

Emerging benefits of Fc fusion technology in the context of recombinant factor VIII replacement therapy

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Abstract

Although the primary reason for recombinant factor VIII Fc fusion protein (rFVIII_{FC}) development was to reduce treatment burden associated with routine prophylaxis, new evidence suggests additional benefits of Fc fusion technology in the treatment of people with haemophilia A. Preclinical research has been utilized to characterize the potential immunomodulatory properties of rFVIII_{FC}, including an ability to reduce inflammation and induce tolerance to factor VIII. This has since been expanded into clinical research in immune tolerance induction (ITI) with rFVIII_{FC}, results of which suggest the potential for rapid tolerization in first-time ITI patients and therapeutic benefit in patients undergoing rescue ITI. The potential for improved joint health through the anti-inflammatory properties of rFVIII_{FC} has also been suggested. In addition, a new avenue of research into the role of rFVIII_{FC} in promoting bone health in patients with haemophilia A, potentially through reduced osteoclast formation, has yielded encouraging results that support further study. This review summarizes the existing preclinical and clinical studies of immunomodulation and tolerization with rFVIII_{FC}, as well as studies in joint and bone health, to elucidate the potential benefits of rFVIII_{FC} in haemophilia A beyond the extension of factor VIII half-life.

KEYWORDS

bone resorption (MeSH terms), FVIII, haemophilia A, immune tolerance, immunomodulation, inflammation, prophylaxis

1 | INTRODUCTION

Prophylactic factor replacement is a well-established standard of care for patients with severe haemophilia A,^{1,2} and routine prophylaxis with factor VIII (FVIII) has been shown to reduce the frequency of bleed episodes, prevent joint damage and improve health-related quality of life.^{3,4} However, optimal prophylaxis with conventional FVIII products requires frequent intravenous injections, a substantial burden that can negatively impact treatment adherence.⁵

Several strategies have been employed to extend the half-life of FVIII in recombinant FVIII (rFVIII) products, thereby reducing

the frequency of infusions.⁶ Fusion of the Fc domain of human immunoglobulin G (IgG) to a therapeutic protein is an established approach that leverages an existing physiologic pathway.^{7,8} Binding to the neonatal Fc receptor (FcRn), which is expressed in endothelial cells lining the vasculature, protects both endocytosed IgG and Fc fusion proteins from lysosomal degradation and cycles them back into the circulation.^{7,8} Recombinant FVIII Fc fusion protein (rFVIII_{FC}), which comprises a single molecule of rFVIII fused to the Fc domain of human IgG1, was the first extended half-life FVIII approved in the European Union (Elocta[®], Sobi) and the United States (Eloctate[®], Sanofi) for the prophylaxis and treatment of bleeding in patients

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with haemophilia A of all age groups. The extended circulating half-life of rFVIII Fc has been confirmed,^{9,10} and long-term efficacy and safety of rFVIII Fc have been extensively documented in clinical trials^{9,11-13} and in real-world studies.¹⁴⁻¹⁷

The immunoregulatory properties of the Fc domain of IgG are an additional, albeit less well-known, characteristic of Fc fusion proteins.¹⁸ Decades ago, it was observed that coupling of haptens to IgG could induce antigen-specific tolerance,¹⁹ which was later understood to be mediated by the Fc domain^{20,21} as well as the F(ab')₂ fragment.²² IgG or Fc fusion proteins bound to the FcRn are diverted away from antigen presentation compartments, preventing an immune response.¹⁸ Later, hapten-IgG fusion proteins were also shown to induce hapten-specific tolerance²³ and intravenously administered immunoglobulin to enhance the suppressive activity of regulatory T cells (Treg)—known for their contribution to the maintenance of immunologic self-tolerance.^{22,24} Since then, two T-cell epitopes, termed Tregitopes, have been identified in the Fc region of IgG1 that are capable of activating Tregs, tipping the resulting immune response towards tolerance.²⁵ Consequently, Fc fusion proteins developed and utilized clinically to date appear to have low immunogenicity.⁷

An improved understanding of the impact of the immunomodulatory properties of Fc fusion proteins in patients with haemophilia A is just now emerging,¹⁸ as well as the potential of rFVIII Fc to tackle some of the other key unmet needs of this therapy area beyond circulating half-life, such as immune tolerance induction, joint health and potentially bone health. The objective of this review was to summarize these potential broad effects of Fc fusion technology that have multiple mechanisms of action and go beyond the original intention of extended FVIII half-life.

2 | OVERVIEW OF THE rFVIII Fc MOLECULE AND ITS PHARMACOKINETIC (PK) PROFILE

rFVIII Fc is a recombinant fusion protein comprising a single molecule of rFVIII covalently fused to the Fc domain of human IgG1.⁸ It is produced in a human cell line using recombinant DNA technology, without the use of any animal-derived components.^{8,26} The active form of the FVIII component of rFVIII Fc is structurally and

functionally comparable with native FVIII, allowing rFVIII Fc to bind normally to von Willebrand factor (VWF) and phospholipids.²⁷ Clinical studies have confirmed that rFVIII Fc displays improved clearance-related PK parameters compared with conventional human rFVIII, with a 1.5-times longer terminal half-life on average (Table 1).^{9,10}

3 | IMMUNOMODULATORY PROPERTIES OF rFVIII Fc

The development of inhibitors is a serious treatment-related complication in patients with haemophilia A,²⁸ occurring in up to 30% of patients with severe haemophilia A.^{29,30} Immune tolerance induction (ITI), involving the regular infusion of FVIII to induce FVIII antigen-specific tolerance, is the only strategy currently available to eradicate inhibitors in patients with high titres (>5 Bethesda units).^{28,31} While current ITI is successful in approximately 70% of patients with inhibitors, the treatment may take 1-2 years and is burdensome to patients and caregivers.²⁸ Given the preliminary preclinical and clinical evidence, rFVIII Fc has the potential to address the current unmet need for ITI treatments that achieve faster responses.

3.1 | Preclinical evidence

Several key preclinical studies have been fundamental in characterizing the immunomodulatory properties of Fc fusion proteins in haemophilia A, laying the groundwork for further clinical assessments. Early evidence of Fc fusion protein immunomodulation in haemophilia A came from Lei and Scott in 2005.³² They transduced B cells with a fusion IgG containing the immunogenic A2 and C2 FVIII domains and demonstrated the induction of FVIII-specific tolerance in both naïve and FVIII-immunized haemophilia A mice, likely dependent on Tregs.³² Later, both Culina et al and Gupta et al demonstrated that the transplacental transfer of Fc-fused antigens induces an increase of thymic and peripherally derived Tregs in an antigen-specific manner.^{33,34} Furthermore, in a preclinical model of haemophilia A, Gupta et al³⁴ found that transplacental transfer of Fc-fused immunodominant A2 and C2 FVIII domains

TABLE 1 Clearance-related pharmacokinetic parameters for rFVIII Fc (Eloctate[®], Sanofi Genzyme) compared with a conventional rFVIII product (Advate[®], Shire) in the Phase 3 A-LONG study in patients ≥ 12 y of age⁹

	rFVIII Fc (n = 28)	rFVIII (n = 28)	P value
t _{1/2} (h)	19.0	12.4	<.001
CL (mL/h/kg)	2.0 (1.7-2.2)	3.0 (2.7-3.4)	<.001
AUC/dose (IU × h/dL per IU/kg)	51.2 (45.0-58.4)	32.9 (29.3-36.9)	<.001
Time to 1 IU/dL (1%) FVIII trough level above baseline (days)	4.9 (4.4-5.5)	3.3 (3.0-3.7)	<.001

Note: Data presented are geometric mean (95% confidence interval).

Abbreviations: AUC/dose, dose-normalized area under the curve; CL, systemic clearance; FVIII, factor VIII; rFVIII, recombinant factor VIII; rFVIII Fc, recombinant factor VIII Fc fusion protein; t_{1/2}, terminal half-life.

TABLE 2 Preclinical studies evaluating Fc fusion proteins in models of haemophilia A

First author, year	Model	Key findings	Conclusion
Gupta 2015 ³⁴	Maternofoetal transfer of chimeric A2Fc and C2Fc proteins in HaemA mice	<ul style="list-style-type: none"> • Transplacental delivery of A2Fc- and C2Fc-induced Tregs and reduced total anti-FVIII IgG titres • Proliferation of CD4+CD25- Tregs from FVIII-primed mice and the antibody response against FVIII upon replacement therapy were reduced by splenic Tregs from mice treated transplacentally with A2Fc plus C2Fc, as compared with Tregs from IgG1-treated mice 	<ul style="list-style-type: none"> • Transplacental transfer of Fc-fused A2 and C2 FVIII domains induced tolerance to FVIII in the progeny, attributable to FVIII-specific Tregs
Krishnamoorthy 2016 ³⁵	Evaluation of immune response to rFVIII-Fc in comparison with BDD-rFVIII and FL-rFVIII (FL-rFVIII) in HaemA mice	<ul style="list-style-type: none"> • rFVIII-Fc at therapeutically relevant doses was less immunogenic and resulted in less inhibitor formation compared with FL-rFVIII and BDD-rFVIII • rFVIII-Fc induced FVIII-specific tolerance • rFVIII-Fc promoted the expression of cytokines associated with tolerance, prevented the expression of inflammatory cytokines and led to upregulation of tolerance-related markers (eg FOXP3, CD25 and PD-1) • Disruption of Fc interactions with either FcRn or Fcγ receptors diminished tolerance induction, suggesting involvement of these pathways 	<ul style="list-style-type: none"> • At therapeutically relevant doses, rFVIII-Fc was less immunogenic than FL-rFVIII and BDD-rFVIII, promoted phenotypic Treg development and promoted a tolerogenic splenic microenvironment in HaemA mice • Mechanistically, this tolerogenic effect was partly mediated by the Fc receptors Fcγ and FcRn
Kis-Toth 2018 ³⁶	In vitro treatment of human monocyte-derived macrophages with rFVIII-Fc	<ul style="list-style-type: none"> • rFVIII-Fc interacts with human monocyte-derived macrophages via their FcRs, which initiates signalling without classical proinflammatory cell activation • rFVIII-Fc-treated macrophages exhibit specific gene expression pattern indicating a shift in phenotype 	<ul style="list-style-type: none"> • rFVIII-Fc induces an FcR-dependent macrophage polarization to an alternatively activated Mox/M2 phenotype with antioxidant characteristics

Abbreviations: Ag, antigen; BDD, B-domain-deleted; FcR, Fc receptor; FcRn, neonatal Fc receptor; Fc γ , Fc γ receptor; FL, full length; FVIII, factor VIII; HaemA, haemophilia A; IgG, immunoglobulin G; rFVIII, recombinant factor VIII; rFVIII-Fc, recombinant factor VIII Fc fusion protein; Teff, effector T cell and Treg, regulatory T cell;

induced tolerance to FVIII in the progeny, attributable to FVIII-specific Tregs; however, the role of Fc in the induction of tolerance was not investigated (Table 2).

A key study for rFVIII-Fc came from Krishnamoorthy et al,³⁵ who reported decreased levels of inhibitor formation after rFVIII-Fc treatment of haemophilia A mice with therapeutically relevant doses, compared with rFVIII treatment (Table 2). The reduced immunogenicity of rFVIII-Fc was attributed to the development of regulatory T cells and a tolerogenic environment, potentially mediated by the interaction of the Fc domain of rFVIII-Fc with the Fc receptors (FcR) on antigen-presenting cells. The authors proposed that a combination of the FcRn and Fc γ receptor signalling pathways, and potentially Fc Tregitopes, contributed to rFVIII-Fc-mediated activation of tolerogenic pathways.

Additional work has since been undertaken to characterize the interaction between rFVIII-Fc and antigen-presenting cells. An in vitro study published by Kis-Toth et al³⁶ showed that rFVIII-Fc,

but not rFVIII, elicits an alternatively activated regulatory macrophage phenotype, further elucidating on the mechanisms of potential rFVIII-Fc immunomodulatory properties (Table 2). rFVIII-Fc was shown to induce human monocyte-derived macrophages to polarize to a Mox/M2 phenotype in vitro, without need for a danger signal or interleukin 4. This report suggests that rFVIII-Fc drives regulatory macrophage polarization in an FcR-dependent way, resulting in an antioxidant state and anti-inflammatory molecular profile.

3.2 | Clinical evidence

ITI with rFVIII-Fc has been examined in case reports,³⁷⁻³⁹ small cohort studies^{40,41} and in prospective studies and chart reviews (some of which are still ongoing; NCT03951103) (Table 3).⁴²⁻⁴⁴ The definition of success with ITI should be taken into account when interpreting the data. While successful tolerization is typically defined as achieving

TABLE 3 Clinical studies and case reports evaluating ITI with rFVIII Fc

First author, year	Study design	Patient(s)	rFVIII Fc ITI dose	ITI outcomes	Safety outcomes
Groomes 2016 ³⁷	Case report	15-mo-old with severe haemophilia A who received prior ITI with Kogenate and Xyntha	50 IU/kg TIW	Inhibitor titre decreased from 11 to 0.7 BU/mL over 308 d	NR
Batsuli 2019 ³⁹	Case report	12-y-old with severe haemophilia A and four prior ITI attempts; concomitant treatment with emicizumab	100 IU/kg TIW	Inhibitor titre decreased from a historical peak titre of 198 BU/mL to <0.6 BU/mL after 37 wk of rFVIII Fc treatment	NR
Malec 2016 ³⁸	Case series	Children with severe haemophilia A undergoing their first ITI treatment (n = 2) or who failed prior ITI (n = 1)	100 IU/kg QOD; 200 IU/kg TIW/QOD	3/3 achieved negative inhibitor status in 11, 4 and 12 wk; no patients had recurrence of detectable inhibitor at mean follow-up of 13–14 mo	No patients experienced treatment complications
Carcao 2019 ⁴²	Non-interventional, retrospective chart review	Children with severe haemophilia A undergoing their first ITI treatment or who have failed previous ITI attempts	43 IU/kg TIW to 200 IU/kg/d	Interim results (8 November 2018): 9/10 first-time ITI patients tolerated in median (range) of 30 (3–99) weeks; 9/18 rescue ITI patients achieved Bethesda negativity with 3 transitioned to prophylaxis	No adverse events reported
Nagao 2019 (FACTS study) ⁴⁰	Prospective, multicentre, observational study	Adolescents and children with haemophilia A undergoing their first ITI treatment (n = 5) or who have failed previous ITI attempts (n = 3)	39.7 IU/kg TIW to 227.3 IU/kg/d	Interim results (October 2018): Decreased inhibitor titre to <1 BU/mL in 2/5 first-time ITI patients at 6 mo; ITI ongoing in all patients	No severe adverse events reported
Abraham 2018 ⁴¹	Prospective, multicentre, observational study	Patients with haemophilia A (N = 38)	50 IU/kg TIW to 200 IU/kg/d	Interim results: 17/38 achieved negative inhibitor status in a median (range) of 20 (10–60) weeks	NR
Malec 2019 (verITI-8 study) ⁴³	Open-label, single-arm, interventional, multicentre, Phase 4 study	Patients with severe haemophilia A undergoing their first ITI treatment (N = 16)	200 IU/kg/d	Interim results (23 January 2019): 6/15 tolerated in median (range) of 11.7 (9.9–12.3) weeks; ITI ongoing in 8/15; ITI failure in 1/15	No adverse events related to rFVIII Fc reported
ReITrate study ⁴⁴	Open-label, single-arm, interventional, multicentre, Phase 4 study	Patients with severe haemophilia A who have failed previous ITI attempts	200 IU/kg/d	NA (study ongoing)	NA (study ongoing)
NCT03951103	Observational, retrospective and prospective chart review	Patients with haemophilia A who have been, or who are currently, treated with rFVIII Fc for ITI (N = ~50)	NR	NA (study ongoing)	NA (study ongoing)
NCT04303572 (The Hemophilia Inhibitor Eradication Trial)	Open-label, randomized, interventional, multicentre, Phase 3 study	Male adults or children >4 mo of age with severe haemophilia A and current or past high-responding FVIII inhibitors	Eloctate 100 IU/kg QOD with or without emicizumab 1.5 mg/kg QOD	NA (planned start in June 2020)	NA (planned start in June 2020)

Abbreviations: BU, Bethesda unit; ITI, immune tolerance induction; NA, not available; NR, not reported; QOD, every other day; TIW, three times per week.

negative Bethesda titre (≤ 0.6 BU) as well as normal FVIII recovery ($\geq 66\%$) and half-life (≥ 6 hours),²⁸ a minority of studies have described negative Bethesda titres alone as a successful outcome.

Case reports from Groomes et al and Malec et al demonstrate successful ITI with rFVIIIc and the potential to achieve tolerance with a regimen of shorter duration and at a lower dose frequency than ITI utilizing conventional half-life products.^{37,38} In addition, there is a case report of successful ITI with rFVIIIc after multiple failed attempts in the setting of emicizumab prophylaxis.³⁹

Carcao et al reported outcomes from a non-interventional, retrospective chart review of patients with severe haemophilia A and high-titre inhibitors treated with rFVIIIc for ITI,^{42,45} the most recent analysis of which includes 28 patients.⁴² Nine of 10 first-time ITI patients were tolerized with a median time to tolerization of 30 (range 3-99) weeks. Nine of 18 rescue ITI patients achieved Bethesda negativity, and of these, 3 patients were tolerized at 21.7, 35.0 and 101.1 weeks, respectively, and transitioned to rFVIIIc prophylaxis. Despite the majority of patients in this study having poor-risk features for ITI success, rFVIIIc use demonstrated a rapid decrease in inhibitor titres and rapid tolerization in first-time ITI patients and showed therapeutic benefit in patients undergoing rescue ITI. Nagao et al and Abraham et al have both reported interim results from observational analyses with promising outcomes (Table 3).^{40,41}

Two prospective open-label, single-arm, interventional, multicentre studies are now underway to examine the efficacy of rFVIIIc in ITI (Table 3). The verITI-8 study (NCT03093480) will assess the time to tolerization with rFVIIIc over a 48-week study period in 16 patients undergoing primary ITI treatment.⁴³ Interim results show that, as of the 23 January 2019 data cut-off, 6 out of 15 patients were successfully tolerized in a median of 11.7 (range 9.9-12.3) weeks. rFVIIIc ITI is ongoing in 8 patients, and 1 patient failed to achieve ITI success by Week 48.⁴³ The ReITrate study (NCT03103542) will assess the rate of ITI success using rFVIIIc over a 60-week study period in 16 patients undergoing rescue ITI therapy having failed at least one previous ITI attempt.⁴⁴

A 5-year observational chart review (NCT03951103) in an estimated 50 patients who have been, or who are currently, treated with rFVIIIc for ITI is also ongoing in Europe and the Middle East (Table 3). The Hemophilia Inhibitor Eradication Trial (NCT04303572), planned to start in June 2020, is an open-label, randomized, interventional, multicentre, 48-week Phase 3 trial that will compare weekly rFVIIIc ITI plus weekly emicizumab with weekly rFVIIIc ITI alone to eradicate inhibitor formation in approximately 90 patients (Table 3).

It is encouraging that in clinical trials evaluating a prophylaxis regimen in previously treated patients, no major signals of immunogenicity for rFVIIIc have been observed.^{9,12,13} For instance, there was no inhibitor development, nor any reports of anaphylaxis or serious vascular thrombotic events, or deaths or serious adverse events considered related to rFVIIIc. However, it should be acknowledged that previously treated patients are at a lower risk of developing an alloimmune response to exogenous FVIII as compared with previously untreated patients (PUPs).³⁰

Results from clinical trials in PUPs, a population for whom clinical data are currently limited, are expected to answer key questions on

the immunomodulatory properties of this molecule. In a case report of two cousins with severe haemophilia A and the same high-risk genotype, the patient treated with rFVIIIc developed low-titre inhibitors not requiring ITI whereas the patient treated with full-length rFVIII developed high-titre inhibitors requiring ITI within a similar timeframe.⁴⁶ The tolerogenic potential of rFVIIIc was offered as a possible explanation for the findings of this report. The Phase 3 PUPs A-LONG trial (NCT02234323) investigating the safety and efficacy of rFVIIIc in PUPs with haemophilia A was recently completed (September 2019) and is the only study, to date, of rFVIIIc in this population.⁴⁷ This was an international, open-label, interventional single-group, multicentre, 3-year study in 108 PUPs aged up to 5 years, the primary outcome for which was the number of participants with inhibitor development over the duration of the study. Data from an interim analysis (N = 95), presented recently, indicate that rFVIIIc was well tolerated and efficacious for prevention of bleeds in a PUP population in which 80% of subjects had a high-risk genotype. While the development of inhibitors in 31% of subjects is within the expected range in PUPs with severe haemophilia A (ie 25%-40%⁴⁸⁻⁵⁰), there was a low risk for development of high-titre inhibitors (15% in subjects with ≥ 10 exposure days or who had an inhibitor).

It has been suggested that activation of the immune system (ie via bleeds, infection, trauma and vaccination) must be minimized at the time of first FVIII exposure to achieve tolerance in PUPs.^{51,52} This approach will be assessed in The Hemophilia Inhibitor Prevention Trial (NCT04303559), a multicenter, randomized, controlled Phase 3 trial that compares weekly rFVIIIc with weekly emicizumab in PUPs with severe haemophilia A, 4 months of age or older.

4 | JOINT AND BONE HEALTH

While haemophilic arthropathy is the leading cause of morbidity in haemophilia A,⁵³ osteopenia (low bone mineral density) and osteoporosis are highly prevalent,^{54,55} and fracture rates are also elevated.^{56,57} In a retrospective cohort of 382 patients, there was a significantly greater relative risk of fracture in patients with haemophilia A and haemophilia B compared with the control population (relative risk: 10.7, 95% confidence interval: 8.2-14.1; $P < .0001$), with risk increasing with age and with haemophilia severity.⁵⁷ The exact aetiology behind both the elevated osteopenia and increased fracture risk, and the interplay between these comorbidities, has not yet been elucidated; nevertheless, it is clear that joint and bone health in patients with haemophilia A remains a significant unmet need beyond successful prevention of joint bleeds.⁵⁸

4.1 | Anti-inflammatory properties of rFVIIIc: Clinical evidence in joint health

The largest body of evidence to date on the impact of rFVIIIc prophylaxis on joint health in patients with haemophilia A comes from a post hoc analysis of A-LONG and Kids A-LONG studies and

the ASPIRE extension study.^{59–61} Joint health showed continued improvement, as reflected by changes in Hemophilia Joint Health Score in children and modified Hemophilia Joint Health Score (mHJHS) in adolescents and adults (domains include swelling, pain, strength, flexion and extension loss), from A-LONG/Kids A-LONG baseline to ASPIRE Year 2.^{59,61} Resolution of at least one target joint (defined as ≤ 2 spontaneous bleed episodes in 12 consecutive months) was achieved in 99.6% of evaluable patients in A-LONG/ASPIRE ($n = 235$) and 100% of evaluable patients in Kids A-LONG ($n = 9$). Improvements in joint health and 100% target joint resolution were also reported in patients who did not reach optimum haemostatic control (ie had an annualized bleed rate in the top 25%) in the first year of treatment with rFVIII-Fc in the A-LONG study and who continued prophylaxis with rFVIII-Fc in ASPIRE.⁶⁰ The mechanism behind this improvement in joint health and target joint health resolution with prolonged rFVIII-Fc treatment most likely relates to low bleed rates. Changes in mHJHS that contributed most to improvements in total score were swelling, range of motion and strength.⁵⁹ The contribution of increased adherence to prophylaxis and the potential anti-inflammatory properties of the Fc-fused rFVIII (described earlier) remain to be fully elucidated.⁵⁹

Interestingly, a prospective study in 26 adult patients with haemophilia A and haemophilia B with arthropathic joints receiving various factor replacement therapies found that acute painful bleed episodes were not associated with expected PK parameters (ie not associated with more time spent below certain clotting factor thresholds in the days preceding the bleed). However, joint bleeds were found to be associated with prominent vascularity changes, indicating that vascular remodelling and leakiness—probably a result of previous joint bleeds—may contribute to joint bleeds. Given that the patients receiving rFVIII-Fc experienced fewer painful episodes than did those receiving conventional products, the potential anti-inflammatory properties of rFVIII-Fc were proposed to play a role.⁶²

4.2 | Impact of rFVIII-Fc on osteoclast formation: Preclinical evidence in bone health

The role of FVIII and/or thrombin in bone health is currently unclear. While animal models support a direct role for FVIII in bone remodelling, whereby RANKL (which increases bone breakdown) expression is decreased by FVIII treatment, bone formation may also be indirectly affected by FVIII via thrombin generation.⁵⁸ It is, therefore, interesting to note the findings of a recent in vitro study presented by Rajani et al,⁶³ where rFVIII-Fc, but not FVIII, was shown to inhibit monocyte-derived osteoclast formation, suggesting a potential effect on bone resorption in patients with haemophilia. In this study, the bone resorption capabilities of rFVIII-Fc-treated, monocyte-derived osteoclasts were compromised compared with untreated, human IgG1- or rFVIII-treated cells. Furthermore, gene and protein expression of rFVIII-Fc-treated cells showed upregulation of NRF2 pathway-related molecules

and subsequent downregulation of molecules involved in osteoclast formation and function.⁶³ These findings lend support to further study of the potential benefits of rFVIII-Fc for bone health in patients with haemophilia A.

5 | CONCLUSION

Further to its prolonged half-life compared with conventional rFVIII products, the Fc domain of rFVIII-Fc may confer additional benefits to patients with haemophilia A owing to its immunomodulatory and non-immunomodulatory effects. Based on the available preclinical and clinical evidence, these may include reduced immunogenicity and inflammation, and an enhanced ability to induce tolerance. An additional emerging role for rFVIII-Fc in promoting bone health, potentially through reduced osteoclast activation and, consequently, bone resorption, lends further support to the belief that the Fc fragment of rFVIII-Fc confers benefits, beyond those originally intended in the molecule's design, that have yet to be fully characterized. The Fc domain continues to be incorporated into novel haemophilia therapeutics, such as BIVV001 (rFVIII-Fc-VWF-XTEN), a new class of FVIII replacement.⁶⁴ Whether these emerging benefits of the Fc fragment persist in highly bio-engineered proteins remain to be elucidated.

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REFERENCES

1. Fischer K, Ljung R. Primary prophylaxis in haemophilia care: guideline update 2016. *Blood Cells Mol Dis*. 2017;67:81–85.
2. Srivastava A, Brewer AK, Mauser-Bunschoten EP, et al. Guidelines for the management of hemophilia. *Haemophilia*. 2013;19:e1–47.
3. Manco-Johnson MJ, Abshire TC, Shapiro AD, et al. Prophylaxis versus episodic treatment to prevent joint disease in boys with severe hemophilia. *N Engl J Med*. 2007;357:535–544.



4. Usuba K, Price VE, Blanchette V, et al. Impact of prophylaxis on health-related quality of life of boys with hemophilia: an analysis of pooled data from 9 countries. *Res Pract Thromb Haemost.* 2019;3:397-404.
5. Thornburg CD, Duncan NA. Treatment adherence in hemophilia. *Patient Prefer Adherence.* 2017;11:1677-1686.
6. Graf L. Extended half-life factor VIII and factor IX preparations. *Transfus Med Hemother.* 2018;45:86-91.
7. Rath T, Baker K, Dumont JA, et al. Fc-fusion proteins and FcRn: structural insights for longer-lasting and more effective therapeutics. *Crit Rev Biotechnol.* 2015;35:235-254.
8. Mancuso ME, Mannucci PM. Fc-fusion technology and recombinant FVIII and FIX in the management of the hemophilias. *Drug Des Devel Ther.* 2014;8:365-371.
9. Mahlangu J, Powell JS, Ragni MV, et al. Phase 3 study of recombinant factor VIII Fc fusion protein in severe hemophilia A. *Blood.* 2014;123:317-325.
10. Powell JS, Josephson NC, Quon D, et al. Safety and prolonged activity of recombinant factor VIII Fc fusion protein in hemophilia A patients. *Blood.* 2012;119:3031-3037.
11. Nolan B, Mahlangu J, Perry D, et al. Long-term safety and efficacy of recombinant factor VIII Fc fusion protein (rFVIII-Fc) in subjects with haemophilia A. *Haemophilia.* 2016;22:72-80.
12. Young G, Mahlangu J, Kulkarni R, et al. Recombinant factor VIII Fc fusion protein for the prevention and treatment of bleeding in children with severe hemophilia A. *J Thromb Haemost.* 2015;13:967-977.
13. Nolan B, Mahlangu J, Pabinger I, et al. Recombinant factor VIII Fc fusion protein for the treatment of severe haemophilia A: final results from the ASPIRE extension study. *Haemophilia.* 2020;26:494-502.
14. Peyvandi F, Garagiola I, Boscarino M, Ryan A, Hermans C, Makris M. Real-life experience in switching to new extended half-life products at European haemophilia centres. *Haemophilia.* 2019;25:946-952.
15. Scott M, Xiang H, Collins PW, Hay CR. The effect of switching to rFVIII-Fc on treatment patterns and annualised bleed rate before and after: a within-patient comparison from the UK National Haemophilia Database. *Haemophilia.* 2018;24:3-196.
16. Tagliaferri A, Matichecchia A, Rivolta GF, et al. Optimising prophylaxis outcomes and costs in haemophilia patients switching to recombinant FVIII-Fc: a single-centre real-world experience. *Blood Transfus.* 2019;1:11.
17. Wang C, Young G. Clinical use of recombinant factor VIII Fc and recombinant factor IX Fc in patients with haemophilia A and B. *Haemophilia.* 2018;24:414-419.
18. Blumberg RS, Lillicrap D, the IgG Fc Immune Tolerance Group. Tolerogenic properties of the Fc portion of IgG and its relevance to the treatment and management of hemophilia. *Blood.* 2018;131:2205-2214.
19. Borel Y. Haptens bound to self IgG induce immunologic tolerance, while when coupled to syngeneic spleen cells they induce immune suppression. *Immunol Rev.* 1980;50:71-104.
20. Waldschmidt TJ, Borel Y, Vitetta ES. The use of haptened immunoglobulins to induce B cell tolerance in vitro. The roles of hapten density and the Fc portion of the immunoglobulin carrier. *J Immunol.* 1983;131:2204-2209.
21. Baxevanis CN, Ioannides CD, Reclos GJ, Papamichail M. Evidence for distinct epitopes on human IgG with T cell proliferative and suppressor function. *Eur J Immunol.* 1986;16:1013-1016.
22. Ephrem A, Chamat S, Miquel C, et al. Expansion of CD4⁺CD25⁺ regulatory T cells by intravenous immunoglobulin: a critical factor in controlling experimental autoimmune encephalomyelitis. *Blood.* 2008;111:715-722.
23. Zambidis ET, Scott DW. Epitope-specific tolerance induction with an engineered immunoglobulin. *Proc Natl Acad Sci U S A.* 1996;93:5019-5024.
24. Kessel A, Ammuri H, Peri R, et al. Intravenous immunoglobulin therapy affects T regulatory cells by increasing their suppressive function. *J Immunol.* 2007;179:5571-5575.
25. De Groot AS, Moise L, McMurry JA, et al. Activation of natural regulatory T cells by IgG Fc-derived peptide "Tregitopes". *Blood.* 2008;112:3303-3311.
26. McCue J, Kshirsagar R, Selvitelli K, et al. Manufacturing process used to produce long-acting recombinant factor VIII Fc fusion protein. *Biologicals.* 2015;43:213-219.
27. Leksa N, Pearse B, Goodman A, et al. Identification of FIXa- and FX-specific antibodies for the generation of bispecific antibodies with FVIIIa-like activity. *Res Pract Thromb Haemost.* 2017;1:1-1451.
28. Valentino LA, Kempton CL, Kruse-Jarres R, Mathew P, Meeks SL, Reiss UM. US Guidelines for immune tolerance induction in patients with haemophilia A and inhibitors. *Haemophilia.* 2015;21:559-567.
29. Witmer C, Young G. Factor VIII inhibitors in hemophilia A: rationale and latest evidence. *Ther Adv Hematol.* 2013;4:59-72.
30. Ing M, Gupta N, Teyssandier M, et al. ABIRISK consortium. Immunogenicity of long-lasting recombinant factor VIII products. *Cell Immunol.* 2016;301:40-48.
31. Osooli M, Berntorp E. Inhibitors in haemophilia: what have we learned from registries? A systematic review. *J Intern Med.* 2015;277:1-15.
32. Lei TC, Scott DW. Induction of tolerance to factor VIII inhibitors by gene therapy with immunodominant A2 and C2 domains presented by B cells as Ig fusion proteins. *Blood.* 2005;105:4865-4870.
33. Culina S, Gupta N, Boisgard R, et al. Materno-fetal transfer of preproinsulin through the neonatal Fc receptor prevents autoimmune diabetes. *Diabetes.* 2015;64:3532-3542.
34. Gupta N, Culina S, Meslier Y, et al. Regulation of immune responses to protein therapeutics by transplacental induction of T cell tolerance. *Sci Transl Med.* 2015;7:275ra21.
35. Krishnamoorthy S, Liu T, Drager D, et al. Recombinant factor VIII Fc (rFVIII-Fc) fusion protein reduces immunogenicity and induces tolerance in hemophilia A mice. *Cell Immunol.* 2016;301:30-39.
36. Kis-Toth K, Rajani GM, Simpson A, et al. Recombinant factor VIII Fc fusion protein drives regulatory macrophage polarization. *Blood Adv.* 2018;2:2904-2916.
37. Groomes CL, Gianferante DM, Crouch GD, Parekh DS, Scott DW, Lieuw K. Reduction of factor VIII inhibitor titers during immune tolerance induction with recombinant factor VIII-Fc fusion protein. *Pediatr Blood Cancer.* 2016;63:922-924.
38. Malec LM, Journeycake J, Ragni MV. Extended half-life factor VIII for immune tolerance induction in haemophilia. *Haemophilia.* 2016;22:e552-e554.
39. Batsuli G, Zimowski KL, Tickle K, Meeks SL, Sidonio RF Jr. Immune tolerance induction in paediatric patients with haemophilia A and inhibitors receiving emicizumab prophylaxis. *Haemophilia.* 2019;25:789-796.
40. Nagao A, Nagae C, Moritani K, et al. Real-world data of immune tolerance induction using recombinant factor VIII Fc fusion protein for in hemophilia A patients with inhibitors in Japan: observational Fc Adolescent and Children Treatment Study (FACTs) first interim reports. *Res Pract Thromb Haemost.* 2019;3:1-891.
41. Abraham A, Apte S, Shamukhaiah C, et al. Outcome of immune tolerance induction using an extended half-life clotting factor concentrate—recombinant factor VIII Fc (Eloctate™)—a report from India. *Blood.* 2018;132:2494.
42. Carcao M, Shapiro A, Hwang N, et al. Real-world data of immune tolerance induction (ITI) using recombinant factor VIII Fc fusion protein (rFVIII-Fc) in subjects with severe hemophilia A with inhibitors at high risk for ITI failure. *Haemophilia.* 2019;25:3-77.
43. Malec L, Carcao M, Jain N, et al. rFVIII-Fc for first-time immune tolerance induction (ITI) therapy: interim results from the global, prospective verITI-8 study. Paper presented at: 27th Congress of the

- International Society on Thrombosis and Haemostasis (ISTH), July 6-10, 2019; Melbourne, Australia.
44. Königs CMS, Jain N, Lethagen S. Study design for reiterate—a prospective study of rescue ITI with recombinant factor VIII Fc (rFVIII-Fc) in patients who have failed previous ITI attempts. *Haemophilia*. 2018;24:32-135.
 45. Carcao M, Shapiro A, Staber JM, et al. Recombinant factor VIII Fc fusion protein for immune tolerance induction in patients with severe haemophilia A with inhibitors—a retrospective analysis. *Haemophilia*. 2018;24:245-252.
 46. Ragni MV, Alabek M, Malec LM. Inhibitor development in two cousins receiving full-length factor VIII (FVIII) and FVIII-Fc fusion protein. *Haemophilia*. 2016;22:e462-464.
 47. Königs C, Liesner R, Ozelo MC, et al. Incidence of inhibitors in previously untreated patients with severe haemophilia A treated with rFVIII-Fc: the PUPs A-LONG study. *Haemophilia*. 2019;25:25-34.
 48. Darby SC, Keeling DM, Spooner RJD, et al. The incidence of factor VIII and factor IX inhibitors in the hemophilia population of the UK and their effect on subsequent mortality, 1977-99. *J Thromb Haemost*. 2004;2:1047-1054.
 49. Gouw SC, van der Bom JG, Marijke van den Berg H. Treatment-related risk factors of inhibitor development in previously untreated patients with hemophilia A: the CANAL cohort study. *Blood*. 2007;109:4648-4654.
 50. Peyvandi F, Mannucci PM, Garagiola I, et al. A randomized trial of factor VIII and neutralizing antibodies in hemophilia A. *N Engl J Med*. 2016;374:2054-2064.
 51. Ragni MV, Malec LM. Design of the INHIBIT trial: preventing inhibitors by avoiding 'danger', prolonging half-life and promoting tolerance. *Expert Rev Hematol*. 2014;7:747-755.
 52. Ragni MV, George LA. The national blueprint for future factor VIII inhibitor clinical trials: NHLBI State of the Science (SOS) Workshop on factor VIII inhibitors. *Haemophilia*. 2019;25:581-589.
 53. Knobe K, Berntorp E. Haemophilia and joint disease: pathophysiology, evaluation, and management. *J Comorb*. 2011;1:51-59.
 54. Kempton CL, Antun A, Antonucci DM, et al. Bone density in haemophilia: a single institutional cross-sectional study. *Haemophilia*. 2014;20:121-128.
 55. Gerstner G, Damiano ML, Tom A, et al. Prevalence and risk factors associated with decreased bone mineral density in patients with haemophilia. *Haemophilia*. 2009;15:559-565.
 56. Caviglia H, Landro ME, Galatro G, Candela M, Neme D. Epidemiology of fractures in patients with haemophilia. *Injury*. 2015;46:1885-1890.
 57. Gay ND, Lee SC, Liel MS, Sochacki P, Recht M, Taylor JA. Increased fracture rates in people with haemophilia: a 10-year single institution retrospective analysis. *Br J Haematol*. 2015;170:584-586.
 58. Samuelson Bannow B, Recht M, Négrier C, et al. Factor VIII: long-established role in haemophilia A and emerging evidence beyond haemostasis. *Blood Rev*. 2019;35:43-50.
 59. Oldenburg J, Kulkarni R, Srivastava A, et al. Improved joint health in subjects with severe haemophilia A treated prophylactically with recombinant factor VIII Fc fusion protein. *Haemophilia*. 2018;24:77-84.
 60. Kulkarni R, Shapiro A, Pasi KJ, et al. Improved hemostasis and joint health over time in a subset of patients who did not reach optimal hemostatic control in the first year of recombinant factor VIII Fc fusion protein (rFVIII-Fc) therapy. *Res Pract Thromb Haemost*. 2019;3:1-189.
 61. Oldenburg J, Pasi KJ, Pabinger I, et al. Improvements in joint health during long-term use of recombinant factor VIII Fc fusion protein prophylaxis in subjects with haemophilia A. *Haemophilia*. 2019;25:1-154.
 62. Zhou JY, Barnes RFW, Foster G, Iorio A, Cramer TJ, von Drygalski A. Joint bleeding tendencies in adult patients with hemophilia: it's not all pharmacokinetics. *Clin Appl Thromb Hemost*. 2019;25:1076029619862052.
 63. Rajani GM, Lin Y, Kis-Toth K, Peters R, Salas J. Recombinant factor VIII Fc fusion protein inhibits inflammatory osteoclast formation in vitro. *Res Pract Thromb Haemost*. 2019;3:1-228.
 64. Seth Chhabra E, Liu T, Kulman J, et al. BIVV001, a new class of factor VIII replacement for hemophilia A that is independent of von Willebrand factor in primates and mice. *Blood*. 2020;135:1484-1496.

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