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Original Research Article

# Oral administration of dibutyryl adenosine cyclophosphate improved growth performance in weaning piglets by enhancing lipid fatty acids metabolism



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### ABSTRACT

Dibutyryl adenosine cyclophosphate (dbcAMP-Ca), an analog of cyclic adenosine monophosphate (cAMP), plays greater roles in regulating physiological activities and energy metabolism than cAMP. The aim of this study was to investigate the effect of oral administration of dbcAMP-Ca on growth performance and fatty acids metabolism in weaning piglets. A total of 14 early weaning piglets (7  $\pm$  1 d of age,  $3.31 \pm 0.09$  kg, Landrace  $\times$  Large White  $\times$  Duroc) were randomly divided into 2 groups: control group and dbcAMP-Ca group, and the piglets received 7 mL of 0.9% NaCl or 1.5 mg dbcAMP-Ca dissolved in 7 mL of 0.9% NaCl per day for 10 d, respectively. The results showed that the average daily gain (ADG) increased by 109.17% (P < 0.05) in the dbcAMP-Ca group compared with the control group. Besides, dbcAMP-Ca significantly decreased blood high density lipoprotein cholesterol (HDLC) concentration (P < 0.05) and significantly increased blood low density lipoprotein cholesterol (LDLC) concentration (P < 0.05) compared with the control group. Further, liver C18:2n6t content significantly increased in dbcAMP-Ca group (P < 0.05) compared with the control group. With the increase of C18:2n6t content, the mRNA expression levels of peroxisome proliferator-activated receptor  $\alpha$  (PPAR $\alpha$ ) and hormone sensitive glycerol three lipase (HSL), of which genes are related to lipid metabolism, were also significantly increased (P < 0.05 or P < 0.01). All of the results indicated that dbcAMP-Ca improved the ADG, which was probably done by regulating fatty acids metabolism in the liver of weaning piglets.

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# 1. Introduction

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Neonatal piglets face heavy challenge to adapt to the shift between intrauterine and extrauterine environments because of weak gut absorptive capacity, low immunity and adaptability, etc (Tanghe et al., 2014; Wang et al., 2017). Normally, weaning usually occurred in an early period at around 21 d of age. However, under the integrated production, weaning time of piglets gets earlier and earlier. Weaning in piglets may lead to worse situation and result in weaning stress in piglets, thus may affect their health and welfare with a decline in feed intake and be vulnerable to disease (Duan et al., 2015).

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Milk lipids are the main sources of energy for sucking piglets. An earlier study has found that respiratory entropy of newborn piglets was reduced after birth which indicated that piglets used large amounts of fatty acids to provide energy (Hales, 1997). A former study also showed that lipids in the milk provided nearly 50% of the energy for suckling piglets (Hobbins, 1997). However, it has been reported that the activity of pancreatic lipase increased with age but weaning made it sharply decline (Aumaitre and Corring, 1978; Cera et al., 1990). Therefore, fatty acid, 1 of the 3 major nutrients, plays significant roles in growth, metabolism and physiological functions in newborn mammals because of their considerable energy needs and defective dietary capacity (Gruppuso et al., 1994; Hardy and Kleinman, 1994; Goodyer et al., 2001). Herein, the decomposition and utilization of fatty acids are of great significance to newborn piglets.

Interestingly, cyclic adenosine monophosphate (cAMP), which has been shown to mediate the hormonal regulation of lipid metabolism (Butcher et al., 1968; Gagelin et al., 1999), is vital in regulating and utilizing fatty acids (Luiken et al., 2002; Madsen et al., 2008). Moreover, dibutyryl adenosine cyclophosphate calcium (dbcAMP-Ca, Fig. 1), an analog of cAMP, can regulate the lipid metabolism remarkably in growing and finishing pigs (Gao et al., 2004), and affect the differentiation of sheep inguinal preadipocytes (Kong et al., 2017). However, little is known about how dbcAMP affects the metabolism of fatty acids and the growth performance in weaning piglets. Therefore, the present study was intended to seek the effect of oral administration of dbcAMP-Ca on growth performance and lipid fatty acids metabolism in weaning piglets.

# 2. Materials and methods

### 2.1. Animals and treatments

The animal experiment was approved by the Protocol Management and Review Committee of the institute of Subtropical Agriculture, Chinese Academy of Science. Pigs were cared for and slaughtered according to the guidelines of the institute of Subtropical Agriculture on Animal Care, Changsha, China. Dibutyryl adenosine cyclophosphate (dbcAMP-Ca) ( $C_{18}H_{23}N_5O_8PCa$ ,



Fig. 1. The structural formula of dibutyryl adenosine cyclophosphate (dbcAMP-Ca).

molecular weight 507.00 g/mol, purity 98.00%) was provided by Meiya Haian pharmaceutical Co., Ltd (Haian 226600, China).

Fourteen 7-day-old weaning piglets (Landrace × Large White × Duroc) with mean body weight at  $3.31 \pm 0.09$  kg were randomly divided into 2 groups: control group and dbcAMP-Ca group. The piglets received 7 mL of 0.9% NaCl or 1.5 mg dbcAMP-Ca dissolved in 7 mL of 0.9% NaCl by oral administration at indicated times per day for 10 days, respectively. Ingredients and nutrient levels of the basal milk are shown in Table 1. All the piglets were fed by artificial breast feeder and had *ad libitum* access to water and the basal milk.

#### 2.2. Samples collection

Before slaughter, 5 mL blood samples were collected from the jugular vein, and plasma samples were obtained by centrifugation at 3,000  $\times$  g for 10 min at 4 °C, followed by being immediately stored at -80 °C for later lipid profiles analysis (Wu et al., 2016). Liver samples were taken from each animal, followed by being flash frozen in liquid nitrogen and stored at -80 °C prior to RNA isolation and at -20 °C for fatty acid analysis, respectively.

# 2.3. Fatty acids analysis in liver of piglets

The extraction of fatty acids from 500 mg of the liver tissue and the methylation were performed. The concentration of individual fatty acids was quantified according the peak area and expressed as a percentage of total fatty acids by gas chromatography (GC-2010, Shimadzu Corp, Japan) as previously described (Tan et al., 2009; Raj et al., 2010).

#### 2.4. RNA extraction and cDNA synthesis

About 100 mg of the liver tissue was pulverized in liquid nitrogen. Total RNA was isolated from the homogenate using TRIzol reagent (Invitrogen, CA, USA). The concentration of total RNA was quantified spectrophotometrically (NanoDrop ND-1000; Thermo Fisher Scientific, DE, USA) at 260 nm, and the ratio of 260 nm to 280 nm was used to assess RNA quality, then cDNA synthesis was carried out using a PrimeScript RT reagent Kit With gDNA Eraser (TaKaRa, Dalian, China). Primers (Table 2) were designed by Primer 5.0 based on GenBank (http://www.ncbi.nlm.nih.gov/pubmed/), and Oligo Synthesis was conducted by Sangon Biotech (Shanghai, China).  $\beta$ -actin was chosen as a reference gene.

Table 1
Ingredients (%) and nutrient levels (%) of the basal milk (air-dry basis).

Ingredients	Content	Nutrient levels	Content
Skimmed milk powder	85.0	DE, MJ/kg	14.65
Dried whey	5.0	CP	20.50
Glucose	2.5	Ca	0.70
Plasma proteins	3.5	Total P	0.60
Premix <sup>1</sup>	4.0	Lys	1.45
Total	100.0	Met	0.48
		Try	0.29

<sup>1</sup> The premix provided the following for per kg of the basal milk: vitamin  $A_1$  500 IU, vitamin  $D_3$  200 IU, vitamin E 85 IU, D-pantothenic acid 35 mg, vitamin  $B_2$  12 mg, folic acid 1.5 mg, nicotinic acid 35 mg, vitamin B 13.5 mg, vitamin  $B_6$  2.5 mg, biotin 0.2 mg, vitamin  $B_{12}$  0.05 mg, copper (as copper sulfate) 15 mg, ferrum (as ferrous sulfate) 100 mg, manganese (as manganese sulfate) 20 mg, iodate (as calcium iodate) 1.0 mg, selenium (as sodium selenite) 0.35 mg, cobalt (as cobalt sulfate) 0.2 mg, and chromium (as chromium picolinate) 0.2 mg.

# Table 2 Primer design parameters.

	•		
Genes	Primers	Sequence (5' to 3')	Size, bp
β-actin	Forward	TGCGGGACATCAAGGAGAAG	216
	Reverse	AGTTGAAGGTGGTCTCGTGG	
FAS	Forward	GTCCTGCTGAAGCCTAACTC	206
	Reverse	TCCTTGGAACCGTCTGTG	
PPARα	Forward	GCTATCATTTGGTGCGGAGAC	139
	Reverse	GGAGTTTGGGGAAGAGAAAGAC	
CPT-1α	Forward	CCATCAAAACTGCCTTCCTTAG	118
	Reverse	AGCGAGTGTGCCAGATACAAA	
CPT-1β	Forward	TTCCGCCAAACCTTGAAACT	100
	Reverse	GGACACAGATAGCCCAGACTTT	
ΑССα	Forward	TCCCAGTGCAAGCAGTATG	211
	Reverse	TGCCAATCCACACGAAGAC	
HSL	Forward	GAAGGGAGAGCTATGGCACC	130
	Reverse	CTCACACTCTCCAAGCCCAG	

FAS = fatty acid syntheses;  $PPAR\alpha =$  peroxisome proliferator-activated receptor  $\alpha$ ;  $CPT-1\alpha =$  carnitine palmitoyl transferase  $1\alpha$ ;  $CPT-1\beta =$  carnitine palmitoyl transferase  $1\beta$ ;  $ACC\alpha =$  acetyl-coA carboxylase  $\alpha$ ; HSL = hormone sensitive glycerol three lipase.

#### 2.5. Statistical analysis

The data are presented as means  $\pm$  SEM. All statistical analyses were performed using SPSS 17.0 software (SPSS Inc., Chicago, IL). The differences among groups were evaluated using independent *t* test. *P* < 0.05 and *P* < 0.01 were considered statistically significant and extremely significant, respectively.

# 3. Results

# 3.1. Growth performance

As shown in Fig. 2, compared with the control group, dbcAMP-Ca increased the final weight gain by 12.06% (P > 0.05) and average daily gain (ADG) by 109.17% (P < 0.05) in early weaning piglets.

# 3.2. Blood lipid profiles

As shown in Table 3, low density lipoprotein cholesterol (LDLC) concentration increased while high density lipoprotein cholesterol (HDLC) concentration decreased in dbcAMP-Ca treated piglets compared with the control group (P < 0.05). Triglyceride (TG) and total cholesterol (TC) concentrations in dbcAMP-Ca group decreased by 3.64% and 11.11%, respectively (P > 0.05) compared to the control group.

#### 3.3. Liver fatty acids composition

As shown in Table 4, compared with the control group, the liver content of C12:0 decreased by 57.89% (P < 0.05), and those of

able 3			
lood lipid	profiles (mmol/L) of piglets	(n =	7).

Item	Groups <sup>1</sup>		P-value
	Control	dbcAMP-Ca	
TC	$2.47 \pm 0.15$	$2.38 \pm 0.12$	0.65
TG	$0.36 \pm 0.03$	$0.32 \pm 0.04$	0.56
HDLC	$1.21 \pm 0.07^{*}$	$1.03 \pm 0.04$	0.04
LDLC	$0.80 \pm 0.07$	$0.93 \pm 0.07^{*}$	0.03
T-bil	$20.07 \pm 2.06$	$21.39 \pm 4.76$	0.80
D-bil	$2.54 \pm 0.50$	$1.97 \pm 0.88$	0.51
TBA	$24.17 \pm 6.57$	$14.07 \pm 2.69$	0.18

dbcAMP-Ca = dibutyryl adenosine cyclophosphate; TC = total cholesterol; TG = triglyceride; HDLC = high density lipoprotein cholesterol; LDLC = low density lipoprotein cholesterol; T-bil = total bilirubin; D-bil = bilirubin direct; TBA = total bile acid.

 $^*$  Indicates a significant difference (P < 0.05) between the control group and dbcAMP-Ca group.

<sup>1</sup> Control group: a basal diet; dbcAMP-Ca group: the basal diet supplemented with 1.5 mg/d of dbcAMP-Ca.

monounsaturated fatty acid (MUFA) C18:1n9t and C18:1n9c decreased by 12.50% and 5.14% (P > 0.05) in dbcAMP-Ca group, respectively. Meanwhile, the liver content of C18:3n3 decreased by 29.41% in dbcAMP-Ca group (P > 0.05), and that of C18:2n6t increased by 30.00% in dbcAMP-Ca group (P < 0.05) compared with the control group.

#### 3.4. mRNA expression levels of lipid metabolism related genes

To further confirm the role of lipid metabolism in the liver, the mRNA expression levels of the lipid metabolism related genes, fatty acid syntheses (*FAS*), hormone sensitive glycerol three lipase (*HSL*), acetyl-coA carboxylase  $\alpha$  (*ACC* $\alpha$ ), carnitine palmitoyl transferase 1 $\alpha$  (*CPT*-1 $\alpha$ ), carnitine palmitoyl transferase 1 $\beta$  (*CPT*-1 $\beta$ ), peroxisome proliferator-activated receptor  $\alpha$  (*PPAR* $\alpha$ ), were detected by qRT-PCR. As shown in Fig. 3, compared with the control group, dbcAMP-Ca significantly increased the mRNA expression level of *PPAR* $\alpha$  (*P* < 0.05) and extremely significantly increased that of *HSL* (*P* < 0.01) in the liver of piglets. However, there were no differences between the control group and dbcAMP-Ca group for the mRNA expression levels of *FAS*, *ACC* $\alpha$ , *CPT*-1 $\alpha$  and *CPT*-1 $\beta$ .

# 4. Discussion



**Fig. 2.** Effect of dibutyryl adenosine cyclophosphate (dbcAMP-Ca) on the final body weight (A) and average daily gain (B) of weaning piglets (n = 7). \* indicates a significant difference (P < 0.05) between the control group and dbcAMP-Ca group.

Dibutyryl adenosine cyclophosphate (dbcAMP-Ca), as an analog of cAMP, exerts effects via stimulating cAMP signaling pathway (Arnold et al., 2010), such as cell proliferation and differentiation, hormones release and regulation (Boghaert et al., 1991; Chrenek et al., 2010, 2013). And former studies have also found that

**Table 4** Long chain fatty acid content (%) in liver (n = 7).

Item	Groups <sup>1</sup>		P-value
	Control	dbcAMP-Ca	
C10:0	$0.02 \pm 0.003$	$0.02 \pm 0.003$	0.69
C12:0	$0.19 \pm 0.03$	$0.08 \pm 0.03^{*}$	0.03
C14:0	$0.78 \pm 0.07$	$0.55 \pm 0.27$	0.09
C15:0	$0.09 \pm 0.01$	$0.07 \pm 0.01$	0.34
C16:0	$15.34 \pm 0.44$	$15.73 \pm 0.56$	0.59
C17:0	0.39 ± 0.03	$0.41 \pm 0.03$	0.75
C18:0	$21.24 \pm 0.61$	$22.21 \pm 1.08$	0.47
C20:0	$0.04\pm0.00$	$0.04 \pm 0.00$	0.61
C16:1	0.91 ± 0.21	$0.93 \pm 0.37$	0.94
C17:1	$0.04 \pm 0.01$	$0.03 \pm 0.00$	0.23
C18:1n9t	$0.08 \pm 0.02$	$0.07 \pm 0.04$	0.79
C18:1n9c	17.73 ± 0.98	$16.77 \pm 0.75$	0.45
C18:2n6t	$0.01 \pm 0.003$	$0.013 \pm 0.008^{*}$	0.04
C18:2n6c	15.95 ± 0.37	$16.07 \pm 0.67$	0.88
C18:3n6	$0.16 \pm 0.02$	$0.17 \pm 0.02$	0.76
C18:3n3	$0.17 \pm 0.02$	$0.12 \pm 0.05$	0.14
C20:1	0.31 ± 0.03	$0.28 \pm 0.03$	0.47
C20:3n6	$2.45 \pm 0.20$	$2.53 \pm 0.32$	0.83
C20:4n6	17.67 ± 0.37	$17.36 \pm 0.82$	0.76
C22:6n6	$6.40 \pm 0.30$	$6.49 \pm 0.23$	0.82
SFA	$39.12 \pm 0.6$	$40.00 \pm 0.70$	0.28
MUFA	$17.82 \pm 0.98$	$16.85 \pm 0.76$	0.44
PUFA	$80.52 \pm 0.86$	77.48 ± 1.11	0.08

 $^*$  Indicates a significant difference (P < 0.05) between the control group and dbcAMP-Ca group.

<sup>1</sup> Control group: a basal diet; dbcAMP-Ca group: the basal diet supplemented with 1.5 mg/d of dbcAMP-Ca.



**Fig. 3.** Effect of dibutyryl adenosine cyclophosphate (dbcAMP-Ca) on the mRNA expression levels of lipid metabolism related genes in liver (n = 7). *PPAR* $\alpha$  = peroxisome proliferator-activated receptor  $\alpha$ ; *FAS* = fatty acid syntheses; *CPT*-1 $\alpha$  = carnitine palmitoyl transferase 1 $\alpha$ ; *CPT*-1 $\beta$  = carnitine palmitoyl transferase 1 $\beta$ ; *ACC* $\alpha$  = acetyl-coA carboxylase  $\alpha$ ; *HSL* = hormone sensitive glycerol three lipase. \* and \*\* indicate significant difference (P < 0.05) and extreamly significant difference (P < 0.05) and extreamly significant difference (P < 0.01) between the control group and dbcAMP-Ca group, respectively.

growth hormone could be better stimulated through Ca  $^{2+}$  and cAMP-dependent interactive mechanism thus enhance the production performance of animals (Sartin et al., 1996; Pahan et al., 1998). In this study, the supplement of dbcAMP-Ca significantly increased the ADG and promoted the growth of early weaning piglets, which might be caused by the interactive effect of Ca<sup>2+</sup> and dbcAMP.

Lipid, as a kind of necessary substance for animals, plays a vital role in maintaining cell structure and function (Smith et al., 2003; Liu et al., 2017). Blood lipid concentrations were regarded as the status of dynamic lipid absorption and nutritional in animals and humans (Li et al., 2016). Notably, increasing levels of blood constituents are associated with the increasing of dietary nutrients (Brungardt, 1963; Sink et al., 1973). In this regard, the current result of the elevated blood concentrations of LDLC and HDLC was influenced by dbcAMP-Ca treatment when compared with the control group. In our results, LDLC increased while HDLC decreased, which seemingly shows lipid metabolic disturbance and it might confer the risk for cardiovascular disease according to the former studies (Gupta and Rajagopal, 2007; Shin, 2009). However, for the fast growing piglets, high concentrations of LDLC and low HDLC may represent a high level of nutrition, which is in line with the significant change in the ADG. Furthermore, fatty acids composition in the liver also reflects the changes in fatty acid metabolism. The contents of polyunsaturated fatty acid (PUFA) and MUFA decreased whereas the content of saturated fatty acid (SFA) increased in dbcAMP-Ca group when compared with the control group, and this could be explained by the high LDLC and low HDLC in blood. According to the former studies, the proportion of PUFA is affected by many factors, such as synthesis rate, and mutual conversion (Enser et al., 2000). Besides, PUFA can be oxidized to supply energy to the organism (Tebbey et al., 1994; Clarke, 2000). Our study indicated that the supplement of dbcAMP-Ca could promote the oxidation of PUFA and might provide energy for piglets to meet the requirements of growth and development. For the further researches on lipid metabolism in the liver, we detected the mRNA expression levels of lipid metabolism related genes. Hormone sensitive glycerol three lipase is a rate-limited enzyme in triglyceride decomposition which cleaves fatty acids from triglycerides and diglycerides (Watt, 2013). Peroxisome proliferator-activated receptor  $\alpha$  (*PPAR* $\alpha$ ) has been regarded as the major regulators of lipid metabolism (Aiuwon et al., 2003). Triglyceride (TG) is synthesized via FAS catalyzing acetyl coenzyme A and malonate coenzyme A. Moreover, it is the major form required for body fat deposition (Semenkovich, 1997; Yan et al., 2002). In this experiment, dbcAMP-Ca increased the mRNA expression levels of *PPAR* $\alpha$  and *HSL* in the liver, which indicated that the addition of dbcAMP-Ca mainly promoted lipolysis and inhibited lipid deposition in the liver, thereby promoted the usage of milk lipids, and thus provided more energy for suckling piglets (Luiken et al., 2002; Madsen et al., 2008; Jia et al., 2018).

Taken together, oral administration of dbcAMP-Ca can significantly increase the weight gain of piglets and affect blood HDLC and LDLC concentrations, decrease the content of PUFA and enhance metabolism of fatty acids in the liver, which may be through the decomposition of lipids by *PPAR* $\alpha$  and *HSL* in the liver to provide more energy to ensure the healthy growth of piglets.

# 5. Conclusions

Conclusively, the present study suggests that dbcAMP-Ca has a significant effect on the growth performance mainly by its regulation effects on lipid metabolism. In the future, more well-designed researches will be needed to investigate the effects of dbcAMP-Ca on early weaning piglets.

#### **Conflicts of interest**

We declare that we have no financial and personal relationships with other people or organizations that can inappropriately influence our work, there is no professional or other personal interest of any nature or kind in any product, service and/or company that could be construed as influencing the content of this paper.

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