





Draft Genome Sequence of *Mariprofundus micogutta* Strain ET2

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ABSTRACT *Mariprofundus micogutta* strain ET2 was isolated in 2014 from a deepsea hydrothermal field on the Bayonnaise Knoll of the Izu-Ogasawara arc. Here, we report its draft genome, which comprises 2,497,805 bp and contains 2,417 predicted coding sequences.

ariprofundus micogutta ET2 (=KCTC15556^T=JCM30585^T) is a microaerophilic and chemolithoautotrophic mesophile that grows by oxidizing ferrous iron. The stalkforming type strain *M. ferrooxydans* PV-1 and the non-stalk-forming strain *M. ferrooxydans* JV-1 were the first reported isolates belonging to the genus Mariprofundus (1). Since that initial isolation, strains belonging to the class "Candidatus (Ca.) Zetaproteobacteria" have been discovered throughout the Pacific and Atlantic oceans, mainly from deep-sea hydrothermal fields (2). Recently, *M. ferrooxydans* strains were successfully isolated from a salt marsh of the Great Salt Bay, Newcastle, Maine (3), and the Spillway Area of the Loihi Seamount (4). Here, we report the genetic features of *M. micogutta* strain ET2, which were determined using draft genome sequencing.

Strain ET2 was isolated from sediment of a 772-m-deep hydrothermal field on the Bayonnaise Knoll, Izu-Ogasawara Arc. A pure culture was obtained by a dilution-toextinction series with artificial seawater (ASW) medium, as previously described (5). Strain ET2 was grown in ASW medium at 25°C with filament formation. Genomic DNA was extracted from cell pellets using a PowerMax Soil DNA isolation kit (Mo Bio, Carlsbad, CA, USA). A sequencing library was prepared with a Nextera DNA prep kit (Illumina, Inc., San Diego, CA, USA) and subsequently sequenced using an Illumina MiSeq version 3 reagent kit (600 cycles) with 300-bp paired-end reads on the Illumina MiSeq platform. A total of 17.6 million reads were produced, yielding 7.5 Gb of data. Read quality was assessed with FastQC (https://www.bioinformatics.babraham.ac.uk/ projects/fastqc). Quality-controlled reads were used to assemble the draft genome with SPAdes version 3.5.0 (6), resulting in 59 contigs with an N_{50} value of 105,204 bp (the largest contig was 249,769 bp); the assembled draft genome was 2,497,805 bp in size with a G+C content of 48.75%. Annotation was carried out using Prokka version 1.11 (7), with 2,417 coding sequences (CDSs), 6 rRNAs, and 49 tRNAs being identified. Average nucleotide identity (ANI) comparisons with other M. ferrooxydans genomes, including strains PV-1 and M34, revealed ANIs of 95.89% and 95.8%, respectively. This confirms the placement of M. micogutta strain ET2 in the genus Mariprofundus of the class "Ca. Zetaproteobacteria." Similar to strains PV-1 and M34, strain ET2 encodes genes for growth by chemoautotrophy and motility via a flagellum, although motility was not observed in laboratory experiments and observations (5). A complete set of CDSs for carrying out carbon fixation using the Calvin-Benson-Bassham cycle was identified. Unlike other members of the class "Ca. Zetaproteobacteria", M. micogutta strain ET2 has both aa3- and cbb3-type oxidase genes. The genome includes eight

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chemotaxis (*che*) genes and four methyl-accepting chemotaxis proteins, which were previously shown to play important roles in adaptation to deep-sea vent environments (8). Moreover, since the cobalt-zinc-cadmium resistance protein (*czc*) is present, it is considered to exhibit heavy-metal tolerance (9). The outer membrane cytochrome Cyc2 (10), a possible key gene for catalyzing iron oxidation, was detected. Genes for the putative outer membrane Fe oxidase MtoA (11, 12) were not found in the ET2 genome, like other *Mariprofundus* species.

Accession number(s). The whole-genome shotgun project reported here was deposited at DDBJ/EMBL/GenBank under the accession number BDFD00000000, and the 59 contigs generated were deposited under the accession numbers BDFD01000001 to BDFD01000059.

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REFERENCES

- Emerson D, Rentz JA, Lilburn TG, Davis RE, Aldrich H, Chan C, Moyer CL. 2007. A novel lineage of proteobacteria involved in formation of marine Fe-oxidizing microbial mat communities. PLoS One 2:e667. https://doi.org/10.1371/journal.pone.0000667.
- Scott JJ, Breier JA, Luther GW, III, Emerson D. 2015. Microbial iron mats at the Mid-Atlantic Ridge and evidence that zetaproteobacteria may be restricted to iron-oxidizing marine systems. PLoS One 10:e0119284. https://doi.org/10.1371/journal.pone.0119284.
- McBeth JM, Little BJ, Ray RI, Farrar KM, Emerson D. 2011. Neutrophilic iron-oxidizing "Zetaproteobacteria" and mild steel corrosion in nearshore marine environments. Appl Environ Microbiol 77:1405–1412. https://doi.org/10.1128/AEM.02095-10.
- McAllister SM, Davis RE, McBeth JM, Tebo BM, Emerson D, Moyer CL. 2011. Biodiversity and emerging biogeography of the neutrophilic ironoxidizing *Zetaproteobacteria*. Appl Environ Microbiol 77:5445–5457. https://doi.org/10.1128/AEM.00533-11.
- Makita H, Tanaka E, Mitsunobu S, Miyazaki M, Nunoura T, Uematsu K, Takaki Y, Nishi S, Shimamura S, Takai K. 2017. Mariprofundus micogutta sp. nov., a novel iron-oxidizing zetaproteobacterium isolated from a deep-sea hydrothermal field at the Bayonnaise Knoll of the Izu-Ogasawara arc, and a description of Mariprofundales ord. nov. and Zetaproteobacteria classis nov. Arch Microbiol 199:335–346. https://doi.org/10.1007/s00203 -016-1307-4
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new

- genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. https://doi.org/10.1093/bioinformatics/btu153.
- Nakagawa S, Takaki Y, Shimamura S, Reysenbach A, Takai K, Horikoshi K. 2007. Deep-sea vent ε-proteobacterial genomes provide insights into emergence of pathogens. Proc Natl Acad Sci U S A 104:12146–12150. https://doi.org/10.1073/pnas.0700687104.
- Goldberg M, Pribyl T, Juhnke S, Nies DH. 1999. Energetics and topology of CzcA, a cation/proton antiporter of the resistance-nodulation-cell division protein family. J Biol Chem 274:26065–26070. https://doi.org/ 10.1074/jbc.274.37.26065.
- Barco RA, Emerson D, Sylvan JB, Orcutt BN, Jacobson Meyers M, Ramírez GA, Zhong JD, Edwards KJ. 2015. New insight into microbial iron oxidation as revealed by the proteomic profile of an obligate iron-oxidizing chemolithoautotroph. Appl Environ Microbiol 81:5927–5937. https://doi .org/10.1128/AEM.01374-15.
- Liu J, Wang Z, Belchik SM, Edwards MJ, Liu C, Kennedy DW, Merkley ED, Lipton MS, Butt JN, Richardson DJ, Zachara JM, Fredrickson JK, Rosso KM, Shi L. 2012. Identification and characterization of MtoA: a decaheme c-type cytochrome of the neutrophilic Fe(II)-oxidizing bacterium Sideroxydans lithotrophicus ES-1. Front Microbiol 3:37. https://doi.org/10.3389/fmicb .2012.00037.
- Chiu BK, Kato S, McAllister SM, Field EK, Chan CS. 2017. Novel pelagic iron-oxidizing zetaproteobacteria from the Chesapeake Bay oxic-anoxic transition zone. Front Microbiol 8:1280. https://doi.org/10.3389/fmicb .2017.01280.

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