

# Predictors of Visceral Leishmaniasis Relapse in HIV-Infected Patients: A Systematic Review

Gláucia F. Cota<sup>1,3</sup>\*, Marcos R. de Sousa<sup>2</sup>, Ana Rabello<sup>1</sup>

1 Post-Graduate Program in Health Sciences, René Rachou Institute, Fundação Oswaldo Cruz, Belo Horizonte, Minas Gerais, Brazil, 2 Post-Graduate Program in Adult Health Sciences, Federal University of Minas Gerais, Belo Horizonte, Minas Gerais, Brazil, 3 Eduardo de Menezes Hospital, Fundação Hospitalar do Estado de Minas Gerais-FHEMIG, Belo Horizonte, Minas Gerais, Brazil

#### **Abstract**

**Background and Objectives:** Visceral leishmaniasis (VL) is a common complication in AIDS patients living in *Leishmania*-endemic areas. Although antiretroviral therapy has changed the clinical course of HIV infection and its associated illnesses, the prevention of VL relapses remains a challenge for the care of HIV and *Leishmania* co-infected patients. This work is a systematic review of previous studies that have described predictors of VL relapse in HIV-infected patients.

**Review Methods:** We searched the electronic databases of MEDLINE, LILACS, and the Cochrane Central Register of Controlled Trials. Studies were selected if they included HIV-infected individuals with a VL diagnosis and patient follow-up after the leishmaniasis treatment with an analysis of the clearly defined outcome of prediction of relapse.

Results: Eighteen out 178 studies satisfied the specified inclusion criteria. Most patients were males between 30 and 40 years of age, and HIV transmission was primarily via intravenous drug use. Previous VL episodes were identified as risk factors for relapse in 3 studies. Two studies found that baseline CD4+ T cell count above 100 cells/mL was associated with a decreased relapse rate. The observation of an increase in CD4+ T cells at patient follow-up was associated with protection from relapse in 5 of 7 studies. Meta-analysis of all studies assessing secondary prophylaxis showed significant reduction of VL relapse rate following prophylaxis. None of the five observational studies evaluating the impact of highly active antiretroviral therapy use found a reduction in the risk of VL relapse upon patient follow-up.

Conclusion: Some predictors of VL relapse could be identified: a) the absence of an increase in CD4+ cells at follow-up; b) lack of secondary prophylaxis; and c) previous history of VL relapse. CD4+ counts below 100 cells/mL at the time of primary VL diagnosis may also be a predictive factor for VL relapse.

Citation: Cota GF, de Sousa MR, Rabello A (2011) Predictors of Visceral Leishmaniasis Relapse in HIV-Infected Patients: A Systematic Review. PLoS Negl Trop Dis 5(6): e1153. doi:10.1371/journal.pntd.0001153

Editor: Charles L. Jaffe, Hebrew University-Hadassah Medical School, Israel

Received December 10, 2010; Accepted March 7, 2011; Published June 7, 2011

**Copyright:** © 2011 Cota et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** Ana Rabelo received financial support from CNPq (National Counsel of Technological and Scientific Development) and Oswaldo Cruz Foundation - FIOCRUZ. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

\* E-mail: glauciacota@uol.com.br

#### Introduction

Visceral leishmaniasis (VL) and human immunodeficiency virus (HIV) co-infection has emerged as a serious disease pattern [1,2]. HIV infection increases the risk of developing VL by 100 to 2,320 times in endemic areas [3,4] and, on the other hand, VL promotes the clinical progression of HIV disease and the development of AIDS-defining conditions [5]. Both infections switch the predominantly cellular immunological response from Th1 to Th2 through complex cytokine mediated mechanisms leading to a synergistic detrimental effect on the cellular immune response [6,7,8]. Other important findings related to HIV-Leishmania co-infection is a reduction in therapeutic response and high rate of relapse, which is the clinical deterioration after clinical improvement, observed in 25-61% of patients [9,10,11,12]. Although the term recurrence has also been used as synonym for relapse, recurrence applies to the finding of a parasite repeatedly. It is important to emphasize that neither of these two terms distinguishes parasitological persistence from re-infection.

The poor therapeutic outcome, the high rate of relapse, the poliparasitic nature of VL in HIV-infected persons, as well as the atypical manifestations of the disease and the impaired access to health-care resources make HIV-infected individuals prone to enlarge the number of human reservoirs [13]. This concern is of utmost importance in Asia, where HIV and *Leishmania* coinfections are increasingly being reported in countries that are also facing parasite resistance to antimonial drugs [14].

Recent changes in the epidemiological patterns of HIV and *Leishmania* infections are likely to lead to a greater degree of overlap and greater risk of co-infection and they justify increased alertness. From a global epidemiologic viewpoint, two parallel trends are alarming: the ruralization of the HIV pandemic and the urbanization and spread of VL [1,15]. World Health Organization (WHO) [16] reports that the public health impact of leishmaniasis worldwide has been grossly underestimated for many years because notification was compulsory in only 32 of the 88 countries where 350 million people were at risk. The reported global incidence of co-infection is likely underestimated either because

# **Author Summary**

Visceral leishmaniasis (VL) is the most serious form of an insect-transmitted parasitic disease prevalent in 70 countries. The disease is caused by species of the L. donovani complex found in different geographical regions. These parasites have substantially different clinical, drug susceptibility and epidemiological characteristics. According to data from the World Health Organization, the areas where HIV-Leishmania co-infection is distributed are extensive. HIV infection increases the risk of developing VL, reduces the likelihood of a therapeutic response, and greatly increases the probability of relapse. A better understanding of the factors promoting relapses is essential; therefore we performed a systematic review of articles involving all articles assessing the predictors of VL relapse in HIVinfected individuals older than 14 years of age. Out of 178 relevant articles, 18 met the inclusion criteria and in total, data from 1017 patients were analyzed. We identified previous episodes of VL relapse, CD4+ lymphocyte count fewer than 100 cells/mL at VL diagnosis, and the absence of an increase in CD4+ counts at follow-up as major factors associated with VL relapse. Knowledge of relapse predictors can help to identify patients with different degrees of risk, facilitate and direct prophylaxis choices, and aid in patient counseling.

VL has not been included in the list of AIDS related opportunistic infection in all endemic areas.

Before the widespread use of antiretroviral therapy, such coinfection was common in Europe [5]. The co-infection is now becoming proportionately more prominent in areas with poor access to antiretrovirals, such as Africa. In areas where it is available, highly active antiretroviral therapy (HAART) has changed the course of the HIV/AIDS epidemic and the outcome of associated opportunistic infections. However, evidence of relapse rate reduction in patients using HAART is conflicting [17]. This work is a systematic review of studies describing the predictors of VL relapse in HIV-infected patients.

### **Methods**

#### Search Strategy and Selection Criteria

This review was conducted on all papers published before July, 31, 2010. To ensure scientific rigour, the Preferred Reporting of Systematic Reviews and Meta-Analysis (PRISMA) guidelines [18] were used for systematic data synthesis. Studies were identified by a Medline/PubMed search using a combination of terms that has been shown to maximize sensitivity [19]. The search terms used are shown in Figure 1. The LILACS and Cochrane databases were used for literature review using a Boolean combination of search terms. Additional reports were located using a manual search of references from retrieved papers. Two independent reviewers (GFC and MRS) initially checked the lists of titles and abstracts identified by this search to determine whether an article contained relevant data. Studies were considered eligible if they were presented in an original article, examined HIV-infected individuals over 14 years of age with a VL diagnosis, included follow-up after the leishmaniasis treatment and clearly analyzed predictors of relapse.

There were no restrictions on the publication language, date of publication, use of secondary prophylaxis, or duration of follow-up in the study. We excluded studies evaluating fewer than ten cases and studies evaluating mixed populations of HIV-infected and uninfected subjects unless separated results for HIV patients were identified. The selected articles were read in full to confirm eligibility.

Data were extracted directly from the full-length articles into structured tables containing all of the descriptive variables and relevant outcomes. The following information was extracted: country and period of enrollment; sample size; clinical characteristics of the included patients; study design; the number of excluded patients if specified; statistical analyses utilized; duration of follow-up and number of subjects lost to follow-up; outcome of interest; prognostic variables assessed in each study and quality of the regression model [20,21,22]. When data were available tests required for completion of the tables were performed. To summarize the results regarding secondary prophylaxis, the software Comprehensive Meta-Analysis Version 2.2.048 was used.

Figure 1. Terms used in Medline/PubMed search. doi:10.1371/journal.pntd.0001153.q001



### **Results**

Our selection process is illustrated in Figure 2. Of 178 articles, 136 were excluded because they did not meet the inclusion criteria following reading of titles and/or abstracts. Twenty more articles were excluded after reading the entire article: six analyzed less than ten patients [23,24,25,26,27,28,29], one was a review [30], and thirteen did not evaluate the risk on relapse of different predictors [3,28,31,32,33,34,35,36,37,38,39,40,41]. Four studies [42,43,44,45] were excluded because they included cases published elsewhere [10,46,47]. Thus, 18 studies (Table S1) satisfied the specified inclusion and exclusion criteria and constituted the basis of this investigation.

#### Studies and Patients

Table S1 summarizes the characteristics of the 1017 patients encompassed by the 18 included studies. The year of study publication ranged from 1989 to 2008. The design of 8 of the studies examined was prospective. Fourteen studies were reported

in Spain, two in Italy, and one in Ethiopia and one in France. Eight studies had an enrollment period exclusively after 1996, when HAART became available. Twelve papers stated the proportions of patients receiving HAART involving two nucleoside reverse transcriptase inhibitors and one or two protease inhibitors or non-nucleosides reverse transcriptase inhibitors at VL diagnosis or at relapse or both.

A large proportion of the patients in these studies (87.7%) were male and most were young adults; the median or mean ages reported varied from 27 to 37 years (Table S2). In the 14 studies in which patients' presumed transmission route was known, 72.3% (420/581) of the infections were likely due to intravenous drug use. The median CD4+ T lymphocyte count ranged from 11 to 82 cells/mL. Most patients had an AIDS-defining condition [48] at the time of VL diagnosis (332/572, 58% of patients).

In the majority of the studies, the diagnosis of VL was established by direct demonstration of amastigotes (by cytological study of Wright stains) or by the observation of promastigote growth in samples cultured in specific media. In one study [49],

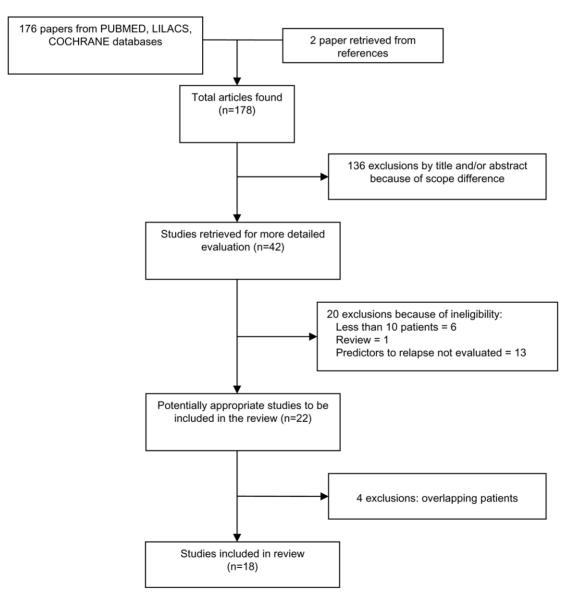


Figure 2. Study selection process. doi:10.1371/journal.pntd.0001153.g002



the VL diagnosis was supported either by positive results from *Leishmania*-specific PCR (polymerase chain reaction) of peripheral blood or bone marrow samples. Three studies [47,50,51] also included patients diagnosed by serologic tests (direct agglutination, indirect immunofluorescence or rK-39 dipsticks).

The drug used in the treatment of the primary episode of VL was reported for 89% of the treated patients. Of this total, 73.4% of cases (733 patients) were treated with pentavalent antimonial drugs, 12.4% with amphotericin B deoxycholate (124 patients), and 2.1% (21 patients) received amphotericin in lipid formulations. A minority of patients (1.2%) received pentamidine isethionate and three papers included patients treated with miltefosine [47] or unconventional regimens such as a combination of allopurinol with an azole compound [50,52]. A test of cure (staining with Giemsa stain and parasite culture or PCR) at the end of treatment was carried out in 8 of 18 studies. In most of these studies, this control was performed for patients whose clinical response was uncertain. Secondary prophylaxis for leishmaniasis was reported in eleven studies.

Three studies explored the impact of mono or dual antiretroviral therapy at VL diagnosis [47] or during the follow-up [50,53] on relapse. Only one [47] of these studies demonstrated a reduction in relapse rate compared with patients who did not undergo retroviral therapy. Similarly, only one [49] of four studies [10,49,51,54] that followed patients on HAART at VL diagnosis reported a reduction in relapse rate. HAART use on follow-up has also been studied in relation to risk of relapse and none of the five [9,51,52,54,55] studies showed reduction on VL relapse rate.

Two studies [52,54] that evaluated VL prophylaxis without specifying the drug used noted a significant reduction in relapse. In a report of ten cases, Bossolasco et al. [55] showed that the relapse rate in patients groups with and without prophylaxis were 60% and 100%, respectively, but this difference did not reach statistical significance. Three studies evaluated specific prophylactic regimens (antimony compounds [46,50] and liposomal amphotericin [50]) and demonstrated reduction on VL relapse. Although the confidence intervals did not reach statistical significance, another author [56] concluded that lipid-complexed amphotericin prophylaxis also reduced the relapse rate. Finally, Laguna et al. [57]

showed a trend towards (p = 0.08) a reduction in VL relapse rate following treatment with pentamidine prophylaxis. A meta-analysis of results from all studies evaluating the impact of secondary prophylaxis is shown in Figure 3. This analysis could consistently demonstrate that secondary prophylaxis reduces VL relapse rate.

CD4+ lymphocyte count at VL diagnosis and follow-up has been studied in relation to risk of relapse. Nine articles [10,11,12,46,50,51,52,55,58] compared CD4+ lymphocyte cell counts at VL diagnosis between relapsing and non-relapsing patients as a continuous variable. Neither of these studies showed significant differences between these two groups. On the other hand, two studies [47,49] that compared relapse rate between patients with CD4+ count at VL diagnosis as a dichotomic variable (above and below than 100 cell/mL) noted that the arms with higher CD4+ counts had lower relapse rate. Similarly, an increase in CD4+ lymphocyte count at follow-up was protective against VL relapse in 5 of 7 studies [10,11,49,55,58]. In another study [12], univariate analyses of CD4+ counts at follow-up revealed a trend towards a reduction in relapse (p = 0.09).

Other variables explored in relation to relapse are shown in Table S3. Factors such as age, route of HIV transmission, history of intravenous drug use, HIV viral load at VL diagnosis, various clinical findings, specific anti-Leishmania treatments given, time from VL diagnosis to the introduction of protease inhibitor therapy, HAART compliance, the presence of an AIDS-defining disease before VL diagnosis and the presence of serum anti-Leishmania antibodies were not substantially different between relapsing and non-relapsing patients. Tuberculosis co-infection [47], hepatitis C virus co-infection [49] and an incomplete course of VL treatment [52] were evaluated in multivariate analysis and showed a statistically significant association of these conditions with the occurrence of relapse. Previous VL episodes were identified as risk factors for relapse in 3 studies, two of which were multivariate analyses.

# Prognostic Variables and Statistical Analysis

The statistical quality and the presentation of methods and results in many studies were poor. In nine studies, the Kaplan-Meier method was used in a univariate survival analysis to

# **Meta Analysis**

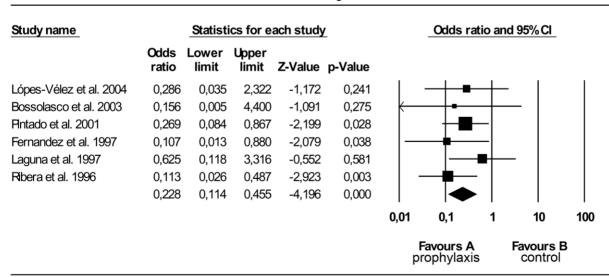


Figure 3. Meta-analysis of secondary prophylaxis results. Footnote:  $l^2 = 0\%$  Egger test for publication bias was negative, p = 0.76. doi:10.1371/journal.pntd.0001153.g003



analyzed VL relapse. Three prospective studies and two retrospective cohort studies employed Cox regressions for multivariate analysis of independent predictors. One study randomized patients to compare prophylaxis (liposomal amphotericin versus no treatment) and performed multivariate analysis to compare relapse rates by logistic regression, including some predictors as covariates. None of these six studies mentioned collinearity assessment (i.e., a high degree of correlation between 2 predictive variables) or developed a risk score for relapse based on their multivariable results. Also, none of the multivariate analyses reported a goodness-of-fit test of their models. Other studies analyzed isolated relapse predictors by univariate association tests in series of prospective or retrospective cases or in intervention studies.

#### Discussion

The present study is the first systematic review of predictors of VL relapse in HIV-infected patients. Our main conclusions are that VL relapse in HIV-infected patients receiving HAART is high and that secondary prophylaxis provides some protective effect but does not completely prevent the occurrence of relapse. We found that patients who did not relapse showed significantly higher CD4+ count at follow-up than patients with relapsing course. Our analysis also suggests that CD4+ count greater than 100 cell/mL at VL diagnosis reduces the odds of relapse. Unlike other opportunistic infections there are some reports of VL relapse in patients with a CD4+ count greater than 200 cell/mL in Ethiopia, and rarely in Europe [9]. This evidence shows that factors other than a CD4+ cell increase are involved in VL control. A threshold for safely discontinuing of secondary prophylaxis has not been established because of these uncertainties.

Most cases reported showed severe reductions in T cells. It could indicate that VL affects HIV-1 patients who exhibit a significant disturbance of cellular immunity; however, VL by itself may reduce CD4+ lymphocyte counts [59]. On the other hand, a CD4+ count greater than 100 cell/mL at VL diagnosis is a potential protective factor against relapse, although the analysis of this beneficial effect may be complicated by the immunosuppression of many the patients included in the studies. When analyzing the CD4+ count range and number of patients with CD4+ counts of greater than 100 cell/mL in the two studies [47,49] demonstrating an association between higher baseline CD4+ counts and reduced VL relapse, it is possible to speculate that studies that did not demonstrate an influence of CD4+ cells had few patients with CD4+ counts of greater than 100 cell/mL. Studies using animal models reported that CD4+ cells are responsible for the initial control of parasite proliferation and dissemination [60]. Thus, a low initial CD4+ count might allow a wide dissemination of the parasite throughout the phagocytic mononuclear system at the beginning of infection, increasing the number of sites that could harbor quiescent parasites (so-called "sanctuaries") [61].

Relapses of VL are suggested to occur mainly in individuals with poor responses to antiretroviral treatment who have no improvement in CD4+ counts [11,12,58,62], with a few exceptions [9,47]. The evolution of patients who acquire VL and thereafter show a significant increase in CD4+ counts while on HAART is currently receiving attention [47 50 51 52]. It has already been established that the outcome of VL is not influenced by humoral immunity but appears to be regulated by CD4+ T helper cells with different patterns of cytokine activity [63]. Protective immunity can be attributed to T helper (Th)-1 cells, whereas Th-2 cell responses produce IL-4 and IL-10 and facilitate the intracellular survival of the parasite [64]. It might be expected

that highly active antiretroviral drug combinations would favor an immunological shift from type 2 to type 1 cytokines in HIV-infected individuals. However, increased CD4+ values in peripheral blood and lymphoid tissues as a result of antiretroviral therapy may have negligible effects on cytokine production during the first 24 weeks [65]. In addition, patients on HAART show an initial increase in the CD4+ memory subset, whereas naive CD4+ cells consistently increase only after 1 year [66].

It is known that HIV patients who are receiving HAART have fewer opportunistic infections and recent data shows that there has been a decline in the incidence of VL after the introduction of HAART [41,54,67,68,69]. HAART seems to be insufficient to prevent VL relapse. Studies in patients receiving HAART showed a relapse rate similar to other studies performed in the pre-HAART era. Only one [49] observational study noted a reduction in the relapse rate among patients on HAART at VL diagnosis. None of the studies reported a statistically significant difference in VL relapse between patients receiving and not receiving HAART on follow-up. These disappointing results so far disagree with a statistically significant association between improvement of CD4+ count at follow-up and reduction of VL relapse. They may be due to the small sample sizes of the studies performed, poor patient adherence to antiviral therapy or insufficient immune response. One possibility to be explored in the future is the role of cytokines [70] influencing the control of VL independently of the CD4+ lymphocyte. The heterogeneity of zymodemes that exhibit different degrees of virulence or parasite burden could contribute to the differences observed in the host immune response and clinical evolution [9]. HAART increases CD4+ count thus influencing the control of VL, but may not be enough in this complex scenario created by the coinfection HIV and Leishmania. Fernandéz-Cotarelo et al. [54] and others [41] have shown a decrease in the number of new episodes of VL in HIV-infected patients receiving HAART but also a tendency toward VL relapse. According to these authors the high rates of relapse could be explained by the increased patient survival that results from effective antiretroviral therapy.

Previous episodes of VL were strongly associated with relapse. Also in agreement with the immune-inflammatory theory, it was hypothesized that the enhancement of the Th-2 response following one early relapse could complicate or prevent the later control of *Leishmania* infection [54].

Secondary prophylaxis seemed to only partially protect against relapse. Some of studies that observed a reduction in VL relapse following the use of secondary prophylaxis had few patients on HAART, which may not reflect the current reality. Data analysis suggests that the small sample sizes and heterogeneity of regimens used make the results less robust. Nevertheless, the evaluation of these studies through meta-analysis indicates a clear benefit of secondary prophylaxis in reducing VL relapse. Based on six studies whose data were available, the average relapse rate in patients not receiving secondary prophylaxis was 67%, while in the secondary prophylaxis arm, the relapse rate was 31%. Given this difference, the estimated total sample size needed for a study with 80% power would be 70 patients. Three out of the six studies examining secondary prophylaxis were not able to demonstrate statistical significance, possibly because of small sample sizes. It is important to emphasize that despite the heterogeneity of prophylaxis regimens used; statistical results are positively homogeneous in meta-analysis.

Thresholds for safe discontinuation of secondary prophylaxis for Spanish patients have been suggested to be CD4+ counts of 200 [71] and 350 cells/mL [11]. Differently of the European experience, one Ethiopian study [47] has shown that many patients suffering relapse (11 from 39 cases) had a CD4+ count above 200 cells/mL before relapse. These data may suggest that *L. donovani*, the predominant

causative agent of VL in east Africa and south Asia, is a more virulent and anthroponotic species than L. infantum. Another plausible explanation for this difference may be the influence of other variables that can affect the host immune response such as nutritional status and the presence of other infections and co-morbidities.

It has been postulated that the maintenance of an undetectable viral load protects against the development of VL [17] and that a high viral load could predict a weak response to antiparasitic treatment [12] although there are contradictory reports on this point [54,72]. None of the papers reviewed here linked HIV load by PCR at VL diagnosis with relapse. On the other hand HIV load by PCR at follow-up was statistically related to relapse in one [58] of four studies that evaluated this variable in a univariate analysis. These observations support the idea that a sustained immunological response is more important than a virological response to cure VL in HIV-infected patients.

It is important to note that a wide range of therapeutic drugs were utilized for the treatment of VL in the studies we have reviewed. There was no notable difference in the relapse rate with regard to specific VL treatment used (all analyzed in univariate analysis); however only four studies explored this association and most of them included a limited number of patients and only two [11,73] involved randomly assigned patients. Few comparative clinical studies have been conducted of the efficacy of treatment for HIV–VL co-infection outside the Mediterranean area. In some instances [74,75], the development of drug resistance could contribute to therapeutic failure and the relapsing course observed in HIV-infected patients. These observations do not allow us to refute the influence of anti-parasite treatment on relapse outcome.

#### **Study Limitations**

Although we have made an extensive review, our analysis includes studies with different definitions of cure and different lengths of follow-up. Cure is seldom defined parasitologically in these studies and the difference between treatment failure and relapse is arbitrary in some studies. It is possible that some episodes of relapse in the group of patients in which parasitological cure were not documented by bone marrow examination were treatment failures rather than relapses. Moreover, re-infection was not distinguished from relapse in any paper. There is a high degree of heterogeneity in the evaluated populations as shown by the wide range of reported mortality (6.5% to 83.8%), treatment failure (0 to 47.6%) and relapse rates (20% to 70%). These studies included patients with different degrees of immunosuppression, and different treatment and prophylaxis regimens. Also, there are differences in the study designs, the types of statistical methods used and the prognostic variables included in analysis. These variations may have resulted in patient selection bias or low statistical power, thus hampering a meta-analysis of all studied predictors of relapse. In spite of these limitations, we believe that the meta-analysis results of secondary prophylaxis are consistent, considering the available evidence. In addition, the quality of published reports was heterogeneous and usually poor. Despite these limitations, this review may assist clinicians in making decisions and may also help in the design of future studies.

# References

- Desjeux P, Alvar J (2003) Leishmania/HIV co-infections: epidemiology in Europe. Ann Trop Med Parasitol 97 Suppl 1: 3–15.
- Desjeux P, Meert JP, Piot B, Alwar J, Medrano FJ, et al. (2000) Leishmania/ HIV co-infection, South Western Europe 1990–98. Geneva: World Health Organization, Report No.:42.
- Lopez-Velez R, Perez-Molina JA, Guerrero A, Baquero F, Villarrubia J, et al. (1998) Clinicoepidemiologic characteristics, prognostic factors, and survival analysis of patients coinfected with human immunodeficiency virus and

### Conclusion

The results of this systematic review suggest there are identifiable predictive factors of VL relapse, such as previous episodes of VL relapse and lack of recovery of CD4+ lymphocyte numbers after primary visceral leishmaniasis. HAART did not produce the anticipated decrease in the incidence of VL relapses and more data is needed in order to better assess the evolution of VL in the HAART era. In contrast, secondary prophylaxis was shown to be protective against relapse. CD4+ count below 100 cells/mL at the time of VL primary diagnosis is a potential predictor of relapse.

Based on these observations, a high-risk population might be identified and such patients might then be eligible for secondary prophylaxis. Strong surveillance will certainly contribute to improved data quality for decision-makers in this complex scenario. Randomized trials to compare the efficacy of different drugs and their role either in treatment or in prophylaxis are required.

## **Supporting Information**

**Table S1 Parasitological control.** Identification of *Leishmania* amastigotes by direct examination or by isolation of promastigotes in culture of tissue samples dAmB: amphotericin B deoxycholate LAmB: liposomal amphotericin LipAmB: amphotericin B lipid complex. PA: Pentavalent antimonial compounds **SD**: standard deviation **IRQ**: interquartile range  $\tilde{\mathbf{x}}$ : median  $\boldsymbol{\mu}$ : mean. (DOC)

Table S2 VL: Visceral leishmaniasis Parasitological confirmation: identification of Leishmania amastigotes by direct examination or by isolation of promastigotes in culture of tissue samples #Serology confirmation: Leishmania direct agglutination positive \$Biologic confirmation: identification of Leishmania amastigotes by direct examination or by isolation of promastigotes in culture of tissue samples or Leishmania-specific PCR on peripheral blood/bone marrow dAmB: amphotericin B deoxycholate LAmB: liposomal amphotericin B LipAmB: amphotericin B lipid complex PA: Pentavalent antimonial compounds **Hemo**: transfusion route **IDU**: intravenous drug user **HETERO**: heterosexual contacts **HOMO**: men who have sex with men **sexual**: heterosexual or homosexual contacts **SD**: standard deviation IRQ: interquartile range # if the information was available x̃: median μ: mean. (DOC)

**Table S3 Yes**: positive association **No**: negative association **VL**: Visceral leishmaniasis **HAART**: highly active antiretroviral therapy **HVC**: hepatitis C virus \* multivariate analysis. (DOC)

#### **Author Contributions**

Conceived and designed the experiments: GFC MRdS AR. Performed the experiments: GFC MRdS. Analyzed the data: GFC MRdS. Wrote the paper: GFC MRdS. Paper review: AR.

- Leishmania in an area of Madrid, Spain. Am J Trop Med Hyg 58(4): 436-43
- Gradoni L, Scalone A, Gramiccia M, Troiani M (1996) Epidemiological surveillance of leishmaniasis in HIV-1-infected individuals in Italy. AIDS (London, England) 10(7): 785–91.
- Alvar J, Aparicio P, Aseffa A, Den Boer M, Canavate C, et al. (2008) The relationship between leishmaniasis and AIDS: the second 10 years. Clin Microbiol Rev 21(2): 334–59.



- Sinha PK, Bimal S, Singh SK, Pandey K, Gangopadhyay DN, et al. (2006) Pre-& post-treatment evaluation of immunological features in Indian visceral leishmaniasis (VL) patients with HIV co-infection. Indian J Med Res 123: 197–202.
- Olivier M, Badaro R, Medrano FJ, Moreno J (2003) The pathogenesis of Leishmania/HIV co-infection: cellular and immunological mechanisms. Ann Trop Med Parasitol 97 Suppl 1: 79–98.
- Tremblay M, Olivier M, Bernier R (1996) Leishmania and the pathogenesis of HIV infection. Parasitol Today 12(7): 257–61.
- Villanueva JL, Alarcon A, Bernabeu-Wittel M, Cordero E, Prados D, et al. (2000) Prospective evaluation and follow-up of European patients with visceral leishmaniasis and HIV-1 coinfection in the era of highly active antiretroviral therapy. Eur J Clin Microbiol Infect Dis 19(10): 798–801.
- Molina I, Falco V, Crespo M, Riera C, Ribera E, et al. (2007) Efficacy of liposomal amphotericin B for secondary prophylaxis of visceral leishmaniasis in HIV-infected patients. J Antimicrob Chemother 60(4): 837–42.
- Berenguer J, Cosin J, Miralles P, Lopez JC, Padilla B (2000) Discontinuation of secondary anti-Leishmania prophylaxis in HIV-infected patients who have responded to highly active antiretroviral therapy. AIDS (London, England) 14(18): 2946–8.
- Casado JL, López-Velez R, Pintado V, Quereda C, Antela A, et al. (2001) Relapsing visceral leishmaniasis in HIV-infected patients undergoing successful protease inhibitor therapy. Eur J Clin Microbiol Infect Dis 20(3): 202–5.
- Paredes R, Munoz J, Diaz I, Domingo P, Gurgui M, et al. (2003) Leishmaniasis in HIV infection. J Postgrad Med 49(1): 39–9.
- Sundar S (2001) Drug resistance in Indian visceral leishmaniasis. Trop Med Int Health 6(11): 849–54.
- Rabello A, Orsini M, Disch J (2003) Leishmania/HIV co-infection in Brazil: an appraisal. Ann Trop Med Parasitol 97 Suppl 1: 17–28.
- World Health Organization (2009) Leishmaniasis: background information. A brief history of the disease. Available: www.who.int/leishmaniasis/en/. Accessed 2011 Feb 18
- Jimenez-Exposito MJ, Alonso-Villaverde C, Sarda P, Masana L (1999) Visceral leishmaniasis in HIV-infected patients with non-detectable HIV-1 viral load after highly active antiretroviral therapy. AIDS (London, England) 13(1): 152–3.
- Liberati A, Altman DG, Tetzlaff J, Mulrow C, Gotzsche PC, et al. (2009) The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanation and elaboration. PLoS Med 6: e100100
- Wilczynski NL, Haynes RB (2004) Developing optimal search strategies for detecting clinically sound prognostic studies in MEDLINE: an analytic survey. BMC Med 2: 23–27.
- Manchikanti L, Datta S, Smith HS, Hirsch JA (2009) Evidence-based medicine, systematic reviews, and guidelines in interventional pain management: part 6.
   Systematic reviews and meta-analyses of observational studies. Pain Physician 12(5): 819–50.
- Groenwold RH, Rovers MM (2010) The Catch-22 of appraisals on the quality of observational studies. J Clin Epidemiol 63(10): 1059–60.
- Malta M, Cardoso LO, Bastos FI, Magnanini MM, da Silva CM (2010) STROBE initiative: guidelines on reporting observational studies. Rev Saude Publica 44(3): 559–65.
- Cabie A, Matheron S, Lepretre A, Bouchaud O, Deluol AM, et al. (1992) Visceral leishmaniasis in HIV infection. A totally opportunistic infection. Presse Med 21(35): 1658–62.
- Peters BS, Fish D, Golden R, Evans DA, Bryceson AD, et al. (1990) Visceral leishmaniasis in HIV infection and AIDS: clinical features and response to therapy. Q.J Med 77: 1101–11.
- Altes J, Salas A, Riera M, Udina M, Galmes A, et al. (1991) Visceral leishmaniasis: another HIV-associated opportunistic infection? Report of eight cases and review of the literature. AIDS (London, England) 5(2): 201–7.
- Davidson RN, Di Martino L, Gradoni L, Giacchino R, Russo R, et al. (1994) Liposomal amphotericin B (AmBisome) in Mediterranean visceral leishmaniasis: a multi-centre trial. Q.J Med 87(2): 75–81.
- Davidson RN, Russo R (1994) Relapse of visceral leishmaniasis in patients who
  were coinfected with human immunodeficiency virus and who received
  treatment with liposomal amphotericin B. Clin Infect Dis 19(3): 560.
- 28. Rosenthal E, Marty P, Poizot-Martin I, Reynes J, Pratlong F, et al. (1995) Visceral leishmaniasis and HIV-1 co-infection in southern France. Trans R Soc Trop Med Hyg 89(2): 159–62.
- Ramos A, Portero JL, Gazapo T, Yebra M, Portero F, et al. (1998) Visceral leishmaniasis in immunocompromised patients. An Med Interna 15(6): 301–4.
- Pintado V, López-Vélez R (2001) HIV-associated visceral leishmaniasis. Clin Microbiol Infect 7(6): 291–300.
- Agostoni C, Dorigoni N, Malfitano A, Caggese L, Marchetti G, et al. (1998) Mediterranean leishmaniasis in HIV-infected patients: epidemiological, clinical, and diagnostic features of 22 cases. Infection 26(2): 93–9.
- 32. Cruz I, Canavate C, Rubio JM, Morales MA, Chicharro C, et al. (2002) A nested polymerase chain reaction (Ln-PCR) for diagnosing and monitoring Leishmania infantum infection in patients co-infected with human immunode-ficiency virus. Trans R Soc Trop Med Hyg 96 Suppl 1: S185–9.
- d'Ettorre G, Ceccarelli G, Carnevalini M, Forcina G, Zaffiri L, et al. (2006)
   Central role of interleukin-15 in human immunodeficiency virus (HIV)-infected patients with visceral leishmaniasis. Acta Trop 99(1): 83–7.

- Daher EF, Fonseca PP, Gerhard ES, Leitao TM, Silva Junior GB (2009) Clinical and epidemiological features of visceral leishmaniasis and HIV co-infection in fifteen patients from Brazil. J Parasitol 95(3): 652–5.
- Hurissa Z, Gebre-Silassic S, Hailu W, Tefera T, Lalloo DG, et al. (2010) Clinical characteristics and treatment outcome of patients with visceral leishmaniasis and HIV co-infection in northwest Ethiopia. Trop Med Int Health 15(7): 848–55.
- 36. Antinori S, Calattini S, Longhi E, Bestetti G, Piolini R, et al. (2007) Clinical use of polymerase chain reaction performed on peripheral blood and bone marrow samples for the diagnosis and monitoring of visceral leishmaniasis in HIV-infected and HIV-uninfected patients: a single-center, 8-year experience in Italy and review of the literature. Clin Infect Dis 44(12): 1602–10.
- Fernandez-Guerrero ML, Robles P, Rivas P, Mojer F, Muniz G, et al. (2004)
   Visceral leishmaniasis in immunocompromised patients with and without AIDS: a comparison of clinical features and prognosis. Acta Trop 90(1): 11–6.
- Russo R, Nigro L, Panarello G, Montineri A (2003) Clinical survey of Leishmania/HIV co-infection in Catania, Italy: the impact of highly active antiretroviral therapy (HAART). Ann Trop Med Parasitol 97 Suppl 1: 149–55.
- Ritmeijer K, Veeken H, Melaku Y, Leal G, Amsalu R, et al. (2001) Ethiopian visceral leishmaniasis: generic and proprietary sodium stibogluconate are equivalent; HIV co-infected patients have a poor outcome. Trans R Soc Trop Med Hyg 95(6): 668–72.
- Lachaud L, Dereure J, Chabbert E, Reynes J, Mauboussin JM, et al. (2000) Optimized PCR using patient blood samples for diagnosis and follow-up of visceral Leishmaniasis, with special reference to AIDS patients. J Clin Microbiol 38(1): 236–40.
- Tortajada C, Perez-Cuevas B, Moreno A, Martinez E, Mallolas J, et al. (2002) Highly active antiretroviral therapy (HAART) modifies the incidence and outcome of visceral leishmaniasis in HIV-infected patients. J Acquir Immune Defic Syndr 30(3): 364–6.
- Ribera E, Cucurull E, Ocana I, Vallespi T, Gasser I, et al. (1995) [Visceral leishmaniasis in patients with HIV infection]. Enferm Infect Microbiol Clin 13(2): 73–9.
- Riera C, Fisa R, Lopez P, Ribera E, Carrio J, et al. (2004) Evaluation of a latex agglutination test (KAtex) for detection of Leishmania antigen in urine of patients with HIV-Leishmania coinfection: value in diagnosis and post-treatment follow-up. Eur J Clin Microbiol Infect Dis 23(12): 899–904.
- Riera C, Fisa R, Ribera E, Carrio J, Falco V, et al. (2005) Value of culture and nested polymerase chain reaction of blood in the prediction of relapses in patients co-infected with *Leishmania* and human immunodeficiency virus. Am J Trop Med Hyg 73(6): 1012–5.
- Ritmeijer K, Dejenie A, Assefa Y, Hundie TB, Mesure J, et al. (2006) A comparison of miltefosine and sodium stibogluconate for treatment of visceral leishmaniasis in an Ethiopian population with high prevalence of HIV infection. Clin Infect Dis 43(3): 357-64
- Ribera E, Ocana I, de Otero J, Cortes E, Gasser I, et al. (1996) Prophylaxis of visceral leishmaniasis in human immunodeficiency virus-infected patients. Am J Med 100(5): 496–501.
- ter Horst R, Collin SM, Ritmeijer K, Bogale A, Davidson RN (2008) Concordant HIV infection and visceral leishmaniasis in Ethiopia: the influence of antiretroviral treatment and other factors on outcome. Clin Infect Dis 46(11): 1702–9.
- 1993 revised classification system for HIV infection and expanded surveillance case definition for AIDS among adolescents and adults. MMWR Recomm Rep1992 41(RR-17): 1–19.
- Bourgeois N, Lachaud L, Reynes J, Rouanet I, Mahamat A, et al. (2008) Longterm monitoring of visceral leishmaniasis in patients with AIDS: relapse risk factors, value of polymerase chain reaction, and potential impact on secondary prophylaxis. J Acquir Immune Defic Syndr 48(1): 13–9.
- Pintado VM-RP, Rivera ML, Moreno S, Bouza E (2001) Visceral leishmaniasis in HIV-infected and non-HIV-infected patients: a comparative study. Medicine (Baltimore) 80: 54–73.
- Pizzuto M, Piazza M, Senese D, Scalamogna C, Calattini S, et al. (2001) Role of PCR in diagnosis and prognosis of visceral leishmaniasis in patients coinfected with human immunodeficiency virus type 1. J Clin Microbiol 39(1): 357–361.
- 52. Pasquau F, Ena J, Sanchez R, Cuadrado JM, Amador C, et al. (2005) Leishmaniasis as an opportunistic infection in HIV-infected patients: determinants of relapse and mortality in a collaborative study of 228 episodes in a Mediterranean region. Eur J Clin Microbiol Infect Dis 24(6): 411–8.
- Laguna F, Adrados M, Alvar J, Soriano V, Valencia ME, et al. (1997) Visceral leishmaniasis in patients infected with the human immunodeficiency virus. Eur J Clin Microbiol Infect Dis 16(12): 898–903.
- 54. Fernandéz Cotarelo MJ, Abellan Martinez J, Guerra Vales JM, Martinez Sanchez P, Rodrigo Gomez De La Barcena M, et al. (2003) Effect of highly active antiretroviral therapy on the incidence and clinical manifestations of visceral leishmaniasis in human immunodeficiency virus-infected patients. Clin Infect Dis 37(7): 973–7.
- Bossolasco S, Gaiera G, Olchini D, Gulletta M, Martello L, et al. (2003) Realtime PCR assay for clinical management of human immunodeficiency virusinfected patients with visceral leishmaniasis. J Clin Microbiol 41(11): 5080–4.
- López-Vélez R, Videla S, Marquez M, Boix V, Jimenez-Mejias ME, et al. (2004) Amphotericin B lipid complex versus no treatment in the secondary prophylaxis of visceral leishmaniasis in HIV-infected patients. J Antimicrob Chemother 53(3): 540–3.



- Delgado Fernandéz M, Garcia Ordonez MA, Martos Perez F, Reguera Iglesias JM, Jimenez Onate F, et al. (1997) [The clinical and evolutional characteristics of visceral leishmaniasis in patients with HIV infection]. An Med Interna 14(10): 506–10.
- Mira JA, Corzo JE, Rivero A, Macias J, De Leon FL, et al. (2004) Frequency of visceral leishmaniasis relapses in human immunodeficiency virus-infected patients receiving highly active antiretroviral therapy. Am J Trop Med Hyg 70(3): 298–301.
- Berman J (2006) Visceral leishmaniasis in the New World & Africa. Indian J Med Res 123: 289–94.
- Schilling S, Glaichenhaus N (2001) T cells that react to the immunodominant Leishmania major LACK antigen prevent early dissemination of the parasite in susceptible BALB/c mice. Infect Immun 69(2): 1212–4.
- Lachaud L, Chabbert E, Dubessay P, Dereure J, Lamothe J, et al. (2002) Value of two PCR methods for the diagnosis of canine visceral leishmaniasis and the detection of asymptomatic carriers. Parasitology 125: 197–207.
- López-Vélez R, Casado JL, Pintado V (2001) Decline of a visceral leishmaniasis epidemic in HIV-infected patients after the introduction of highly active antiretroviral therapy (HAART). Clin Microbiol Infect 7(7): 394–5.
- Kemp M, Kurtzhals JA, Kharazmi A, Theander TG (1993) Interferon-gamma and interleukin-4 in human *Leishmania* donovani infections. Immunol Cell Biol 71: 583–7.
- Lehn M, Weiser WY, Engelhorn S, Gillis S, Remold HG (1989) IL-4 inhibits H2O2 production and antileishmanial capacity of human cultured monocytes mediated by IFN-gamma. J Immunol 143(9): 3020

  –4.
- Zanussi S, Simonelli C, Bortolin MT, D'Andrea M, Crepaldi C, et al. (1999) Immunological changes in peripheral blood and in lymphoid tissue after treatment of HIV-infected subjects with highly active anti-retroviral therapy (HAART) or HAART+IL-2. Clin Exp Immunol 116(3): 486–92.
- Mezzaroma I, Carlesimo M, Pinter É, Alario C, Sacco G, et al. (1999) Longterm evaluation of T-cell subsets and T-cell function after HAART in advanced stage HIV-1 disease. AIDS (London, England) 13(10): 1187–93.

- 67. Kaplan JE, Hanson D, Dworkin MS, Frederick T, Bertolli J, et al. (2000) Epidemiology of human immunodeficiency virus-associated opportunistic infections in the United States in the era of highly active antiretroviral therapy. Clin Infect Dis 30 Suppl 1: S5–14.
- 68. del Giudice P, Mary-Krause M, Pradier C, Grabar S, Dellamonica P, et al. (2002) Impact of highly active antiretroviral therapy on the incidence of visceral leishmaniasis in a French cohort of patients infected with human immunode-ficiency virus. J Infect Dis 186(9): 1366–70.
- Palella FJ, Jr., Delaney KM, Moorman AC, Loveless MO, Fuhrer J, et al. (1998)
   Declining morbidity and mortality among patients with advanced human immunodeficiency virus infection. HIV Outpatient Study Investigators. N Engl J Med 338(13): 853–60.
- Sinha PK, Pandey K, Bhattacharya SK (2005) Diagnosis & management of Leishmania/HIV co-infection. Indian J Med Res 121: 407–414.
- Soriano V, Dona C, Rodriguez-Rosado R, Barreiro P, Gonzalez-Lahoz J (2000)
  Discontinuation of secondary prophylaxis for opportunistic infections in HIVinfected patients receiving highly active antiretroviral therapy. AIDS (London,
  England) 14(4): 383–6.
- Berhe N, Wolday D, Hailu A, Abraham Y, Ali A, et al. (1999) HIV viral load and response to antileishmanial chemotherapy in co-infected patients. AIDS (London, England) 13(14): 1921–5.
- Laguna F, López-Vélez R, Pulido F, Salas A, Torre-Cisneros J, et al. (1999) Treatment of visceral leishmaniasis in HIV-infected patients: a randomized trial comparing meglumine antimoniate with amphotericin B. Spanish HIV-Leishmania Study Group. AIDS (London, England) 13(9): 1063–9.
- 74. Faraut-Gambarelli F, Piarroux R, Deniau M, Giusiano B, Marty P, et al. (1997) In vitro and in vivo resistance of *Leishmania infantum* to meglumine antimoniate: a study of 37 strains collected from patients with visceral leishmaniasis. Antimicrob Agents Chemother 41(4): 827–30.
- Carrio J, Riera C, Gallego M, Ribera E, Portus M (2001) In vitro susceptibility
  of *Leishmania infantum* to meglumine antimoniate in isolates from repeated
  leishmaniasis episodes in HIV-coinfected patients. J Antimicrob Chemother
  47(1): 120–1.

