



Complete Genome Sequence of the Environmental Burkholderia pseudomallei Sequence Type 131 Isolate MSHR1435, Associated with a Chronic Melioidosis Infection

Jason W. Sahl,^a Mark Mayo,^b ^bErin P. Price,^{b,*} ^bDerek S. Sarovich,^{b,*} Mirjam Kaestli,^b Talima Pearson,^a Charles H. D. Williamson,^a Roxanne Nottingham,^a Krystal Sheridan,^a David M. Wagner,^a Bart J. Currie,^b ^bPaul Keim^a

^aPathogen and Microbiome Institute, Northern Arizona University, Flagstaff, Arizona, USA ^bMenzies School of Health Research, Darwin, Australia

ABSTRACT The *Burkholderia pseudomallei* isolate MSHR1435 is a fully virulent environmental sequence type 131 (ST131) isolate that is epidemiologically associated with a 17.5-year chronic melioidosis infection. The completed genome will serve as a reference for studies of environmental ecology, virulence, and chronic *B. pseudomallei* infections.

Jurkholderia pseudomallei is the causative agent of melioidosis, a usually acute $\check{\mathbf{D}}$ disease, with \sim 50% of patients presenting with pneumonia and with an overall mortality rate of 10 to 40% (1). Chronic melioidosis (defined as symptoms being present for more than 2 months before diagnosis) accounts for around 10% of presentations (2). The longest continuous chronic melioidosis infection to be reported is patient 314 (P314) from the Darwin Prospective Melioidosis Study (2), who was initially diagnosed in June 2000 with a sputum culture-positive infection and who remains sputum culture positive as of August 2017 (3). Analysis of 3 clinical P314 genomes (MSHR1655, MSHR1043, and MSHR6686) deposited in GenBank (4) demonstrated substantial mutations and genome deletions with attenuated virulence, explaining the persisting chronic carriage without clinical deterioration (3). Here, we describe the completed genome sequence of MSHR1435, isolated in 2002 from water samples near the home of P314 in the Northern Territory, Australia. The close genetic similarity between the P314 clinical genomes and MSHR1435 suggests that this strain reflects B. pseudomallei environmental strains that were involved in the initial infection in P314. Clinical strains show mutations in genes associated with virulence, suggesting that MSHR1435 represents a model strain to better understand the genomic mechanisms behind B. pseudomallei pathogenesis in this unique-to-date chronic infection scenario.

Total genomic DNA was extracted using the DNeasy blood and tissue kit from Qiagen, with an attempt to limit mechanical shearing. For PacBio sequencing, approximately 10 μ g of DNA was fragmented to 10 to 20 kbp using the g-TUBE apparatus (Covaris). The sequencing library was constructed using the SMRTbell template prep kit 1.0 and according to the PacBio 20-kb library protocol. Sequencing was performed on the PacBio RS II instrument in one single-molecule real-time (SMRT) cell (version 3) for 6 h. Illumina libraries were constructed with 300-bp inserts using standard methods. The genome was assembled with Canu version 1.3 (5), with 6 iterations of Pilon version 1.22 (6) to correct small indels, resulting in 2 contigs representing the 2 chromosomes in *B. pseudomallei*. The genome was reoriented with Circlator version 1.4.0 (7) and annotated with the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) version 4.3 (8).

The chromosomes of MSHR1435 were 4,019,555 (chromosome 1) and 3,258,775 (chromosome 2) nucleotides (nt) in size, with an average GC content of 67.9%. PGAP

Received 6 February 2018 Accepted 13 February 2018 Published 15 March 2018

Citation Sahl JW, Mayo M, Price EP, Sarovich DS, Kaestli M, Pearson T, Williamson CHD, Nottingham R, Sheridan K, Wagner DM, Currie BJ, Keim P. 2018. Complete genome sequence of the environmental *Burkholderia pseudomallei* sequence type 131 isolate MSHR1435, associated with a chronic melioidosis infection. Genome Announc 6:e00072-18. https://doi .org/10.1128/genomeA.00072-18.

Copyright © 2018 Sahl et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Paul Keim, paul.keim@nau.edu.

* Present address: Erin P. Price, University of the Sunshine Coast, Maroochydore, Australia; Derek S. Sarovich, University of the Sunshine Coast, Maroochydore, Australia. predicted 6,946 coding genes, 4 complete *rrn* operons, and 60 tRNAs. A screen of virulence genes previously identified in *B. pseudomallei* using the large-scale BLAST score ratio (LS-BSR) (9) pipeline demonstrated the presence of the *Burkholderia thailandensis*-like flagellum and chemotaxis (BTFC) gene cluster (10), lipopolysac-charide (LPS) genotype A (11), an intact *wcbR* gene associated with capsular polysaccharide synthesis (12), the filamentous hemagglutinin gene, *fhaB3*, associated with positive blood cultures (13), and intact type III (14) and type VI secretion systems (15). A study of stepwise mutations in clinical strains from P314 compared to MSHR1435 will provide insight into the genomic mechanisms associated with attenuated virulence and chronic colonization in *B. pseudomallei*.

Accession number(s). This whole-genome assembly has been deposited at DDBJ/ ENA/GenBank under accession numbers CP025264 and CP025265. Raw data were deposited in the Sequence Read Archive (SRA) for both PacBio (SRR6413814) and Illumina (SRR5314868) read data.

ACKNOWLEDGMENTS

We acknowledge Vanessa Theobald and Glenda Harrington at Menzies for assistance with *B. pseudomallei* collection and culture.

This project was funded under Defense Threat Reduction Agency (DTRA) contract HDTRA1-17-1-0051 and supported by the Australian National Health and Medical Research Council (project grants 1046812, 1098337, and 1131932 [the HOT NORTH initiative]).

REFERENCES

- 1. Wiersinga WJ, Currie BJ, Peacock SJ. 2012. Melioidosis. N Engl J Med 367:1035–1044. https://doi.org/10.1056/NEJMra1204699.
- Currie BJ, Ward L, Cheng AC. 2010. The epidemiology and clinical spectrum of melioidosis: 540 cases from the 20 year Darwin prospective study. PLoS Negl Trop Dis 4:e900. https://doi.org/10.1371/journal.pntd .0000900.
- Price EP, Sarovich DS, Mayo M, Tuanyok A, Drees KP, Kaestli M, Beckstrom-Sternberg SM, Babic-Sternberg JS, Kidd TJ, Bell SC, Keim P, Pearson T, Currie BJ. 2013. Within-host evolution of *Burkholderia pseudomallei* over a twelve-year chronic carriage infection. mBio 4:e00388 -13. https://doi.org/10.1128/mBio.00388-13.
- Benson DA, Karsch-Mizrachi I, Clark K, Lipman DJ, Ostell J, Sayers EW. 2012. GenBank. Nucleic Acids Res 40:D48–D53. https://doi.org/10.1093/ nar/gkr1202.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. Genome Res 27:722–736. https://doi .org/10.1101/gr.215087.116.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. https://doi.org/10.1371/journal .pone.0112963.
- Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. Genome Biol 16:294. https://doi.org/10.1186/s13059-015-0849-0.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https:// doi.org/10.1093/nar/gkw569.
- Sahl JW, Caporaso JG, Rasko DA, Keim P. 2014. The large-scale blast score ratio (LS-BSR) pipeline: a method to rapidly compare genetic content

between bacterial genomes. PeerJ 2:e332. https://doi.org/10.7717/peerj .332.

- Tuanyok A, Auerbach RK, Brettin TS, Bruce DC, Munk AC, Detter JC, Pearson T, Hornstra H, Sermswan RW, Wuthiekanun V, Peacock SJ, Currie BJ, Keim P, Wagner DM. 2007. A horizontal gene transfer event defines two distinct groups within *Burkholderia pseudomallei* that have dissimilar geographic distributions. J Bacteriol 189:9044–9049. https://doi.org/10 .1128/JB.01264-07.
- Tuanyok A, Stone JK, Mayo M, Kaestli M, Gruendike J, Georgia S, Warrington S, Mullins T, Allender CJ, Wagner DM, Chantratita N, Peacock SJ, Currie BJ, Keim P. 2012. The genetic and molecular basis of O-antigenic diversity in *Burkholderia pseudomallei* lipopolysaccharide. PLoS Negl Trop Dis 6:e1453. https://doi.org/10.1371/journal.pntd.0001453.
- DeShazer D, Waag DM, Fritz DL, Woods DE. 2001. Identification of a Burkholderia mallei polysaccharide gene cluster by subtractive hybridization and demonstration that the encoded capsule is an essential virulence determinant. Microb Pathog 30:253–269. https://doi.org/10 .1006/mpat.2000.0430.
- Sarovich DS, Price EP, Webb JR, Ward LM, Voutsinos MY, Tuanyok A, Mayo M, Kaestli M, Currie BJ. 2014. Variable virulence factors in *Burk-holderia pseudomallei* (melioidosis) associated with human disease. PLoS One 9:e91682. https://doi.org/10.1371/journal.pone.0091682.
- Warawa J, Woods DE. 2005. Type III secretion system cluster 3 is required for maximal virulence of *Burkholderia pseudomallei* in a hamster infection model. FEMS Microbiol Lett 242:101–108. https://doi.org/10.1016/j .femsle.2004.10.045.
- Burtnick MN, Brett PJ, Harding SV, Ngugi SA, Ribot WJ, Chantratita N, Scorpio A, Milne TS, Dean RE, Fritz DL, Peacock SJ, Prior JL, Atkins TP, Deshazer D. 2011. The cluster 1 type VI secretion system is a major virulence determinant in *Burkholderia pseudomallei*. Infect Immun 79: 1512–1525. https://doi.org/10.1128/IAI.01218-10.