



# Comparison of DNA Methylation in *Vibrio vulnificus* Cells Grown in Human Serum with Those Grown in Seawater

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**ABSTRACT** The chromosomal methylation statuses of the highly virulent *Vibrio vulnificus* strain CMCP6 grown in human serum and in seawater are compared here. Growth in seawater resulted in ~4 times as much methylation as that in human serum, primarily N<sup>4</sup>-methylcytosines.

The virulence of *Vibrio vulnificus* is poorly understood, and the only definitive virulence factor is the presence of a capsule (1–3). DNA methylation has been implicated in the virulence of several other bacterial species (4–7). Changes in methylation sites may enable bacteria to respond to changes in their environment by altering gene expression (4, 8, 9). We hypothesized that the transition from an aquatic habitat to infection in a human could require altered DNA methylation, which we explored by growing the bacterium in human serum amended with iron (to mimic hemochromatosis) and in seawater (10).

Here, we have sequenced the methylome of the highly virulent and well-studied *V. vulnificus* strain CMCP6 (NCBI:txid216895), which was originally isolated from an infected patient in Korea (11). Bacteria were grown to late exponential phase in brain heart infusion broth, pelleted, and resuspended in phosphate-buffered saline at pH 7.5 (repeated once to wash cells). Normal pooled human serum (MP Biomedicals) was amended with ferric citrate to a final concentration of 0.896 mM Fe (12). Seawater (pH 7.8; salinity, 2‰) was collected from Hudson Beach, Florida, and sterilized using a hollow fiber ultrafilter (Rexeed 25S). Cultures were established at a starting concentration of 10<sup>7</sup> CFU/ml in 10 ml of serum or 40 ml of seawater and were shaken for 210 min at 37°C and 30°C, respectively. DNA was extracted with a blood and cell culture DNA minikit (Qiagen).

Preparation of 10-kb libraries and sequencing were performed at the National Center for Genome Resources in Santa Fe, New Mexico. Each sample was sequenced on two single-molecule real-time (SMRT) cells using PacBio RS II P5-C3 chemistry. *De novo* genome assembly proceeded with all four SMRT cells using RS\_HGAP\_Assembly.3 in SMRT Analysis 2.3.0 with default parameters, resulting in >1.3 million mapped subreads (average subread length, 2,053 bp) and eight contigs (six spurious) (13). Two contigs corresponding to chromosomes 1 and 2 (14) with a total length of 5,199,228 nucleotides (46.75% GC content; 499.3× coverage; quality value [QV], 48.7) were obtained. The assembled genome was 72,430 nucleotides and 72,532 nucleotides longer than the *V. vulnificus* CMCP6 genomes in the Integrated Microbial Genomes and Microbiomes system and NCBI, respectively.

Methylated bases and motifs were identified in the *de novo* assembly using RS\_Modification\_and\_Motif\_Analysis with default parameters (modification QV, 30). All identified methylated motifs had ≥70-fold or 180-fold coverage in human serum and seawater, respectively (Table 1). Seawater treatment resulted in ~4 times as many methylated bases (primarily N<sup>4</sup>-methylcytosine) as human serum did (447,389 versus 142,369, respectively).

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**TABLE 1** Motifs in human serum and seawater detected using SMRT Analysis v2.3.0

Motif <sup>a</sup>	Type	Percent modified in:		No. of motifs in genome	Motif score for:	
		Human serum	Seawater		Human serum	Seawater
GATC	m6A	98.74	98.89	44,386	111.96	239.94
GCCAN <sub>9</sub> TCC	m6A	98.32	98.32	537	109.05	242.73
GGAN <sub>9</sub> TGGC	m6A	97.39	97.58	537	109.26	239.93
AKGYAVYW	m6A	18.80	NA <sup>b</sup>	6,283	48.53	NA
AKGYASYW	m6A	NA	27.58	3,985	NA	68.71
ADDRGCAD	m6A	18.54	22.16	2,956	47.74	69.04
CSNNNNNG	m4C	6.67	NA	264,532	47.59	NA
CWGNNVNG	m4C	3.98	NA	50,788	43.90	NA

<sup>a</sup> Modified bases are bolded as follows: **A**, N<sup>6</sup>-methyladenine (m6A); **C**, N<sup>4</sup>-methylcytosine (m4C). The methylated bases on the complementary strand are underlined.

<sup>b</sup> NA, not applicable.

**Data availability.** These whole-genome sequencing data have been deposited in the Sequence Read Archive under the project accession number [PRJNA503483](https://www.ncbi.nlm.nih.gov/sra/PRJNA503483). The GenBank genome accession numbers are [CP037931](https://www.ncbi.nlm.nih.gov/nuccore/CP037931) and [CP037932](https://www.ncbi.nlm.nih.gov/nuccore/CP037932) for chromosomes 1 and 2, respectively. The accession numbers for the raw sequencing reads of *V. vulnificus* CMCP6 are [SAMN10360788](https://www.ncbi.nlm.nih.gov/sra/SAMN10360788) and [SAMN10360789](https://www.ncbi.nlm.nih.gov/sra/SAMN10360789) for human serum and seawater, respectively.

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## REFERENCES

- Gauthier JD, Jones MK, Thiaville P, Joseph JL, Swain RA, Krediet CJ, Gulig PA, Teplitski M, Wright AC. 2010. Role of GacA in virulence of *Vibrio vulnificus*. *Microbiology* 156:3722–3733. <https://doi.org/10.1099/mic.0.043422-0>.
- Wright AC, Simpson LM, Oliver JD, Morris JG, Jr. 1990. Phenotypic evaluation of acapsular transposon mutants of *Vibrio vulnificus*. *Infect Immun* 58:1769–1773.
- Moreno ML, Landgraf M. 1998. Virulence factors and pathogenicity of *Vibrio vulnificus* strains isolated from seafood. *J Appl Microbiol* 84: 747–751. <https://doi.org/10.1046/j.1365-2672.1998.00404.x>.
- Anjum A, Brathwaite KJ, Aidley J, Connerton PL, Cummings NJ, Parkhill J, Connerton I, Bayliss CD. 2016. Phase variation of a type IIG restriction-modification enzyme alters site-specific methylation patterns and gene expression in *Campylobacter jejuni* strain NCTC11168. *Nucleic Acids Res* 44:4581–4594. <https://doi.org/10.1093/nar/gkw019>.
- Chao MC, Zhu S, Kimura S, Davis BM, Schadt EE, Fang G, Waldor MK. 2015. A cytosine methyltransferase modulates the cell envelope stress response in the cholera pathogen. *PLoS Genet* 11:e1005666. <https://doi.org/10.1371/journal.pgen.1005666>.
- Robertson GT, Reisenauer A, Wright R, Jensen RB, Jensen A, Shapiro L, Roop RM, II. 2000. The *Brucella abortus* CcrM DNA methyltransferase is essential for viability, and its overexpression attenuates intracellular replication in murine macrophages. *J Bacteriol* 182:3482–3489. <https://doi.org/10.1128/jb.182.12.3482-3489.2000>.
- Cohen NR, Ross CA, Jain S, Shapiro RS, Gutierrez A, Belenky P, Li H, Collins JJ. 2016. A role for the bacterial GATC methylome in antibiotic stress survival. *Nat Genet* 48:581–586. <https://doi.org/10.1038/ng.3530>.
- Anton BP, Harhay GP, Smith TPL, Blom J, Roberts RJ. 2016. Comparative methylome analysis of the occasional ruminant respiratory pathogen *Bibersteinia trehalosi*. *PLoS One* 11:e0161499. <https://doi.org/10.1371/journal.pone.0161499>.
- Manso AS, Chai MH, Atack JM, Furi L, De Ste Croix M, Haigh R, Trappetti C, Ogunniyi AD, Shewell LK, Boitano M, Clark TA, Korlach J, Blades M, Mirkes E, Gorban AN, Paton JC, Jennings MP, Oggioni MR. 2014. A random six-phase switch regulates pneumococcal virulence via global epigenetic changes. *Nat Commun* 5:5055. <https://doi.org/10.1038/ncomms6055>.
- Williams TC, Blackman ER, Morrison SS, Gibas CJ, Oliver JD. 2014. Transcriptome sequencing reveals the virulence and environmental genetic programs of *Vibrio vulnificus* exposed to host and estuarine conditions. *PLoS One* 9:e114376. <https://doi.org/10.1371/journal.pone.0114376>.
- Kim YR, Lee SE, Kim CM, Kim SY, Shin EK, Shin DH, Chung SS, Choy HE, Progulsk-Fox A, Hillman JD, Handfield M, Rhee JH. 2003. Characterization and pathogenic significance of *Vibrio vulnificus* antigens preferentially expressed in septicemic patients. *Infect Immun* 71:5461–5471. <https://doi.org/10.1128/iai.71.10.5461-5471.2003>.
- Alice AF, Naka H, Crosa JH. 2008. Global gene expression as a function of the iron status of the bacterial cell: influence of differentially expressed genes in the virulence of the human pathogen *Vibrio vulnificus*. *Infect Immun* 76:4019–4037. <https://doi.org/10.1128/IAI.00208-08>.
- Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <https://doi.org/10.1038/nmeth.2474>.
- Kim HU, Kim SY, Jeong H, Kim TY, Kim JJ, Choy HE, Yi KY, Rhee JH, Lee SY. 2011. Integrative genome-scale metabolic analysis of *Vibrio vulnificus* for drug targeting and discovery. *Mol Syst Biol* 7:460. <https://doi.org/10.1038/msb.2010.115>.