

Modulating Microglial Cells for **Promoting Brain Recovery and** Repair

Dirk M. Hermann 1 * and Matthias Gunzer 2

¹Department of Neurology, University of Duisburg-Essen, University Hospital Essen, Essen, Germany, ²Institute of Experimental Immunology and Imaging, University of Duisburg-Essen, University Hospital Essen, Essen, Germany

Representing the brain's innate immune cells that interact vividly with blood-derived opportunities for promoting neurological recovery in human patients upon brain injury.

Keywords: blood-derived immune cell, brain injury, brain ischemia, neuroimmunology, neurodegeneration,

immune cells and brain parenchymal cells, microglia set the stage for successful brain remodeling and repair in the aftermath of brain damage. With the development of pharmacological colony-stimulating factor-1 receptor inhibitors, which allow inhibiting or depleting microglial cells, and of transgenic mice, allowing the inducible depletion of microglial cells, experimental tools have become available for studying roles of microglia in neurodegenerative and neurorestorative processes. These models open fundamental insights into roles of microglia in controlling synaptic plasticity in the healthy and the injured brain. Acting as a switch from injury to repair, microglial cells might open

neuroplasticity, neurorepair

Immune responses play a central role in modulating brain injury and recovery postinjury (Anrather and Iadecola, 2016; Jayaraj et al., 2019). In the injured brain, complex cellular and molecular mechanisms are triggered, including the release of inflammatory cytokines and alarmins by damaged cells (Bianchi, 2007; Roth et al., 2018), glial activation (Neumann et al., 2015; Manrique-Castano et al., 2020), and the brain invasion of leukocytes belonging to the innate and adaptive immune systems (Gelderblom et al., 2009; Neumann et al., 2015). Immune cell trafficking across the blood-brain barrier is mediated by adhesion molecules on cerebral endothelial cells including intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 (Lopes Pinheiro et al., 2016). In the injured brain, both peripheral leukocytes and resident microglia have been shown to accumulate in evolving brain lesions (Neumann et al., 2015; Perez-de-Puig et al., 2015).

Microglial cells, which are the brain's innate immune cells, vividly interact with brain-invading leukocytes in the injured brain (Neumann et al., 2006, 2008, 2015), controlling their brain access and activity. Microglial cells also communicate with brain endothelial cells, maintaining the integrity of the brain microvasculature, and, specifically, the blood-brain barrier, as well as immune cell access to the injured brain (Dudvarski Stankovic et al., 2016). Microvascular protection by

1

OPEN ACCESS

Edited by:

Ertuarul Kilic Istanbul Medipol University, Turkey

Reviewed by:

Hulva Karatas. Hacettepe University, Turkey Taha Kelestemur. Istanbul Medipol University, Turkey

*Correspondence:

Dirk M. Hermann dirk.hermann@uk-essen.de

Specialty section:

This article was submitted to Cellular Neuropathology, a section of the journal Frontiers in Cellular Neuroscience

Received: 10 November 2020 Accepted: 07 December 2020 Published: 11 January 2021

Citation:

Hermann DM and Gunzer M (2021) Modulating Microglial Cells for Promoting Brain Recovery and Repair. Front. Cell. Neurosci. 14:627987. doi: 10.3389/fncel.2020.627987 microglia is enabled by direct effects of microglia on endothelial cells, e.g., by inducing their phagocytosis or stabilizing endothelial cells by secretion of vascular endothelial growth factor (Dudvarski Stankovic et al., 2016). In organotypic brain slices *ex vivo* and experimental models of ischemic stroke *in vivo*, activated microglial cells were shown to engulf and phagocytose leukocytes (Neumann et al., 2008, 2018; Otxoa-de-Amezaga et al., 2019).

These dynamic responses of microglial cells in the injured brain were identified following advances in two fields, namely, in: (a) brain imaging and image analysis; and (b) pharmacological microglial deactivation and/or depletion. Brain imaging techniques facilitating the analysis of microglial cells are two-photon microscopy, which allows real-time imaging of cell physiology and pathology in vivo in the injured brain (Davalos et al., 2005; Neumann et al., 2015), and confocal microscopy combined with morphological image analysis, which via the segmentation, skeletonization, and threedimensional reconstruction of microglial cells allows the evaluation of microglial activation in responses to injury and therapy (Heindl et al., 2018; Manrique-Castano et al., 2020). Innovations of brain imaging greatly promoted progress in our understanding of the contribution of microglial cells to brain recovery processes.

With the emergence of pharmacological colony-stimulating factor-1 receptor (CSF1R) inhibitors, which allow inhibiting and depleting microglial cells (Elmore et al., 2014; Waisman et al., 2015; Olmos-Alonso et al., 2016) and of the CX3CR1-CreER-R26iDTR mouse, in which microglia depletion can efficiently be induced by diphtheria toxin delivery (Parkhurst et al., 2013; Waisman et al., 2015), experimental tools have become available for assessing the role of microglial cells in neurodegenerative processes. Microglial depletion studies using a CSF1R inhibitor revealed that reactive microglia efficiently eliminate leukocytes from ischemic brain tissue (Otxoa-de-Amezaga et al., 2019). Microglia deactivation and depletion by long-term treatment with the CSF1R inhibitor increased brain leukocyte numbers, and microglial depletion enlarged brain infarcts (Otxoa-de-Amezaga et al., 2019). The combined evidence of these studies revealed that upon brain injury microglia set the stage for successful brain remodeling and repair.

Following these seminal works, our understanding of the role of microglial cells in brain remodeling and repair has strongly expanded in the last 2 years. Studies recently published in *Frontiers of Cellular Neuroscience* revealed how microglial cells push the balance toward remodeling

REFERENCES

Anrather, J., and Iadecola, C. (2016). Inflammation and stroke: an overview. *Neurotherapeutics* 13, 661–670. doi: 10.1007/s13311-016-0483-x

Bernardino, L., Volonté, C., Passani, M. B., and Ferreira, R. (2020). Dual role of microglia in health and disease: pushing the balance towards repair. Front. Cell. Neurosci. 14:259. doi: 10.3389/fncel.2020.00259

Bianchi, M. E. (2007). DAMPs, PAMPs and alarmins: all we need to know about danger. J. Leukoc. Biol. 81, 1–5. doi: 10.1189/jlb. 0306164 and repair upon brain injury (Bernardino et al., 2020). Particularly noteworthy is the description of exosomes (Vaz et al., 2019), microparticles (Grimaldi et al., 2019), and secreted growth factors (Fuentes-Santamaría et al., 2019; Myhre et al., 2019; Wlodarczyk et al., 2019) as mediators of neuronal recovery induced by microglial cells. The role of microglia in controlling neurotransmission (Fuentes-Santamaría et al., 2019), neuronal myelination (Wlodarczyk et al., 2019), and synaptic plasticity (Gunner et al., 2019; Fuentes-Santamaría et al., 2019; Nguyen et al., 2020) was outlined. Further studies evaluated age and sex factors influencing microglial responses (Lively et al., 2018). These findings exemplify that manipulation of a distinct cell type allows modulating recovery processes in a clinically meaningful way.

Many open questions remain, e.g., with respect to: (a) bystanders of the induced degeneration of microglial cells in the living brain; (b) side effects of microglial deactivation or depletion on blood-derived immune cells, specifically of monocytes, which also carry CSF1R; and (c) the rapid repopulation of microglia following genetic or pharmacological microglia depletion. Some aspects of effects of microglial cells on neuronal and, specifically, synaptic plasticity still remain unresolved. These questions include whether or to which degree synaptic plasticity is influenced by synaptic pruning or nonphagocytic processes (Cheadle et al., 2020). Acting as a switch from injury to repair, microglial cells potently influence neurological recovery processes. Whether this strategy holds its promises in human patients still has to be shown.

DATA AVAILABILITY STATEMENT

Requests to access these datasets should be directed to dirk.hermann@uk-essen.de.

AUTHOR CONTRIBUTIONS

DH wrote the draft. Both authors revised and finalized it. All authors contributed to the article and approved the submitted version.

FUNDING

This research was supported by the German Research Foundation (Deutsche Forschungsgemeinschaft; HE3173/11-1, HE3173/12-1, HE3173/13-1, GU769/10-1 and GU769/15-1).

Cheadle, L., Rivera, S. A., Phelps, J. S., Ennis, K. A., Stevens, B., Burkly, L. C., et al. (2020). Sensory experience engages microglia to shape neural connectivity through a non-phagocytic mechanism. *Neuron* 108, 451.e9–468.e9. doi: 10.1016/j.neuron.2020.08.002

Davalos, D., Grutzendler, J., Yang, G., Kim, J. V., Zuo, Y., Jung, S., et al. (2005).
ATP mediates rapid microglial response to local brain injury in vivo. Nat. Neurosci. 8, 752–758. doi: 10.1038/nn1472

Dudvarski Stankovic, N., Teodorczyk, M., Ploen, R., Zipp, F., and Schmidt, M. H. H. (2016). Microglia-blood vessel interactions: a doubleedged sword in brain pathologies. Acta Neuropathol. 131, 347–363. doi:10.1007/s00401-015-1524-y

- Elmore, M. R., Najafi, A. R., Koike, M. A., Dagher, N. N., Spangenberg, E. E., Rice, R. A., et al. (2014). Colony-stimulating factor 1 receptor signaling is necessary for microglia viability, unmasking a microglia progenitor cell in the adult brain. *Neuron* 82, 380–397. doi: 10.1016/j.neuron.2014. 02.040
- Fuentes-Santamaría, V., Alvarado, J. C., Rodríguez-de la Rosa, L., Juiz, J. M., and Varela-Nieto, I. (2019). Neuroglial involvement in abnormal glutamate transport in the cochlear nuclei of the Igf1—/— mouse. Front. Cell. Neurosci. 13:67. doi: 10.3389/fncel.2019.00067
- Gelderblom, M., Leypoldt, F., Steinbach, K., Behrens, D., Choe, C. U., Siler, D. A., et al. (2009). Temporal and spatial dynamics of cerebral immune cell accumulation in stroke. Stroke 40, 1849–1857. doi: 10.1161/STROKEAHA.108. 534503
- Grimaldi, A., Serpe, C., Chece, G., Nigro, V., Sarra, A., Ruzicka, B., et al. (2019). Microglia-derived microvesicles affect microglia phenotype in glioma. Front. Cell. Neurosci. 13:41. doi: 10.3389/fncel.2019.00041
- Gunner, G., Cheadle, L., Johnson, K. M., Ayata, P., Badimon, A., Mondo, E., et al. (2019). Sensory lesioning induces microglial synapse elimination via ADAM10 and fractalkine signaling. Nat. Neurosci. 22, 1075–1088. doi: 10.1038/s41593-019-0419-y
- Heindl, S., Gesierich, B., Benakis, C., Llovera, G., Duering, M., and Liesz, A. (2018). Automated morphological analysis of microglia after stroke. Front. Cell. Neurosci. 12:106. doi: 10.3389/fncel.2018.00106
- Jayaraj, R. L., Azimullah, S., Beiram, R., Jalal, F. Y., and Rosenberg, G. A. (2019). Neuroinflammation: friend and foe for ischemic stroke. J. Neuroinflammation 16:142. doi: 10.1186/s12974-019-1516-2
- Lively, S., Wong, R., Lam, D., and Schlichter, L. C. (2018). Sex- and development-dependent responses of rat microglia to pro- and anti-inflammatory stimulation. Front. Cell. Neurosci. 12:433. doi: 10.3389/fncel.2018. 00433
- Lopes Pinheiro, M. A., Kooij, G., Mizee, M. R., Kamermans, A., Enzmann, G., Lyck, R., et al. (2016). Immune cell trafficking across the barriers of the central nervous system in multiple sclerosis and stroke. *Biochim. Biophys. Acta* 1862, 461–471. doi: 10.1016/j.bbadis.2015.10.018
- Manrique-Castano, D., Dzyubenko, E., Borbor, M., Vasileiadou, P., Roll, L., Kleinschnitz, C., et al. (2020). Tenascin-C preserves microglia surveillance and restricts leukocyte and, more specifically, T cell infiltration of the ischemic brain. *Brain Behav. Immun.* 1591, 32366–32367. doi: 10.1016/j.bbi.2020. 10.016
- Myhre, C. L., Thygesen, C., Villadsen, B., Vollerup, J., Ilkjær, L., Krohn, K. T., et al. (2019). Microglia express insulin-like growth factor-1 in the hippocampus of aged APPswe/PS1∆E9 transgenic mice. Front. Cell. Neurosci. 13:308. doi: 10.3389/fncel.2019.00308
- Neumann, J., Gunzer, M., Gutzeit, H. O., Ullrich, O., Reymann, K. G., and Dinkel, K. (2006). Microglia provide neuroprotection after ischemia. FASEB J. 20, 714–716. doi: 10.1096/fj.05-4882fje
- Neumann, J., Henneberg, S., von Kenne, S., Nolte, N., Muller, A. J., Schraven, B., et al. (2018). Beware the intruder: real time observation of infiltrated neutrophils and neutrophil-microglia interaction during stroke in vivo. PLoS One 13:e0193970. doi: 10.1371/journal.pone.0193970
- Neumann, J., Riek-Burchardt, M., Herz, J., Doeppner, T. R., König, R., Hütten, H., et al. (2015). Very-late-antigen-4 (VLA-4)-mediated brain invasion

- by neutrophils leads to interactions with microglia, increased ischemic injury and impaired behavior in experimental stroke. *Acta Neuropathol.* 129, 259–277. doi: 10.1007/s00401-014-1355-2
- Neumann, J., Sauerzweig, S., Rönicke, R., Gunzer, F., Dinkel, K., Ullrich, O., et al. (2008). Microglia cells protect neurons by direct engulfment of invading neutrophil granulocytes: a new mechanism of CNS immune privilege. *J. Neurosci.* 28, 5965–5975. doi: 10.1523/JNEUROSCI.0060-08.2008
- Nguyen, P. T., Dorman, L. C., Pan, S., Vainchtein, I. D., Han, R. T., Nakao-Inoue, H., et al. (2020). Microglial remodeling of the extracellular matrix promotes synapse plasticity. *Cell* 182, 388–403. doi: 10.1016/j.cell.2020.05.050
- Olmos-Alonso, A., Schetters, S. T., Sri, S., Askew, K., Mancuso, R., Vargas-Caballero, M., et al. (2016). Pharmacological targeting of CSF1R inhibits microglial proliferation and prevents the progression of Alzheimer's-like pathology. *Brain* 139, 891–907. doi: 10.1093/brain/awv379
- Otxoa-de-Amezaga, A., Miró-Mur, F., Pedragosa, J., Gallizioli, M., Justicia, C., Gaja-Capdevila, N., et al. (2019). Microglial cell loss after ischemic stroke favors brain neutrophil accumulation. *Acta Neuropathol.* 137, 321–341. doi: 10.1007/s00401-018-1954-4
- Parkhurst, C. N., Yang, G., Ninan, I., Savas, J. N., Yates, J. R., III, Lafaille, J. J., et al. (2013). Microglia promote learning-dependent synapse formation through brain-derived neurotrophic factor. *Cell* 155, 1596–1609. doi: 10.1016/j.cell. 2013.11.030
- Perez-de-Puig, I., Miro-Mur, F., Ferrer-Ferrer, M., Gelpi, E., Pedragosa, J., Justicia, C., et al. (2015). Neutrophil recruitment to the brain in mouse and human ischemic stroke. *Acta Neuropathol.* 129, 239–257. doi: 10.1007/s00401-014-1381-0
- Roth, S., Singh, V., Tiedt, S., Schindler, L., Huber, G., Geerlof, A., et al. (2018). Brain-released alarmins and stress response synergize in accelerating atherosclerosis progression after stroke. Sci. Transl. Med. 10:eaao1313. doi: 10.1126/scitranslmed.aao1313
- Vaz, A. R., Pinto, S., Ezequiel, C., Cunha, C., Carvalho, L. A., Moreira, R., et al. (2019). Phenotypic effects of wild-type and mutant SOD1 expression in N9 murine microglia at steady state, inflammatory and immunomodulatory conditions. Front. Cell. Neurosci. 13:109. doi: 10.3389/fncel.2019.00109
- Waisman, A., Ginhoux, F., Greter, M., and Bruttger, J. (2015). Homeostasis of microglia in the adult brain: review of novel microglia depletion systems. *Trends Immunol.* 36, 625–636. doi: 10.1016/j.it.2015.08.005
- Wlodarczyk, A., Benmamar-Badel, A., Cédile, O., Jensen, K. N., Kramer, I., Elsborg, N. B., et al. (2019). CSF1R stimulation promotes increased neuroprotection by CD11c+ microglia in EAE. Front. Cell. Neurosci. 12:523. doi: 10.3389/fncel.2018.00523

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Hermann and Gunzer. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.