

Review

Metabolomics: A Tool to Understand the Impact of Genetic Mutations in Amyotrophic Lateral Sclerosis

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Abstract: Metabolomics studies performed in patients with amyotrophic lateral sclerosis (ALS) reveal a set of distinct metabolites that can shed light on the pathological alterations taking place in each individual. Metabolites levels are influenced by disease status, and genetics play an important role both in familial and sporadic ALS cases. Metabolomics analysis helps to unravel the differential impact of the most common ALS-linked genetic mutations (as *C9ORF72, SOD1, TARDBP,* and *FUS*) in specific signaling pathways. Further, studies performed in genetic models of ALS reinforce the role of TDP-43 pathology in the vast majority of ALS cases. Studies performed in differentiated cells from ALS-iPSC (induced Pluripotent Stem Cells) reveal alterations in the cell metabolism that are also found in ALS models and ultimately in ALS patients. The development of metabolomics approaches in iPSC derived from ALS patients allow addressing and ultimately understanding the pathological mechanisms taking place in any patient. Lately, the creation of a "patient in a dish" will help to identify patients that may benefit from specific treatments and allow the implementation of personalized medicine.

Keywords: ALS; metabolomics; genetics; iPSC; personalized medicine

1. Introduction

Amyotrophic lateral sclerosis (ALS) is the most common motor neuron disease in adults, with an incidence of around two per 100,000 persons/year [1]. ALS is characterized by the progressive loss of both upper and lower motor neurons, and patients usually die after 3–5 years of symptoms onset. The pathogenesis of ALS is only partially understood, with several studies highlighting that aberrant RNA metabolism and protein aggregation, glutamatergic toxicity, oxidative stress, and deficits in axonal transport can lead to motor neuron death. Among all the hypotheses put forward to explain the disease, the major role of genetics is worldwide accepted with more than 50 genes currently linked to familial forms of ALS (fALS) that account approximately for 10% of ALS cases [2]. Most of ALS causative genes are linked to the pathogenic mechanisms described before [3]. Interestingly, in the other 90% of ALS cases (sporadic, sALS with no familial hereditary component involved), similar pathogenic mechanisms are observed, and a non-negligible proportion of these cases presents deleterious mutations in the same genes [4].

Such pathogenic mechanisms produce metabolic alterations in each affected cell (as the main target in ALS, motor neurons, but also surrounding cells such as astrocytes) that eventually reflect in the whole organism. For example, it is well known that most of the ALS patients present a hypermetabolic state [5–8], although a recent meta-analysis showed some controversy in this matter [9].



"Omics" approaches are capable of detecting these alterations in the whole organism. In this sense, metabolomics is capable of investigate the whole metabolome, and the metabolomics profile can highlight a particular metabolic pathway that is under or over-regulated in the patient or a group of patients. Moreover, metabolomics analyses reveal pathways that are commonly altered in sporadic patients with no genetic mutation detected and in known genetic-linked ALS cases, highlighting common pathogenic mechanisms. Further, it can help to identify subgroups of ALS patients—with and without mutations—that could benefit from a particular therapy. This knowledge is important to (1) identify biomarkers for ALS; (2) define homogeneous subgroups at the metabolomics level; and (3) identify patients with alterations in metabolism that could be compensated by targeted therapy.

In this review, we will discuss the most common mutations linked to ALS and present the studies that report metabolic alterations associated with these genetic findings in patients and models of ALS. As metabolomics identifies targets that might be attractive for therapeutic manipulation, we will also discuss some studies that evaluated the possible therapeutic effect of targeting and compensating the alterations found in the metabolome. Finally, studies performed in induced Pluripotent Stem Cells (iPSC) derived from ALS patients reproduce the same metabolic alterations found in the patient. The development of iPSC from ALS patients will help to create the "ALS patient in a dish", to study drug response and to increase our understanding about the pathological mechanisms linked to each case.

2. Overview of ALS Genetics

Almost all familial cases of ALS are inherited in an autosomal dominant manner [2]. Genome-wide association studies (GWAS) and next-generation sequencing techniques allowed the analysis of very large datasets, contributing to the identification of around 50 potentially causative or disease-modifying genes linked to ALS. Among these, four genes are more frequently associated with ALS [10]: these include variants of *C9ORF72*, *SOD1*, *TARDBP*, and *FUS*, which account for approximately 55.5% of fALS and 7.4% of sALS cases in Europe [11]. Low-frequency variants could contribute significantly to these apparent sporadic cases, but low-allele frequencies may not be captured by current genotyping arrays, and effect sizes may not be large enough to detect it by linkage analysis in families [12]. Moreover, relatively rare but potentially deleterious variants can hinder the determination of their pathogenicity [11].

2.1. C9ORF72

Hexanucleotide repeat expansions (GGGGCC) in the intron 1 of chromosome 9 open reading frame 72 (*C9ORF72*) gene are the most common inherited cause of ALS in Europe, accounting for 34% of fALS and 5% of sALS (Table 1) [4]. While the normal gene presents 5 to 10 copies of these hexanucleotide repeats, the *C9ORF72* mutation found in ALS patients presents hundreds to thousands of repeats [13,14]. Although the exact role of the C9ORF72 protein is not well understood yet, some reports suggest its involvement in the autophagy–lysosome pathway [15–17]. ALS patients present decreased levels of C9ORF72 protein and mRNA, leading to the hypothesis that a loss of function of this protein could be pathological. However, several studies report that the knockdown of *C9ORF72* in mouse and ALS-derived iPSC does not cause cell degeneration [18–23]. Two other proposed pathological mechanisms involve the gain of function: first, RNA toxicity induced by sense, and secondly, anti-sense RNA that sequesters RNA-binding proteins and form foci in the cells [18,24,25]. Second, repeat-associated non-ATG translation of dipeptide repeat proteins (DPRs) inclusions cause toxicity to neurons [26–32].

2.2. SOD1

The protein SOD1 (or Cu/Zn-superoxide dismutase 1) plays an important antioxidant role in the cell. Located in the cytoplasm and in the intermembrane space of the mitochondria (mSOD1), this protein neutralizes the superoxide species produced during the cellular respiration [33]. The involvement of *SOD1* mutations in ALS was first described in 1993 [34]. Since then, studies point out that *SOD1*

mutations account for 15%–20% of fALS and around 2% of sALS cases [4], but there is controversy regarding if all mutations are indeed pathogenic [35] (Table 1). Around 185 disease-associated variations in the *SOD1* gene have been identified (the majority being missense mutations) [36]. Disease duration, severity, and site of onset (bulbar or limb) differ significantly and depend on the variants involved (variants and their phenotypes were reviewed elsewhere [11]).

2.3. TARDBP

Transactive Response DNA-binding protein 43 (TDP-43) is a protein of 43 kDa encoded by the *TARDBP* gene. TDP-43 binds to pyrimidine-rich DNA and RNA motifs and is implicated in multiple steps of transcriptional and posttranscriptional regulation. To date, at least 48 pathogenic variants in *TARDBP* have been associated with ALS [37]. Even if mutations in *TARDBP* account for 3% of fALS and around 1.5% of sALS (Table 1), 97% of all ALS cases present degenerated motor neurons containing cytoplasmic aggregates rich in TDP-43, making these aggregates the key hallmark for ALS. Furthermore, TDP-43 aggregates were described in fALS caused by mutations in other 19 genes (as *SOD1* and *C9ORF72*, for example) [38]. These aggregates consist of aberrantly phosphorylated and ubiquitinated full-length TDP-43, as well as 35- and 25-kDa C-terminal fragments of the protein [39,40]. While much has been discussed whether TDP-43-associated neurotoxicity is due to a potential loss of function or gain of function, more and more research point to the central role of TDP-43 aggregates in mediating the motor neuron death observed in ALS patients [38].

2.4. FUS

Fused in sarcoma (FUS) is a ubiquitously expressed RNA binding protein found aggregated in ALS patients with pathogenic variants in the *FUS* gene. The first variants were identified in 2009 and are frequently associated with early onset and juvenile ALS [41–45]. Today, more than 50 autosomal dominant variants were identified (from missense mutations to nonsense mutations) [37]. Interestingly, FUS cytoplasmic aggregates are only found in FUS-ALS patients, and these patients do not present TDP-43 aggregates [38,42]. As TDP-43, FUS is predominantly a nuclear protein that can shuttle between the nucleus and the cytoplasm [46], and many of the pathogenic variants are found within the nuclear localization signal of the FUS protein, leading to the redistribution of FUS to the cytoplasm [47,48] (Table 1).

Gene	Function of Coded Protein	fALS/sALS Cases (%)	Alteration in Metabolome	Model
C9ORF72	Autophagy–lysosome pathway	34/5	↓HDL	FTLD [49]
SOD1	Antioxidant	15-20/2	↓ aminoacids ↓ aminoacids; ↑ glycolysis ↓ glutamate ↑ putrescine and spermidine; ↓ hydroxyproline ↑ creatinine	ALS patients [50] NSC-34 cells [51] Motor neuron/ astrocytes cultures [52] mice [53] ALS patients [53]
TARDBP	RNA metabolism	3/1.5	↓ carnitine and beta-hydroxybutyrate ↑ phosphoenolpyruvate and pyruvate ↑ fatty acids	Drosophila [54] Drosophila [55] HEK293T cells [56]
FUS	RNA metabolism	2.4/0.16 [57] 3.8 [58] 4.1 [42]	none	iPSC-derived motor neurons [59]
SNP rs1985243	not described	-	↑ gamma-glutamylphenylalanine	ALS patients [60]

Table 1. Metabolomics findings from studies performed in genes associated with amyotrophic lateral sclerosis (ALS) cases.

fALS: familial ALS; FTLD: frontotemporal lobar degeneration; sALS: sporadic ALS; SNP: single nucleotide polymorphisms.

3. Metabolomics and Genetic-Linked ALS: A Way into the Targeted Treatment

Metabolomics evaluates a vast array of metabolites in a biological sample, thus reflecting the overall metabolome of an individual at a given time. For example, metabolites levels are influenced by food and lifestyle choices, the environment, disease mechanisms, and exposure to drugs or pharmacological treatments [61]. With this in mind, it is logical to hypothesize that the genetic background of an individual may play a role in the metabolic profile. Further, it can help to identify diseases subtypes according to the different genetic background and mutations involved in the pathogenic process (Figure 1). Metabolomics can also be used to follow drug response, as metabolites will reflect how the organism responds to selected drug candidates. Moreover, as metabolomics reflects alterations in the physiology, it can also show which pathways are dysfunctional and need to be compensated, opening new perspectives for metabolomics to be applied with therapeutic purposes. Some clinical trials applied metabolomics with these purposes are also discussed below.



Figure 1. "Omics" approaches analyze the whole set of molecules in an organism, and the metabolome of an individual is ultimately influenced by the genetic background. By combining genotyping and metabolomics approaches, it is possible to identify subgroups of patients and design a targeted therapy for each case. Finally, combination of genetics, metabolomics, and targeted therapy will boost the development of a personalized medicine. The figure was designed using image templates from Servier Medical Art (https://smart.servier.com/image-set-download/).

3.1. C9ORF72

Despite its wide involvement in fALS, few metabolomics studies have been conducted in *C9ORF72* models. A metabolomics study performed on sera of patients with *C9ORF72* mutation (only patients with frontotemporal lobar degeneration (FTLD), no ALS patients), revealed lower HDL cholesterol in *C9ORF72* patients versus non-mutated patients [49]. The pathogenicity of hexanucleotide repeat expansions in *C9ORF72* has been mainly studied in Drosophila and zebrafish models. However, the emergence of the use of iPSC in ALS research opens the perspective of using cellular models from patients' cells. Interestingly, it was shown that iPSC-derived motor neurons from C9ORF72-linked ALS/FTD patients present higher levels of free fatty acids and liquid droplets than controls, indicating a dysfunctional lipid metabolism [62]. Although no study so far correlated lipid metabolism and the

onset of behavioral and cognitive symptoms in ALS/FTLD patients, it is possible that the alterations in lipid metabolism are associated with the appearance of such symptoms. For example, dysfunction in lipid metabolism was linked to cognitive impairment in several types of dementia and Alzheimer's disease [63–66]. Further studies should be conducted in ALS/FTLD patients carrying the *C9ORF72* repeat expansions—even from the asymptomatic phase—to improve our understanding about the pathological mechanisms taking place.

3.2. SOD1

One striking example of metabolome profile linked to a genetic mutation was reported in the study of Wuolikainen and co-workers [50]. They analyzed the metabolomics profile in the cerebrospinal fluid (CSF) of ALS patients and found a different signature of metabolites in ALS patients bearing different *SOD1* mutations (including S105L, D101G, A89V, G93S, I113F, and D90A) in comparison to patients without *SOD1* mutations (both familial and sporadic cases). Further, they described that both homozygous and heterozygous carriers of the D90A mutation presented a very distinct metabolic signature, behaving as a separate group. Interestingly, cases with this homozygous mutation disclosed a slowly progressing lower-limb onset phenotype, with mean survival of around 14 years [67]. This distinct profile was marked by a decrease in amino acids in the CSF, as for example arginine, lysine, glutamine, ornithine, serine, and threonine [50].

As the first mutated gene described in genetic ALS, mutant SOD1 is still widely used in genetic ALS models, especially in cellular cultures and rodent models. Thus, metabolomics studies are looking for a better understanding of pathophysiological mechanisms of ALS, which are widely used SOD1 models. Several metabolomics studies performed in cellular models bearing mutations in the SOD1 gene highlighted a disturbance of energy metabolism. Valbuena and co-workers observed an increased aerobic glycolysis and an amino acid deficit in NSC-34 cells expressing mutant SOD1-G93A [51]. The same research team also showed a reduction in glutamate levels in co-cultures of spinal neurons and astrocytes expressing SOD1-G93A, and metabolic alterations related with oxidative stress, involving an important role of neuron-astrocyte relation in ALS pathophysiology [52]. This alteration of glutamate metabolism, which is associated with disturbance of the TCA (tricarboxylic acid) cycle, was also observed in a co-culture model expressing mutant SOD1-G93A [68]. To further explore the glutamate hypothesis in ALS, astrocytes and motor neurons bearing SOD1-G93A mutation were exposed to glutamate, revealing an alteration of the cellular shuttling of lactate between astrocytes and motor neurons [69]. Metabolic alterations observed in cellular models expressing mutant SOD1 were confirmed by metabolomics studies performed on animal models such as SOD1 transgenic mice expressing mutants G86R or G93A [53,70–72].

Lipids are another class of molecules that are found profoundly altered in ALS patients and can be analyzed trough metabolomics—and more specifically, lipidomics—studies. One study reported a very distinct lipidomics profile in the CSF of ALS patients compared to healthy controls. From the most discriminant lipids, several phosphatidylcholines and sphingomyelins were reported as higher in ALS patients, whereas triglycerides levels were decreased in the CSF of ALS patients. Interestingly, some phosphatidylcholines were also determined as discriminant in the lipidomics profile obtained from the cerebral cortex of the ALS mouse model SOD1-G93A [53,73]. These studies reinforce the notion that even in the presence of pathological mutations, several mechanisms associated with ALS pathology are common between sporadic and familial cases of ALS. Other classes of lipids are also reported to be altered in the plasma of ALS patients, such as triglycerides and fatty acids (reviewed by Gonzalez De Aguilar [74]). Cholesterol seems to be altered in ALS patients, and some studies report hypercholesterolemia at the time of diagnosis [75–79]. It was also shown that the administration of acetyl-L-carnitine, which supports the transport of fatty acids into mitochondria for being used as energy substrate, slowed down the worsening of motor symptoms in ALS patients [80].

3.3. TARDBP

Although metabolomics studies were not performed yet in ALS patients bearing mutations in the TARDBP gene, in vitro and in vivo studies shed light on the metabolic alterations that could be related with TDP-43 pathology in these models. In a Drosophila model expressing wild type or a mutation associated with ALS (TDP-43-G298S) specifically in motor neurons, metabolomics analysis revealed alterations in the carnitine pathway. They demonstrated an accumulation of carnitine conjugated long-chain fatty acids (as myristoylcarnitine, palmitoylcarnitine, oleoylcarnitine, and linoleoylcarnitine), leading to a decrease in carnitine levels. They also documented a significant reduction in beta-hydroxybutyrate, which is a ketone precursor and a key product of lipid beta-oxidation. These metabolic changes point to defects in the carnitine shuttle, which is required for long-chain fatty acid import into mitochondria and subsequent breakdown by lipid beta-oxidation ultimately leading to ATP production. When they bypassed these deficits by supplementing animals with a mixture of medium-chain fatty acids at varying concentrations (notably, coconut oil), they observed an amelioration in the motor symptoms associated with TDP-43 wild-type and G298S expression [54]. In a cellular model of wild-type TDP-43, metabolomics analysis found alterations in several unsaturated and saturated fatty acids of monounsaturated fatty acids (MUFA), polyunsaturated fatty acids (PUFA), and saturated fatty acids, among others [56]. Another study also performed in the same Drosophila model reported alterations in glucose metabolism, with increase in phosphoenolpyruvate and pyruvate in animals expressing wild-type or mutant TDP-43. Interestingly, when TDP-43 overexpression was knocked down, pyruvate levels return to control levels [55]. Together, these studies demonstrate a clear alteration in the energetic metabolism of ALS patients associated with TDP-43 pathology.

3.4. FUS

A recent metabolomics study performed on motor neurons bearing FUS mutation does not reveal a difference in metabolism due to the presence of FUS mutants [59]. Using iPSC-derived motor neurons from ALS patients carrying *FUS* mutations (R521H or P525L) and their CRISPR/Cas9-corrected counterparts, they highlighted a metabolic switch when cells differentiate to functional motor neurons but did not observe any disturbance of energy metabolism in mutant *FUS* cells compared to controls. It would be interesting to continue this study further by integrating astrocytes or other glial cells.

Ultimately, metabolomics reflects the biochemical activities taking place in an individual in a specific time point, and genetic factors play a key role in the biochemical pathways and ultimately in the production of metabolites [49] (Figure 1). For example, some GWAS applied to metabolic phenotypes generated a wide database of genetically determined metabolites [81–84]. Very recently, a study used this database in a Mendelian randomization study, combining genomics and metabolomics. They detected 18 metabolites that might have a causal role in the development of ALS and suggest that a dysfunction in the glutathione pathway is involved with ALS pathogenesis. In the same study, they investigated single nucleotide polymorphisms (SNP) in ALS patients and identified several as being genetic variants contributing to ALS pathology. The SNP most associated with ALS was rs1985243, and the authors hypothesize that it might contribute to the development of ALS through affecting the levels of gamma-glutamylphenylalanine [60]. Knowing this, it is possible to follow the alterations in any specific genetic linked case since the non-symptomatic phase and shed a light, at any giving time point, on the biochemical alterations that led the individual to start to experience disease-associated symptoms. This knowledge could help to identify altered metabolites that predict the alterations in molecular processes early in the disease course, and it could further be exploited in non-genetic cases.

One critical feature for the application of metabolomics in combination with genetic analysis in the clinical practice is standardization. Different techniques used in the metabolomics analysis, different methodologies for sample preparation, and even differences in samples storage can induce artificial differences in the metabolome of individuals that are not a real reflect of alterations induced by the disease or the genetic background. This lack of standardization makes it impossible to perform comparisons between studies and different ALS cohorts, and it is probably one of the main problems researchers face when identifying biomarkers for ALS diagnosis or prognosis.

4. Combination of Omics and Patient-Derived iPSC: The Future of ALS Research

The development of iPSC reprogrammed from human cells constitutes the ultimate tool when studying diverse pathologies, including neurodegenerative diseases. Nowadays, it is possible to create the "patient in a dish": iPSC derived from skin fibroblasts or peripheral blood mononuclear cells (PBMC)—or even cells collected from the urine—can be differentiated into virtually all cell types to model disease pathology and study drug response and drug toxicity [85–87]. Several research groups started to investigate the pathological mechanisms associated with ALS in iPSC obtained from patients and observe that motor neurons and glial cells differentiated from ALS-iPSC present the same pathological hallmarks as found in post-mortem tissue. Furthermore, they brought to light several disease mechanisms that are common to sporadic and familial or genetic-linked ALS [88–91].

Although metabolomics analysis in motor neurons or glial cells derived from ALS-iPSC has not been reported yet, several groups investigated the metabolic changes occurring in cells derived from ALS patients when compared to control cells. For example, alterations in the energetic metabolism were also reported in iPSC-derived astrocytes from sporadic and ALS patients bearing the expansions in the C9ORF72 gene. In these cells, researchers reported defects in adenosine, fructose, and glycogen metabolism [92,93]. The reduction in glycogen metabolism was attributed to decreased mRNA and protein expression of both enzymes glycogen phosphorylase and phosphoglucomutase. They also reported disruptions in the membrane transport of mitochondrial-specific energy substrates [93], which supports the supplementation of ALS patients with drugs that increase the transport of energetic subtracts to the mitochondria, such as the example of acetyl-L-carnitine [80]. Defects in adenosine use as an energy substrate were linked to a reduction in the levels of adenosine deaminase, which is the enzyme that converts adenosine to inosine. Bypassing this defect with inosine treatment restored the metabolism of astrocytes and also inhibited their toxicity when co-cultured with motor neurons. Remarkably, these alterations were also found in some sporadic ALS patients who also presented a decrease in the levels of both mRNA and protein expression of adenosine deaminase [92]. This study also points to another role of inosine treatment: more than increasing the levels of the endogenous antioxidant uric acid, inosine can also be used as an alternative energetic substrate.

Metabolomics analysis performed by different studies showed uric acid to be decreased in several neurodegenerative diseases, such as Parkinson's disease, Huntington's disease, and ALS [94–98]. Low uric acid was significantly decreased in the serum of ALS patients compared to control subjects and was inversely associated with an all-cause mortality risk [99]. Interestingly, low urate levels in the plasma were related to a higher risk of developing ALS, years before the onset of symptoms [97]. Preclinical studies showed the protective effect of uric acid [100,101], and uric acid in ALS patients is associated with its important antioxidant properties [102–104]. A recent clinical trial evaluated the effects of elevating the levels of urate in the blood of ALS patients by treating them with inosine [105]. Indeed, results recently published demonstrated that the urate levels rose to the targeted levels (7–8 mg/dL) after 6 weeks of treatment, without serious adverse effects observed. Although several biomarkers for oxidative stress and DNA damage were decreased in treated patients, predictions of ALSFRS-R did not change from baseline [105]. Even so, this pilot clinical trial offers hope, as inosine treatment was only performed during 12 weeks, and more prolonged clinical trials with a proper number of patients could reveal important therapeutic effects of inosine. Interestingly, this clinical trial followed the levels of uric acid in treated patients, as both metabolomics markers for treatment strategy and follow up of drug response [106]. Together with data obtained from C9ORF72-iPSC [92,93], these data reinforce the need of a better designed clinical trial on the effects of inosine—with, for example, the inclusion of two exclusive arms: one of sporadic ALS patients and another only composed by patients bearing the C9ORF72 expansions.

Other studies pointed to alterations in the metabolism of cells bearing TDP-43 mutations that are also observed in sporadic cases of ALS presenting cytoplasmic TDP-43 aggregates in motor neurons. iPSC-derived motor neurons from ALS patients bearing a *TARDBP* mutation (G298S) present an increase in the mRNA for two isoforms of the enzyme phosphofructokinase-1 (PFK1), PFKM, and PFKP, which are considered to be rate limiting and control the rate of glycolysis [107]. Remarkably, the same alterations were reported in the Drosophila fly model bearing the same mutation in the *TARDBP* gene (TDP-43-G298S) [55]. Furthermore, PFKM and PFKP transcripts were also increased in spinal cord tissue obtained from sporadic ALS cases with confirmed TDP-43 pathology [55], reinforcing the existence of common pathological mechanisms in sporadic and genetic-linked ALS and that TDP-43 pathology plays a key role in these pathological mechanisms.

Finally, as iPSC derived from ALS patients represent well the pathological characteristics of each patient, these cells could be developed from each patient with the aim to shed light on the pathological alterations taking place in every individual. For example, in the study of Allen et al., not all sporadic ALS patients presented deficits in adenosine deaminase, but these deficits were observed in all C9ORF72 patients. This study strengthens the notion of ALS as a highly heterogeneous disease and the need to develop individual therapeutic strategies, culminating with the establishment of a personalized medicine. Of note, metabolomics techniques are evolving so to be performed in single cells, allowing the study of a single cell metabolome. Some examples of techniques that are sensitive enough to be applied in small volumes and lower concentrations of metabolites are mass spectrometry, matrix-assisted laser desorption ionization (MALDI), and live single-cell mass spectrometry (LSC-MS) [108–113]. However, as discussed previously, these methods and techniques need to be standardized to reveal real common alterations among the different ALS subtypes and move the field forward.

5. Conclusions

The development of metabolomics analysis in ALS patients with different genetic mutations can help understand the metabolic pathways that are altered in such patients and help understand the pathological mechanisms involved in each case, as ALS is a highly heterogeneous disease. Genetics combined to metabolomics studies can help identify subgroups of patients that could benefit from a particular therapy. Furthermore, metabolomics studies unravel pathological mechanisms that are common to genetic and non-genetic cases and emphasize the impact of TDP-43 pathology in ALS. By unraveling the pathways that are altered in each genetic-linked ALS case, we can envisage a metabolomics-oriented therapy. The combination of metabolomics and iPSC-derived cells from ALS patients will help better understand the pathological mechanisms taking place in each patient and will pave the way to the implementation of a personalized medicine.

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References

- Chio, A.; Logroscino, G.; Traynor, B.J.; Collins, J.; Simeone, J.C.; Goldstein, L.A.; White, L.A. Global epidemiology of amyotrophic lateral sclerosis: A systematic review of the published literature. *Neuroepidemiology* 2013, 41, 118–130. [CrossRef] [PubMed]
- Alsultan, A.A.; Waller, R.; Heath, P.R.; Kirby, J. The genetics of amyotrophic lateral sclerosis: Current insights. Degener. Neurol. Neuromuscul. Dis. 2016, 6, 49–64. [CrossRef] [PubMed]
- 3. Maurel, C.; Dangoumau, A.; Marouillat, S.; Brulard, C.; Chami, A.; Hergesheimer, R.; Corcia, P.; Blasco, H.; Andres, C.R.; Vourc'h, P. Causative genes in amyotrophic lateral sclerosis and protein degradation pathways: A link to neurodegeneration. *Mol. Neurobiol.* **2018**, *55*, 6480–6499. [CrossRef] [PubMed]

- Zou, Z.Y.; Zhou, Z.R.; Che, C.H.; Liu, C.Y.; He, R.L.; Huang, H.P. Genetic epidemiology of amyotrophic lateral sclerosis: A systematic review and meta-analysis. *J. Neurol. Neurosurg. Psychiatry* 2017, *88*, 540–549. [CrossRef] [PubMed]
- 5. Dupuis, L.; Pradat, P.F.; Ludolph, A.C.; Loeffler, J.P. Energy metabolism in amyotrophic lateral sclerosis. *Lancet Neurol.* **2011**, *10*, 75–82. [CrossRef]
- 6. Ferri, A.; Coccurello, R. What is "Hyper" in the ALS hypermetabolism? *Mediat. Inflamm.* 2017, 2017, 7821672. [CrossRef]
- Steyn, F.J.; Ioannides, Z.A.; van Eijk, R.P.A.; Heggie, S.; Thorpe, K.A.; Ceslis, A.; Heshmat, S.; Henders, A.K.; Wray, N.R.; van den Berg, L.H.; et al. Hypermetabolism in ALS is associated with greater functional decline and shorter survival. *J. Neurol. Neurosurg. Psychiatry* 2018, *89*, 1016–1023. [CrossRef]
- 8. Jesus, P.; Fayemendy, P.; Nicol, M.; Lautrette, G.; Sourisseau, H.; Preux, P.M.; Desport, J.C.; Marin, B.; Couratier, P. Hypermetabolism is a deleterious prognostic factor in patients with amyotrophic lateral sclerosis. *Eur. J. Neurol.* **2018**, *25*, 97–104. [CrossRef]
- 9. Brito, M.D.; da Silva, G.F.G.; Tilieri, E.M.; Araujo, B.G.; Calio, M.L.; Rosenstock, T.R. Metabolic alteration and amyotrophic lateral sclerosis outcome: A systematic review. *Front. Neurol.* **2019**, *10*, 1205. [CrossRef]
- Boylan, K.B.; Glass, J.D.; Crook, J.E.; Yang, C.; Thomas, C.S.; Desaro, P.; Johnston, A.; Overstreet, K.; Kelly, C.; Polak, M.; et al. Phosphorylated neurofilament heavy subunit (pNF-H) in peripheral blood and CSF as a potential prognostic biomarker in amyotrophic lateral sclerosis. *J. Neurol. Neurosurg. Psychiatry* 2013, 84, 467–472. [CrossRef]
- 11. Mejzini, R.; Flynn, L.L.; Pitout, I.L.; Fletcher, S.; Wilton, S.D.; Akkari, P.A. ALS Genetics, mechanisms, and therapeutics: Where are we now? *Front. Mol. Neurosci.* **2019**, *13*, 1310. [CrossRef] [PubMed]
- 12. Manolio, T.A.; Collins, F.S.; Cox, N.J.; Goldstein, D.B.; Hindorff, L.A.; Hunter, D.J.; McCarthy, M.I.; Ramos, E.M.; Cardon, L.R.; Chakravarti, A.; et al. Finding the missing heritability of complex diseases. *Nature* **2009**, *461*, 747–753. [CrossRef]
- 13. DeJesus-Hernandez, M.; Mackenzie, I.R.; Boeve, B.F.; Boxer, A.L.; Baker, M.; Rutherford, N.J.; Nicholson, A.M.; Finch, N.A.; Flynn, H.; Adamson, J.; et al. Expanded GGGGCC hexanucleotide repeat in noncoding region of *C9ORF72* causes chromosome 9p-linked FTD and ALS. *Neuron* **2011**, *72*, 245–256. [CrossRef]
- 14. Renton, A.E.; Majounie, E.; Waite, A.; Simon-Sanchez, J.; Rollinson, S.; Gibbs, J.R.; Schymick, J.C.; Laaksovirta, H.; van Swieten, J.C.; Myllykangas, L.; et al. A hexanucleotide repeat expansion in *C9ORF72* is the cause of chromosome 9p21-linked ALS-FTD. *Neuron* **2011**, *72*, 257–268. [CrossRef]
- Nassif, M.; Woehlbier, U.; Manque, P.A. The enigmatic role of C9ORF72 in autophagy. *Front. Mol. Neurosci.* 2017, 11, 442. [CrossRef] [PubMed]
- 16. Liu, Y.; Wang, J. C9orf72-dependent lysosomal functions regulate epigenetic control of autophagy and lipid metabolism. *Autophagy* **2019**, *15*, 913–914. [CrossRef] [PubMed]
- Farg, M.A.; Sundaramoorthy, V.; Sultana, J.M.; Yang, S.; Atkinson, R.A.; Levina, V.; Halloran, M.A.; Gleeson, P.A.; Blair, I.P.; Soo, K.Y.; et al. C9ORF72, implicated in amytrophic lateral sclerosis and frontotemporal dementia, regulates endosomal trafficking. *Hum. Mol. Genet.* 2014, 23, 3579–3595. [CrossRef] [PubMed]
- Donnelly, C.J.; Zhang, P.W.; Pham, J.T.; Haeusler, A.R.; Mistry, N.A.; Vidensky, S.; Daley, E.L.; Poth, E.M.; Hoover, B.; Fines, D.M.; et al. RNA toxicity from the ALS/FTD *C9ORF72* expansion is mitigated by antisense intervention. *Neuron* 2013, *80*, 415–428. [CrossRef]
- 19. Koppers, M.; Blokhuis, A.M.; Westeneng, H.J.; Terpstra, M.L.; Zundel, C.A.; Vieira de Sa, R.; Schellevis, R.D.; Waite, A.J.; Blake, D.J.; Veldink, J.H.; et al. C9orf72 ablation in mice does not cause motor neuron degeneration or motor deficits. *Ann. Neurol.* **2015**, *78*, 426–438. [CrossRef]
- 20. Burberry, A.; Suzuki, N.; Wang, J.Y.; Moccia, R.; Mordes, D.A.; Stewart, M.H.; Suzuki-Uematsu, S.; Ghosh, S.; Singh, A.; Merkle, F.T.; et al. Loss-of-function mutations in the *C9ORF72* mouse ortholog cause fatal autoimmune disease. *Sci. Transl. Med.* **2016**, *8*, 347ra393. [CrossRef]
- 21. Jiang, J.; Zhu, Q.; Gendron, T.F.; Saberi, S.; McAlonis-Downes, M.; Seelman, A.; Stauffer, J.E.; Jafar-Nejad, P.; Drenner, K.; Schulte, D.; et al. Gain of toxicity from ALS/FTD-linked repeat expansions in *C9ORF72* Is alleviated by antisense oligonucleotides targeting GGGGCC-containing RNAs. *Neuron* **2016**, *90*, 535–550. [CrossRef] [PubMed]

- 22. O'Rourke, J.G.; Bogdanik, L.; Yanez, A.; Lall, D.; Wolf, A.J.; Muhammad, A.K.; Ho, R.; Carmona, S.; Vit, J.P.; Zarrow, J.; et al. *C9orf72* is required for proper macrophage and microglial function in mice. *Science* **2016**, 351, 1324–1329. [CrossRef] [PubMed]
- 23. Sudria-Lopez, E.; Koppers, M.; de Wit, M.; van der Meer, C.; Westeneng, H.J.; Zundel, C.A.; Youssef, S.A.; Harkema, L.; de Bruin, A.; Veldink, J.H.; et al. Full ablation of C9orf72 in mice causes immune system-related pathology and neoplastic events but no motor neuron defects. *Acta Neuropathol.* 2016, *132*, 145–147. [CrossRef] [PubMed]
- 24. Peters, O.M.; Cabrera, G.T.; Tran, H.; Gendron, T.F.; McKeon, J.E.; Metterville, J.; Weiss, A.; Wightman, N.; Salameh, J.; Kim, J.; et al. Human *C9ORF72* hexanucleotide expansion reproduces RNA foci and dipeptide repeat proteins but not neurodegeneration in BAC transgenic mice. *Neuron* **2015**, *88*, 902–909. [CrossRef]
- Lee, Y.B.; Chen, H.J.; Peres, J.N.; Gomez-Deza, J.; Attig, J.; Stalekar, M.; Troakes, C.; Nishimura, A.L.; Scotter, E.L.; Vance, C.; et al. Hexanucleotide repeats in ALS/FTD form length-dependent RNA foci, sequester RNA binding proteins, and are neurotoxic. *Cell Rep.* 2013, *5*, 1178–1186. [CrossRef]
- Kwon, I.; Xiang, S.; Kato, M.; Wu, L.; Theodoropoulos, P.; Wang, T.; Kim, J.; Yun, J.; Xie, Y.; McKnight, S.L. Poly-dipeptides encoded by the *C9orf72* repeats bind nucleoli, impede RNA biogenesis, and kill cells. *Science* 2014, 345, 1139–1145. [CrossRef]
- May, S.; Hornburg, D.; Schludi, M.H.; Arzberger, T.; Rentzsch, K.; Schwenk, B.M.; Grasser, F.A.; Mori, K.; Kremmer, E.; Banzhaf-Strathmann, J.; et al. *C9orf72* FTLD/ALS-associated Gly-Ala dipeptide repeat proteins cause neuronal toxicity and Unc119 sequestration. *Acta Neuropathol.* 2014, *128*, 485–503. [CrossRef]
- 28. Mizielinska, S.; Gronke, S.; Niccoli, T.; Ridler, C.E.; Clayton, E.L.; Devoy, A.; Moens, T.; Norona, F.E.; Woollacott, I.O.C.; Pietrzyk, J.; et al. *C9orf72* repeat expansions cause neurodegeneration in *Drosophila* through arginine-rich proteins. *Science* **2014**, *345*, 1192–1194. [CrossRef]
- 29. Wen, X.; Tan, W.; Westergard, T.; Krishnamurthy, K.; Markandaiah, S.S.; Shi, Y.; Lin, S.; Shneider, N.A.; Monaghan, J.; Pandey, U.B.; et al. Antisense proline-arginine RAN dipeptides linked to C9ORF72-ALS/FTD form toxic nuclear aggregates that initiate in vitro and in vivo neuronal death. *Neuron* 2014, 84, 1213–1225. [CrossRef]
- 30. Lee, K.H.; Zhang, P.; Kim, H.J.; Mitrea, D.M.; Sarkar, M.; Freibaum, B.D.; Cika, J.; Coughlin, M.; Messing, J.; Molliex, A.; et al. C9orf72 dipeptide repeats impair the assembly, dynamics, and function of membrane-less organelles. *Cell* **2016**, *167*, 774–788 e717. [CrossRef]
- 31. Lin, Y.; Mori, E.; Kato, M.; Xiang, S.; Wu, L.; Kwon, I.; McKnight, S.L. Toxic PR poly-dipeptides encoded by the C9orf72 repeat expansion target LC domain polymers. *Cell* **2016**, *167*, 789–802 e712. [CrossRef] [PubMed]
- 32. Hao, Z.; Liu, L.; Tao, Z.; Wang, R.; Ren, H.; Sun, H.; Lin, Z.; Zhang, Z.; Mu, C.; Zhou, J.; et al. Motor dysfunction and neurodegeneration in a *C9orf72* mouse line expressing poly-PR. *Nat. Commun.* **2019**, *10*, 2906. [CrossRef] [PubMed]
- 33. Fontecave, M.; Graslund, A.; Reichard, P. The function of superoxide dismutase during the enzymatic formation of the free radical of ribonucleotide reductase. *J. Biol. Chem.* **1987**, *262*, 12332–12336. [PubMed]
- 34. Rosen, D.R.; Siddique, T.; Patterson, D.; Figlewicz, D.A.; Sapp, P.; Hentati, A.; Donaldson, D.; Goto, J.; O'Regan, J.P.; Deng, H.X.; et al. Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis. *Nature* **1993**, *362*, 59–62. [CrossRef]
- 35. Felbecker, A.; Camu, W.; Valdmanis, P.N.; Sperfeld, A.D.; Waibel, S.; Steinbach, P.; Rouleau, G.A.; Ludolph, A.C.; Andersen, P.M. Four familial ALS pedigrees discordant for two SOD1 mutations: Are all SOD1 mutations pathogenic? *J. Neurol. Neurosurg. Psychiatry* **2010**, *81*, 572–577. [CrossRef]
- 36. Yamashita, S.; Ando, Y. Genotype-phenotype relationship in hereditary amyotrophic lateral sclerosis. *Transl. NeuroDegener.* **2015**, *4*, 13. [CrossRef]
- 37. Lattante, S.; Rouleau, G.A.; Kabashi, E. *TARDBP* and *FUS* mutations associated with amyotrophic lateral sclerosis: Summary and update. *Hum. Mutat.* **2013**, *34*, 812–826. [CrossRef]
- 38. Hergesheimer, R.C.; Chami, A.A.; de Assis, D.R.; Vourc'h, P.; Andres, C.R.; Corcia, P.; Lanznaster, D.; Blasco, H. The debated toxic role of aggregated TDP-43 in amyotrophic lateral sclerosis: A resolution in sight? *Brain* **2019**, *142*, 1176–1194. [CrossRef]
- Arai, T.; Hasegawa, M.; Akiyama, H.; Ikeda, K.; Nonaka, T.; Mori, H.; Mann, D.; Tsuchiya, K.; Yoshida, M.; Hashizume, Y.; et al. TDP-43 is a component of ubiquitin-positive tau-negative inclusions in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *BioChem. Biophys. Res. Commun.* 2006, 351, 602–611. [CrossRef]

- Neumann, M.; Sampathu, D.M.; Kwong, L.K.; Truax, A.C.; Micsenyi, M.C.; Chou, T.T.; Bruce, J.; Schuck, T.; Grossman, M.; Clark, C.M.; et al. Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. *Science* 2006, *314*, 130–133. [CrossRef]
- 41. Kwiatkowski, T.J., Jr.; Bosco, D.A.; Leclerc, A.L.; Tamrazian, E.; Vanderburg, C.R.; Russ, C.; Davis, A.; Gilchrist, J.; Kasarskis, E.J.; Munsat, T.; et al. Mutations in the *FUS/TLS* gene on chromosome 16 cause familial amyotrophic lateral sclerosis. *Science* **2009**, *323*, 1205–1208. [CrossRef] [PubMed]
- 42. Vance, C.; Rogelj, B.; Hortobagyi, T.; De Vos, K.J.; Nishimura, A.L.; Sreedharan, J.; Hu, X.; Smith, B.; Ruddy, D.; Wright, P.; et al. Mutations in FUS, an RNA processing protein, cause familial amyotrophic lateral sclerosis type 6. *Science* **2009**, *323*, 1208–1211. [CrossRef] [PubMed]
- 43. Zou, Z.Y.; Cui, L.Y.; Sun, Q.; Li, X.G.; Liu, M.S.; Xu, Y.; Zhou, Y.; Yang, X.Z. De novo *FUS* gene mutations are associated with juvenile-onset sporadic amyotrophic lateral sclerosis in China. *NeuroBiol. Aging* **2013**, *34*, 1312.e1–1312.e8. [CrossRef]
- 44. Hubers, A.; Just, W.; Rosenbohm, A.; Muller, K.; Marroquin, N.; Goebel, I.; Hogel, J.; Thiele, H.; Altmuller, J.; Nurnberg, P.; et al. De novo *FUS* mutations are the most frequent genetic cause in early-onset German ALS patients. *NeuroBiol. Aging* **2015**, *36*, 3117.e1–3117.e6. [CrossRef] [PubMed]
- 45. Gromicho, M.; Oliveira Santos, M.; Pinto, A.; Pronto-Laborinho, A.; De Carvalho, M. Young-onset rapidly progressive ALS associated with heterozygous FUS mutation. *Amyotroph. Lateral Scler. Front. Degener.* **2017**, *18*, 451–453. [CrossRef] [PubMed]
- 46. Zinszner, H.; Sok, J.; Immanuel, D.; Yin, Y.; Ron, D. TLS (FUS) binds RNA in vivo and engages in nucleo-cytoplasmic shuttling. *J. Cell Sci.* **1997**, *110 Pt 15*, 1741–1750.
- Niu, C.; Zhang, J.; Gao, F.; Yang, L.; Jia, M.; Zhu, H.; Gong, W. FUS-NLS/Transportin 1 complex structure provides insights into the nuclear targeting mechanism of FUS and the implications in ALS. *PLoS ONE* 2012, 7, e47056. [CrossRef]
- Vance, C.; Scotter, E.L.; Nishimura, A.L.; Troakes, C.; Mitchell, J.C.; Kathe, C.; Urwin, H.; Manser, C.; Miller, C.C.; Hortobagyi, T.; et al. ALS mutant FUS disrupts nuclear localization and sequesters wild-type FUS within cytoplasmic stress granules. *Hum. Mol. Genet.* 2013, *22*, 2676–2688. [CrossRef]
- 49. Jaaskelainen, O.; Solje, E.; Hall, A.; Katisko, K.; Korhonen, V.; Tiainen, M.; Kangas, A.J.; Helisalmi, S.; Pikkarainen, M.; Koivisto, A.; et al. Low serum high-density lipoprotein cholesterol levels associate with the *C9orf72* repeat expansion in frontotemporal lobar degeneration patients. *J. Alzheimers Dis.* **2019**, *72*, 127–137. [CrossRef]
- 50. Wuolikainen, A.; Andersen, P.M.; Moritz, T.; Marklund, S.L.; Antti, H. ALS patients with mutations in the *SOD1* gene have an unique metabolomic profile in the cerebrospinal fluid compared with ALS patients without mutations. *Mol. Genet. Metab.* **2012**, *105*, 472–478. [CrossRef]
- 51. Valbuena, G.N.; Rizzardini, M.; Cimini, S.; Siskos, A.P.; Bendotti, C.; Cantoni, L.; Keun, H.C. Metabolomic analysis reveals increased aerobic glycolysis and amino acid deficit in a cellular model of amyotrophic lateral sclerosis. *Mol. Neurobiol.* **2016**, *53*, 2222–2240. [CrossRef]
- 52. Valbuena, G.N.; Tortarolo, M.; Bendotti, C.; Cantoni, L.; Keun, H.C. Altered metabolic profiles associate with toxicity in SOD1^{G93A} astrocyte-neuron co-cultures. *Sci. Rep.* **2017**, *7*, 50. [CrossRef]
- 53. Patin, F.; Corcia, P.; Vourc'h, P.; Nadal-Desbarats, L.; Baranek, T.; Goossens, J.F.; Marouillat, S.; Dessein, A.F.; Descat, A.; Madji Hounoum, B.; et al. Omics to explore amyotrophic lateral sclerosis evolution: The central role of arginine and proline metabolism. *Mol. Neurobiol.* **2017**, *54*, 5361–5374. [CrossRef]
- 54. Manzo, E.; O'Conner, A.G.; Barrows, J.M.; Shreiner, D.D.; Birchak, G.J.; Zarnescu, D.C. Medium-chain fatty acids, beta-hydroxybutyric acid and genetic modulation of the carnitine shuttle are protective in a drosophila model of ALS based on TDP-43. *Front. Mol. NeuroSci.* **2018**, *11*, 182. [CrossRef]
- 55. Manzo, E.; Lorenzini, I.; Barrameda, D.; O'Conner, A.G.; Barrows, J.M.; Starr, A.; Kovalik, T.; Rabichow, B.E.; Lehmkuhl, E.M.; Shreiner, D.D.; et al. Glycolysis upregulation is neuroprotective as a compensatory mechanism in ALS. *Elife* **2019**, *8*. [CrossRef]
- 56. Lanznaster, D.; Bourgeais, J.; Bruno, C.; Hergesheimer, R.C.; Thepault, R.A.; Vourc'h, P.; Corcia, P.; Andres, C.R.; Herault, O.; Blasco, H. TDP-43-mediated toxicity in HEK293T Cells: A fast and reproducible protocol to be employed in the search of new therapeutic options against amyotrophic lateral sclerosis. *Cells* 2019, *9*, 68. [CrossRef]

- 57. Drepper, C.; Herrmann, T.; Wessig, C.; Beck, M.; Sendtner, M. C-terminal FUS/TLS mutations in familial and sporadic ALS in Germany. *NeuroBiol. Aging* **2011**, *32*, 548.e1–548.e4. [CrossRef]
- Chio, A.; Restagno, G.; Brunetti, M.; Ossola, I.; Calvo, A.; Mora, G.; Sabatelli, M.; Monsurro, M.R.; Battistini, S.; Mandrioli, J.; et al. Two Italian kindreds with familial amyotrophic lateral sclerosis due to FUS mutation. *NeuroBiol. Aging* 2009, 30, 1272–1275. [CrossRef]
- 59. Vandoorne, T.; Veys, K.; Guo, W.; Sicart, A.; Vints, K.; Swijsen, A.; Moisse, M.; Eelen, G.; Gounko, N.V.; Fumagalli, L.; et al. Differentiation but not ALS mutations in FUS rewires motor neuron metabolism. *Nat. Commun.* **2019**, *10*, 4147. [CrossRef]
- 60. Yang, L.; Lv, X.; Du, H.; Wu, D.; Wang, M. Causal effects of serum metabolites on amyotrophic lateral sclerosis: A Mendelian randomization study. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2020**, *97*, 109771. [CrossRef]
- 61. Menni, C.; Zierer, J.; Valdes, A.M.; Spector, T.D. Mixing omics: Combining genetics and metabolomics to study rheumatic diseases. *Nat. Rev. Rheumatol.* **2017**, *13*, 174–181. [CrossRef]
- Liu, Y.; Wang, T.; Ji, Y.J.; Johnson, K.; Liu, H.; Johnson, K.; Bailey, S.; Suk, Y.; Lu, Y.N.; Liu, M.; et al. A C9orf72-CARM1 axis regulates lipid metabolism under glucose starvation-induced nutrient stress. *Genes Dev.* 2018, 32, 1380–1397. [CrossRef]
- 63. Ruan, Q.; D'Onofrio, G.; Sancarlo, D.; Greco, A.; Yu, Z. Potential fluid biomarkers for pathological brain changes in Alzheimer's disease: Implication for the screening of cognitive frailty. *Mol. Med. Rep.* **2016**, *14*, 3184–3198. [CrossRef]
- 64. Kim, S.H.; Yang, J.S.; Lee, J.C.; Lee, J.Y.; Lee, J.Y.; Kim, E.; Moon, M.H. Lipidomic alterations in lipoproteins of patients with mild cognitive impairment and Alzheimer's disease by asymmetrical flow field-flow fractionation and nanoflow ultrahigh performance liquid chromatography-tandem mass spectrometry. *J. Chromatogr. A* **2018**, *1568*, 91–100. [CrossRef]
- 65. Costa, A.C.; Joaquim, H.P.G.; Forlenza, O.; Talib, L.L.; Gattaz, W.F. Plasma lipids metabolism in mild cognitive impairment and Alzheimer's disease. *World J. Biol. Psychiatry* **2019**, *20*, 190–196. [CrossRef]
- 66. Sarrafpour, S.; Ormseth, C.; Chiang, A.; Arakaki, X.; Harrington, M.; Fonteh, A. Lipid metabolism in late-onset Alzheimer's disease differs from patients presenting with other dementia phenotypes. *Int. J. Environ. Res. Public Health* **2019**, *16*, 1995. [CrossRef]
- 67. Andersen, P.M.; Forsgren, L.; Binzer, M.; Nilsson, P.; Ala-Hurula, V.; Keranen, M.L.; Bergmark, L.; Saarinen, A.; Haltia, T.; Tarvainen, I.; et al. Autosomal recessive adult-onset amyotrophic lateral sclerosis associated with homozygosity for Asp90Ala CuZn-superoxide dismutase mutation. A clinical and genealogical study of 36 patients. *Brain* **1996**, *119 Pt 4*, 1153–1172. [CrossRef]
- Veyrat-Durebex, C.; Corcia, P.; Piver, E.; Devos, D.; Dangoumau, A.; Gouel, F.; Vourc'h, P.; Emond, P.; Laumonnier, F.; Nadal-Desbarats, L.; et al. Disruption of TCA cycle and glutamate metabolism identified by metabolomics in an in vitro model of amyotrophic lateral sclerosis. *Mol. Neurobiol.* 2016, *53*, 6910–6924. [CrossRef]
- Madji Hounoum, B.; Mavel, S.; Coque, E.; Patin, F.; Vourc'h, P.; Marouillat, S.; Nadal-Desbarats, L.; Emond, P.; Corcia, P.; Andres, C.R.; et al. Wildtype motoneurons, ALS-Linked SOD1 mutation and glutamate profoundly modify astrocyte metabolism and lactate shuttling. *Glia* 2017, 65, 592–605. [CrossRef]
- 70. Patin, F.; Baranek, T.; Vourc'h, P.; Nadal-Desbarats, L.; Goossens, J.F.; Marouillat, S.; Dessein, A.F.; Descat, A.; Hounoum, B.M.; Bruno, C.; et al. combined metabolomics and transcriptomics approaches to assess the IL-6 blockade as a therapeutic of ALS: Deleterious alteration of lipid metabolism. *Neurotherapeutics* 2016, 13, 905–917. [CrossRef]
- 71. Henriques, A.; Croixmarie, V.; Bouscary, A.; Mosbach, A.; Keime, C.; Boursier-Neyret, C.; Walter, B.; Spedding, M.; Loeffler, J.P. Sphingolipid metabolism is dysregulated at transcriptomic and metabolic levels in the spinal cord of an animal model of amyotrophic lateral sclerosis. *Front. Mol. Neurosci.* **2017**, *10*, 433. [CrossRef]
- Valbuena, G.N.; Cantoni, L.; Tortarolo, M.; Bendotti, C.; Keun, H.C. spinal cord metabolic signatures in models of fast- and slow-progressing SOD1^{G93A} amyotrophic lateral sclerosis. *Front. Mol. Neurosci.* 2019, 13, 1276. [CrossRef]
- 73. Blasco, H.; Veyrat-Durebex, C.; Bocca, C.; Patin, F.; Vourc'h, P.; Kouassi Nzoughet, J.; Lenaers, G.; Andres, C.R.; Simard, G.; Corcia, P.; et al. Lipidomics reveals cerebrospinal-fluid signatures of ALS. *Sci. Rep.* **2017**, *7*, 17652. [CrossRef]

- 74. Gonzalez De Aguilar, J.L. Lipid biomarkers for amyotrophic lateral sclerosis. *Front. Neurol.* **2019**, *10*, 284. [CrossRef]
- Dupuis, L.; Corcia, P.; Fergani, A.; Gonzalez De Aguilar, J.L.; Bonnefont-Rousselot, D.; Bittar, R.; Seilhean, D.; Hauw, J.J.; Lacomblez, L.; Loeffler, J.P.; et al. Dyslipidemia is a protective factor in amyotrophic lateral sclerosis. *Neurology* 2008, *70*, 1004–1009. [CrossRef]
- 76. Dedic, S.I.; Stevic, Z.; Dedic, V.; Stojanovic, V.R.; Milicev, M.; Lavrnic, D. Is hyperlipidemia correlated with longer survival in patients with amyotrophic lateral sclerosis? *Neurol. Res.* **2012**, *34*, 576–580. [CrossRef]
- 77. Ikeda, K.; Hirayama, T.; Takazawa, T.; Kawabe, K.; Iwasaki, Y. Relationships between disease progression and serum levels of lipid, urate, creatinine and ferritin in Japanese patients with amyotrophic lateral sclerosis: A cross-sectional study. *Intern. Med.* 2012, *51*, 1501–1508. [CrossRef]
- Rafiq, M.K.; Lee, E.; Bradburn, M.; McDermott, C.J.; Shaw, P.J. Effect of lipid profile on prognosis in the patients with amyotrophic lateral sclerosis: Insights from the olesoxime clinical trial. *Amyotroph. Lateral Scler. Front. Degener.* 2015, *16*, 478–484. [CrossRef]
- 79. Delaye, J.B.; Patin, F.; Piver, E.; Bruno, C.; Vasse, M.; Vourc'h, P.; Andres, C.R.; Corcia, P.; Blasco, H. Low IDL-B and high LDL-1 subfraction levels in serum of ALS patients. *J. Neurol. Sci.* 2017, *380*, 124–127. [CrossRef]
- Beghi, E.; Pupillo, E.; Bonito, V.; Buzzi, P.; Caponnetto, C.; Chio, A.; Corbo, M.; Giannini, F.; Inghilleri, M.; Bella, V.L.; et al. Randomized double-blind placebo-controlled trial of acetyl-L-carnitine for ALS. *Amyotroph. Lateral Scler. Front. Degener.* 2013, 14, 397–405. [CrossRef]
- Gieger, C.; Geistlinger, L.; Altmaier, E.; Hrabe de Angelis, M.; Kronenberg, F.; Meitinger, T.; Mewes, H.W.; Wichmann, H.E.; Weinberger, K.M.; Adamski, J.; et al. Genetics meets metabolomics: A genome-wide association study of metabolite profiles in human serum. *PLoS Genet.* 2008, *4*, e1000282. [CrossRef]
- Suhre, K.; Shin, S.Y.; Petersen, A.K.; Mohney, R.P.; Meredith, D.; Wagele, B.; Altmaier, E.; CardioGram; Deloukas, P.; Erdmann, J.; et al. Human metabolic individuality in biomedical and pharmaceutical research. *Nature* 2011, 477, 54–60. [CrossRef]
- Kettunen, J.; Tukiainen, T.; Sarin, A.P.; Ortega-Alonso, A.; Tikkanen, E.; Lyytikainen, L.P.; Kangas, A.J.; Soininen, P.; Wurtz, P.; Silander, K.; et al. Genome-wide association study identifies multiple loci influencing human serum metabolite levels. *Nat. Genet.* 2012, 44, 269–276. [CrossRef]
- 84. Shin, S.Y.; Fauman, E.B.; Petersen, A.K.; Krumsiek, J.; Santos, R.; Huang, J.; Arnold, M.; Erte, I.; Forgetta, V.; Yang, T.P.; et al. An atlas of genetic influences on human blood metabolites. *Nat. Genet.* **2014**, *46*, 543–550. [CrossRef]
- 85. Bellin, M.; Marchetto, M.C.; Gage, F.H.; Mummery, C.L. Induced pluripotent stem cells: The new patient? *Nat. Rev. Mol. Cell Biol.* **2012**, *13*, 713–726. [CrossRef]
- 86. Kumar, D.; Anand, T.; Kues, W.A. Clinical potential of human-induced pluripotent stem cells: Perspectives of induced pluripotent stem cells. *Cell Biol. Toxicol.* **2017**, *33*, 99–112. [CrossRef]
- Gaignerie, A.; Lefort, N.; Rousselle, M.; Forest-Choquet, V.; Flippe, L.; Francois-Campion, V.; Girardeau, A.; Caillaud, A.; Chariau, C.; Francheteau, Q.; et al. Urine-derived cells provide a readily accessible cell type for feeder-free mRNA reprogramming. *Sci. Rep.* 2018, *8*, 14363. [CrossRef]
- Burkhardt, M.F.; Martinez, F.J.; Wright, S.; Ramos, C.; Volfson, D.; Mason, M.; Garnes, J.; Dang, V.; Lievers, J.; Shoukat-Mumtaz, U.; et al. A cellular model for sporadic ALS using patient-derived induced pluripotent stem cells. *Mol. Cell NeuroSci.* 2013, *56*, 355–364. [CrossRef]
- 89. Bohl, D.; Pochet, R.; Mitrecic, D.; Nicaise, C. Modelling and treating amyotrophic lateral sclerosis through induced-pluripotent stem cells technology. *Curr. Stem Cell Res. Ther.* **2016**, *11*, 301–312. [CrossRef]
- Fujimori, K.; Ishikawa, M.; Otomo, A.; Atsuta, N.; Nakamura, R.; Akiyama, T.; Hadano, S.; Aoki, M.; Saya, H.; Sobue, G.; et al. Modeling sporadic ALS in iPSC-derived motor neurons identifies a potential therapeutic agent. *Nat. Med.* 2018, 24, 1579–1589. [CrossRef]
- Sun, X.; Song, J.; Huang, H.; Chen, H.; Qian, K. Modeling hallmark pathology using motor neurons derived from the family and sporadic amyotrophic lateral sclerosis patient-specific iPS cells. *Stem Cell Res. Ther.* 2018, 9, 315. [CrossRef] [PubMed]
- 92. Allen, S.P.; Hall, B.; Castelli, L.M.; Francis, L.; Woof, R.; Siskos, A.P.; Kouloura, E.; Gray, E.; Thompson, A.G.; Talbot, K.; et al. Astrocyte adenosine deaminase loss increases motor neuron toxicity in amyotrophic lateral sclerosis. *Brain* **2019**, *142*, 586–605. [CrossRef] [PubMed]

- Allen, S.P.; Hall, B.; Woof, R.; Francis, L.; Gatto, N.; Shaw, A.C.; Myszczynska, M.; Hemingway, J.; Coldicott, I.; Willcock, A.; et al. *C9orf72* expansion within astrocytes reduces metabolic flexibility in amyotrophic lateral sclerosis. *Brain* 2019, 142, 3771–3790. [CrossRef] [PubMed]
- Lawton, K.A.; Brown, M.V.; Alexander, D.; Li, Z.; Wulff, J.E.; Lawson, R.; Jaffa, M.; Milburn, M.V.; Ryals, J.A.; Bowser, R.; et al. Plasma metabolomic biomarker panel to distinguish patients with amyotrophic lateral sclerosis from disease mimics. *Amyotroph. Lateral Scler. Front. Degener.* 2014, 15, 362–370. [CrossRef]
- 95. Kori, M.; Aydin, B.; Unal, S.; Arga, K.Y.; Kazan, D. Metabolic biomarkers and neurodegeneration: A pathway enrichment analysis of Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis. *Omics* **2016**, *20*, 645–661. [CrossRef]
- Huang, X.; Ng, S.Y.; Chia, N.S.; Acharyya, S.; Setiawan, F.; Lu, Z.H.; Ng, E.; Tay, K.Y.; Au, W.L.; Tan, E.K.; et al. Serum uric acid level and its association with motor subtypes and non-motor symptoms in early Parkinson's disease: PALS study. *Park. Relat. Disord.* 2018, 55, 50–54. [CrossRef]
- Bjornevik, K.; Zhang, Z.; O'Reilly, E.J.; Berry, J.D.; Clish, C.B.; Deik, A.; Jeanfavre, S.; Kato, I.; Kelly, R.S.; Kolonel, L.N.; et al. Prediagnostic plasma metabolomics and the risk of amyotrophic lateral sclerosis. *Neurology* 2019, 92, e2089–e2100. [CrossRef]
- Corey-Bloom, J.; Haque, A.; Aboufadel, S.; Snell, C.; Fischer, R.S.; Granger, S.W.; Granger, D.A.; Thomas, E.A. Uric acid as a potential peripheral biomarker for disease features in Huntington's patients. *Front. Mol. Neurosci.* 2020, *14*, 73. [CrossRef]
- 99. Zhang, F.; Zhang, Q.; Ke, Y.; Hao, J.; Lu, L.; Lu, N.; Chen, X. Serum uric acid levels in patients with amyotrophic lateral sclerosis: A meta-analysis. *Sci. Rep.* **2018**, *8*, 1100. [CrossRef]
- 100. Bakshi, R.; Xu, Y.; Mueller, K.A.; Chen, X.; Granucci, E.; Paganoni, S.; Sadri-Vakili, G.; Schwarzschild, M.A. Urate mitigates oxidative stress and motor neuron toxicity of astrocytes derived from ALS-linked SOD1^{G93A} mutant mice. *Mol. Cell. Neurosci.* 2018, 92, 12–16. [CrossRef]
- 101. Zhang, C.; Yang, Y.; Liang, W.; Wang, T.; Wang, S.; Wang, X.; Wang, Y.; Jiang, H.; Feng, H. Neuroprotection by urate on the mutant hSOD1-related cellular and Drosophila models of amyotrophic lateral sclerosis: Implication for GSH synthesis via activating Akt/GSK3beta/Nrf2/GCLC pathways. *Brain Res. Bull.* 2019, 146, 287–301. [CrossRef] [PubMed]
- 102. Keizman, D.; Ish-Shalom, M.; Berliner, S.; Maimon, N.; Vered, Y.; Artamonov, I.; Tsehori, J.; Nefussy, B.; Drory, V.E. Low uric acid levels in serum of patients with ALS: Further evidence for oxidative stress? *J. Neurol. Sci.* 2009, 285, 95–99. [CrossRef] [PubMed]
- 103. Nagase, M.; Yamamoto, Y.; Miyazaki, Y.; Yoshino, H. Increased oxidative stress in patients with amyotrophic lateral sclerosis and the effect of edaravone administration. *Redox Rep.* 2016, 21, 104–112. [CrossRef] [PubMed]
- 104. Wang, Z.; Bai, Z.; Qin, X.; Cheng, Y. Aberrations in oxidative stress markers in amyotrophic lateral sclerosis: A systematic review and meta-analysis. *Oxidative Med. Cell. Longev.* **2019**, 2019, 1712323. [CrossRef]
- 105. Nicholson, K.; Chan, J.; Macklin, E.A.; Levine-Weinberg, M.; Breen, C.; Bakshi, R.; Grasso, D.L.; Wills, A.M.; Jahandideh, S.; Taylor, A.A.; et al. Pilot trial of inosine to elevate urate levels in amyotrophic lateral sclerosis. *Ann. Clin. Transl. Neurol.* 2018, *5*, 1522–1533. [CrossRef]
- 106. Lanznaster, D.; de Assis, D.R.; Corcia, P.; Pradat, P.F.; Blasco, H. Metabolomics biomarkers: A strategy toward therapeutics improvement in ALS. *Front. Neurol.* **2018**, *9*, 1126. [CrossRef]
- 107. Tanner, L.B.; Goglia, A.G.; Wei, M.H.; Sehgal, T.; Parsons, L.R.; Park, J.O.; White, E.; Toettcher, J.E.; Rabinowitz, J.D. Four Key Steps Control Glycolytic Flux in Mammalian Cells. *Cell Syst.* 2018, 7, 49–62 e48. [CrossRef]
- 108. Laiko, V.V.; Baldwin, M.A.; Burlingame, A.L. Atmospheric pressure matrix-assisted laser desorption/ionization mass spectrometry. *Anal. Chem.* 2000, 72, 652–657. [CrossRef]
- Emara, S.; Amer, S.; Ali, A.; Abouleila, Y.; Oga, A.; Masujima, T. Single-cell metabolomics. *Adv. Exp. Med. Biol.* 2017, 965, 323–343. [CrossRef]
- Chappell, L.; Russell, A.J.C.; Voet, T. Single-cell (Multi)omics technologies. *Ann. Rev. Genom. Hum. Genet.* 2018, 19, 15–41. [CrossRef]
- Qi, M.; Philip, M.C.; Yang, N.; Sweedler, J.V. Single cell neurometabolomics. ACS Chem. NeuroSci. 2018, 9, 40–50. [CrossRef] [PubMed]

- 112. Germeys, C.; Vandoorne, T.; Bercier, V.; Van Den Bosch, L. Existing and emerging metabolomic tools for ALS research. *Genes* **2019**, *10*, 1011. [CrossRef] [PubMed]
- 113. Shrestha, B. Single-cell metabolomics by mass spectrometry. *Methods Mol. Biol.* **2020**, 2064, 1–8. [CrossRef] [PubMed]



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