**Research** Article

# miR-149-3p Is a Potential Prognosis Biomarker and Correlated with Immune Infiltrates in Uterine Corpus Endometrial Carcinoma

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Background. Endocrine disruption is an important factor in the development of endometrial cancer. Expression of miR-149-3p is observed in some cancer types, while its role in uterine corpus endometrial carcinoma (UCEC) is unclear. Methods. The clinical and genomic data and prognostic information on UCEC were obtained for patients from the TCGA database. The Kruskal-Wallis test, Wilcoxon signed-rank test, and logistic regression were used to analyze the relationship between clinical characteristics and miR-149-3p expression. Kaplan-Meier survival curve analysis was used to study the influence of miR-149-3p expression and miR-149-3p target genes on the prognosis of UCEC patients. The TargetScan, PicTar, Gene Ontology (GO), and Kyoto Encyclopedia of Genes and Genomes (KEGG) analyses were used to determine the involvement of miR-149-3p target genes in function. Immune infiltration analysis was used to analyze the functional involvement of miR-149-3p. QRT-PCR was used to validate the expression of miR-149-3p in UCEC cell lines. Results. High expression of miR-149-3p in UCEC was significantly associated with age (P < 0.001), histological type (P < 0.001), histological grade (P < 0.001), tumor invasion (P = 0.014), and radiation therapy (P = 0.011). High miR-149-3p expression predicted poorer overall survival (OS) (HR: 2.56; 95% CI: 1.64–4.00; P < 0.001), progression-free interval (PFI) (HR: 1.85; 95% CI: 1.29–2.65; P = 0.001), and disease-specific survival (DSS) (HR: 2.33; 95% CI: 1.37–3.99; P = 0.002). Low expressions of miR-149-3p target genes, including ADCYAP1R1, CGNL1, CHST3, CYGB, DNAH9, ESR1, HHIP, HIC1, HOXD11, IGF1, INMT, LSP1, MTMR10, NFIC, PLCE1, RARA, SNTN, SPRYD3, and ZBTB7A, were associated with poor OS in UCEC. MiR-149-3p may be involved in the occurrence and development of UCEC via pathways including PI3K-Akt signaling pathway, Ras signaling pathway, AGE-RAGE signaling pathway in diabetic complications, focal adhesion, and MAPK signaling pathway. miR-149-3p may inhibit the function of CD8 T cells, cytotoxic cells, eosinophils, iDC, mast cells, neutrophils, NK CD56bright cells, NK CD56dim cells, pDC, T cells, T helper cells, TFH, Th17 cells, and Treg. miR-149-3p was significantly upregulated in UCEC cell lines compared with endometriotic stromal cells. Conclusion. High expression of miR-149-3p was significantly associated with poor survival in UCEC patients. It may be a promising biomarker of prognosis and response to immunotherapy for UCEC patients.

## 1. Introduction

Uterine corpus endometrial carcinoma (UCEC) is one of the three main gynaecological malignancies, the incidence of which increases over time. Endometrial cancer is an epithelial tumor of the endometrium and a malignant tumor of the female reproductive system with a high incidence, posing a serious threat to the health of women worldwide [1, 2]. It currently ranks second in the incidence of gynaecological cancers in developing countries, behind cervical cancer [3]. According to the pathogenesis and biological behavior characteristics of endometrial cancer, endometrial cancer can be divided into estrogen-dependent (type I) and nonestrogen-dependent (type II), and most endometrial cancers belong to the former, which means endometrial cancer is closely related to endocrine. The etiological mechanisms of UCEC have not been fully elucidated. Risk factors for UCEC that have been identified include unstable postmenopausal or perimenopausal oestrogen levels, obesity, infertility, diabetes, hypertension, and family history [4]. Early clinical signs in patients with UCEC include irregular vaginal bleeding in the postmenopausal or perimenopausal period, pelvic cramps, and abdominal pain. The current lack of effective early diagnosis of UCEC has resulted in many UCEC patients missing the best time for treatment, and the poor prognosis of UCEC patients, especially those with metastases after surgery and radiotherapy, is also one of the important issues to be addressed in the current treatment of UCEC [5]. Although progress has been made in early detection and treatment, a considerable number of advanced UCEC cases have been diagnosed [6, 7]. With the development of gene microarray technology, increasing research data are available to help resolve the complex pathogenesis of UCEC and monitor the disease progress. Therefore, systematic bioinformatic analysis of UCEC-related disease data allows for a rapid search for biomarkers that can be used for UCEC prognosis.

MicroRNAs (miRNAs) are a unique class of endogenous and small noncoding RNAs that are approximately 18 to 25 nucleotides in length. They alter gene expression at the posttranscriptional level primarily through complete or incomplete base pairing with the 3' untranslated region (3'UTR) of their target mRNAs. Translational repression and mRNA degradation are the 2 main pathways through which miRNAs direct gene regulation [8]. Many studies have shown that different miRNAs are abnormally expressed in different tumors and participate in tumor formation and growth as oncogenes or oncogenes [9]. There is substantial evidence that miRNAs are stably detectable in serum and plasma and have the potential to be noninvasive biomarkers for the diagnosis and prognosis of various cancers. miRNAs offer an innovative idea for screening and detection of cancer patients [10-12].

In oral squamous cell carcinoma, reduced levels of miRNA-149-3p lead to malignant progression and predict poor prognosis [13]. Increased miR-149-3p expression significantly inhibited the proliferation, migration, and invasion of bladder cancer cells [14]. Only a few studies mentioned that the lnc HOXB-AS1 was upregulated in endometrial cancer and spongy miR-149-3p upregulated Wnt10b [15]. However, the expression of miR-149-3p in UCEC and its relation to clinical features has not been well studied.

The present study examined the expression of miR-149-3p in UCECs using an online database and analyzed the relationship between expression levels and clinical characteristics. A survival curve was drawn to analyze the relationship between miR-149-3p expression level and overall survival (OS). Important contributions of miR-149-3p target genes to function were identified by TargetScan, PicTari, Gene Ontology (GO), and Kyoto Gene and Genome Encyclopedia (KEGG) analyses. The functionally significant involvement of miR-149-3p was analyzed by immune infiltration analysis. QRT-PCR was used to validate the expression of miR-149-3p in UCEC cell lines. The results of this study could provide new prognostic biomarkers for UCEC.

## 2. Patients and Methods

2.1. Clinical Information. The analysis was carried out according to references [16, 17]. R (version 3.6.3) was used for statistical analysis and visualization. The R package was the base R package. The molecule was hsa-miR-149-3p (MIMAT0004609). The grouping condition was median. The disease was UCEC. Data were obtained from miRNAseq data from level 3 BCGSC miRNA Profiling in the TCGA (https://portal.gdc.cancer.gov/) UCEC project. The miR-NAseq data in RPM (reads per million mapped reads) format were log2-transformed.

#### 2.2. Differential Expression of miR-149-3p

*Unpaired Samples.* The analysis was carried out according to reference [18]. R (version 3.6.3) was used for statistical analysis and visualization. ggplot2 (version 3.3.3) was used for visualization. The molecule was hsa-miR-149-3p. The disease was UCEC. Data were miRNAseq data from level 3 BCGSC miRNA Profiling in TCGA UCEC. The miRNAseq data in RPM format were log2 transformed. Data were unfiltered.

*ROC Curves*. The analysis was carried out according to reference [18]. R (version 3.6.3) was used for statistical analysis and visualization. The R packages were the pROC package (version 1.17.0.1) (for analysis) and the ggplot2 package (version 3.3.3) (for visualization). The molecule was hsa-miR-149-3p. The clinical variable is normal versus tumor. The disease was endometrial cancer. Data were miRNAseq data from level 3 BCGSC miRNA Profiling in the TCGA UCEC project. The miRNAseq data in RPM format were log2 transformed. Data are unfiltered.

2.3. Correlation of miR-149-3p Expression with Clinical Features. Correlation analysis of gene expression with clinical characteristics was carried out according to reference [16]. R (version 3.6.3) was used for statistical analysis and visualization. ggplot2 (version 3.3.3) was used for visualization. The molecule was hsa-miR-149-3p. Clinical variables were age, histological type, tumor invasion, and radiation therapy. The disease was UCEC. Data were miRNAseq data and clinical data from level 3 BCGSC miRNA Profiling in the TCGA UCEC project. The miRNAseq data in RPM format were log2 transformed. Data filtering conditions included removing controls/normal (not all items have controls/ normal) and keeping clinical information available.

Logistics analysis was carried out according to references [18, 19]. R (version 3.6.3) was used for statistical analysis and visualization. R package was mainly a basic package. The statistical method was a dichotomous logistic model. The independent variable was hsa-miR-149-3p. The type of independent variable was low high dichotomous. The disease

was UCEC. Data were miRNAseq data from level 3 BCGSC miRNA Profiling in the TCGA UCEC project. The miR-NAseq data in RPM format were log2 transformed. Data filtering conditions include removal of control/normal (not all items have control/normal) and retention of clinical information.

2.4. Prognostic Value of miR-149-3p Expression in UCEC Patients. The Kaplan-Meier method analysis was carried out according to references [16, 17]. R (version 3.6.3) was used for statistical analysis and visualization. The survinier package (version 0.4.9) was used for visualization. The survival package (version 3.2-10) was used for statistical analysis of survival data. The molecule was hsa-miR-149-3p. Subgroups were 0–50 and 50–100. Prognostic types were OS, PFI, and DSS. The disease was UCEC. Data were miRNAseq data from level 3 BCGSC miRNA Profiling in the TCGA UCEC project. The miRNAseq data in RPM format were log2 transformed. Supplementary data were prognostic data from reference [20]. Data filtering conditions included removal of control/normal (not all items had control/normal) and retention of clinical information.

COX regression analysis was carried out according to references [19-21]. R (version 3.6.3) was used for statistical analysis and visualization. The survival package (version 3.2-10) was used for statistical analysis of survival data. Statistical method was the Cox regression module. The prognosis type was OS. Included variables were clinical stage, primary therapy outcome, age, weight, height, BMI, histological type, residual tumor, histologic grade, tumor invasion (%), menopause status, hormones therapy, diabetes, radiation therapy, surgical approach, and hsa-miR-149-3p. The disease was UCEC. Data were miRNAseq data from level 3 BCGSC miRNA Profiling in the TCGA UCEC project. The miRNAseq data in RPM format were log2 transformed. Supplementary data were prognostic data from reference [20]. Data filtering included removal of control/normal (not all items have control/normal) and retention of clinical information.

Forest plot: software was R (version 3.6.3). R package was the ggplot2 package.

Nomogram plot analysis was carried out according to literature [19, 20]. R package was rms package and survival package. The prognosis type was OS. Included variables were clinical stage, primary therapy outcome, age, histological type, residual tumor, tumor invasion, radiation therapy, and miR-149-3p.

2.5. Predicted Putative Targets of miR-149-3p. miR-149-3p targets were obtained from Database TargetScan, miRanda, TarBase, miRTarBase, miR2Disease, miRecords, and miR-Walk [22–25]. UCEC mRNA expression media files are downloaded from the website (https://bioinfo.life.hust.edu. cn/miR\_path/download.html). The UCEC prognosis-related genes were analyzed according to reference [16]. R (version 3.6.3) was used for statistical analysis and visualization. The surviner package (version 0.4.9) was used for statistical

analysis of survival data. The statistical method was Cox. The subgroups were 0–50 versus 50–100. The prognosis type was OS. The disease was UCEC. Data were miRNAseq data from level 3 BCGSC miRNA Profiling in the TCGA UCEC project. The miRNAseq data were log2 transformed in RPM format. Supplementary data were prognostic data from reference [20]. Data filtering criteria included removal of control/normal (not all items had control/normal) and retention of clinical information. The common genes among miR-149-3p target genes, UCEC-downregulated genes, and UCEC prognosis-related genes were analyzed according to the Venn diagram.

2.6. Relationship between miR-149-3p Target Gene Expression and Prognosis. The analysis was performed according to reference [18]. R (version 3.6.3) was used for statistical analysis and visualization. The survminer package (version 0.4.9) was used for visualization. The survival package (version 3.2-10) was used for statistical analysis of survival data. Molecules were ADCYAP1R1, CGNL1, CHST3, CYGB, DNAH9, ESR1, HHIP, HIC1, HOXD11, IGF1, INMT, LSP1, MTMR10, NFIC, PLCE1, RARA, SNTN, SPRYD3, and ZBTB7A. Subgroups were 0-50 and 50-100. Prognostic types were OS, PFI, and DSS. The disease was UCEC. Data were RNAseq data in level 3 HTSeq-FPKM format from the TCGA UCEC project. RNAseq data in FPKM format were converted to TPM (transcripts per million reads) format and log2 transformed. Supplementary data were prognostic data from reference [20]. Data filtering conditions included removal of control/normal (not all items had control/normal) and retention of clinical information.

2.7. GO and KEGG Analyses of miR-149-3p Targets. The Database for Annotation, Visualization, and Integrated Discovery (DAVID) can provide a comprehensive set of functional annotation tools to facilitate understanding of the biological significance behind a large number of genes. GO and KEGG analyses were performed on the targets of miR-149-3p using the DAVID database (https://david.ncifcrf. gov/) [26–28]. GO and KEGG enrichment pathways (adjusted *P* value less than 0.05) were considered significant categories.

2.8. Immune Infiltration Analysis by ssGSEA. The analysis was performed according to reference [18]. R (version 3.6.3) was used for statistical analysis and visualization. The R package was the GSVA package (version 1.34.0) [29]. The immunoinfiltration algorithm was ssGSEA (built-in algorithm of the GSVA package). The molecule was hsa-miR-149-3p. Immune cells were aDC (activated DC), B cells, CD8 T cells, cytotoxic cells, DC, eosinophils, iDC (immature DC), macrophages, mast cells, neutrophils, NK CD56bright cells, NK CD56dim cells, NK cells, pDC (plasmacytoid DC), T cells, T helper cells, Tcm (T central memory), Tem (T effector memory), Tfh (T follicular helper), Tgd (T gamma delta), Th1 cells, Th17 cells, Th2 cells, and Treg. The disease

was endometrial cancer. Data were miRNAseq data from level 3 BCGSC miRNA Profiling in the TCGA UCEC project. The miRNAseq data in RPM format were log2 transformed. The data filtering condition was to remove control/normal (not all items had control/normal). Markers for 24 immune cells were obtained from reference [30].

2.9. QRT-PCR. Human endometriotic stromal cells (ESC) have been isolated from endometriotic tissue. Human UCEC cells Ishikawa and KLE were obtained from our laboratory. KLE cells were grown in F12. Ishikawa cells were grown in RPMI-1640. 10% fetal bovine serum was added to the medium to maintain the cell state. Add 1% antibiotics to the soil to prevent contamination. Cells were grown in a 37°C incubator containing 5% CO<sub>2</sub>. The miR-149-3p levels in the ESC, Ishikawa, and KLE cell lines were identified by qRT-PCR. The specific steps were performed according to reference [31]. The primer sequences used are as follows: U6, Forward: CTCGCTTCGGCAGCACA, **Reverse:** AACGCTTCACGAATTTGCGT; miR-149-3p, Forward: 5'-ACAGGGGAGGGACGGGGG-3', Reverse: 5'-CAGTG-CAGGGTCCGAGGTATT-3'.

2.10. Statistical Analysis. Statistical analysis was carried out according to reference [18]. All statistical analyses were performed using R (v.3.6.3). The Wilcoxon rank-sum test, chi-square test, and Fisher exact test were used to analyze the relationship between clinical characteristics and miR-149-3p. P values less than 0.05 were considered statistically significant.

### 3. Results

3.1. Clinical Characteristics. A total of 546 patients were analyzed in the present study (Table 1). The clinical stage included 341 Stage I (62.5%), 49 Stage II (9%), 128 Stage III (23.4%), and 28 Stage IV (5.1%). The primary therapy outcome included 20 PD (4.2%), 6 SD (1.3%), 12 PR (2.5%), and 437 CR (92%). The race included 20 Asian (4%), 109 Black or African American (21.8%), and 372 White (74.3%). The age included 205 patients (≤60, 55.5%) and 338 patients (>60, 44.5%). The height included 244 patients (≤160) (47%) and 275 patients (>160) (53%). The weight included 240 patients (≤80) (45.9%) and 283 patients (>80) (54.1%). The BMI included 209 patients ( $\leq$ 30) (40.6%) and 306 patients (>30) (59.4%). The histological type included 409 endometrioid (74.9%), 24 mixed (4.4%), and 113 serous (20.7%). The residual tumor included 371 R0 (90.9%), 22 R1 (5.4%), and 15 R2 (3.7%). The histologic grade included 98 G1 (18.3%), 121 G2 (22.6%), and 316 G3 (59.1%). Tumor invasion (%) included 260 patients (<50) (55.3%) and 210 patients ( $\geq$ 50) (44.7%). The menopause status included 34 pre (6.8%), 17 peri (3.4%), and 449 post (89.8%). The hormone therapy included 297 no (86.6%) and 46 yes (13.4%). The diabetes included 323 no (72.6%) and 122 yes (27.4%). The radiation therapy included 278 no (53.4%) and 243 yes (46.6%). The surgical approach included 207

minimally invasive (39.6%) and 316 open (60.4%). The age range was 57 to 71 years, with a median of 64 years.

3.2. miR-149-3p Expression is Correlated with Poor Clinical Characteristics of UCEC. Genomic data were screened from the TCGA database to obtain data for 546 UCEC tissues and 33 normal tissue samples. miR-149-3p expression level in UCEC tissues  $(0.427 \pm 0.515, n = 546)$  was significantly higher than that in normal tissues  $(0.135 \pm 0.159, n = 33)$  (P = 0.007) (Figure 1(a)). The area under curve (AUC) of miR-149-3p was 0.634 (Figure 1(b)), suggesting that miR-149-3p could be used as a promising biomarker to differentiate UCEC from nontumor tissue.

The characteristics of UCEC patients are shown in Table 1, in which 546 primary UCEC with both clinical and gene expression data were collected from the TCGA database. According to the mean value of the relative miR-149-3p expression, the patients with UCEC were divided into high (n = 273) and low (n = 273) expression groups. As shown in Figure 2 and Table 1, miR-149-3p was significantly related to age (P < 0.001), histological type (P < 0.001), histological grade (P < 0.001), tumor invasion (P = 0.014), and radiation therapy (P = 0.011). As shown in Table 2, miR-149-3p was significantly related to primary therapy outcome (HR: 0.501; 95% CI: 0.247-0.983; P = 0.048), age (HR: 2.006; 95% CI: 1.411–2.863; *P* < 0.001), histological type (HR: 0.311; 95% CI: 0.204–0.470; *P* < 0.001), histologic grade (HR: 0.302; 95% CI: 0.183-0.485; P < 0.001), tumor invasion (HR: 1.608; 95% CI: 1.116-2.324; P < 0.011), and radiation therapy (HR: 1.595; 95% CI: 1.129–2.259; *P* = 0.008).

3.3. Role of miR-149-3p in UCEC Patient Survival. The association between miR-149-3p expression and OS of patients with UCEC was evaluated by Kaplan-Meier analysis, which indicated that the expression of miR-149-3p was correlated with poor OS (HR: 2.56; 95% CI: 1.64-4.00; P < 0.001), PFI (HR: 1.85; 95% CI: 1.29–2.65; P = 0.001), and DSS (HR: 2.33; 95% CI: 1.37-3.99; P = 0.002) of UCEC patients (Figure 3). The above data indicated that miR-149-3p is a prognostic factor, and high miR-149-3p level is associated with poor OS. As shown in Table 3, high expression levels of miR-149-3p were associated with worse OS (HR: 2.56; 95% CI: 1.64–4.00; *P* < 0.001), clinical stage (HR: 3.774; 95% CI: 2.484–5.732; P < 0.001), primary therapy outcome (HR: 0.125; 95% CI: 0.075–0.208; *P* < 0.001), age (HR: 2.002; 95% CI: 1.236–3.242; P = 0.005), histological type (HR: 0.366; 95% CI: 0.241–0.557; *P* < 0.001), residual tumor (HR: 3.081; 95% CI: 1.725–5.503; *P* < 0.001), histologic grade (HR: 22.795; 95% CI: 3.127-163.794; P < 0.001), tumor invasion (HR: 3.124; 95% CI: 1.909-5.112; P < 0.001), and radiation therapy (HR: 0.586; 95% CI: 0.375-0.914; P = 0.018). As shown in Table 3 and Figure 4, clinical stage (HR: 3.774; 95% CI: 2.484–5.732; *P* < 0.001), primary therapy outcome (HR: 0.125; 95% CI: 0.075–0.208; P < 0.001), radiation therapy (HR: 3.774; 95% CI: 2.484–5.732; *P* < 0.001), and hsa-miR-149-3p (HR: 0.125; 95% CI: 0.075-0.208; P < 0.001) were independently correlated with OS in multivariate analysis. The above data indicated that miR-149-3p is a prognostic

TABLE 1: Correlation between miR-149-3p expression and clinical characteristics in UCEC.

Characteristic	Levels	Overall	Low expression of hsa- miR-149-3p	High expression of hsa- miR-149-3p	Р
n		546	273	273	
	Stage I	341 (62.5%)	182 (33.3%)	159 (29.1%)	
	Stage II	49 (9%)	21 (3.8%)	28 (5.1%)	
Clinical stage, n (%)	Stage III	128 (23.4%)	56 (10.3%)	72 (13.2%)	0.208
	Stage IV	28 (5.1%)	14 (2.6%)	14 (2.6%)	
	PD	20 (4.2%)	7 (1.5%)	13 (2.7%)	
Primary therapy outcome,	SD	6 (1.3%)	4 (0.8%)	2 (0.4%)	
n (%)	PR	12 (2.5%)	3 (0.6%)	9 (1.9%)	0.080
(70)	CR	437 (92%)	235 (49.5%)	202 (42.5%)	
	Asian	20 (4%)	14 (2.8%)	6 (1.2%)	
	Black or African	20 (470)	14 (2.070)	0 (1.270)	
Race, <i>n</i> (%)	American	109 (21.8%)	49 (9.8%)	60 (12%)	0.118
	White	372 (74.3%)	185 (36.9%)	187 (37.3%)	
	≤60	205 (37.8%)	125 (23%)	80 (14.7%)	
Age, <i>n</i> (%)	>60	338 (62.2%)	148 (27.3%)	190 (35%)	< 0.001
	≥00 ≤80	240 (45.9%)	113 (21.6%)	127 (24.3%)	
Weight, <i>n</i> (%)	≥80 >80			· · · ·	0.133
	≥80 ≤160	283 (54.1%)	153 (29.3%)	130 (24.9%)	
Height, <i>n</i> (%)	>160	244 (47%)	127 (24.5%)	117 (22.5%)	0.616
		275 (53%)	136 (26.2%)	139 (26.8%)	
BMI, <i>n</i> (%)	≤30 >30	209 (40.6%)	104 (20.2%)	105 (20.4%)	0.743
	>50 Endometrioid	306 (59.4%)	158 (30.7%)	148 (28.7%)	
$\mathbf{U}$ istological true $\mathbf{u}$ (0/)	Mixed	409 (74.9%)	233 (42.7%)	176 (32.2%)	<0.001
Histological type, n (%)		24 (4.4%)	9 (1.6%) 21 (5.7%)	15 (2.7%)	< 0.001
	Serous	113 (20.7%)	31 (5.7%)	82 (15%)	
$\mathbf{D} = \frac{1}{2} \mathbf{J} = \frac{1}{2} \mathbf{J}$	RO	371 (90.9%)	192 (47.1%)	179 (43.9%)	0 422
Residual tumor, $n$ (%)	R1	22 (5.4%)	10 (2.5%)	12 (2.9%)	0.432
	R2	15 (3.7%)	10 (2.5%)	5 (1.2%)	
	G1	98 (18.3%)	72 (13.5%)	26 (4.9%)	.0.001
Histologic grade, n (%)	G2	121 (22.6%)	78 (14.6%)	43 (8%)	< 0.001
	G3	316 (59.1%)	121 (22.6%)	195 (36.4%)	
Tumor invasion (%), $n$	<50	260 (55.3%)	152 (32.3%)	108 (23%)	0.014
(%)	≥50	210 (44.7%)	98 (20.9%)	112 (23.8%)	
	Pre	34 (6.8%)	21 (4.2%)	13 (2.6%)	0.000
Menopause status, n (%)	Peri	17 (3.4%)	9 (1.8%)	8 (1.6%)	0.360
	Post	449 (89.8%)	221 (44.2%)	228 (45.6%)	
Hormones therapy, <i>n</i> (%)	No	297 (86.6%)	158 (46.1%)	139 (40.5%)	0.806
177 (7)	Yes	46 (13.4%)	23 (6.7%)	23 (6.7%)	
Diabetes, n (%)	No	323 (72.6%)	165 (37.1%)	158 (35.5%)	0.475
2 100000, 11 (10)	Yes	122 (27.4%)	57 (12.8%)	65 (14.6%)	
Radiation therapy, <i>n</i> (%)	No	278 (53.4%)	157 (30.1%)	121 (23.2%)	0.011
	Yes	243 (46.6%)	109 (20.9%)	134 (25.7%)	0.011
Surgical approach, n (%)	Minimally invasive	207 (39.6%)	113 (21.6%)	94 (18%)	0.197
<b>o 11</b>	Open	316 (60.4%)	153 (29.3%)	163 (31.2%)	
Age, median (IQR)		64 (57, 71)	62 (55, 69)	66 (59, 72)	< 0.001

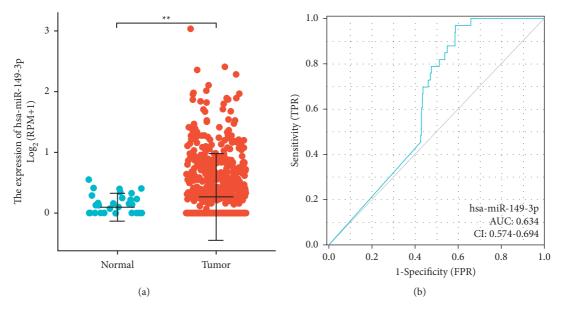


FIGURE 1: Expression of miR-149-3p in 537 UCEC and 33 normal tissues. (a) miR-149-3p relative expression levels (expressed in transcripts per million, TPM) were significantly higher in UCEC tissues ( $3.469 \pm 0.0805$ ) compared with normal tissues ( $2.70 \pm 0.151$ ). (b) ROC. Significance markers: \* \* *P* < 0.01.

factor, and increased miR-149-3p is associated with poor OS. A nomogram was constructed to predict the 1-, 3-, and 5year survival probability of UCEC patients by combing the expression of miR-149-3p with clinical variables, as shown in Figure 5.

3.4. Relationship between miR-149-3p Target Genes and Survival of UCEC Patients. There were 1756 miR-149-3p target genes, 4627 UCEC-downregulated genes, and 1102 UCEC prognosis-related genes in Figure 6. As shown in Figure 4 and Table 4, the 19 common genes included ADCYAP1R1, CGNL1, CHST3, CYGB, DNAH9, ESR1, HHIP, HIC1, HOXD11, IGF1, INMT, LSP1, MTMR10, NFIC, PLCE1, RARA, SNTN, SPRYD3, and ZBTB7A.

3.5. GO and KEGG Analyses of miR-149-3p Target Genes. The shared genes are involved in the biological process, including regulation of cation transmembrane transport, regulation of metal ion transport, extracellular matrix organization, regulation of ion transmembrane transport, and morphogenesis of a branching structure; cellular components, including main axon, collagen-containing extracellular matrix, membrane raft, membrane microdomain, membrane region, and postsynaptic membrane; molecular functions, including growth factor binding, extracellular matrix structural constituent, platelet-derived growth factor binding, SMAD binding, extracellular matrix structural constituent conferring tensile strength (Figure 7). The miR-149-3p target genes are involved in PI3K-Akt signaling pathway, Ras signaling pathway, AGE-RAGE signaling pathway in diabetic complications, focal adhesion, MAPK signaling pathway, proteoglycans in cancer, hypertrophic cardiomyopathy, cholinergic synapse, cocaine addiction, and protein digestion and absorption (Figure 8).

3.6. The Correlation between miR-149-3p Expression and Immune Infiltration. As shown in Figures 9 and 10 and Table 5, miR-149-3p expression was negatively correlated with that of CD8 T cells (P = 0.005), cytotoxic cells (P < 0.001), eosinophils (P < 0.001), iDC (P < 0.001), mast cells (P = 0.002), neutrophils (P < 0.001), NK CD56bright cells (P < 0.001), NK CD56dim cells (P < 0.001), pDC (P < 0.001), T cells (P < 0.001), T helper cells (P = 0.004), TFH (P = 0.024), Th17 cells (P = 0.006), and Treg (P < 0.001).

3.7. Validation of miR-149-3p Expression in Cell Lines. The expression of miR-149-3p in Ishikawa was significantly higher than that in ESC  $(2.608 \pm 0.253 \text{ versus } 0.874 \pm 0.233,$ 

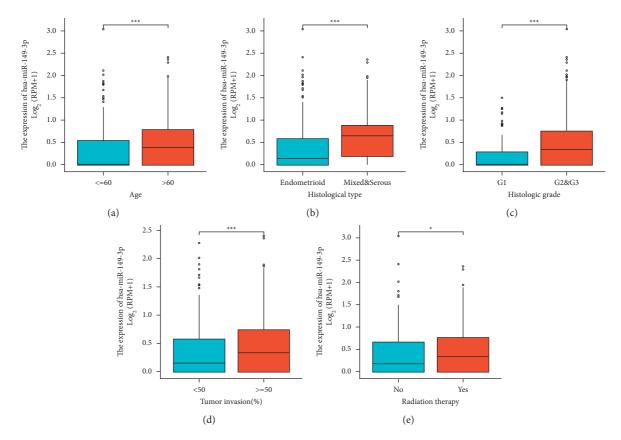


FIGURE 2: Association with miR-149-3p expression and clinical stage in UCEC. (a) Age, (b) histological grade, (c) histological grade, (d) tumor invasion, and (e) radiation therapy. Significance markers: \*P < 0.05; \*\*\*P < 0.001.

TABLE 2: miR-149-3p expression correlated with clinical and pathological features (logistic regression analysis).

Characteristics	Total (N)	Odds ratio (OR)	P value
Clinical stage (stage III and stage IV versus stage I and stage II)	546	1.334 (0.919–1.940)	0.130
Primary therapy outcome (CR versus PD, SD, and PR)	475	0.501 (0.247-0.983)	0.048
Race (White versus Asian and Black or African American)	501	0.965 (0.646-1.441)	0.861
Age (>60 versus ≤60)	543	2.006 (1.411-2.863)	< 0.001
Weight (>80 versus ≤80)	523	0.756 (0.535-1.067)	0.112
Height (>160 versus ≤160)	519	1.109 (0.786-1.567)	0.555
BMI (>30 versus ≤30)	515	0.928 (0.652-1.319)	0.676
Histological type (endometrioid versus mixed and serous)	546	0.311 (0.204-0.470)	< 0.001
Residual tumor (R0 versus R1 and R2)	408	1.097 (0.557-2.182)	0.789
Histologic grade (G1 versus G2 and G3)	535	0.302 (0.183-0.485)	< 0.001
Tumor invasion (%) (≥50 versus <50)	470	1.608 (1.116-2.324)	0.011
Menopause status (pre versus peri and post)	500	0.603 (0.288-1.219)	0.166
Hormones therapy (yes versus no)	343	1.137 (0.609-2.123)	0.686
Diabetes (yes versus no)	445	1.191 (0.785-1.810)	0.412
Radiation therapy (yes versus no)	521	1.595 (1.129-2.259)	0.008
Surgical approach (open versus minimally invasive)	523	1.281 (0.902-1.822)	0.168

P < 0.001) (Figure 11). The expression of miR-149-3p in KLE was significantly higher than that in ESC (1.823  $\pm$  0.055 versus 0.874  $\pm$  0.233, P < 0.001) (Figure 11). These results suggested that miR-149-3p was significantly upregulated in UCEC cell lines compared with endometriotic stromal cells.

## 4. Discussion

The development of type I endometrial cancer is associated with continuous estrogen stimulation of the endometrium without progestin antagonism. The endometrium lacks progesterone antagonism, and continuous stimulation by

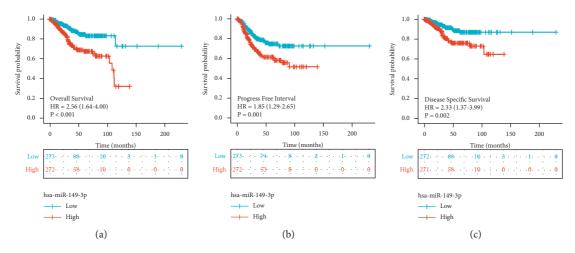


FIGURE 3: The relationship between the expression level of miR-149-3p and prognosis. (a) OS, overall survival; (b) PFI, progress-free interval; (c) DSS, disease survival specific.

TABLE 3: Univariate and multivariate Cox regression analysis of OS and clinical features in UCEC patients.

Characteristics	Total	Univariate analy			
Characteristics		Univariate analysis		Multivariate analysis	
	(N)	HR (95% CI)	P value	HR (95% CI)	P value
Clinical stage (stage I and stage II versus stage III and stage IV)	545	3.774 (2.484–5.732)	<0.001	3.912 (1.711-8.946)	0.001
Primary therapy outcome (PD, SD, and PR versus CR)	475	0.125 (0.075-0.208)	< 0.001	0.299 (0.130-0.689)	0.005
Age ( $\leq 60$ versus >60)	543	2.002 (1.236-3.242)	0.005	1.944 (0.868-4.355)	0.106
Weight (≤80 versus >80)	522	1.032 (0.676-1.576)	0.885		
Height (≤160 versus >160)	518	1.115 (0.728-1.708)	0.616		
BMI ( $\leq 30$ versus $> 30$ )	514	0.942 (0.616-1.442)	0.784		
Histological type (mixed and serous versus endometrioid)	545	0.366 (0.241-0.557)	< 0.001	1.285 (0.615-2.685)	0.505
Residual tumor (R0 versus R1 and R2)	407	3.081 (1.725-5.503)	< 0.001	1.951 (0.811-4.694)	0.136
Histologic grade (G1 versus G2 and G3)	534	22.795 (3.172–163.794)	0.002	156521500.831 (0.000- Inf)	0.997
Tumor invasion (%) (<50 versus ≥50)	469	3.124 (1.909-5.112)	< 0.001	1.111 (0.532-2.320)	0.779
Menopause status (peri and post versus pre)	499	1.070 (0.466-2.457)	0.874		
Hormones therapy (no versus yes)	343	0.872 (0.412-1.845)	0.72		
Diabetes (no versus yes)	445	1.188 (0.733-1.925)	0.485		
Radiation therapy (no versus yes)	521	0.586 (0.375-0.914)	0.018	0.320 (0.165-0.622)	< 0.001
Surgical approach (minimally invasive versus open)	522	0.731 (0.474-1.127)	0.156		
hsa-miR-149-3p (low versus high)	545	2.559 (1.637-4.002)	< 0.001	2.393 (1.161-4.935)	0.018

estrogen will result in a prolonged state of hyperproliferation, which will further develop into endometrial cancer. MiRNAs regulate the cell cycle and cell differentiation and migration, which may also act as tumor suppressor genes or oncogenes during tumorigenesis and tumor development [32]. MiRNAs are also involved in the development of various cancers and are described in detail in UCEC. MiR-181c affects the growth of estrogen-dependent endometrial cancer cells by targeting PTEN, which may be an effective target for endometrial cancer therapy [33]. MiR-320a exerts antitumor effects on endometrial cancer by regulating IGF-1R, which can be used as a target for endometrial cancer gene therapy [34]. Downregulated miR-29b expression is associated with poor prognosis in endometrial cancer (EC) and contributes to the evaluation of EC prognosis [35]. miRNA-205 has potential clinical utility as a prognostic marker for endometrial cancer [36]. Therefore, detection of miR-149-3p is an ideal candidate for improving the early detection of UCEC.

In the present study, miR-149-3p was significantly correlated with age (P < 0.001), histological type (P < 0.001), histological grade (P < 0.001), tumor invasion (P = 0.014), and radiation therapy (P = 0.011). UCEC expressed more miR-149-3p than normal tissue, especially in patients with age (>60), histological type (mixed and serous), histologic grade (G2 and G3), tumor invasion ( $\geq$ 50%), or radiation therapy (yes). These phenomena suggested that miR-149-3p may be involved in tumor development and promote proliferation. And miR-149-3p was highly expressed in UCEC, and patients with high miR-149-3p expression had poorer OS (HR: 2.56; 95% CI: 1.64–4.00; P < 0.001), PFI (HR: 1.85; 95% CI: 1.29–2.65; P = 0.001), and DSS (HR: 2.33; 95% CI: 1.37–3.99; P = 0.002) of UCEC patients. Low expressions of

Characteristics	Total (N)	HR (95% CI) Multivariate analys	is	P value Multivariate analysis
Clinical stage (Stage I&Stage II vs. Stage III&Stage IV)	545	3.912 (1.711-8.946)		0.001
Primary therapy outcome (PD&SD&PR vs. CR)	475	0.299 (0.13-0.689)	<b>H</b>	0.005
Age (<=60 vs. >60)	543	1.944 (0.86-4.355)	<b>↓</b> ●	0.106
Histological type (Mixed&Serous vs. Endometrioid)	545	1.285 (0.615-2.685)	⊭⊣	0.505
Residual tumor (R0 vs. R1&R2)	407	1.951 (0.81-4.694)	¦ ¦⊷	0.136
Tumor invasion (%) (<50 vs. >=50)	469	1.111 (0.532-2.320)	<b>  </b> −	0.779
Radiation therapy (No vs. Yes)	521	0.320 (0.165-0.622)	4	<0.001
hsa-miR-149-3p (Low vs. High)	545	2.393 (1.61-4.935)	<b> </b> -●	0.018
			0.0 2.5 5.0 7.5	

FIGURE 4: UCEC for multivariate Cox regression analysis of forest plots.

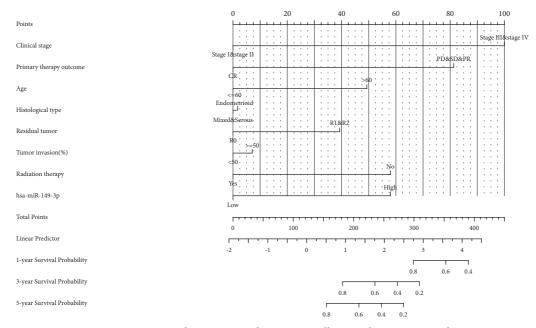


FIGURE 5: Nomogram predicts 1-, 3-, and 5-year overall survival in patients with UCEC.

miR-149-3p target genes, including ADCYAP1R1, CGNL1, CHST3, CYGB, DNAH9, ESR1, HHIP, HIC1, HOXD11, IGF1, INMT, LSP1, MTMR10, NFIC, PLCE1, RARA, SNTN, SPRYD3, and ZBTB7A, were associated with poor OS in UCEC. Lymphovascular space invasion (LVSI) has an independent influence on the poor prognosis of UCEC patients [37–39]. And this index can be used as an important reference indication for adjuvant therapy in endometrial cancer patients after surgery. The data of this study were not available for patients' LVSI data because they were obtained from TCGA UCEC. The correlation between miR-149-3p and LVSI in patients with endometrial cancer in the real world needs to be further investigated.

miR-149-3p inhibits bladder cancer proliferation, migration, and invasion targeting S100A4 [14]. Long noncoding RNA ARAP1-AS1 promotes cervical cancer progression through the regulation of miR-149-3p and

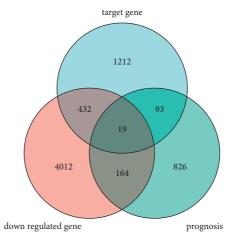


FIGURE 6: Venn diagram of common genes for miR-149-3p target genes, UCEC-downregulated genes, and prognosis-related genes.

TABLE 4: The relationship between miR-149-3p target gene expression level and prognosis.

Gene name	HR	CI	P value
PLCE1	0.49	0.318-0.754	0.001
DNAH9	0.66	0.437-0.997	0.048
ADCYAP1R1	0.655	0.433-0.991	0.045
RARA	0.622	0.408-0.949	0.027
IGF1	0.62	0.409-0.939	0.024
CYGB	0.597	0.394-0.905	0.015
CHST3	0.638	0.421-0.967	0.034
HIC1	0.623	0.412-0.943	0.025
INMT	0.603	0.397-0.916	0.018
CGNL1	0.616	0.406-0.934	0.022
NFIC	0.499	0.326-0.765	0.001
ESR1	0.519	0.339-0.795	0.003
HHIP	0.659	0.437-0.993	0.046
HOXD11	0.65	0.430-0.984	0.042
ZBTB7A	0.613	0.404-0.932	0.022
SPRYD3	0.641	0.423-0.972	0.036
MTMR10	0.617	0.406-0.936	0.023
LSP1	0.503	0.328-0.771	0.002
SNTN	0.613	0.406-0.925	0.020

POU2F2 [40]. miR-149-3p inhibits cell proliferation by targeting AKT2 in oral squamous cell carcinoma [41]. In this study, GO and KEGG analyses showed that the shared genes are involved in PI3K-Akt signaling pathway, Ras signaling pathway, AGE-RAGE signaling pathway in diabetic complications, focal adhesion, MAPK signaling pathway, proteoglycans in cancer, hypertrophic cardiomyopathy, cholinergic synapse, cocaine addiction, and protein digestion and absorption in UCEC. Phosphorylated proteome is involved in smooth muscle tumor growth, and biomimetic analysis revealed ten tumorigenic signaling pathways (FAS signaling pathway, p38 MAPK pathway, VEGF signaling pathway, Rho GTPase regulation of cytoskeleton, integrin signaling pathway, apoptosis signaling pathway, angiogenesis, gonadotropin-releasing hormone receptor pathway, ubiquitin-proteasome pathway, chemokines, and cytokinemediated inflammatory signaling pathways) and four phosphoproteins (HSPA5, HSPB1, HSPD1, and PDRX2) involved in apoptosis and inhibition of cell survival [42]. The relationship between miR-149-3p and phosphoproteins needs further investigation.

Immune cell infiltration has emerged as a new indicator of prognosis for patients with different types of solid tumors [43]. There is heterogeneity in the immune response between tumors [44]. Immune infiltration in UCEC is currently a hot topic, and an understanding of immune infiltration will

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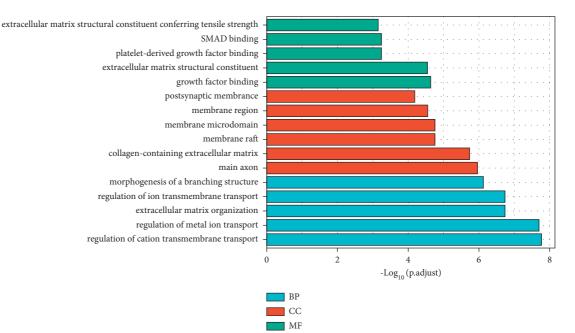


FIGURE 7: GO analysis of miR-149-3p target genes.

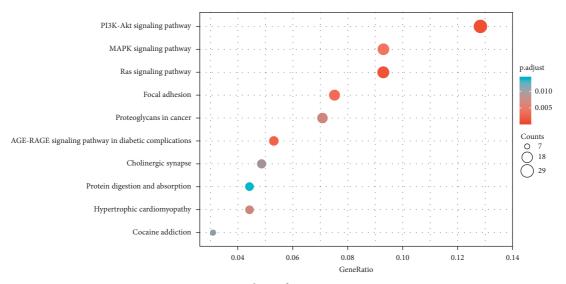


FIGURE 8: KEGG analysis of miR-149-3p target genes.

facilitate the development of immunotherapy for UCEC. The results of this study showed that miR-149-3p expression was correlated with infiltration of CD8 T cells, cytotoxic cells, eosinophils, iDC, mast cells, neutrophils, NK CD56bright cells, NK CD56dim cells, pDC, T cells, T helper cells, TFH, Th17 cells, and Treg in UCEC. It indicated that miR-149-3p

may promote the function of CD8 T cells, cytotoxic cells, eosinophils, iDC, mast cells, neutrophils, NK CD56bright cells, NK CD56dim cells, pDC, T cells, T helper cells, TFH, Th17 cells, and Treg.

This study is limited to the analysis of UCEC tissues in the TCGA database, and further studies, including

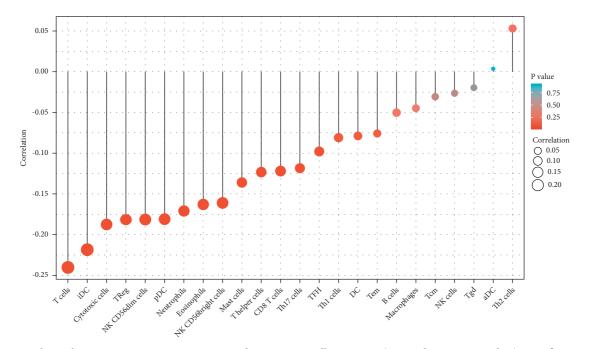


FIGURE 9: Correlation between miR-149-3p expression and 24 immune cells in UCEC (grouped comparison plots). Significance markers: \*P < 0.05; \*\* P < 0.01; \*\* \* P < 0.001.

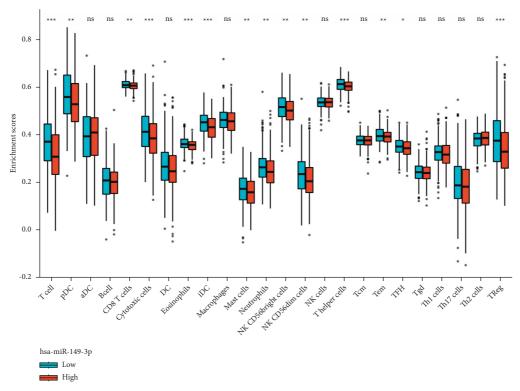


FIGURE 10: Correlation between miR-149-3p expression and 24 immune cells in UCEC (lollipop chart). The size of the dots indicates the absolute value of spearman *r*.

Gene name	Cell type	Correlation coefficient (spearman)	P value (spearman)
hsa-miR-149-3p	aDC	0.004	0.933
hsa-miR-149-3p	B cells	-0.05	0.244
hsa-miR-149-3p	CD8 T cells	-0.122	0.005
hsa-miR-149-3p	Cytotoxic cells	-0.187	< 0.001
hsa-miR-149-3p	DC	-0.079	0.068
hsa-miR-149-3p	Eosinophils	-0.163	< 0.001
hsa-miR-149-3p	iDC	-0.218	< 0.001
hsa-miR-149-3p	Macrophages	-0.045	0.298
hsa-miR-149-3p	Mast cells	-0.136	0.002
hsa-miR-149-3p	Neutrophils	-0.171	< 0.001
hsa-miR-149-3p	NK CD56bright cells	-0.161	< 0.001
hsa-miR-149-3p	NK CD56dim cells	-0.181	< 0.001
hsa-miR-149-3p	NK cells	-0.027	0.539
hsa-miR-149-3p	pDC	-0.181	< 0.001
hsa-miR-149-3p	T cells	-0.24	< 0.001
hsa-miR-149-3p	T helper cells	-0.123	0.004
hsa-miR-149-3p	Ťcm	-0.031	0.476
hsa-miR-149-3p	Tem	-0.076	0.08
hsa-miR-149-3p	TFH	-0.098	0.024
hsa-miR-149-3p	Tgd	-0.019	0.653
hsa-miR-149-3p	Th1 cells	-0.081	0.062
hsa-miR-149-3p	Th17 cells	-0.118	0.006
hsa-miR-149-3p	Th2 cells	0.053	0.221
hsa-miR-149-3p	Treg	-0.182	< 0.001

TABLE 5: Expression of miR-149-3p in relation to immune cells (spearman method).

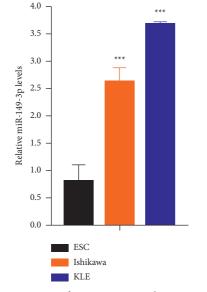


FIGURE 11: Expression of miR-149-3p in human endometriotic stromal cells (ESC), UCEC cell lines Ishikawa and KLE.

serological and clinical data and cellular experiments, are needed to confirm the results.

## 5. Conclusion

miR-149-3p was highly expressed in UCECs and correlated with poorer OS compared with normal tissues. The high miR-149-3p expression in UCEC was significantly associated with age (P < 0.001), histological type (P < 0.001), histological grade (P < 0.001), tumor invasion (P = 0.014),

and radiation therapy (P = 0.011). miR-149-3p may be involved in the occurrence and development of UCEC via pathways, including PI3K-Akt signaling pathway, Ras signaling pathway, AGE-RAGE signaling pathway in diabetic complications, focal adhesion, and MAPK signaling pathway. Expression of miR-149-3p was correlated with immune infiltration in UCEC. This study partially elucidates the role of miR-149-3p in UCEC and provides a promising biomarker for prognosis and immunotherapy response in UCEC patients.

### **Data Availability**

All data generated or analyzed during this study are included in this article. The datasets generated in this study are available from TCGA that provides free resources.

### **Ethical Approval**

TCGA is a public database. Ethical permission was obtained for patients participating in the database. Users can download relevant data for free to conduct research and publish relevant articles. Our research is based on opensource data, so there are no ethical issues.

## **Conflicts of Interest**

The authors declare that they have no conflicts of interest.

#### **Authors' Contributions**

Xiaoyuan Lu and Buze Chen conceived and designed the experiments; Li Jing, Sicong Liu, and Haihong Wang

performed the experiments; Xiaoyuan Lu and Buze Chen analyzed the data; Xiaoyuan Lu, LiJing, and Buze Chen prepared the figures and/or tables; Xiaoyuan Lu, Li Jing, Sicong Liu, Haihong Wang, and Buze Chen drafted the work and revised it critically for important content. Xiaoyuan Lu and Li Jing contributed equally to this work.

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#### References

- T. R. Sponholtz, J. R. Palmer, L. Rosenberg, E. E. Hatch, L. L. Adams-Campbell, and L. A. Wise, "Reproductive factors and incidence of endometrial cancer in U.S. black women," *Cancer Causes & Control*, vol. 28, no. 6, pp. 579–588, 2017.
- [2] L. A. Cantrell and F. Backes, "Highlights from the society of gynecologic oncology 2017 annual meeting on women's cancer," *Gynecologic Oncology*, vol. 145, no. 3, pp. 483–485, 2017.
- [3] R. L. Siegel, K. D. Miller, and A. Jemal, "Cancer statistics, 2017," CA: A Cancer Journal for Clinicians, vol. 67, no. 1, pp. 7–30, 2017.
- [4] E. L. Busch, M. Crous-Bou, J. Prescott et al., "Endometrial cancer risk factors, hormone receptors, and mortality prediction," *Cancer Epidemiology Biomarkers & Prevention*, vol. 26, no. 5, pp. 727–735, 2017.
- [5] J. Stubert and B. Gerber, "Current issues in the diagnosis and treatment of endometrial carcinoma," *Geburtshilfe und Frauenheilkunde*, vol. 76, no. 2, pp. 170–175, 2016.
- [6] Y. Suryo Rahmanto, W. Shen, X. Shi et al., "Inactivation of Arid1a in the endometrium is associated with endometrioid tumorigenesis through transcriptional reprogramming," *Nature Communications*, vol. 11, no. 1, p. 2717, 2020.
- [7] F. Amant, P. Moerman, P. Neven, D. Timmerman, E. Van Limbergen, and I. Vergote, "Endometrial cancer," *The Lancet*, vol. 366, no. 9484, pp. 491–505, 2005.
- [8] Y. Moran, M. Agron, D. Praher, and U. Technau, "The evolutionary origin of plant and animal microRNAs," *Nature Ecology & Evolution*, vol. 1, no. 3, p. 27, 2017.
- [9] S. M. Masud Karim, L. Liu, T. D. Le, and J. Li, "Identification of miRNA-mRNA regulatory modules by exploring collective group relationships," *BMC Genomics*, vol. 17, no. S1, p. 7, 2016.
- [10] K. Zhang, Y.-W. Wang, Y.-Y. Wang et al., "Identification of microRNA biomarkers in the blood of breast cancer patients based on microRNA profiling," *Gene*, vol. 619, pp. 10–20, 2017.
- [11] S. Azarbarzin, M. A. H. Feizi, R. Safaralizadeh, M. Kazemzadeh, and A. Fateh, "The value of MiR-383, an intronic MiRNA, as a diagnostic and prognostic biomarker in intestinal-type gastric cancer," *Biochemical Genetics*, vol. 55, no. 3, pp. 244–252, 2017.
- [12] A. Valentino, P. Reclusa, R. Sirera et al., "Exosomal micro-RNAs in liquid biopsies: future biomarkers for prostate cancer," *Clinical and Translational Oncology*, vol. 19, no. 6, pp. 651–657, 2017.
- [13] Q. Shen, P. Xiong, D. Yang, and L. Chen, "Downregulated microRNA-149-3p triggers malignant development and

predicts worse prognosis in oral squamous cell carcinoma," *Archives of Oral Biology*, vol. 134, Article ID 105336, 2022.

- [14] D. Yang, G. Du, A. Xu, X. Xi, and D. Li, "Expression of miR-149-3p inhibits proliferation, migration, and invasion of bladder cancer by targeting S100A4," *American Journal of Cancer Research*, vol. 7, no. 11, pp. 2209–2219, 2017.
- [15] D. Liu, M. Qiu, L. Jiang, and K. Liu, "Long noncoding RNA HOXB-AS1 is upregulated in endometrial carcinoma and sponged miR-149-3p to upregulate Wnt10b," *Technology in Cancer Research and Treatment*, vol. 19, Article ID 153303382096746, 2020.
- [16] B. Chen, X. Jin, H. Wang, Q. Zhou, G. Li, and X. Lu, "Expression, clinical significance, and prospective pathway signaling of miR-501-3p in ovarian cancer based on database and informatics analysis," *International Journal of General Medicine*, vol. 14, pp. 5193–5201, 2021.
- [17] C. Wei, X. Liu, Q. Wang, Q. Li, and M. Xie, "Identification of hypoxia signature to assess the tumor immune microenvironment and predict prognosis in patients with ovarian cancer," *International Journal of Endocrinology*, vol. 2021, Article ID 4156187, 16 pages, 2021.
- [18] X. Lu, G. Li, S. Liu, H. Wang, Z. Zhang, and B. Chen, "Bioinformatics analysis of KIF1A expression and gene regulation network in ovarian carcinoma," *International Journal* of General Medicine, vol. 14, pp. 3707–3717, 2021.
- [19] H. Xu, H. Wang, G. Li, X. Jin, and B. Chen, "The immunerelated gene ELF3 is a novel biomarker for the prognosis of ovarian cancer," *International Journal of General Medicine*, vol. 14, pp. 5537–5548, 2021.
- [20] J. Liu, T. Lichtenberg, K. A. Hoadley et al., "An integrated TCGA pan-cancer clinical data resource to drive high-quality survival outcome analytics," *Cell*, vol. 173, no. 2, pp. 400–416, 2018.
- [21] Y. Cui, Z. Zhou, Y. Chai, and Y. Zhang, "Upregulated GSDMB in clear cell renal cell carcinoma is associated with immune infiltrates and poor prognosis," *Journal of Immunology Research*, vol. 2021, Article ID 7753553, 22 pages, 2021.
- [22] V. Agarwal, G. W. Nam, J.-W. Bartel, and D. P. Au, "Predicting effective microRNA target sites in mammalian mRNAs," *Elife*, vol. 4, Article ID e05005, 2015.
- [23] H. R. Chiang, L. W. Schoenfeld, J. G. Ruby et al., "Mammalian microRNAs: experimental evaluation of novel and previously annotated genes," *Genes & Development*, vol. 24, no. 10, pp. 992–1009, 2010.
- [24] W. Liu and X. Wang, "Prediction of functional microRNA targets by integrative modeling of microRNA binding and target expression data," *Genome Biology*, vol. 20, no. 1, p. 18, 2019.
- [25] A. Krek, D. Grün, M. N. Poy et al., "Combinatorial microRNA target predictions," *Nature Genetics*, vol. 37, no. 5, pp. 495–500, 2005.
- [26] D. W. Huang, B. T. Sherman, and R. A. Lempicki, "Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources," *Nature Protocols*, vol. 4, no. 1, pp. 44–57, 2009.
- [27] D. W. Huang, B. T. Sherman, and R. A. Lempicki, "Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists," *Nucleic Acids Research*, vol. 37, no. 1, pp. 1–13, 2009.
- [28] M. Zhai, P. Luan, Y. Shi et al., "Identification of three significant genes associated with immune cells infiltration in dysfunctional adipose tissue-induced insulin-resistance of obese patients via comprehensive bioinformatics analysis,"

International Journal of Endocrinology, vol. 2021, Article ID 8820089, 13 pages, 2021.

- [29] S. Hänzelmann, R. Castelo, and J. Guinney, "GSVA: gene set variation analysis for microarray and RNA-seq data," *BMC Bioinformatics*, vol. 14, no. 1, p. 7, 2013.
- [30] G. Bindea, B. Mlecnik, M. Tosolini et al., "Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer," *Immunity*, vol. 39, no. 4, pp. 782–795, 2013.
- [31] X. Lu, G. Li, S. Liu, H. Wang, and B. Chen, "MiR-585-3p suppresses tumor proliferation and migration by directly targeting CAPN9 in high grade serous ovarian cancer," *Journal of Ovarian Research*, vol. 14, no. 1, p. 90, 2021.
- [32] X. Yuan, Y. Zhang, and Z. Yu, "Expression and clinical significance of miR-3615 in hepatocellular carcinoma," *Journal of International Medical Research*, vol. 49, no. 1, Article ID 030006052098154, 2021.
- [33] L. Zhuang, H. Qu, J. Cong, H. Dai, and X. Liu, "MiR-181c affects estrogen-dependent endometrial carcinoma cell growth by targeting PTEN," *Endocrine Journal*, vol. 66, no. 6, pp. 523–533, 2019.
- [34] S. Shu, X. Xu, M. Gao et al., "MicroRNA-320a acts as a tumor suppressor in endometrial carcinoma by targeting IGF-1R," *International Journal of Molecular Medicine*, vol. 43, no. 3, pp. 1505–1512, 2019.
- [35] H. Wang, T.-T. Wang, and X.-P. Lv, "Expression and prognostic value of miRNA-29b in peripheral blood for endometrial cancer," *Future Oncology*, vol. 14, no. 14, pp. 1365–1376, 2018.
- [36] M. Wilczynski, J. Danielska, M. Dzieniecka, B. Szymanska, M. Wojciechowski, and A. Malinowski, "Prognostic and clinical significance of miRNA-205 in endometrioid endometrial cancer," *PLoS One*, vol. 11, no. 10, Article ID e0164687, 2016.
- [37] J. M. Briët, H. Hollema, N. Reesink et al., "Lymphvascular space involvement: an independent prognostic factor in endometrial cancer," *Gynecologic Oncology*, vol. 96, no. 3, pp. 799–804, 2005.
- [38] S. R. Guntupalli, I. Zighelboim, N. T. Kizer et al., "Lymphovascular space invasion is an independent risk factor for nodal disease and poor outcomes in endometrioid endometrial cancer," *Gynecologic Oncology*, vol. 124, no. 1, pp. 31–35, 2012.
- [39] H. Sahin, F. C. Sarioglu, M. Bagci, T. Karadeniz, H. Uluer, and M. Sanci, "Preoperative magnetic resonance volumetry in predicting myometrial invasion, lymphovascular space invasion, and tumor grade: is it valuable in international federation of gynecology and obstetrics stage I endometrial cancer?" *International Journal of Gynecological Cancer*, vol. 28, no. 4, pp. 666–674, 2018.
- [40] L. Zhou and X.-L. Xu, "Long non-coding RNA ARAP1-AS1 facilitates the progression of cervical cancer by regulating miR-149-3p and POU2F2," *Pathobiology*, vol. 88, no. 4, pp. 301–312, 2021.
- [41] Q. Shen, H. Zhu, Q. Lei, L. Chen, D. Yang, and W. Sui, "MicroRNA-149-3p inhibits cell proliferation by targeting AKT2 in oral squamous cell carcinoma," *Molecular Medicine Reports*, vol. 23, no. 3, p. 172, 2020.
- [42] B. Ura, L. Monasta, G. Arrigoni et al., "Phosphoproteins involved in the inhibition of apoptosis and in cell survival in the leiomyoma," *Journal of Clinical Medicine*, vol. 8, no. 5, p. 691, 2019.
- [43] M. Suarez-Carmona, P. Chaorentong, J. N. Kather et al., "CCR5 status and metastatic progression in colorectal

cancer," OncoImmunology, vol. 8, no. 9, Article ID e1626193, 2019.

[44] Y. Dou, E. A. Kawaler, D. Cui Zhou et al., "Proteogenomic characterization of endometrial carcinoma," *Cell*, vol. 180, no. 4, pp. 729–748, 2020.