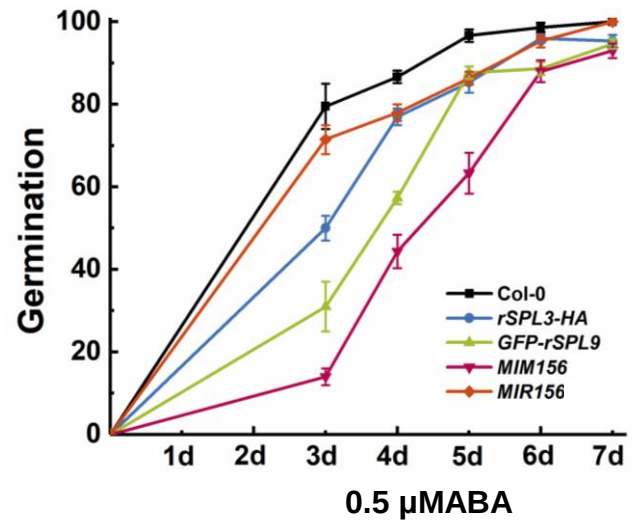
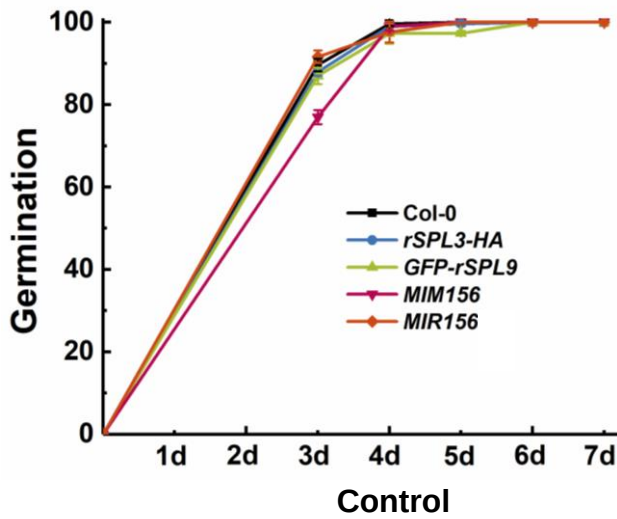
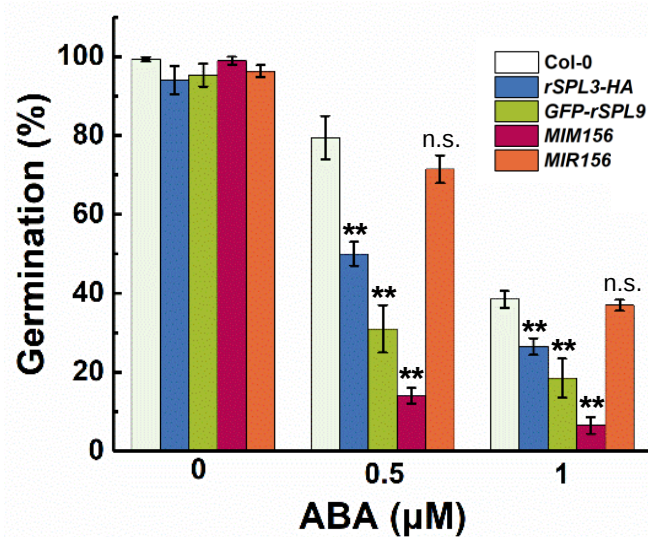


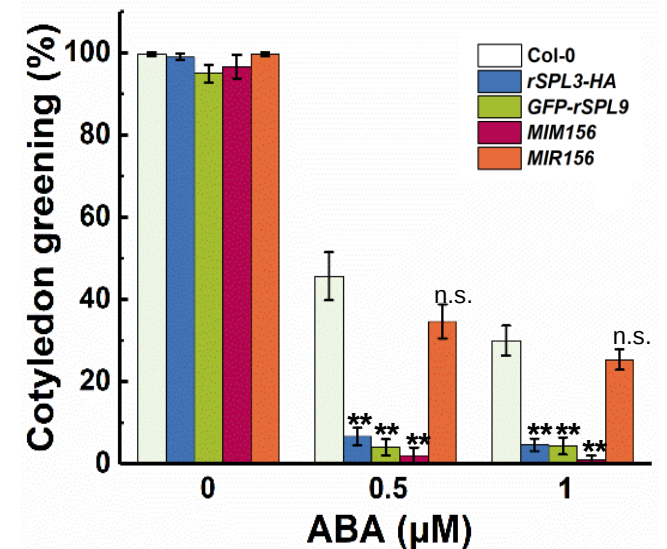
A



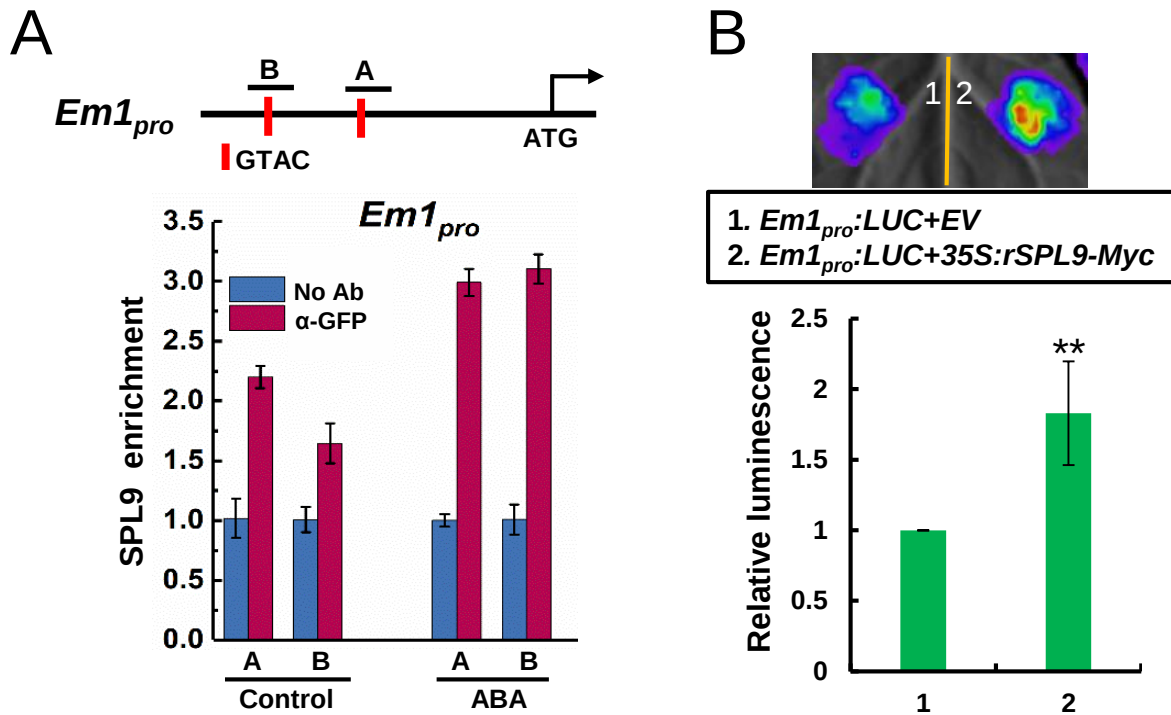
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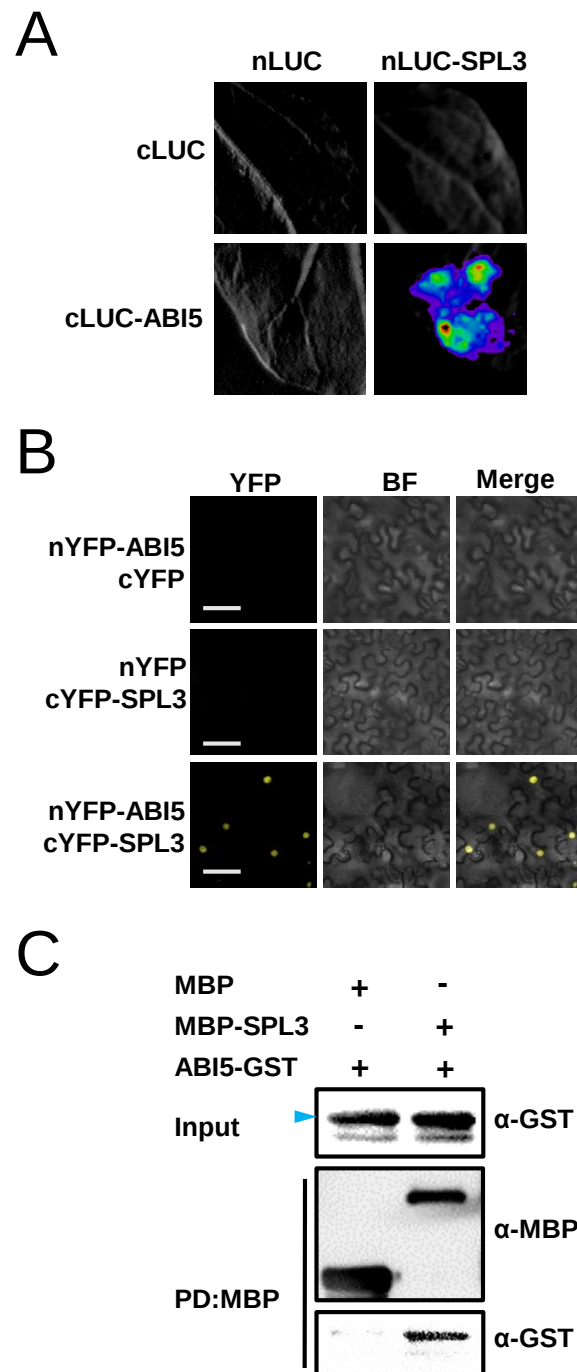
C



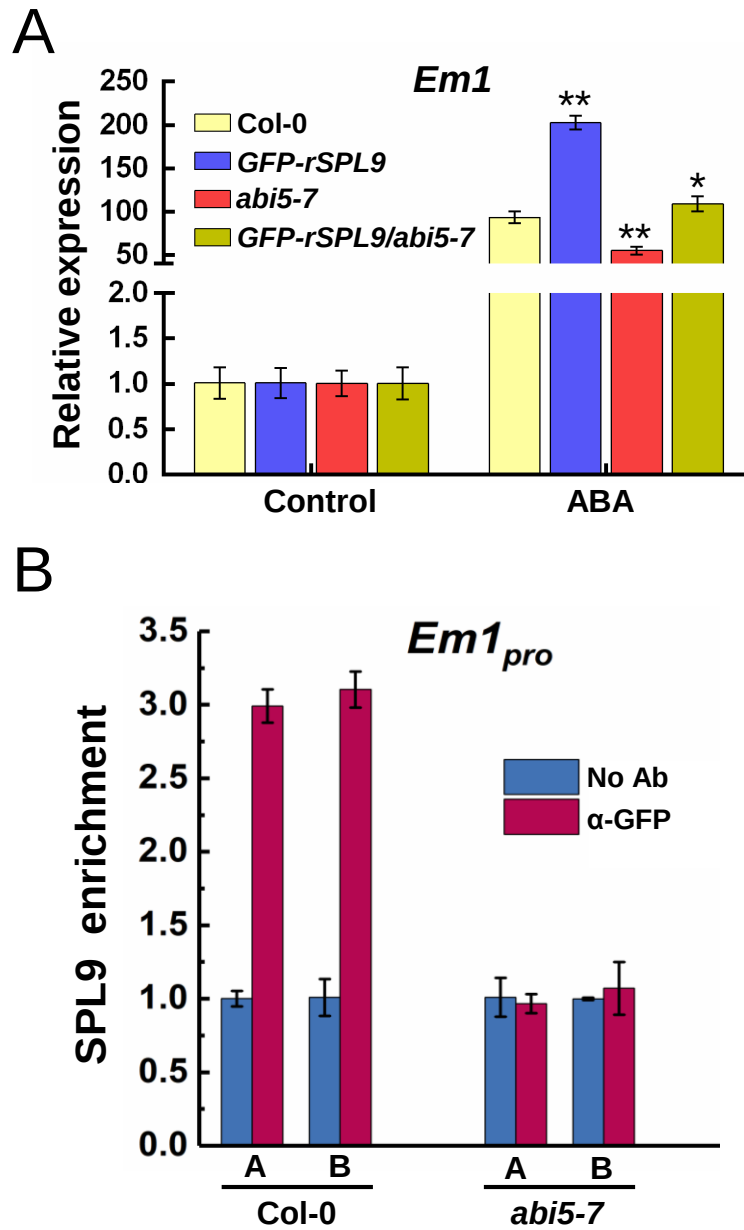
Supplementary Figure 1 | The miR156-targeted SPLs positively regulate ABA responses during seed germination and cotyledon greening. **A** Seed germination percentages of indicated genotypes grown on medium without or with 0.5 μ M ABA during a time course. **B, C** Quantification of seed germination (**B**) and cotyledon greening (**C**) of indicated genotypes in response to different concentration of ABA. Seed germination percentage was recorded at 3 d after the end of stratification and cotyledon-greening percentage was recorded at 5 d after the end. Data shown are mean $\pm$ SD ($n = 3$). At least 100 seeds per genotype were measured in each replicate. ** $P < 0.01$, n.s. indicates no significant difference (Student's t -test).



Supplementary Figure 2 | SPL9 directly activates the transcriptional expression of *Em1*. **A** ChIP-qPCR analysis showing enrichment of SPL9 on the *Em1* promoter region. Red boxes in the upper panel indicate the positions of SBP-box binding core motifs. The 6-d-old *GFP-rSPL9* seedlings treated without (Control) or with 50 μ M ABA for 2 hours were harvested for ChIP assays. Error bars denote $\pm$ SD ($n = 3$). **B** Transient expression assays showing the activation of *Em1* promoter by SPL9. Upper panel shows a representative leaf image, and the column diagram represents the quantification of the relative luminescence intensities ($n = 10$). The mean value in combination 1 was set to 1. ** $P < 0.01$ (Student's *t*-test).

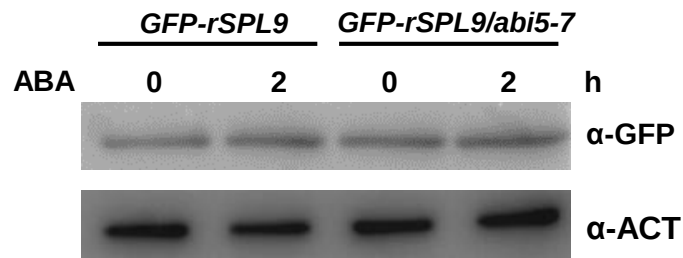


Supplementary Figure 3 | SPL3 directly interacts with ABI5. **A** LCI assays showing that SPL3 interacts with ABI5 in *N. benthamiana* leaves. **B** BiFC assays showing the interaction of SPL3 and ABI5 in *N. benthamiana* leaves. **C** Pull-down assays showing that SPL3 directly interacts with ABI5 *in vitro*. The ABI5-GST fusion proteins could be pulled down by MBP-SPL3 proteins but not MBP alone. Arrowhead indicates specific bands. PD, pull down.

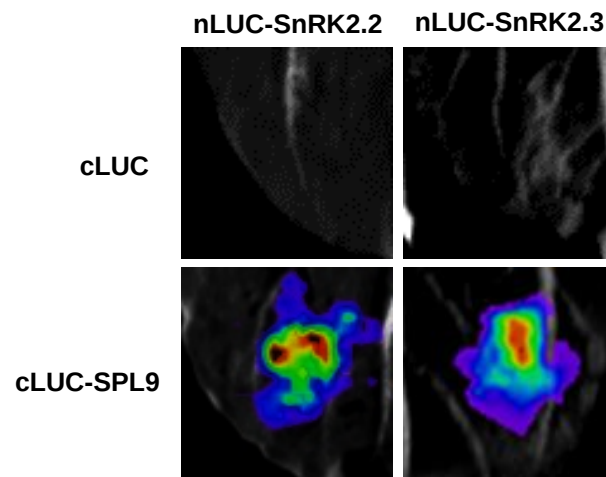


Supplementary Figure 4 | SPL9 activates the expression of *Em1* in an ABI5-dependent manner.

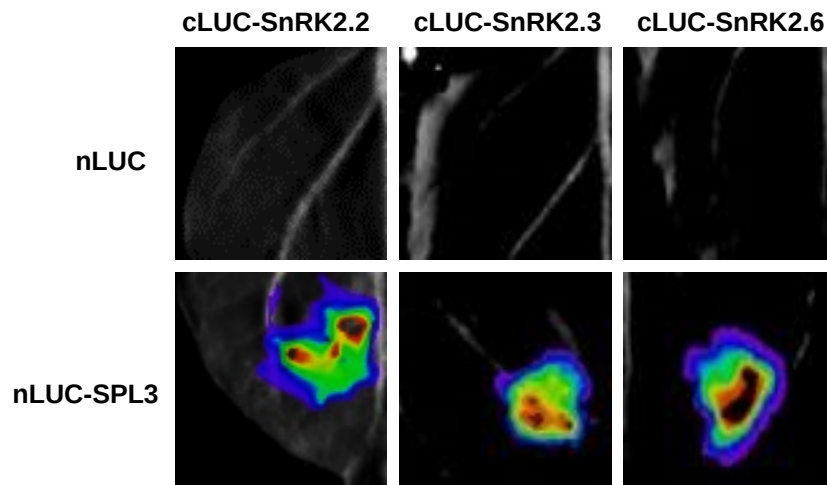
A qRT-PCR assays showing the expression levels of *Em1* in the indicated genotypes in response to ABA treatment. The 4-d-old seedlings were treated without or with 10 μ M ABA for 4 h. The expression levels of *Em1* in untreated seedlings (Control) for each genotype were set to 1. Data are means $\pm$ SD (n = 3). * P < 0.05, ** P < 0.01 (Student's *t*-test). **B** ChIP-qPCR analysis showing that the ABA-induced enrichment of SPL9 on the *Em1* promoter regions is dependent on ABI5. The 6-d-old *GFP-rSPL9* and *GFP-rSPL9/abi5-7* seedlings were treated with 50 μ M ABA for 2 h and then harvested for ChIP assays. Error bars denote $\pm$ SD (n = 3).



Supplementary Figure 5 | The GFP-SPL9 protein levels were comparable between the *GFP-rSPL9* and *GFP-rSPL9/abi5-7* seedlings. Immunoblotting assays showing the protein levels of SPL9 in the indicated 6-d-old seedlings treated without or with 50 μ M ABA for 2 h. Actin was used as a loading control.



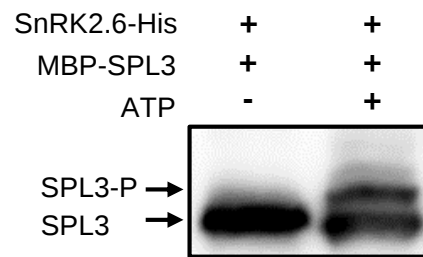
Supplementary Figure 6 | SnRK2s physically interact with SPL9. LCI assays showing that SnRK2s physically interact with SPL9. The nLUC-SnRK2s and cLUC-SPL9 constructs were co-transformed into leaves of *N. benthamiana*. Empty vectors were used as negative controls.



Supplementary Figure 7 | SnRK2s physically interact with SPL3. LCI assays showing that SnRK2s physically interact with SPL3. The cLUC-SnRK2s and nLUC-SPL3 constructs were co-transformed into leaves of *N. benthamiana*. Empty vectors were used as negative controls.

SPL9 : MEMGSNSGPGHGPQAESGGSSSTESSFSGGIMFGQKIYFEDGGGGSGSSSSGGRSNRRVRGG : 63
SPL9 : GSGQSGQIPRCQVEGCGMDLTNAKGYYSRHRVCGVHSKTPKVTVAGIEQRFCQQCSRFBQLPE : 126
SPL9 : FDLEKRSCRRRLAGHNERRRKPQPASLSVLASRYGRIAPSLYENG DAGMNGSFLGNQEIGWPS : 189
SPL9 : SRTLDTRVMRREVSSPSWQINPMNVFSQGSVGGGGTSFSSPEIMDTKLESYKGIGDSNCALSL : 252
SPL9 : LSNPHQPHDNNNNNNNNNNNNNTWRASSGFGPMTVTMAQPPPAPSQHQYLNPFWVFKDNDND : 315
SPL9 : MSFVLNLGRYTEPDNCQISSGTAMGEFELSDHHHQSRRQYMEDENTRAYDSSSHHTNWSL- : 375

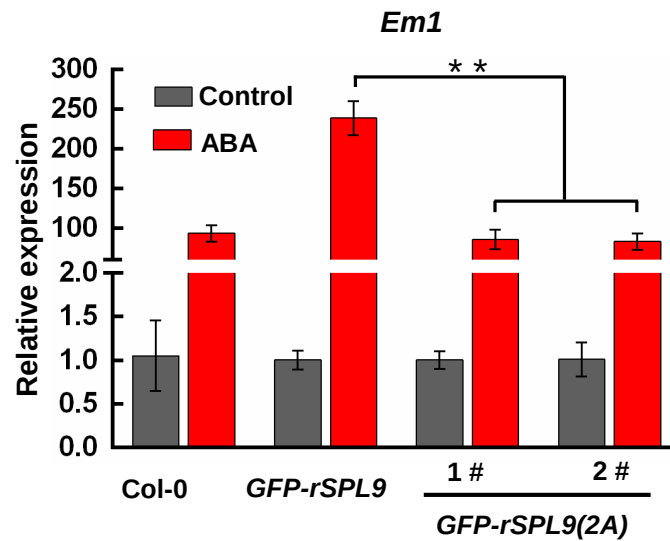
Supplementary Figure 8 | The putative SnRK2.6 phosphorylation sites in SPL9 protein. Blue lines indicate the RXXS/T motifs of SPL9 protein, which contains two putative SnRK2.6 phosphorylation sites Ser203 and Ser281, as shown by red arrows.



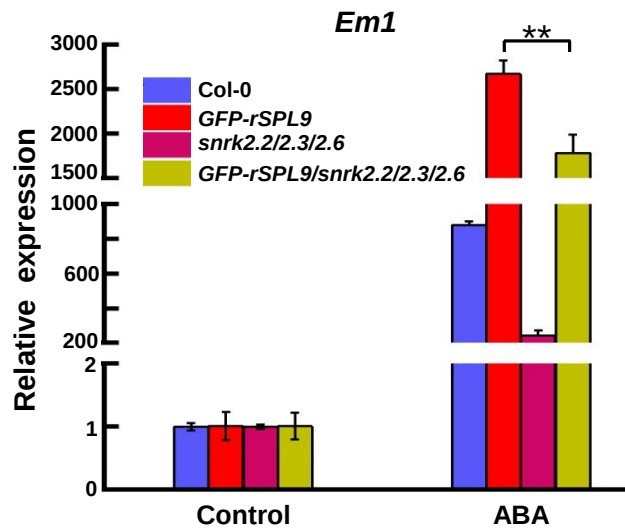
Supplementary Figure 9 | *In vitro* kinase assay showing that SnRK2.6 could phosphorylate SPL3.

Proteins were detected by phos-tag gel with anti-MBP antibody.

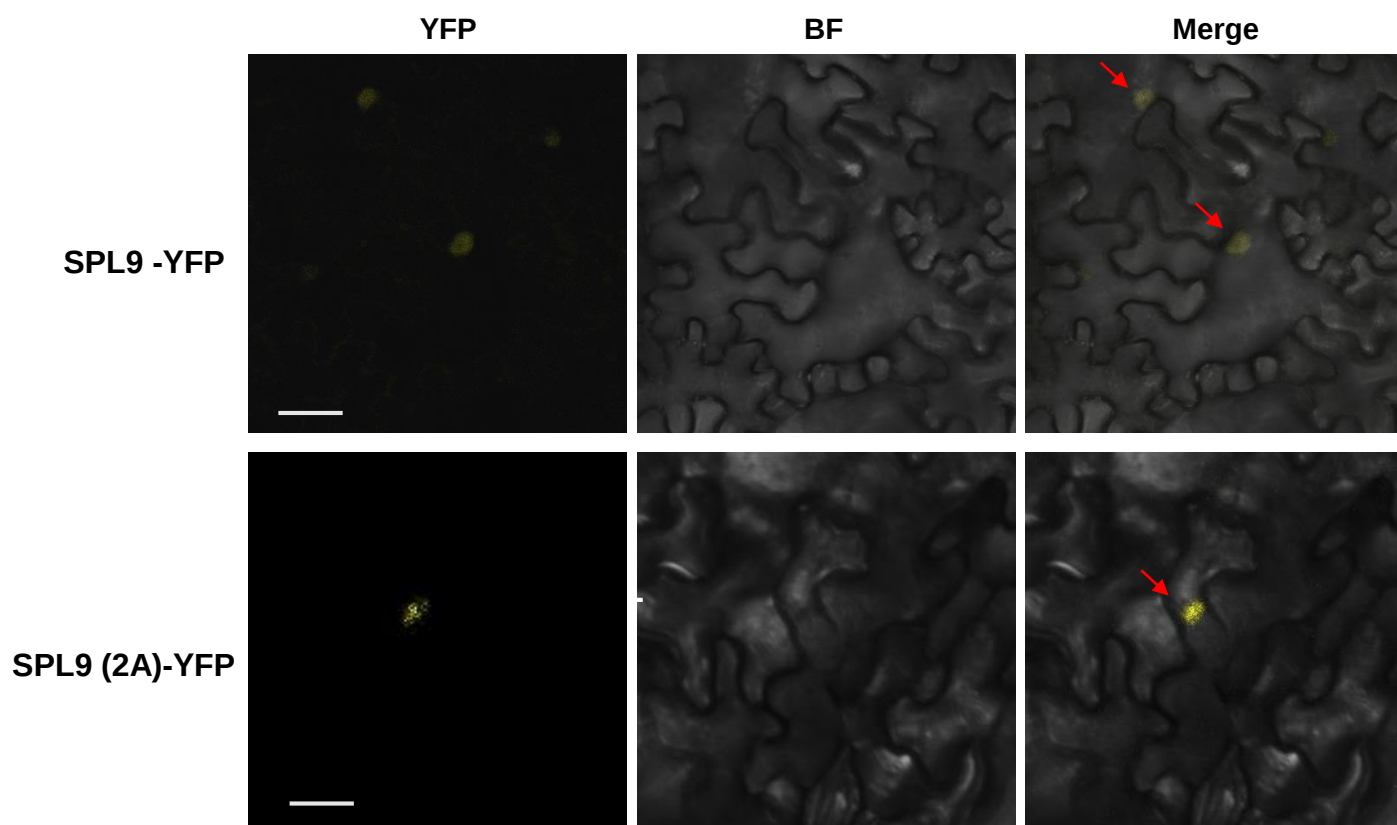
A



B



Supplementary Figure 10 | Phosphorylation by SnRK2s is required for SPL9-activated expression of *Em1*. **A** qRT-PCR assays showing phosphorylation by SnRK2s is required for SPL9-mediated activation of *Em1*. The 6-d-old seedlings were treated without or with 50 μ M ABA for 4 h. The expression of *Em1* in control samples was set to 1 for each genotype. Data are means $\pm$ SD (n = 3). ** $P < 0.01$ (Student's *t*-test). **B** qRT-PCR assays showing the activation of *Em1* by SPL9 is dependent on SnRK2s. The 4-d-old seedlings were treated without or with 50 μ M ABA for 4 h. The expression of *Em1* in control samples was set to 1 for each genotype. Data are means $\pm$ SD (n = 3). ** $P < 0.01$ (Student's *t*-test).



Supplementary Figure 11 | SnRK2s-mediated phosphorylation did not affect the nuclear localization of SPL9. Subcellular localization assays showed that both the SPL9 and SPL9(2A) mutant proteins were localized in nuclei.

Supplementary Table 1 | Primers used for DNA constructs in this study

Primer name	Forward primer (5'-3')	Reverse primer (5'-3')
nLUC-SPL9	GACGAGCTCGGTACCATGGAGATGGGTTCCT ACT	CGAGATCTGGTCGACGAGAGACCAGTTGGT ATGGTGA
cLUC-SPL9	TCCCGGGGCGGTACCATGGAGATGGGTTCCT ACT	GTAGTCCATTTGTTGGAGAGACCAGTTGGTA TGG
nLUC-SPL3	GGACGAGCTCGGTACCATGAGTATGAGAAG AAGCAA	ACGAGATCTGGTCGACGTCAGTTGTGCTTTT CCGCC
cLUC-ABI5	GTCCCGGGGCGGTACCATGGTAACTAGAGA AACGAA	TGTAGTCCATTTGTTGGAGTGGACAACCTCGG GTTCC
nLUC-ABI5	GGGACGAGCTCGGTACCATGGTAACTAGAG AAACGAAGT	TACGAGATCTGGTCGACGAGTGGACAACCTC GGGTTCTC
nLUC-ABI3	GGGACGAGCTCGGTACCATGAAAAGCTTGC ATGTGGCGG	TACGAGATCTGGTCGACTTTAACAGTTTGAG AAGTTGGT
nLUC-ABI4	GGGACGAGCTCGGTACCATGGACCCTTTAGC TTCCCAAC	TACGAGATCTGGTCGACATAGAATTCCCCCA AGATGGGA
nLUC-SnRK2.2	GACGAGCTCGGTACCATGGATCCGGCGACTA ATTCAC	CGAGATCTGGTCGACGAGAGCATAAACTATC TCTCC
nLUC-SnRK2.3	GACGAGCTCGGTACCATGGATCGAGCTCCGG TGACC	CGAGATCTGGTCGACGAGAGCGTAAACTATC TCTCC
nLUC-SnRK2.6	GGGACGAGCTCGGTACCATGGATCGACCAG CAGTGAGTG	TACGAGATCTGGTCGACCATTGCGTACACAA TCTCTCCG
cLUC-SnRK2.2	TCCCGGGGCGGTACCATGGATCCGGCGACTA ATTCA	GTAGTCCATTTGTTGGAGAGCATAAACTATCT CTCC
cLUC-SnRK2.3	TCCCGGGGCGGTACCATGGATCGAGCTCCGG TGACC	GTAGTCCATTTGTTGGAGAGCGTAAACTATC TCTCC
cLUC-SnRK2.6	TCCCGGGGCGGTACCATGGATCGACCAGCA GTGAGT	GTAGTCCATTTGTTGCATTGCGTACACAATCT CTCC
cLUC-SPL9-N	TCCCGGGGCGGTACCATGGAGATGGGTTCCT ACT	TGTAGTCCATTTGTTGGTAACGAGAAGCTAA CACAGAG
cLUC-SPL9-C	GTCCCGGGGCGGTACCATGGGGAGGATCGC ACCTTCGCT	GTAGTCCATTTGTTGGAGAGACCAGTTGGTA TGG
cLUC-ABI5-N	GTCCCGGGGCGGTACCATGGTAACTAGAGA AACGAA	TGTAGTCCATTTGTTGATGTTCTCTAACCACA CCAGCC
cLUC-ABI5-M	GTCCCGGGGCGGTACCATGCCACTAATCCT AAACCTAA	TGTAGTCCATTTGTTGACCATCCACTACTCTT TTCCTT
cLUC-ABI5-C	GTCCCGGGGCGGTACCATGCCAGTGGAGAA AGTAGTGGA	TGTAGTCCATTTGTTGGAGTGGACAACCTCGG GTTCC
ABI5-GST	TGGATCCCCGGAATTCATGGTAACTAGAGAA ACGAAGTT	GGCCGCTCGAGTCGACTTAGAGTGGACAAC TCGGGTTCC
MBP-SPL9	TTCAGAAATTCGGATCCATGGAGATGGGTTC AACT	TTGCCTGCAGGTCGACTCAGAGAGCCCAGT TGGTATGGT
MBP-SPL3	TTCAGAAATTCGGATCCATGAGTATGAGAAGA AGC	TTGCCTGCAGGTCGACGTCAGTTGTGCTTTT CCGCCTT
Em1 _{pro} -LUC	CTTTTAGATAGATGCATTCACAA	TTTTTGAAGAAAAAACACG
Em6 _{pro} -LUC	TGGGATGGTACTATATTGGTG	GGCTCTTTAGTTACTTACACAA
SPL9 _{pro} -GFP	ATTACGAATTCGAGCTCATTTAATACTACTTA AATTTAA	CCGGAATTAAGACTAGTGTGGTTTCTCTT ACTCAGAC
SPL9 _{pro} -rSPL9-GFP	ACGAGCTGTACAGATCTATGGAGATGGGTTC CAACTCGG	CGGCCGCTTTAAGATCTTCAGAGAGACCAGT TGGTATGG
SnRK2.6-Flag	CACGGGGGACTCTAGAATGGATCGACCAGC AGTGAG	CCTTGTAGTCCATGTGACCATTCGCGTACAC AATCTCTC
SPL9-S203A	CAGTGGCGTCACCGTCATGGCAGAT	TGACGCCACTGGCCGCCTCATCAC
SPL9-S281A	AGCTTCTGCAGGTTTTGGCCCGATG	CCTGCAGAAGCTCGCCATGTATTG

Supplementary Table 2 | Constructs used in this study

Construct name	Vector	Description
nLUC-SPL9	p1300-35S-nLUC	LCI
cLUC-SPL9	p1300-35S-cLUC	LCI
nLUC-SPL3	p1300-35S-nLUC	LCI
cLUC-ABI5	p1300-35S-cLUC	LCI
nLUC-ABI5	p1300-36S-nLUC	LCI
nLUC-ABI3	p1300-37S-nLUC	LCI
nLUC-ABI4	p1300-38S-nLUC	LCI
nLUC-SnRK2.2	p1300-39S-nLUC	LCI
nLUC-SnRK2.3	p1300-39S-nLUC	LCI
nLUC-SnRK2.6	p1300-39S-nLUC	LCI
cLUC-SnRK2.2	p1300-39S-cLUC	LCI
cLUC-SnRK2.3	p1300-39S-cLUC	LCI
cLUC-SnRK2.6	p1300-39S-cLUC	LCI
cLUC-SPL9-N	p1300-35S-cLUC	LCI
cLUC-SPL9-C	p1300-35S-cLUC	LCI
cLUC-ABI5-N	p1300-35S-cLUC	LCI
cLUC-ABI5-M	p1300-35S-cLUC	LCI
cLUC-ABI5-C	p1300-35S-cLUC	LCI
ABI5-GST	pGEX4T-1	Pull down
MBP-SPL9	pMAL-c2X	Pull down
MBP-SPL3	pMAL-c2X	Pull down
Em1 _{pro} -LUC	pGWB35	Transcriptional activity assay in <i>Nicotiana</i>
Em6 _{pro} -LUC	pGWB35	Transcriptional activity assay in <i>Nicotiana</i>
35S:rSPL9-Myc	pGWB17	Transcriptional activity assay in <i>Nicotiana</i>
nYFP-ABI5	pEarleygate201-YN	BiFC
cYFP-SPL9	pEarleygate202-YN	BiFC
cYFP-SPL3	pEarleygate202-YN	BiFC
SnRK2.6-His	pCOLD	<i>In vitro</i> phosphorylation and semi- <i>in vivo</i> pull-down
SPL9 _{pro} -GFP-rSPL9	p1305-35S-GFP	Transgenic plant
SPL9 _{pro} -GFP-rSPL9(2A)	p1305-35S-GFP	Transgenic plant
SnRK2.6-Flag	p1300-35S-Flag	Transgenic plant

Supplementary Table 3 | Primers used for qRT-PCR

Primer name	Primer sequence (5'-3')
Em1-Q-F	TTCCTCGCCTCTCCTCCTTTGTGT
Em1-Q-R	TTCCTCGCCTCTCCTCCTTTGTGT
Em6-Q-F	TTCCTCGCCTCTCCTCCTTTGTGT
Em6-Q-R	TCTTGGTCCTGAATTTGGATTCTG
SPL9-Q-F	CAAGG TTCAGTTGGTGGAGGA
SPL9-Q-R	ATGATGAGTAGGACTGGCAGGTG
ACT7-Q-F	TCCATGAAACAAC TTACA ACTCCATCA
ACT7-Q-R	TGAAGAAGCTCGCCATGTATTG

Supplementary Table 4 | Primers used for ChIP-qPCR assays

Primer name	Primer sequence (5'-3')
Em1 _{pro} -A-F	ATTTATCCGAAATATTGCACTTAGA
Em1 _{pro} -A-R	TTAATCTTAATCCATTTTCTGTTTC
Em1 _{pro} -B-F	CTAATCCACACCAGATTCTAACATT
Em1 _{pro} -B-R	TTAGGTCTTTTCAAGTTTGTAGTT
Em6 _{pro} -A-F	GGCAACACACAAAATAATTGC
Em6 _{pro} -A-R	TACGAAGAAGACTATAGTAGCTCGC
Em6 _{pro} -B-F	AATCAATTTGATCGGAACTGAAC
Em6 _{pro} -B-R	TGAAAGTAAATATGAATACCTGAG
Em6 _{pro} -C-F	TAGCATTTGAATCTTTTCAAAGTTG
Em6 _{pro} -C-R	GGCATCGACTTTAAGCAGATT
Em6 _{pro} -D-F	AGGAACACAAATAACTTGCCTCGTA
Em6 _{pro} -D-R	TCTCAATACTACGTGACTACGTCAA
Em6 _{pro} -E-F	ACGTCATAATTAGAAAAAGTCAAAA
Em6 _{pro} -E-R	ATTACATACACACTTCACACTTGC
ACT7 _{pro} -F	CGTTTCGCTTTCCTTAGTGTTAGCT
ACT7 _{pro} -R	CACAACGCATGCTAAACAGATCTAG