

RESEARCH ARTICLE

Interactome overlap between risk genes of epilepsy and targets of anti-epileptic drugs

Yu-Qin Lv¹*, Xing Wang¹*, Yu-Zhuang Jiao², Yan-Hua Wang¹, Na Wang³, Lei Gao^{4*}, Jing-Jun Zhang^{5*}

1 School of Clinical Medicine, Shandong First Medical University & Shandong Academy of Medical Sciences, Jinan, Shandong, China, **2** Shandong Provincial Qianfoshan Hospital, First Affiliated Hospital of Shandong First Medical University, Jinan, Shandong, China, **3** Department of Internal Medicine, Taishan Vocational College of Nursing, Tai'an, Shandong, China, **4** Department of Bioinformatics, School of Life Sciences, Shandong First Medical University & Shandong Academy of Medical Sciences, Tai'an, Shandong, China, **5** Department of Neurology, The second Affiliated Hospital of Shandong First Medical University, Tai'an, Shandong, China

* These authors contributed equally to this work.

* jjzhang63@126.com (JJZ); gaolei_tsmu@163.com (LG)



OPEN ACCESS

Citation: Lv Y-Q, Wang X, Jiao Y-Z, Wang Y-H, Wang N, Gao L, et al. (2022) Interactome overlap between risk genes of epilepsy and targets of anti-epileptic drugs. *PLoS ONE* 17(8): e0272428. <https://doi.org/10.1371/journal.pone.0272428>

Editor: Nien-Pei Tsai, University of Illinois at Urbana-Champaign, UNITED STATES

Received: February 28, 2022

Accepted: July 19, 2022

Published: August 25, 2022

Peer Review History: PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: <https://doi.org/10.1371/journal.pone.0272428>

Copyright: © 2022 Lv et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: DrugBank: <https://go.drugbank.com/>; Interactome data and Network Calculator: <https://github.com/Haoxiang-Qi/Network-Calculator.git>; EWCE and KI dataset:

Abstract

Anti-epileptic drugs have been used for treating epilepsy for decades, meanwhile, more than one hundred genes have been identified to be associated with risk of epilepsy; however, the interaction mechanism between anti-epileptic drugs and risk genes of epilepsy was still not clearly understood. In this study, we systematically explored the interaction of epilepsy risk genes and anti-epileptic drug targets through a network-based approach. Our results revealed that anti-epileptic drug targets were significantly over-represented in risk genes of epilepsy with 17 overlapping genes and $P\text{-value} = 2.2 \times 10^{-16}$. We identified a significantly localized PPI network with 55 epileptic risk genes and 94 anti-epileptic drug target genes, and network overlap analysis showed significant interactome overlap between risk genes and drug targets with $P\text{-value} = 0.04$. Besides, genes from PPI network were significantly enriched in the co-expression network of epilepsy with 22 enriched genes and $P\text{-value} = 1.3 \times 10^{-15}$; meanwhile, cell type enrichment analysis indicated genes in this network were significantly enriched in 4 brain cell types (Interneuron, Medium Spiny Neuron, CA1 pyramidal Neuron, and Somatosensory pyramidal Neuron). These results provide evidence for significant interactions between epilepsy risk genes and anti-epileptic drug targets from the perspective of network biology.

Introduction

Epilepsy is a collective term for a group of syndromes caused by abnormal discharges of the nervous system, and can cause varying degrees of damage to behavior, cognition, and memory. Previous studies have shown that epilepsy genetic factors contributed a lot to the pathogenesis of epilepsy. Approximately 20–30% of epilepsy cases are caused by acquired conditions such as stroke, tumor or head injury, but there are still 70–80% of cases are considered to be associated with one or more genetic factors [1]. During these decades, whole exome sequencing [2],

<https://github.com/NathanSkene/EWCE>; http://www.hjerling-leffler-lab.org/data/scz_singlecell/.

Funding: This research was supported by Medical Health Science and Technology Project of Shandong Provincial Health Commission (2019WS391), Academic Promotion Program of Shandong First Medical University (2019QL013), National Natural Science Foundation of China (32000477), Shandong provincial Natural Science Foundation (ZR2020MC061), Science and Technology Program of Colleges and Universities in Shandong Province (J15LL07) and the planned project of Tai'an Science and Technology Bureau (2017NS0248).

Competing interests: The authors have declared that no competing interests exist.

genome-wide association studies [3], as well as researches using next generation sequencing technology [4] have identified variations of genes such as sodium channel, potassium channel and GABA_A receptor that were clearly associated with multiple epileptic phenotypes [5].

Among treatments for epilepsy, anti-epileptic drug therapy are the most commonly used, in which most drugs exert their effects by regulating excitatory-inhibitory balance of the brain. Currently, more than one hundred of drug targets, which are involved of voltage-gated ion channels [6], γ -aminobutyric acid energy transfer [7], as well as glutamate energy transfer have been identified. However, the response of patients with epilepsy to anti-epileptic drugs varied greatly, which were largely due to genetic variations that affected both pharmacokinetics and pharmacodynamics of anti-epileptic drugs and risk of epilepsy [8]. Although a variety of drug targets and risk genes of epilepsy have been identified, the interaction between anti-epileptic drugs and risk genes of epilepsy were still not fully understood. With the development of system biology and the accumulation of interactome data, network biology has become an effective approach to explore the underlying biological mechanism of diseases by protein-protein interaction (PPI) networks [9], therefore, in this study, we implemented a network-based approach to explore the interactions between risk genes of epilepsy and anti-epileptic drug target genes. Meanwhile, emerging advances of single-cell RNA sequencing (scRNA-seq) in the central nervous system (CNS) have provide exciting molecular insights into understanding the complexity of the brain, as well as disease-relevant mechanisms by identifying novel cellular subtypes [10]. By applying knowledge of the cellular taxonomy of the brain from single-cell RNA sequencing, previous researchers have performed genetic identification of brain cell-types in schizophrenia [11]. In our study, we explored the molecular mechanism and cellular localization of the interaction network between anti-epileptic drug targets and epileptic risk genes by combining single cell sequencing data and network biological analysis. Our analysis may provide insights for understanding of the genetic basis of epilepsy and the development of anti-epileptic drugs.

Materials and methods

The flowchart of our study was shown in Fig 1.

1. Identification of risk genes of epilepsy

We obtained risk genes of epilepsy identified by both common variants and rare variants, of which common variants are extracted from the largest trans-ethnic meta-analyses of genome-wide association studies currently [12], which included 15,212 individuals with epilepsy and 29,677 controls and identified 16 genome-wide significant loci with P -value $< 5.0 \times 10^{-8}$. Rare variants were identified by exome-sequencing under the largest sample size of 1165 cases and 3877 controls, and their mapped genes were considered as monogenic epilepsy genes [12, 13]. Besides, we also included genes supported by literature retrieval and comprehensive databases providing genetic evidence of risk genes in disease (DisGeNET database [14] and MalaCards database [15]).

2. Identification of anti-epileptic drugs targets

DrugBank (<https://go.drugbank.com/>) is a web-based database containing comprehensive molecular information about drugs, their mechanisms of action, interactions, and their targets [16]. To obtain target genes of anti-epileptic drugs, we searched DrugBank database (Version 5.0) with ATC classification (N03 for anti-epileptic drugs), and a total of 47 anti-epileptic drugs with supported publications were collected. Fisher's Exact Test were performed with R

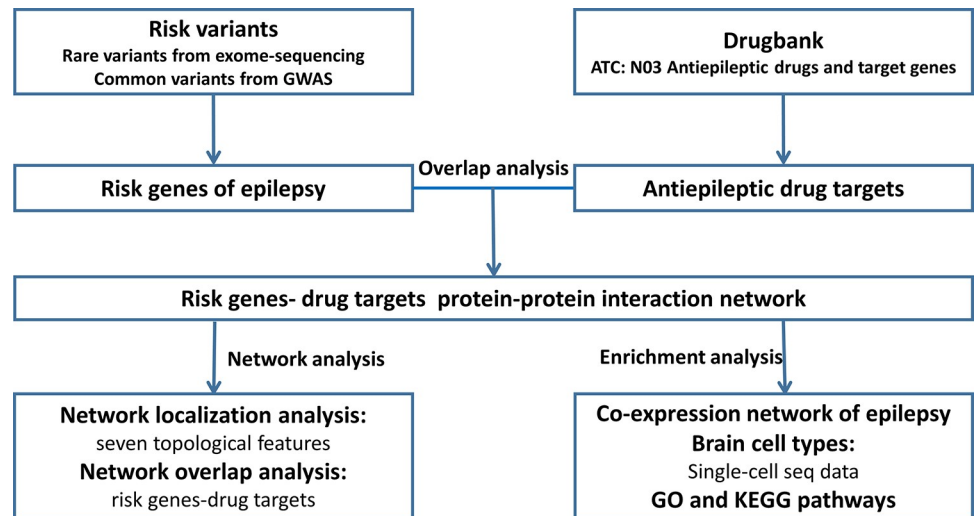


Fig 1. The flowchart of our study. GWAS: Genome-wide association study, GO: Gene Ontology, KEGG: Kyoto Encyclopedia of Genes and Genomes.

<https://doi.org/10.1371/journal.pone.0272428.g001>

(version 4.1.0) to assess whether the targets of anti-epileptic drug were significantly over-represented in epilepsy risk genes.

3. Network topological features of PPI network generated by risk genes of epilepsy and anti-epilepsy drug targets

In order to investigate the network interactions between risk genes of epilepsy and anti-epilepsy drug targets, We first generated a PPI network using risk genes of epilepsy and anti-epilepsy drug targets as input from a comprehensive PPI network data containing 17,252 genes and 471,448 experimental-validated interactions, which combined PPI networks from five databases, including Corum [17], BioPlex [18], CCSB [19], Integral [20] and BioGRID [21], then we calculated seven topological parameters (mean degree, all edges, largest subnetwork, closeness centrality, mean shortest distance, clustering coefficient and betweenness centrality) to evaluate the topological characteristics of this network. To evaluate the significance of these topological characteristics, we carried out permutation test by 5,000 times random sampling with the same number nodes as that in the observed network. This procedure was implemented by the network analysis software Network Calculator (network localization analysis module (<https://github.com/Haoxiang-Qi/Network-Calculator.git>) [22]). Network was visualized by Cytoscape Version 3.8.2 [23].

4. Network overlap analysis between risk genes of epilepsy and anti-epileptic drug targets

In order to evaluate whether there was significant interactions between anti-epileptic drug targets and epileptic risk genes at the network level, we calculated that mean shortest distance within network module of risk gene set of epilepsy (gene set A) as d_A , mean shortest distance within network module of anti-epileptic drugs target genes set (gene set B) as d_B , mean shortest distance between gene set A and gene set B as d_{AB} , then we calculated the network proximity between A and B as $S_{AB} = d_{AB} - (d_A + d_B) / 2$. Using the network overlap analysis module of the Network calculator [22], we evaluated the significance of network overlap by

permutation test of S_AB with a random sampling of 1,000 time using the same number of genes as that in gene set A and B.

5. Enrichment analysis of genes from PPI network in co-expression network of epilepsy

To evaluate whether genes identified by PPI network was enriched in co-expression network of epilepsy, we performed enrichment analysis of genes from our PPI network on a co-expression network of epilepsy including 320 genes, which was identified by gene co-expression network analysis (Weighted Gene Co-expression Network Analysis (WGCNA) [24] and DiffCoEx [25] in the brain reported by a previous study [26]. Fisher's Exact Test were performed with R (version 4.1.0).

6. Expression Weighted Cell Type Enrichment analysis of interaction network analysis between anti-epileptic drug targets and risk genes of epilepsy

To explore whether interaction network between target genes of anti-epileptic and drug risk genes of epilepsy could map on specific brain cell types, we implemented Expression Weighted Cell type Enrichment (EWCE) method, which used single-cell transcriptome dataset to calculate whether the average expression levels of input gene list was significantly stronger than that in randomly generated gene list with the same size as input in each annotated cell type [27]. Moreover, we utilized a superset of brain scRNA-seq data from the Karolinska Institutet (KI, [S1 File](#)) [11, 28–30], which included a total of 9,970 cells annotated with 24 cell types from mouse brain regions of neocortex, hippocampus, hypothalamus, striatum and midbrain, as well as samples enriched for oligodendrocytes, dopaminergic neurons and cortical parvalbuminergic interneurons. Since we used the mouse single-cell transcriptome sequencing data set as the background gene set, we first converted the human interaction network genes into mouse gene form, then we perform EWCE to calculate that significance of expression enrichment for interaction network genes in each brain cell type with 100,000 permutations and Bonferroni adjusted-P-value < 0.05 was considered as significance. R Package EWCE was utilized to perform the analysis and ggplot2 was used to generate graphs [31].

7. Gene ontology and kyoto encyclopedia of genes and genomes enrichment analysis of interaction network between anti-epileptic drug targets and risk genes of epilepsy

We used R Package clusterProfiler [32] to perform functional enrichment analysis for interaction network between anti-epileptic drug targets and risk genes of epilepsy, in which Gene ontology (GO) [33] functional annotation and kyoto encyclopedia of genes and genomes (KEGG) [34] annotation were used and Hypergeometric test was performed, with false discovery rate (FDR) < 0.01 as significance. R package ggplot2 was used to generate graphs [35].

Results

1. Identification of risk genes of Epilepsy

Through literature searches, risk genes of epilepsy were obtained, and the results are indicated in [S1 Table](#). A total of 118 epileptic risk genes including 102 rare variation genes and 16 newly discovered common variation genes were summed up.

2. Identification of targets of anti-epileptic drugs

After searching in DrugBank [16], we retrieved a total of 47 anti-epileptic drugs and 151 targets (S2 Table), of which identified 17 target genes (*CACNA1A*, *CHRNA4*, *CHRNA7*, *GABRA1*, *GABRA2*, *GABRB2*, *GABRG2*, *GRIK1*, *GRIN1*, *GRIN2B*, *KCNQ2*, *KCNQ3*, *SCN1A*, *SCN2A*, *SCN3A*, *SCN8A* and *SCN9A*) were overlapped with risk genes of epilepsy. Fisher’s Exact Test demonstrated a significant over-representation of targets of anti-epileptic drug in the risk genes of epilepsy (odds ratio [OR] = 25.08, $P = 2.2 \times 10^{-16}$).

3. Analysis of PPI network and network characteristics between epilepsy risk genes and anti-epileptic drug target genes

By network construction, we obtained a PPI network containing 247 interactions, including 55 epileptic risk genes and 94 anti-epileptic drug target genes (Fig 2). As shown in Table 1 and Fig 3, by analyzing seven topological characteristics of the PPI network, we identified that six of them were significant, in which all edges, mean degree, largest subnetwork, closeness centrality and clustering coefficient were significantly larger than randomly generated, and mean shortest distance was significantly smaller than randomly generated.

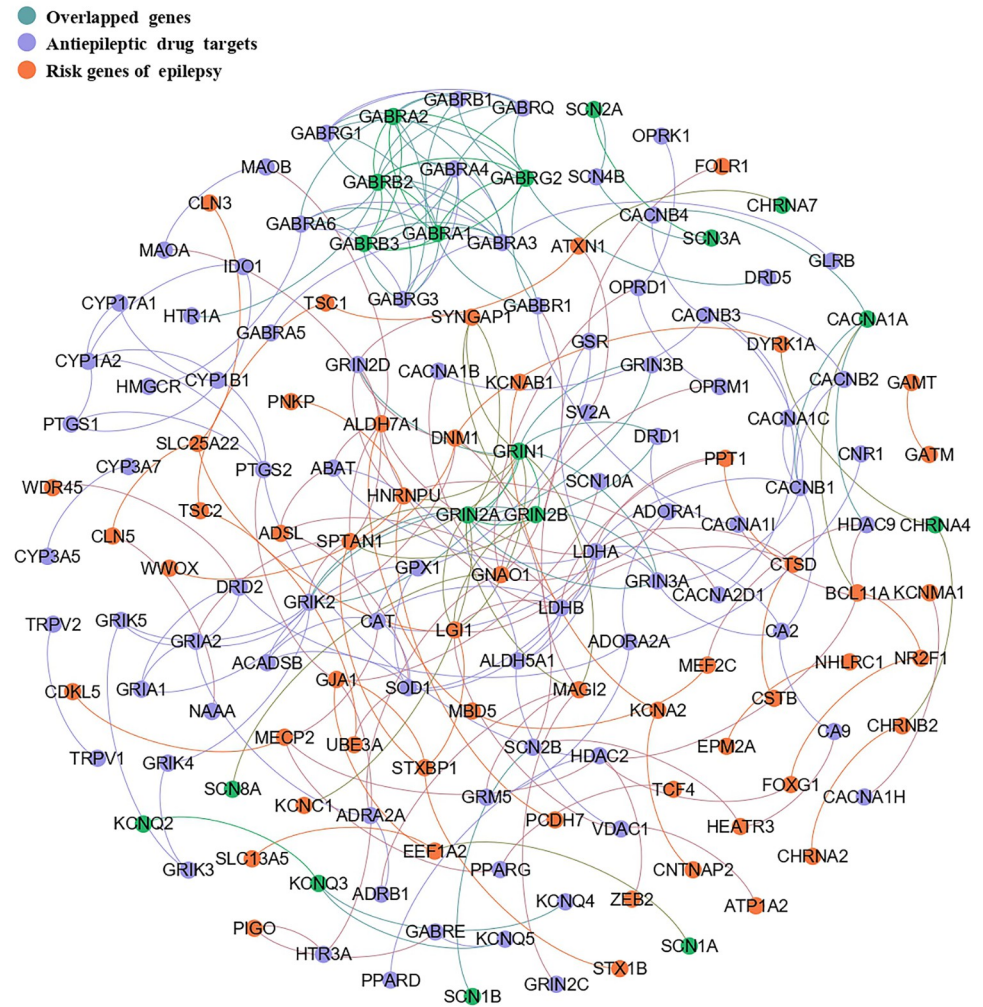


Fig 2. The PPI network of risk genes of epilepsy and anti-epileptic drug target genes.

<https://doi.org/10.1371/journal.pone.0272428.g002>

Table 1. Analysis of network characteristics between epilepsy risk genes and anti-epileptic drug target genes.

Network parameters	Observed	Random_mean	P_value
ALL edges	247	87.648	0.000
Largest subnetwork	133	52.294	0.040
Mean degree	1.05	0.373	0.000
Closeness centrality	0.058	0.017	0.018
Clustering coefficient	0.134	0.028	0.011
Betweenness centrality	0.007	0.028	0.930
Mean shortest distance	1.38	1.655	0.011

<https://doi.org/10.1371/journal.pone.0272428.t001>

4. Network overlap analysis between risk genes of epilepsy and anti-epileptic drug targets

To evaluate whether the network interactions between anti-epileptic drug targets and epileptic risk genes were significant, we performed network overlap analysis and the results showed mean shortest distance of network interactions among risk genes of epilepsy d_A was 1.47, that among anti-epileptic drug target genes d_B was 1.44, that between risk genes of epilepsy and anti-epileptic drug target genes d_{AB} was 1.37, and the network proximity between A and B (S_{AB}) was -0.09, which was significant with P-value of 0.04 calculated by permutation test (Fig 3H), demonstrating a significant interactome overlap between risk genes of epilepsy and anti-epileptic drug target genes.

5. Enrichment analysis in co-expression network of epilepsy

We performed enrichment analysis of 147 genes from our PPI network on 320 genes from a co-expression network of epilepsy [26] (S3 Table), and there were 22 genes (*CACNA1C*, *CACNB2*, *CACNB4*, *CHRN2*, *DNM1*, *EEF1A2*, *GABRA1*, *GABRA3*, *GABRA4*, *GABRB2*, *GABRB3*, *GABRG2*, *GRIN1*, *KCNA2*, *KCNC1*, *KCNQ3*, *SCN1A*, *SCN4B*, *SCN8A*, *SOD1*, *STXBP1*, and *SV2A*) from PPI network were enriched in the co-expression network of epilepsy, and Fisher's Exact Test demonstrated the enrichment was significant (odds ratio [OR] = 11.80, $P = 1.30 \times 10^{-15}$).

6. Brain cell-type enrichment analysis of interaction network between anti-epileptic drug targets and risk genes of epilepsy

To evaluate whether genes in interacted network of anti-epileptic drug targets and risk genes of epilepsy was significantly enriched in specific brain cell types, we performed EWCE in mouse brain scRNA-seq of Karolinska Institute (KI) dataset [11, 28–30]. For KI dataset, among 24 cell types, interacted network between anti-epileptic drug targets and risk genes of epilepsy were significantly enriched in four brain cell types (interneurons, Medium Spiny Neuron, CA1 pyramidal Neuron, and Somatosensory pyramidal Neuron), with Bonferroni-adjusted P-value < 0.05 (Fig 4 and S4 Table).

7. Gene ontology and kyoto encyclopedia of genes and genomes enrichment analysis of interaction network between anti-epileptic drug targets and risk genes of epilepsy

By Gene ontology (GO) and kyoto encyclopedia of genes and genomes (KEGG) pathway enrichment analysis of interaction network between anti-epileptic drug targets and risk genes of epilepsy, a total of 30 and 28 pathways were significantly enriched respectively with

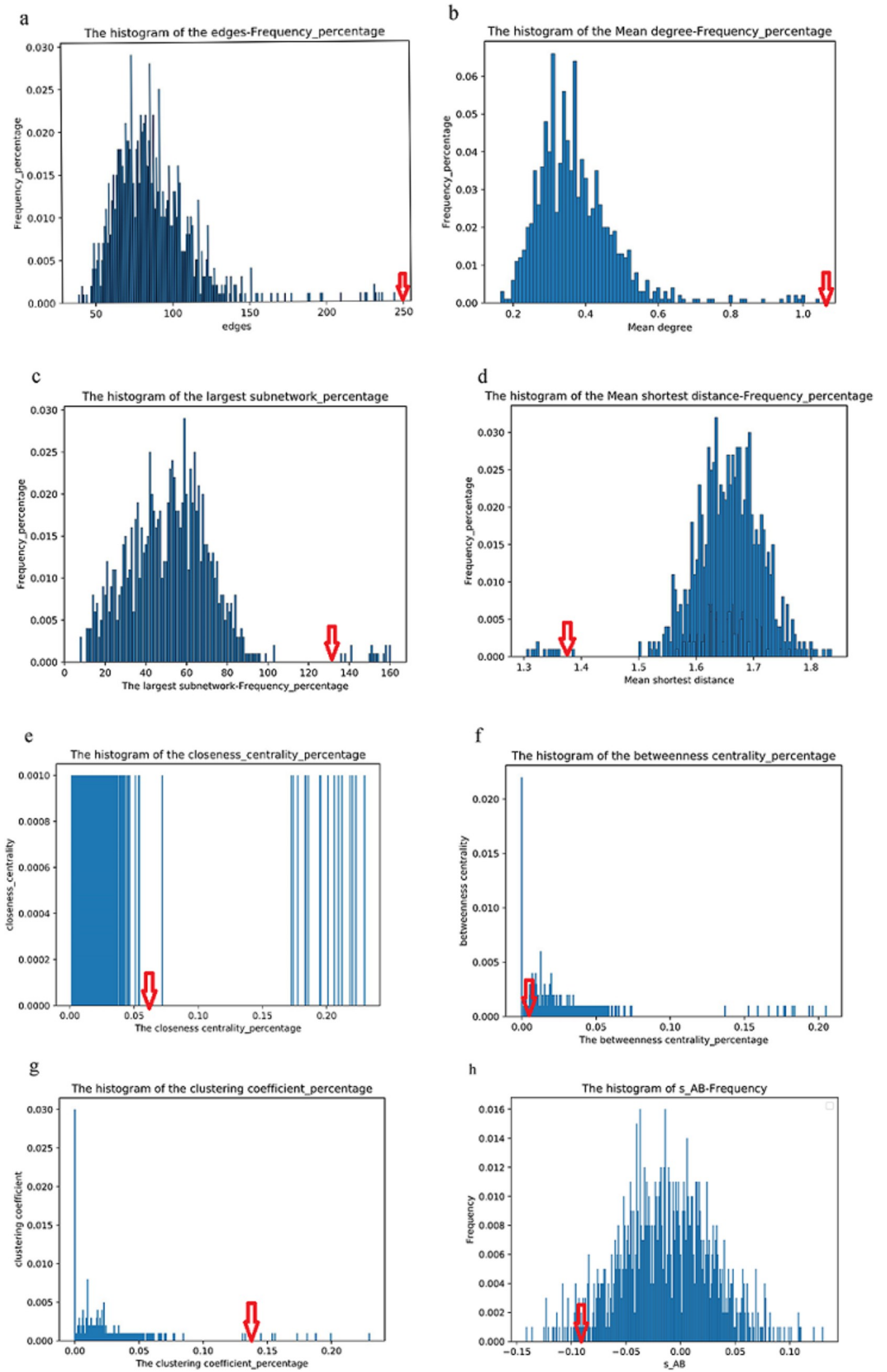


Fig 3. Distribution of random sampling of network topological features.

<https://doi.org/10.1371/journal.pone.0272428.g003>

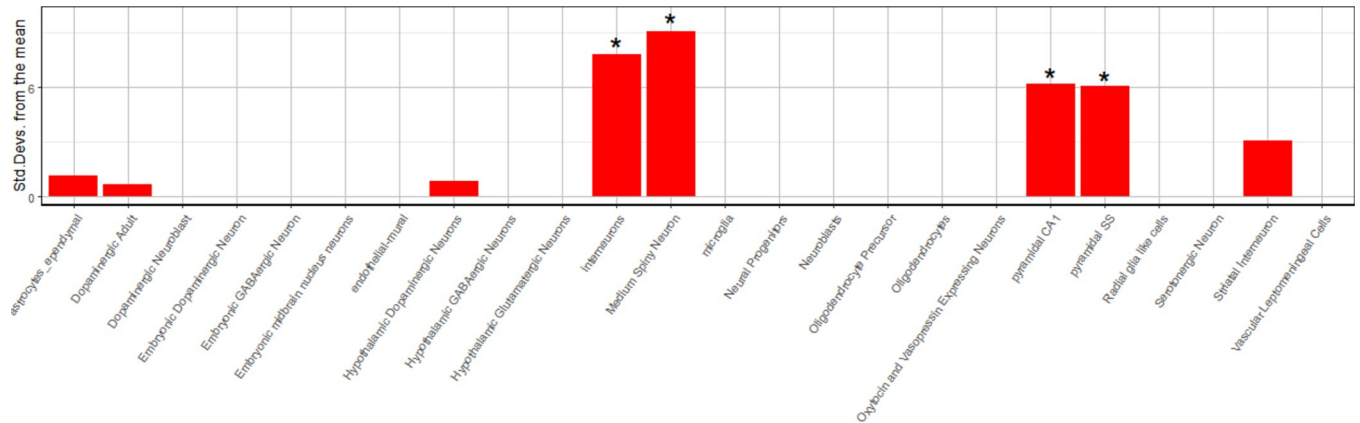


Fig 4. Brain cell-type enrichment analysis of interaction network between anti-epileptic drug targets and risk genes of epilepsy. * Bonferroni adjusted-P-value < 0.05.

<https://doi.org/10.1371/journal.pone.0272428.g004>

FDR < 0.01 (Fig 5). Among these pathways, regulation of postsynaptic membrane potential, regulation of ion trans-membrane transduction, membrane depolarization, postsynaptic chemical synaptic transmission, and GABAergic Synaptic transmission are top significant for GO terms, as well as nicotine addiction, neuroactive ligand-receptor interaction, GABAergic synapse, morphine addiction and retrograde endocannabinoid signaling for KEGG.

Discussion

Epilepsy is a widespread chronic nervous system disease, which affects about 70 million people all over the world [36], during this decade, with the development of sequencing technology, hundreds of risk genes associated with epilepsy have been identified by genetic studies [2], GWAS [12], as well as sequencing [10]. Meanwhile, currently there were more than 40 anti-epileptic drugs based on various target genes used for clinical application [37]. To systematically explore the interactions between risk genes of epilepsy and anti-epileptic drug targets, we used a network-based approach to construct PPI network with interacted risk genes of epilepsy and anti-epileptic drug targets and evaluated the interactome overlap between them. By analyzing seven topological parameters of the PPI network, we identified interactions in the network were significantly higher than randomly generated network with the same size of nodes, similar results were observed in network of schizophrenia and related antipsychotic drugs [38], indicating network constructed by risk genes of epilepsy and anti-epileptic drug targets formed a distinct inner-connected network rather than randomly scattered in the interactome.

To investigate whether there was significant overlap between epilepsy risk genes with anti-epileptic drug targets, we identified 17 overlapped genes (*CACNA1A*, *CHRNA4*, *CHRNA7*, *GABRA1*, *GABRA2*, *GABRB2*, *GABRG2*, *GRIK1*, *GRIN1*, *GRIN2B*, *KCNQ2*, *KCNQ3*, *SCN1A*, *SCN2A*, *SCN3A*, *SCN8A* and *SCN9A*), which showed a significant overlap by Fisher's Exact Test, demonstrating anti-epileptic drug targets were over-represented in risk genes of epilepsy. Meanwhile, we also identified there was significant interactome overlap between them ($P = 0.04$), suggesting the distance between genes of anti-epileptic drug targets and risk genes of epilepsy was significantly closer than the distance among genes in their respective networks. Similar results have been reported in previous studies investigating overlap between the drug targets of antipsychotics and schizophrenia risk genes [38, 39]. These results indicated the genetic overlap between the pathogenesis of epilepsy and the action mechanism of anti-

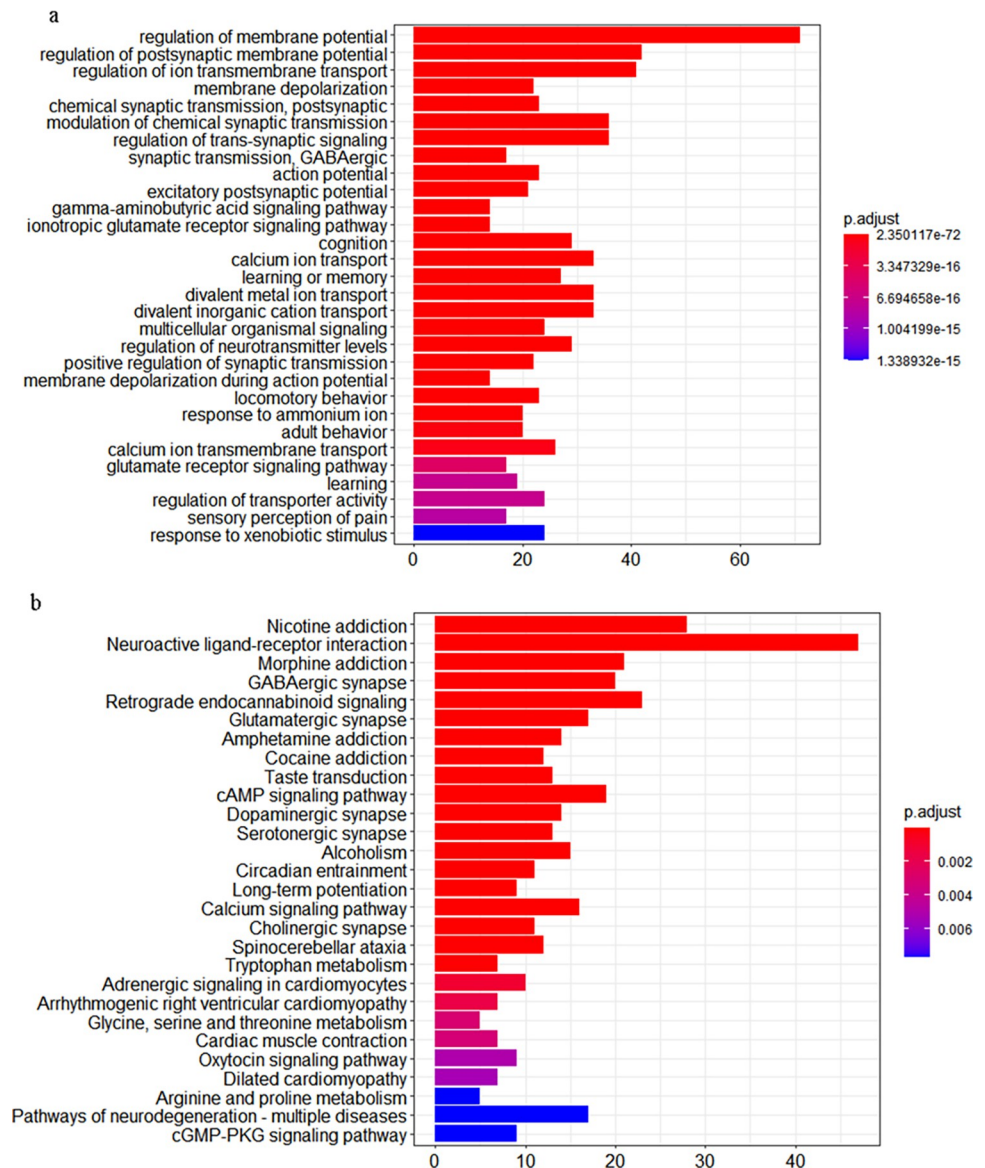


Fig 5. The GO and KEGG pathway enrichment analysis of interaction network between anti-epileptic drug targets and risk genes of epilepsy. *FDR<0.01.

<https://doi.org/10.1371/journal.pone.0272428.g005>

epileptic drugs and provide genetic support evidence for the treatment of epilepsy with anti-epileptic drugs.

Moreover, when we compared genes in this PPI network with genes from a co-expression network of epilepsy [26] (S3 Table), we also identified a significant enrichment of genes in our PPI network in the co-expression network of epilepsy, with 22 gene overlapped, interestingly, all the 22 genes were anti-epileptic drug targets, in which 7 genes were also risk genes (*GABRA1*, *GABRB2*, *GABRG2*, *GRIN1*, *KCNQ3*, *SCN1A*, and *SCN8A*). Voltage-gated sodium channels (VGSCs) play a critical role in generation of action potentials, *SCN1A*, *SCN2A*, *SCN3A*, *SCN8A* and *SCN1B* have been identified to be associated with a spectrum of epilepsy phenotypes and neurodevelopmental disorders [40]. Besides, So far, the most widespread viewpoint considered that GABA_A receptor, as an isomer receptor that binds to GABA, affects

the excitability of nerve cells by stimulating chloride ions influx into the postsynaptic membrane and exerts anti-epileptic effect [41]. It was also reported a personalized therapy in a *GRIN1* mutated girl with intellectual disability and epilepsy [42]. Our results demonstrated that it was important to know the functional effect (Loss-of-function versus Gain-of-function) of a variant for genes which were both risk genes and drug targets to orient therapeutic decisions.

By EWCE in brain scRNA-seq of Karolinska Institute (KI) dataset, including 24 cell types, we found network formed by anti-epileptic drug targets and risk genes of epilepsy were significantly enriched in four brain cell types (interneurons, Medium Spiny Neuron, CA1 pyramidal Neuron, and Somatosensory pyramidal Neuron), These results suggest that the pathogenesis of epilepsy might the result of impaired function of some specific brain cell types [43, 44] and anti-epileptic drugs may play a role in some specific brain cell types [45].

In our study, we explored the network interaction between anti-epileptic drug and risk genes of epilepsy by systematic data collection and integrative analysis. However, there were still some inevitable limitations. First, although we used interactome data by integrating five comprehensive PPI databases, there might still exist interactions between risk genes and drug targets that could not be identified by current interactome data, which are worthy of being explored with the update of interactome data. Second, since the number of cells taken in the single-cell data sets accounts for only a small portion of the whole brain tissue, they may not represent all types of brain cells and need further validation. Third, all the results were obtained by systems biology and network analyses based on data from public databases, which might only reveal underlying mechanisms with currently existing information and need further experimental validation.

Conclusion

In this study, we systematically explored the interaction of epilepsy risk genes and anti-epileptic drug targets through a network-based approach. We identified a significantly localized PPI network with 55 epileptic risk genes and 94 anti-epileptic drug target genes, and network overlap analysis showed significant interactome overlap between risk genes and drug targets. Besides, cell type enrichment analysis indicated genes in this network were significantly enriched in 4 brain cell types (Interneuron, Medium Spiny Neuron, CA1 pyramidal Neuron, and Somatosensory pyramidal Neuron). These results provide evidence for interactions between epilepsy risk genes and anti-epileptic drug targets from the perspective of network biology.

Supporting information

S1 Table. The risks genes of epilepsy.

(DOCX)

S2 Table. The target genes of anti-epileptic drugs.

(DOCX)

S3 Table. Genes of co-expression network of epilepsy.

(DOCX)

S4 Table. Brain cell-types enrichment analysis of interaction network of risk gene of epilepsy and anti-epileptic drug targets.

(DOCX)

S1 File. Raw KI dataset for analysis of cell types enriched by anti-epileptic drugs.
(ZIP)

Author Contributions

Data curation: Yu-Qin Lv, Xing Wang.

Resources: Lei Gao, Jing-Jun Zhang.

Supervision: Lei Gao, Jing-Jun Zhang.

Writing – original draft: Yu-Qin Lv, Xing Wang, Yu-Zhuang Jiao, Yan-Hua Wang, Na Wang.

Writing – review & editing: Yu-Qin Lv, Xing Wang.

References

1. Hildebrand MS, Dahl HH, Damiano JA, Smith RJ, Scheffer IE, Berkovic SF. Recent advances in the molecular genetics of epilepsy. *Journal of medical genetics*. 2013; 50(5):271–9. Epub 2013/03/08. <https://doi.org/10.1136/jmedgenet-2012-101448> PMID: 23468209.
2. Rochtus A, Olson HE, Smith L, Keith LG, El Achkar C, Taylor A, et al. Genetic diagnoses in epilepsy: The impact of dynamic exome analysis in a pediatric cohort. *Epilepsia*. 2020; 61(2):249–58. <https://doi.org/10.1111/epi.16427> PMID: 31957018; PubMed Central PMCID: PMC7404709.
3. International League Against Epilepsy Consortium on Complex E. Genome-wide mega-analysis identifies 16 loci and highlights diverse biological mechanisms in the common epilepsies. *Nat Commun*. 2018; 9(1):5269. <https://doi.org/10.1038/s41467-018-07524-z> PMID: 30531953; PubMed Central PMCID: PMC6288131.
4. Dixit AB, Banerjee J, Srivastava A, Tripathi M, Sarkar C, Kakkar A, et al. RNA-seq analysis of hippocampal tissues reveals novel candidate genes for drug refractory epilepsy in patients with MTL-EHS. *Genomics*. 2016; 107(5):178–88. <https://doi.org/10.1016/j.ygeno.2016.04.001> PMID: 27094248.
5. Helbig I, Scheffer IE, Mulley JC, Berkovic SF. Navigating the channels and beyond: unravelling the genetics of the epilepsies. *The Lancet Neurology*. 2008; 7(3):231–45. Epub 2008/02/16. [https://doi.org/10.1016/S1474-4422\(08\)70039-5](https://doi.org/10.1016/S1474-4422(08)70039-5) PMID: 18275925.
6. Taylor CP, Angelotti T, Fauman E. Pharmacology and mechanism of action of pregabalin: the calcium channel alpha2-delta (alpha2-delta) subunit as a target for antiepileptic drug discovery. *Epilepsy research*. 2007; 73(2):137–50. Epub 2006/11/28. <https://doi.org/10.1016/j.eplepsyres.2006.09.008> PMID: 17126531.
7. Nickels KC, Wirrell EC. Stiripentol in the Management of Epilepsy. *CNS drugs*. 2017; 31(5):405–16. Epub 2017/04/24. <https://doi.org/10.1007/s40263-017-0432-1> PMID: 28434133.
8. Balestrini S, Sisodiya SM. Pharmacogenomics in epilepsy. *Neuroscience letters*. 2018; 667:27–39. Epub 2017/01/14. <https://doi.org/10.1016/j.neulet.2017.01.014> PMID: 28082152; PubMed Central PMCID: PMC5846849.
9. Menche J, Sharma A, Kitsak M, Ghiassian SD, Vidal M, Loscalzo J, et al. Disease networks. Uncovering disease-disease relationships through the incomplete interactome. *Science*. 2015; 347(6224):1257601. Epub 2015/02/24. <https://doi.org/10.1126/science.1257601> PMID: 25700523; PubMed Central PMCID: PMC4435741.
10. Ofengeim D, Giagtzoglou N, Huh D, Zou C, Yuan J. Single-Cell RNA Sequencing: Unraveling the Brain One Cell at a Time. *Trends in molecular medicine*. 2017; 23(6):563–76. Epub 2017/05/16. <https://doi.org/10.1016/j.molmed.2017.04.006> PMID: 28501348; PubMed Central PMCID: PMC5531055.
11. Skene NG, Bryois J, Bakken TE, Breen G, Crowley JJ, Gaspar HA, et al. Genetic identification of brain cell types underlying schizophrenia. *Nature genetics*. 2018; 50(6):825–33. Epub 2018/05/23. <https://doi.org/10.1038/s41588-018-0129-5> PMID: 29785013; PubMed Central PMCID: PMC6477180.
12. Genome-wide mega-analysis identifies 16 loci and highlights diverse biological mechanisms in the common epilepsies. *Nature communications*. 2018; 9(1):5269. Epub 2018/12/12. <https://doi.org/10.1038/s41467-018-07524-z> PMID: 30531953; PubMed Central PMCID: PMC6288131.
13. Ultra-rare genetic variation in common epilepsies: a case-control sequencing study. *The Lancet Neurology*. 2017; 16(2):135–43. Epub 2017/01/20. [https://doi.org/10.1016/S1474-4422\(16\)30359-3](https://doi.org/10.1016/S1474-4422(16)30359-3) PMID: 28102150.

14. Piñero J, Bravo À, Queralt-Rosinach N, Gutiérrez-Sacristán A, Deu-Pons J, Centeno E, et al. DisGeNET: a comprehensive platform integrating information on human disease-associated genes and variants. *Nucleic acids research*. 2017; 45(D1):D833–d9. Epub 2016/12/08. <https://doi.org/10.1093/nar/gkw943> PMID: 27924018; PubMed Central PMCID: PMC5210640.
15. Rappaport N, Twik M, Plaschkes I, Nudel R, Iny Stein T, Levitt J, et al. MalaCards: an amalgamated human disease compendium with diverse clinical and genetic annotation and structured search. *Nucleic acids research*. 2017; 45(D1):D877–d87. Epub 2016/12/03. <https://doi.org/10.1093/nar/gkw1012> PMID: 27899610; PubMed Central PMCID: PMC5210521.
16. Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, et al. DrugBank 5.0: a major update to the DrugBank database for 2018. *Nucleic acids research*. 2018; 46(D1):D1074–d82. Epub 2017/11/11. <https://doi.org/10.1093/nar/gkx1037> PMID: 29126136; PubMed Central PMCID: PMC5753335.
17. Ruepp A, Brauner B, Dunger-Kaltenbach I, Frishman G, Montrone C, Stransky M, et al. CORUM: the comprehensive resource of mammalian protein complexes. *Nucleic acids research*. 2008; 36(Database issue):D646–50. Epub 2007/10/30. <https://doi.org/10.1093/nar/gkm936> PMID: 17965090; PubMed Central PMCID: PMC2238909.
18. Huttlin EL, Ting L, Bruckner RJ, Gebreab F, Gygi MP, Szpyt J, et al. The BioPlex Network: A Systematic Exploration of the Human Interactome. *Cell*. 2015; 162(2):425–40. Epub 2015/07/18. <https://doi.org/10.1016/j.cell.2015.06.043> PMID: 26186194; PubMed Central PMCID: PMC4617211.
19. Rolland T, Taşan M, Charloteaux B, Pevzner SJ, Zhong Q, Sahni N, et al. A proteome-scale map of the human interactome network. *Cell*. 2014; 159(5):1212–26. Epub 2014/11/25. <https://doi.org/10.1016/j.cell.2014.10.050> PMID: 25416956; PubMed Central PMCID: PMC4266588.
20. Orchard S, Ammari M, Aranda B, Breuza L, Briganti L, Broackes-Carter F, et al. The MIntAct project—IntAct as a common curation platform for 11 molecular interaction databases. *Nucleic acids research*. 2014; 42(Database issue):D358–63. Epub 2013/11/16. <https://doi.org/10.1093/nar/gkt1115> PMID: 24234451; PubMed Central PMCID: PMC3965093.
21. Oughtred R, Stark C, Breitkreutz BJ, Rust J, Boucher L, Chang C, et al. The BioGRID interaction database: 2019 update. *Nucleic acids research*. 2019; 47(D1):D529–d41. Epub 2018/11/27. <https://doi.org/10.1093/nar/gky1079> PMID: 30476227; PubMed Central PMCID: PMC6324058.
22. Qi HX, Shen QD, Zhao HY, Qi GZ, Gao L. Network-based analysis revealed significant interactions between risk genes of severe COVID-19 and host genes interacted with SARS-CoV-2 proteins. *Briefings in bioinformatics*. 2021. Epub 2021/09/19. <https://doi.org/10.1093/bib/bbab372> PMID: 34535795.
23. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, et al. Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome research*. 2003; 13(11):2498–504. Epub 2003/11/05. <https://doi.org/10.1101/gr.1239303> PMID: 14597658; PubMed Central PMCID: PMC403769.
24. Langfelder P, Horvath S. WGCNA: an R package for weighted correlation network analysis. *BMC bioinformatics*. 2008; 9:559. Epub 2008/12/31. <https://doi.org/10.1186/1471-2105-9-559> PMID: 19114008; PubMed Central PMCID: PMC2631488.
25. Tesson BM, Breitling R, Jansen RC. DiffCoEx: a simple and sensitive method to find differentially coexpressed gene modules. *BMC bioinformatics*. 2010; 11:497. Epub 2010/10/12. <https://doi.org/10.1186/1471-2105-11-497> PMID: 20925918; PubMed Central PMCID: PMC2976757.
26. Delahaye-Duriez A, Srivastava P, Shkura K, Langley SR, Laaniste L, Moreno-Moral A, et al. Rare and common epilepsies converge on a shared gene regulatory network providing opportunities for novel antiepileptic drug discovery. *Genome biology*. 2016; 17(1):245. Epub 2016/12/14. <https://doi.org/10.1186/s13059-016-1097-7> PMID: 27955713; PubMed Central PMCID: PMC5154105.
27. Skene NG, Grant SG. Identification of Vulnerable Cell Types in Major Brain Disorders Using Single Cell Transcriptomes and Expression Weighted Cell Type Enrichment. *Frontiers in neuroscience*. 2016; 10:16. Epub 2016/02/10. <https://doi.org/10.3389/fnins.2016.00016> PMID: 26858593; PubMed Central PMCID: PMC4730103.
28. Zeisel A, Muñoz-Manchado AB, Codeluppi S, Lönnerberg P, La Manno G, Jureus A, et al. Brain structure. Cell types in the mouse cortex and hippocampus revealed by single-cell RNA-seq. *Science*. 2015; 347(6226):1138–42. Epub 2015/02/24. <https://doi.org/10.1126/science.aaa1934> PMID: 25700174.
29. La Manno G, Gyllborg D, Codeluppi S, Nishimura K, Salto C, Zeisel A, et al. Molecular Diversity of Mid-brain Development in Mouse, Human, and Stem Cells. *Cell*. 2016; 167(2):566–80.e19. Epub 2016/10/08. <https://doi.org/10.1016/j.cell.2016.09.027> PMID: 27716510; PubMed Central PMCID: PMC5055122.
30. Romanov RA, Zeisel A, Bakker J, Girach F, Hellysaz A, Tomer R, et al. Molecular interrogation of hypothalamic organization reveals distinct dopamine neuronal subtypes. *Nature neuroscience*. 2017; 20(2):176–88. Epub 2016/12/20. <https://doi.org/10.1038/nn.4462> PMID: 27991900.

31. Ginestet C. ggplot2: Elegant Graphics for Data Analysis. *J R Stat Soc Ser A-Stat Soc.* 2011; 174:245–. https://doi.org/10.1111/j.1467-985X.2010.00676_9.x WOS:000285969600026.
32. Yu G, Wang LG, Han Y, He QY. clusterProfiler: an R package for comparing biological themes among gene clusters. *Omics: a journal of integrative biology.* 2012; 16(5):284–7. Epub 2012/03/30. <https://doi.org/10.1089/omi.2011.0118> PMID: 22455463; PubMed Central PMCID: PMC3339379.
33. Gene Ontology Consortium: going forward. *Nucleic acids research.* 2015; 43(Database issue):D1049–56. Epub 2014/11/28. <https://doi.org/10.1093/nar/gku1179> PMID: 25428369; PubMed Central PMCID: PMC4383973.
34. Kanehisa M, Furumichi M, Tanabe M, Sato Y, Morishima K. KEGG: new perspectives on genomes, pathways, diseases and drugs. *Nucleic acids research.* 2017; 45(D1):D353–d61. Epub 2016/12/03. <https://doi.org/10.1093/nar/gkw1092> PMID: 27899662; PubMed Central PMCID: PMC5210567.
35. Ito K, Murphy D. Application of ggplot2 to Pharmacometric Graphics. *CPT: pharmacometrics & systems pharmacology.* 2013; 2(10):e79. Epub 2013/10/18. <https://doi.org/10.1038/psp.2013.56> PMID: 24132163; PubMed Central PMCID: PMC3817376.
36. Thijs RD, Surges R, O'Brien TJ, Sander JW. Epilepsy in adults. *Lancet (London, England).* 2019; 393(10172):689–701. Epub 2019/01/29. [https://doi.org/10.1016/S0140-6736\(18\)32596-0](https://doi.org/10.1016/S0140-6736(18)32596-0) PMID: 30686584.
37. Löscher W, Potschka H, Sisodiya SM, Vezzani A. Drug Resistance in Epilepsy: Clinical Impact, Potential Mechanisms, and New Innovative Treatment Options. *Pharmacological reviews.* 2020; 72(3):606–38. Epub 2020/06/17. <https://doi.org/10.1124/pr.120.019539> PMID: 32540959; PubMed Central PMCID: PMC7300324.
38. Kauppi K, Rosenthal SB, Lo MT, Sanyal N, Jiang M, Abagyan R, et al. Revisiting Antipsychotic Drug Actions Through Gene Networks Associated With Schizophrenia. *The American journal of psychiatry.* 2018; 175(7):674–82. Epub 2018/03/03. <https://doi.org/10.1176/appi.ajp.2017.17040410> PMID: 29495895; PubMed Central PMCID: PMC6028303.
39. Ruderfer DM, Charney AW, Readhead B, Kidd BA, Kähler AK, Kenny PJ, et al. Polygenic overlap between schizophrenia risk and antipsychotic response: a genomic medicine approach. *The lancet Psychiatry.* 2016; 3(4):350–7. Epub 2016/02/27. [https://doi.org/10.1016/S2215-0366\(15\)00553-2](https://doi.org/10.1016/S2215-0366(15)00553-2) PMID: 26915512; PubMed Central PMCID: PMC4982509.
40. Musto E, Gardella E, Møller RS. Recent advances in treatment of epilepsy-related sodium channelopathies. *European journal of paediatric neurology: EJPN: official journal of the European Paediatric Neurology Society.* 2020; 24:123–8. Epub 2020/01/01. <https://doi.org/10.1016/j.ejpn.2019.12.009> PMID: 31889633.
41. Diniz TC, Silva JC, de Lima-Saraiva SR, Ribeiro FP, Pacheco AG, de Freitas RM, et al. The role of flavonoids on oxidative stress in epilepsy. *Oxidative medicine and cellular longevity.* 2015; 2015:171756. Epub 2015/02/06. <https://doi.org/10.1155/2015/171756> PMID: 25653736; PubMed Central PMCID: PMC4306219.
42. Papa FT, Mancardi MM, Frullanti E, Fallerini C, Della Chiara V, Zalba-Jadraque L, et al. Personalized therapy in a GRIN1 mutated girl with intellectual disability and epilepsy. *Clin Dysmorphol.* 2018; 27(1):18–20. Epub 2017/12/02. <https://doi.org/10.1097/MCD.0000000000000205> PMID: 29194067.
43. Liu YQ, Yu F, Liu WH, He XH, Peng BW. Dysfunction of hippocampal interneurons in epilepsy. *Neurosci Bull.* 2014; 30(6):985–98. <https://doi.org/10.1007/s12264-014-1478-4> PMID: 25370443; PubMed Central PMCID: PMC5562563.
44. Righes Marafija J, Vendramin Pasquetti M, Calcagnotto ME. GABAergic interneurons in epilepsy: More than a simple change in inhibition. *Epilepsy & behavior: E&B.* 2021; 121(Pt B):106935. Epub 2020/02/10. <https://doi.org/10.1016/j.yebeh.2020.106935> PMID: 32035792.
45. Ghiglieri V, Sgobio C, Patassini S, Bagetta V, Fejtova A, Giampa C, et al. TrkB/BDNF-dependent striatal plasticity and behavior in a genetic model of epilepsy: modulation by valproic acid. *Neuropsychopharmacology.* 2010; 35(7):1531–40. <https://doi.org/10.1038/npp.2010.23> PMID: 20200504; PubMed Central PMCID: PMC3055450.