokine signature. Our observations, although limited due to the small sample size, suggest that there are unique features in the BM of patients with MIS-C that are likely secondary to immuno-dysregulation, and there are notable differences in BM features compared to those reported in sHLH.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

AUTHOR CONTRIBUTIONS

Maria Antonietta De Ioris and Rita De Vito conceived the study. Maria Antonietta De Ioris, Alessia Scarselli, Angela Mastronuzzi, and Claudia Bracaglia drafted the manuscript. Rita De Vito, Maria Antonietta De Ioris, Chiara Agrati, and Claudia Bracaglia performed the data analysis. The remaining authors participated in its experimentation and revision of the manuscript. All authors have read and agreed to the published version of the manuscript.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

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