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Immunophenotypic profile of tumor buds in breast cancer

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Abstract

Background: Tumor buds are associated with lympho-vascular invasion and lymph node metastases leading to the assumption that they are involved in the early metastatic process. Hence, it would be important to know if tumor buds can be targeted with the most widely used targeted therapies in breast cancer (BC) and if changes in hormone and Her2 status occur. The aim of this study was to answer these questions by determining whether hormone receptor (HR) and Her2 status are expressed in the tumor buds of a large cohort of BCs.

Design: We constructed a tumor bud next-generation tissue microarray (ngTMA) consisting of n = 199 BCs of non-special type. Generally, two 1 mm punches were taken from the tumor bud areas in the periphery (PTB) and within the tumor center (ITB). HR and Her2 status was assessed using immunohistochemistry and fluorescence in situ hybridization, respectively. HR status was positive if 1% of tumor bud cells were positive. Her2 status was considered positive if bud cells showed strong complete membranous Her2 over-expression or *Her2* amplification.

Results: Most tumor buds were positive for estrogen (ER) (PTB: 86%; ITB: 88.3) and progesterone receptor (PgR) (PTB: 72%; ITB: 72.8%) and Her2 was positive in: PTB 11.5% and ITB 11%. A difference between the main tumor mass and tumor buds (PTB and ITB) was seen for PgR in 3.5% of cases (n = 7). No differences were seen for ER and Her2 between tumor buds and main tumor mass.

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Authors contribution

LC helped in constructing the budding TMA, the evaluation of tumor markers in tumor buds and drafted the first version of the manuscript and approved final version. BS helped writing the manuscript and gave intellectual input and approved the final manuscript, NC helped in acquiring clinical data and gave intellectual input and approved the final version, ZI helped in study design, TMA construction, gave intellectual input and approved the final manuscript, TC designed the study, helped in TMA construction, evaluated the tumor markers, helped writing the manuscript and approved the final manuscript.

Conflict of interest

None of the authors has a conflict of interest.

Conclusion: Most tumor buds (96.5%) share the same HR and Her2 expression profile of the main tumor mass, implying that tumor buds rely on the same pathways as the main tumor mass and might be equally responsive to targeted therapies.

Keywords

Tumor budding; Breast cancer; Hormone receptors; Her2

1. Introduction

Tumor buds are small cell clusters or single tumor cells, detaching from the main tumor mass. This phenomenon can be seen within the tumor (intra-tumoral budding), or at the tumor periphery (peripheral tumor budding) [22,27,19]. Tumor budding is best characterized in colon cancer [6,19,31] but it is increasingly recognized and described in other tumor types such as e.g. breast, pancreatic-, esophagus-, larynx- and other cancers [16–18,22–25,27]. In breast cancer (BC) and other tumor types, high numbers of tumor buds are associated with lympho-vascular invasion (LVI) and/or lymph node metastasis [16,18,22,27]. Additionally, high numbers of tumor buds are associated with shorter overall and cancer-specific survival in BC [8,18] and this has been described in other tumor types as well [6,15]. The association of vascular invasion and tumor budding led to the assumption that tumor buds are involved in the early metastatic process by undergoing epithelial-mesenchymal transition (EMT) [8,19]. It is well-known that tumor cells undergoing EMT are more invasive and prone to metastasize that can lead to worse overall survival in cancer patients [1,10,23].

Inhibiting tumor cells involved in the early metastatic process would be of great clinical value since metastatic disease remains the major cause of cancer deaths with around 30% of BC patients developing metastasis [4,20]. This indicates that a better understanding of the metastatic process is needed in order to develop novel targeted approaches for highly aggressive and invasive cancer to improve patient outcomes. If tumor buds are involved in the early metastatic process, then it would be advantageous to determine whether and how they can be uniquely targeted. Tumor buds are known to have characteristics of EMT and in breast cancer, estrogen and Her2 overexpression was shown to be involved in EMT [11,13]. However, little is known regarding the expression of markers commonly targeted in breast cancer such as the estrogen and progesterone receptors and Her2 status in tumor buds.

Hence, the aim of this study was to examine the immunophenotypic profile of tumor buds to determine if they are targetable with the most widely used targeted therapies in BC, such as anti-hormonal and anti-Her2 therapy. In the current study, we report the estrogen, progesterone and Her2 receptor status of a large number of tumor buds in BC of non-special type (NST) using a next-generation tissue microarray (ngTMA).

2. Material and methods

Patients:

We selected 199 NST BCs out of our previously described cohort of 356 therapy naïve, unilateral BCs diagnosed in female patients that underwent surgery between 2005 and

2011 at the Inselspital Bern, Switzerland [7]. T category was available for all BCs and N category was available for n = 181 (91%). Tumor grading, estrogen-, progesterone-, and Her2 receptor status (ER, PgR and Her2) and the molecular subtypes according to the St. Gallen 2013 criteria from the main tumor mass was available from our previous studies [7,22]. The median age at diagnosis was 64 years (range: 33–98 years) and clinical information regarding chemotherapy was available for 100 (50.2%) cases; for anti-hormone therapy for n = 102 (51.3%), for anti-her2 therapy in n = 95 (47.7%); and for radiation therapy for n = 140 (70.9%) cases. The clinic-pathological characteristics are shown in Table 1. The study was approved by the ethical commission of the University of Bern (Registration 200/2014).

Next-generation tissue microarray (ngTMA) of tumor buds: The ngTMA was constructed as previously described (3DHistech, Budapest, Hungary) [33]. In brief, pathologists reviewed breast cancer cases using H&E slides [7]. The decision, what block should be used for the peripheral and intra-tumoral bud ngTMA was made by one pathologist (CT). The H&E slides were then scanned and uploaded to the digital platform to perform annotation on the computer screen. Whenever feasible, two areas from peripheral and central tumor buds were punched. We successfully made two 1 mm punches in 199 and 193 of PTB and ITB cases, respectively.

Definition and assessment of tumor buds: We used our previous definition of tumor buds: One isolated tumor cell or a small tumor cell clusters of up to 5 tumor cells [22]. The slides of the tumor bud ngTMA were stained with ER, PgR and Her2 using the same antibodies and conditions as in our previous study [22]. Briefly, any nuclear ER and PgR staining, regardless of intensity, was considered as positive. The cases were then dichotomized into negative and positive cases according to the cut-off of 1% [9]. Her2 status was evaluated according to ASCO/CAP guidelines 2013 [30]. ER and PgR positive tumor cells were estimated in the tumor buds and a positive rate of 1% positive tumor cells was regarded as a positive hormone (HR) status. For HR status any intensity of nuclear staining was regarded as positive and the cases were dichotomized into negative and positive cases according to the cut-off of 1%. A strong, complete membranous staining for Her2, or a *Her2* amplification was considered Her2 positive. Difference in ER and PgR status was defined according to the dichotomized result obtained for the main tumor and the tumor buds or according to the defined positive expression of Her2 status.

Statistics:

We used the Pearson Chi-Square test to calculate significant correlations between categorical variables. A p-value of < 0.05 was considered statistically significant. Analyses were carried out using the IBM SPSS Statistics 22.0 (IBM, Armonk, NY, USA).

3. Results

3.1. Peripheral tumor buds (PTB)

Informative ER, PgR and Her2 results were available for 172 (86.9%), 168 (84.8%), and 156 (78.8%) cases. ER status was positive in 148 (86%) cases. Her2 status was positive in

18 (11.5%). PgR was positive in 121 (72%). No difference in ER or Her2 status was seen between main tumor mass and tumor buds. However there was a difference in receptor status between main tumor mass and buds for PgR in 6 (3.6%; 6/168) cases. All cases showed a positive PgR status in the main tumor mass but were negative in PTBs. Comparing the differences of PgR status with the molecular subtypes of the main tumor mass differences in 2 (2%) of luminal A, and 4 luminal B (Her2-negative) (10%) were observed. No differences in other molecular subtypes were seen.

3.2. Intra-tumoral buds (ITB)

Informative ER, PgR and Her2 results were available for 179 (92.7%), 169 (87.6%), and 173 (89.6%) cases. As for PTB, no difference in ER or Her2 status was seen between main tumor mass and tumor buds. ER status was positive in 158 (88.3%) cases. PgR status was positive in 123 (72.8%) cases, and Her2 status was positive in 19 (11%) cases. A difference in receptor status between main tumor mass and buds was seen again only for PgR in 2 (1.2%; 2/169) cases. The two cases were positive for PgR status in the main tumor mass but negative in ITB. Comparing the differences of PgR status with the molecular subtypes of the main tumor mass differences in 1 luminal B (Her2-negative) (2%) and in 1 luminal B (Her2-positive) (7%) was seen. No differences in other molecular subtypes were seen. The summary of cases that different PgR status is given in Table 2. Examples of positive HR and Her2 status and difference of PgR status are shown in Fig. 1.

3.3. Significant associations of HR status and PTB

ER positive PTB were significantly ($p < 0.05$) associated with the following characteristics of the main tumor: ER positivity, PgR positivity low proliferation ($< 20\%$), low tumor grade, and luminal molecular subtypes.

PgR positive PTB were significantly ($p < 0.05$) associated with the following characteristics of the main tumor: ER positivity, PgR positivity low proliferation ($< 20\%$), low tumor grade, vascular invasion (V1), and luminal molecular subtypes.

3.4. Significant associations of HR status and ITB

ER positive ITB were significantly ($p < 0.05$) associated with the following characteristics of the main tumor: ER positivity, PgR positivity, negative Her2 status, low proliferation ($< 20\%$), low tumor grade tumors, and luminal molecular subtypes.

PgR positive PTB were significantly ($p < 0.05$) associated with the following characteristics of the main tumor: ER positivity, PgR positivity, low proliferation ($< 20\%$), low tumor grade, vascular invasion (V1), and luminal molecular subtypes.

4. Discussion

Targeting BC tumor buds, a tumor cell population regarded as highly migratory and invasive [8] could be crucial in interfering with tumor progression. Tumor buds were shown to have an EMT phenotype e.g. with vimentin, or β -catenin expression and loss of e-cadherin [6,18]. In breast cancer (BC), it was reported that ER and Her2 play a role in EMT. In

experiments with BC cell lines, it was shown that ER disrupts tight-junctions [13] and Her2 overexpression in breast epithelial cell lines with stem cell properties leading to EMT by loss of epithelial markers [11,13]. According to this data, we hypothesized that hormone receptor (HR) and Her2 status might change in tumor buds, which may be important in targeting tumor buds with anti-hormonal or anti-Her2 therapies. However, hormone and Her2 status are highly conserved, and ITB and PTB shared in the majority of cases the HR and Her2 status of the main tumor mass. These results indicate that BC tumor buds seem to rely on the same pathways as the cohesive tumor cells. This assumption, that main tumor mass and tumor buds can share the same profiles, is supported by a recent observation made in colon cancer, which revealed that the driver mutations in the main tumor mass and in tumor buds remained unchanged [3]. According to these results, we argue that tumor buds are equally responsive to anti-hormonal and anti-Her2 therapy as the main tumor mass.

We observed only different results for PgR status, whereas ER and Her2 showed consistent expression between tumor buds and the main tumor mass. Differences in PgR receptor could have different reasons. HR status can be heterogeneous within the tumor and all BCs with different results were heterogeneous for PgR expression and positivity might not have been captured in the tumor bud ngTMA. Although working with TMAs has been regarded as a weakness, TMAs are an excellent high throughput screening tool for protein expression [2,26], and ngTMAs have been shown to be suitable for characterizing tumor buds [33]. However, an underlying biological background of PgR status change cannot be excluded. In vivo experiments in breast cancer cell lines showed that progesterone can lead to inhibition of EMT relevant proteins such as e.g. e-cadherin due to binding with progesterone receptor alpha [34] and in endometrial cancer, it was suggested that the Wnt/beta-catenin pathway is inhibited by progesterone in the presence of progesterone receptor [29]. Hence, loss of PgR would favor EMT and EMT is reported to be a characteristic of tumor buds [18,19]. Loss of PgR expression was reported to be an independent factor for low survival rates in breast cancer [21]. A large cohort with long-term followup and survival data is needed to determine if loss of PgR expression in tumor buds represents a change in tumor biology towards a more aggressive tumor phenotype.

We did not identify any difference, in particular, no change to positive hormone or Her2 status in tumor buds in triple negative BCs (TNBC). ER, PgR and Her2, up-or down regulation, might facilitate EMT and tumor budding but other pathways must play a role. TNBC can be positive for many EMT markers such as e.g. vimentin [5,12] and vimentin was shown to be expressed in tumor buds of BCs [18]. Hence, to elucidate underlying mechanisms of tumor budding in BC and BC subtypes, other pathways and markers need to be considered besides HR and Her2. Other factors such as processes within the tumor microenvironment should be included in this consideration. Several studies showed that the tumor stroma might play an important role in tumor budding formation and inflammatory cells can be associated with tumor buds [8,14,17,28,32], indicating that the complex phenomenon of tumor budding may involve all tumor compartments. At this point, we would argue that tumor buds are most efficiently targeted through their immuno profile. However, in the future, stroma and/or immune cell modulators may prove to be equally efficient.

In conclusion, for the first time, we identified that tumor buds generally reflect the targetable immuno profile of the main tumor mass. Therefore, we assume that tumor buds are equally responsive to anti-hormonal and anti-Her2 therapies. Different PgR status can occur in tumor buds but its significance is unclear and needs further investigation.

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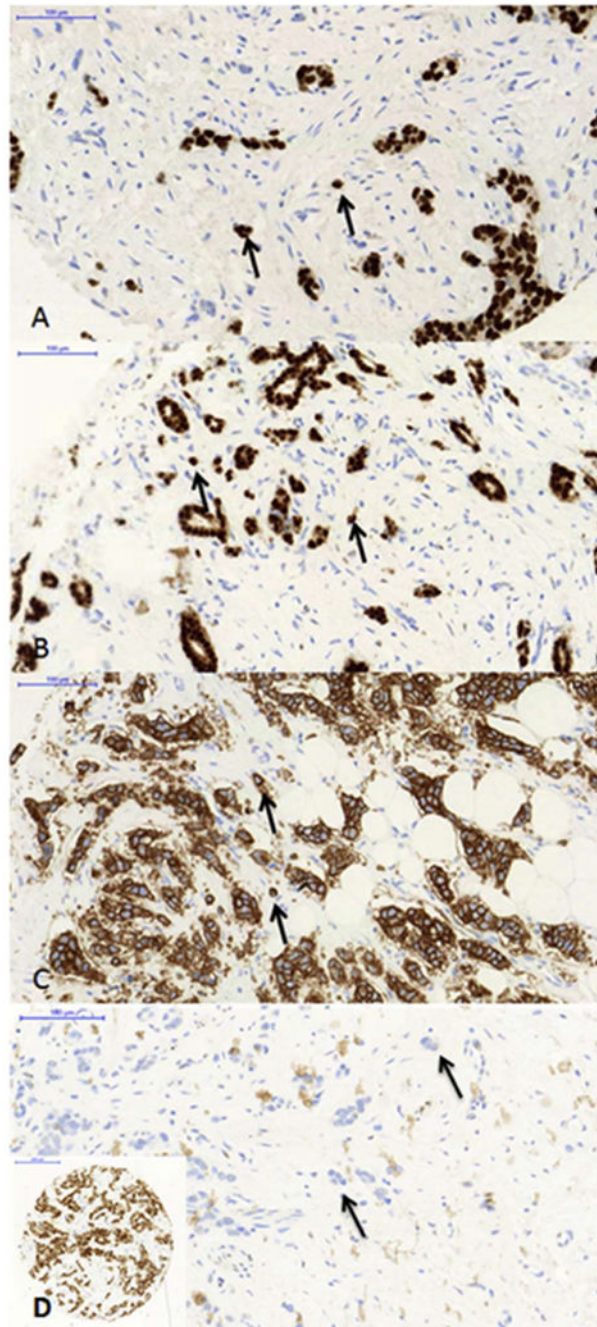


Fig. 1.
Examples of hormone and Her2 expression in tumor buds and BC with difference in PgR status.

A: ER positive breast cancer and positive tumor buds.

B: PgR positive breast cancer and positive tumor buds.

C: Her2 positive breast cancer with strong complete membrane her2 positivity in tumor buds.

D: Breast cancer (ID10) with a positive PgR status of the main tumor mass (inlet) and negative PgR status in PTB.

A-D: Black arrows are pointing towards tumor buds.

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Table 1

Patient characteristics of the whole cohort (n = 199).

Feature		Frequency n (%)
Age at diagnosis (years)	Median (min, max)	64 (33, 98)
Tumor size (centimeter)	Mean (min, max)	2.4 (0.6, 8.4)
ER	positive	165 (82.9)
	negative	34 (17.1)
PgR	positive	142 (71.4)
	negative	57 (28.6)
Her2	negative	177 (88.9)
	positive	21 (10.6)
	No data	1 (0.5)
Nottingham Grad	G1	21 (10.6)
	G2	97 (48.7)
	G3	81 (40.7)
LVI	Yes	82 (41.2)
	No	104 (52.3)
	No data	13 (6.5)
VI	Yes	21 (10.6)
	No	161 (80.9)
	No data	17 (8.5)
Pn1	Yes	26 (13.1)
	No	148 (74.3)
	No data	25 (12.6)
pT	T1	92 (46.2)
	T2	93 (46.8)
	T3	7 (3.5)
	T4	7 (3.5)
pN	N0	84 (42.3)
	N1mi	11 (5.5)
	N1	60 (30.2)
	N2	17 (8.5)
	N3	9 (4.5)
	No data	18 (9.0)
Recurrence	Yes	12 (6.0)
	No data	187 (94)
Molecular subtypes (St. Gallen 2013)	Luminal A	106 (53.3)
	Luminal B (her2 negative)	44 (22.1)
	Luminal B (her2 positive)	14 (7.0)
	Her2 non-luminal	7 (3.5)
	Triple negative	26 (13.1)
	no data	2 (1.0)

Feature		Frequency n (%)
Anti-hormonal therapy	Yes	42 (21.1)
	No	60 (30.2)
	No data	97 (48.7)
Chemotherapy	Yes	37 (18.6)
	No	63 (31.7)
	No data	99 (49.7)
Anti-her2 therapy	Yes	5 (2.5)
	No	90 (45.2)
	No data	104 (52.3)
Radiation therapy	Yes	100 (50.3)
	No	40 (20.1)
	no data	59 (29.6)

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Table 2

Breast cancers with differences of PgR expression.

ID	Main tumor mass	PTB	ITB	Molecular subtype
10	PgR positive	No PgR expression	PgR positive	luminal A
21	PgR positive	No PgR expression	No PgR expression	luminal B (her2-)
43	PgR positive	No PgR expression	PgR positive	luminal B (her2-)
54	PgR positive	No PgR expression	No data available	luminal B (her2-)
248	PgR positive	No PgR expression	PgR positive	luminal A
181	PgR positive	No PgR expression	PgR positive	Luminal B (her2-)
142	PgR positive	PgR positive	No PgR expression	luminal B (her2+)

Color code: red=positive, green=no expression. (For interpretation of the references to colour in this Table note, the reader is referred to the web version of this article.)