



Soil Microbial Biogeography in a Changing World: Recent Advances and Future Perspectives

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ABSTRACT Soil microbial communities are fundamental to maintaining key soil processes associated with litter decomposition, nutrient cycling, and plant productivity and are thus integral to human well-being. Recent technological advances have exponentially increased our knowledge concerning the global ecological distributions of microbial communities across space and time and have provided evidence for their contribution to ecosystem functions. However, major knowledge gaps in soil biogeography remain to be addressed over the coming years as technology and research questions continue to evolve. In this minireview, we state recent advances and future directions in the study of soil microbial biogeography and discuss the need for a clearer concept of microbial species, projections of soil microbial distributions toward future global change scenarios, and the importance of embracing culture and isolation approaches to determine microbial functional profiles. This knowledge will be critical to better predict ecosystem functions in a changing world.

KEYWORDS future perspectives, recent advances, soil microbial biogeography

oils would not exist without the activity and diversity of millions of soil-resident animals and microorganisms. The aims of soil microbial biogeography are to study the ecological distributions of soil microbial diversity, community composition, and functional traits across space and time from regional to global scales. The study of microbial biogeography is essential to better understand the mechanisms that generate and maintain microbial diversity and that regulate key ecosystem processes, such as nutrient cycling, organic matter decomposition, plant productivity, and public health. Thanks to the development of high-throughput sequencing techniques and bioinformatic analyses (Fig. 1A) (1) and to the growing interest in this topic (Fig. 1B), we are now far from the initial stages of microbial biogeography when Baas-Becking (2) proposed that "everything is everywhere, but, the environment selects" (data in Fig. 1 were collected from the Web of Science Core Collection using the keywords described in the figure legend). Today, we know a lot about the ecological drivers of microbial diversity and community composition across different ecosystem types, including oceans (3, 4), soils (5, 6), and freshwater (7, 8) (Fig. 1A). However, in addition to the critical lack of agreement about the concept of "microbial species," very little is still known about the hundreds of thousands of microbe-microbe and plant/animalmicrobe interactions, which presumably control soil biodiversity and ecosystem functions. Moreover, we know little about the future global distributions of soil microbial taxa under global-change scenarios (especially for the less studied protists), which limits our capacity to predict changes in ecosystem function worldwide. Our capacity to predict changes in microbially driven functions is hampered because of the approaches used. Studies focusing on culturing and whole-genome sequencing are needed to reduce these knowledge gaps. In this minireview, we focus on the bioge-

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FIG 1 Numbers of published articles in the field of microbial biogeography over time. (A) Numbers of published articles on different ecosystems. Keywords [(microorganism* or microbe* or bacter* or archae* or fung*) and (spatial distribution* or biogeograph* or geographic distance)] were used to search for papers related to microbial biogeography. Additional keywords representing each ecosystem were added, as follows: soil (soil*), ocean (ocean* or sea* or marine*), inland waterbody (river* or lake* or spring* or stream*), and glacier (ice sheet* or glacier*). The vertical dashed lines indicate the critical time points at which the relevant technologies advanced. (B) Numbers of published articles related to all soils and the soils within different ecosystems. Additional keywords representing each ecosystem were added, as follows: or farmland* or paddy or agricultur* or rice* or wheat or till* or field soil* or corn* or upland* or peanut), grassland (grassland* or prairie* or turgrass or grass* or steppe* or meadow* or herb* or sedge*), and desert (desert*). All data were collected from the Web of Science Core Collection. Asterisks represent all possible word endings used for the searches; e.g., fung* represents, fungus, fungi, and fungal.

ography of soil communities and highlight the knowns and unknowns in the field of soil microbial biogeography, from the constantly changing concept of microbial species to future projections of the soil microbiome, and will highlight the advances required to move this field of knowledge forward.

DISTRIBUTIONS AND ECOLOGICAL DRIVERS OF SOIL MICROBIAL COMMUNITIES AT A GLOBAL SCALE

Studies over the last 2 decades have significantly improved our knowledge of the distributions of soil microbial communities from local, regional, and continental to global scales. From a classic geographical perspective, a negative correlation between distance from the equator and the diversity of plant and animals was largely reported during the last century (9). Similarly, the microbial diversity in marine ecosystems, including bacteria, protists, and planktonic foraminiferans, also exhibited a negative correlation with the global latitudinal gradient (10, 11). However, in soil systems, most studies have not identified the expected trend of soil biodiversity on a global scale. The typical trend of increasing diversity from the poles to the tropics has been partially proven in the Southern Hemisphere. For example, Delgado-Baquerizo et al. (12) found reduced soil bacterial diversity from the equator to Antarctica. However, there is no latitudinal diversity gradient for soil bacteria in the Northern Hemisphere (13, 14). In addition, Bahram et al. (6) found that fungal and bacterial diversity exhibited contrasting patterns across the latitudinal gradient in global topsoils; bacterial, but not fungal, taxonomic diversity was highest in temperate habitats. To date, studies focusing on



classic elevation patterns (9) have mostly failed to find a consistent negative or hump-shaped association between elevation and microbial diversity similar to that reported for plants and animals (15–21). Nevertheless, a few studies have reported a declining diversity of soil microbes with increasing elevation, which is similar to the elevation pattern of plant diversity (12, 22). In addition, different microbial taxa may present distinct elevation patterns. For example, Singh et al. (17) observed a single peak pattern for soil bacterial diversity but a double peak pattern for archaeal diversity. However, most latitudinal and altitudinal studies focused on specific mountains ranges and particular locations (e.g., North America or Australia) and focused on a single group of organisms (e.g., bacteria), hampering our capacity to evaluate the changes in microbial diversity across latitudinal and altitudinal gradients comprehensively. Therefore, future global collaborations in soil sampling and data sharing will be particularly important in soil microbial biogeographic studies.

Environmental properties are the most important drivers of the distribution of soil microbial communities globally (Fig. 2). The seminal work by Fierer and Jackson (13) highlighted the importance of soil pH as a fundamental driver of the distribution of bacterial diversity and community composition across contrasting biomes. Delgado-Baquerizo and Eldridge (23) further identified vegetation type and soil carbon content as universal predictors of the diversity of soil bacteria across global biomes. Fierer (24) recently reviewed the major ecological drivers of bacterial diversity. Regarding fungal communities, Tedersoo et al. (25) revealed the role of climate as the major ecological driver and provided the first global study of fungal biogeography. Other less studied global drivers of bacterial and fungal diversity include paleoclimatic legacies (26) and biological warfare (6). Bahram et al. (6) also found that different diversity responses to precipitation and soil pH contributed to the global niche differentiation of soil bacteria and fungi. However, we know much less about the major ecological drivers of other less studied organisms, such as soil protists, mites, nematodes (27, 28), and viruses (29, 30). For example, Bates et al. (31) suggested that climatic conditions likely controlled the distribution of soil protists globally. Liu et al. (32) demonstrated that the distribution of a T4-type phage community in paddy field soils was affected mainly by geographical distance; however, studies focusing on these phages are still rare. In addition, aboveground-belowground interactions and rhizosphere-microbe associations are major drivers of soil microbial diversity on a large spatial scale (Fig. 2). For example, aboveground plant taxonomic and functional attributes help to explain the distribution patterns of microbial biomass, diversity, and community composition (33-37). However, much less is known about the role of belowground plant traits in driving soil microbial distribution. Recently, Ramirez et al. (38) proposed that range-expanding plant species might predict the composition of belowground microbial communities, possibly in association with the belowground traits of these plants (e.g., roots). Animal behavior can also influence soil microbes. For example, ant colonies and mammalian foraging pits can alter the diversity and community composition of soil microbial communities across eastern Australia (39, 40). In addition to the effects of contemporary environmental factors (e.g., climate, soil, plants, and animals), the effects of historical factors (e.g., climatic legacies) (6, 26), and the characteristics of microorganisms themselves (e.g., body size, the ability to colonize, and adhesion) (41) on microbial distribution should be considered (Fig. 2).

Microorganisms are interdependent (42), resulting in some ecologically important but complex interactions, such as antagonistic, competitive, mutualistic, and predatorprey interactions (43) (Fig. 2). This complexity of the interactions among microbial members has been explored increasingly using network analysis (44, 45). The use of correlation networks in microbial ecology (46) have improved our capacity to quantify the level of microbial co-occurrence patterns, understand the drivers of microbial community assembly (e.g., soil carbon and pH and vegetation types) (47–49), and identify highly connected taxa and keystone species (50, 51) across environmental gradients (48). The field of microbial networks is relatively new and should be developed based on the years of experience in studying plant and animal communities (52,





FIG 2 Diagram of the main research fields in soil microbial biogeography that need to be studied spatially and temporally. Boxes with different-colored headings indicate the different research areas. Some diagram elements were modified from the work of Ladau et al. (93). OTUS, operational taxonomic units.



53). However, we still lack strong evidence of the ecological interpretation that exists in network inference, which needs more experimental verification in the future (54).

LINKING SOIL MICROBIAL BIOGEOGRAPHY TO ECOSYSTEM FUNCTION

Soil biodiversity plays active roles in the regulation of ecological functions and ecosystem services (55–57). A central goal of soil microbial biogeography is to link the distribution of microbial communities with the ecological functions that they support, including single (nutrient cycling, plant productivity, and public health) (58-61) and multiple (ecosystem multifunctionality) functions (56, 57) (Fig. 2). For example, Nelson et al. (61) investigated the global biogeography of microbial N traits (defined as eight N-cycling pathways) and found that some microbial groups seemed to be N-cycling specialists or generalists, suggesting the close relationship between microbial community and N cycling across global soils. In addition, using metagenomic sequencing, Fierer et al. (62) comprehensively surveyed soil microbes and multiple functions in different ecosystems and found that potential soil functional and taxonomic diversity and community composition were highly correlated. Similarly, the richness of bacteria and fungi has been found to drive ecosystem multifunctionality (nutrient cycling, organic matter decomposition, and plant productivity) in regional (63, 64) and global (e.g., dryland) (57) biomes of terrestrial ecosystems. Moreover, microbial community composition was found to regulate the resistance of ecosystem multifunctionality to global change in drylands globally (65). Furthermore, the relationships between soil microbial biodiversity and ecosystem multifunctionality are mediated by climate in the alpine grasslands of the Tibetan Plateau (63). Improving our knowledge of these associations between microbial communities and soil functions is necessary to advance the general prediction for ecological functions at local and global scales. However, further study is needed to better understand how different microbes correlate with soil functions and the underlying mechanisms of how the microbial community drives multiple ecological functions.

Despite the growing amount of data supporting the microbial biodiversity-function relationship, the majority of these studies are based on observational data, and experimental approaches to support the links between microbial taxa and functions are lacking, except in references 56, 64, and 66. For example, experiments have explored the mechanisms of ectomycorrhizal fungi in slowing soil carbon cycling (67), and strictly controlled experiments have been used to directly examine the distribution of trophic links as underlying mechanisms to predict the invasion resistance of plant rootassociated bacterial communities against an invading pathogen and the subsequent reduction of disease incidence (68). Recent experimental work suggests that microbial diversity (64) and microbiome complexity (66) determine ecosystem function. However, very little is known about how specific species contribute to ecosystem functioning in the real world. Future experimental work and global initiatives should focus on isolating and culturing soil microbial taxa and on obtaining information via whole-genome sequencing, proteomics, and metabolomics-based approaches to assign specific functions to specific taxa (Fig. 2) (64, 69–72). This information is critical for identifying soil organisms to promote plant production and fight their pests in the field. More importantly, this information will improve our current classification of soil microbes, of which most species remain unclassified. Global initiatives should support taxonomists aiming to dedicate part of their career to culturing and isolating taxa, a fundamental work that is required to move the field of microbial ecology forward but that remains overlooked, partly because it is time-consuming and does not always result in distinguished publications, which hampers researchers' early careers.

GLOBAL ATLASES OF SOIL BIODIVERSITY AND THEIR FUNCTIONS UNDER GLOBAL-CHANGE SCENARIOS

A major breakthrough in soil microbial biogeography was the recent appearance of the first global atlases of the abundance or biodiversity of bacteria (23, 73), fungi (74), nematodes (75, 76), earthworms (77), mycorrhizal fungi, and N fixer organisms (78),



highlighting the most likely locations containing unknown taxa (79) (Fig. 2). A diverse range of soil characteristics (e.g., soil pH) and climatic conditions has been used to predict and map the global distributions of a myriad of soil organisms at regional (80, 81), national (82, 83), continental (84, 85), and global (25, 73) scales. These efforts have led to the first national atlas of bacterial biodiversity across European Union (EU) member states based on the existing EU-wide soil pH data (84) and the first French national atlas of soil organisms across their territories, an effort which constitutes the foundation for the national conservation of soil biodiversity. Moreover, global initiatives are needed to further investigate how important land uses, such as agriculture (https://www.globalsustainableagriculture.org), regulate the global distributions of soil organisms.

In addition, to map the current distributions of soil microorganisms, extensive knowledge about the global projections of soil biodiversity under global-change scenarios is missing (Fig. 2). Soil microbial communities are strongly shaped by their surrounding environment; for example, soil microbial diversity and composition are sensitive to drought (86, 87), temperature (88), and fertilization (89). Changes in environmental conditions exert a powerful influence on microbial functions. For example, climate is a predominant driver that regulates litter and root decomposition over large spatial scales (90, 91), mainly via its direct influence on the reaction kinetics of decomposition processes and the decomposer community (92). By leveraging the associations between current bacterial distributions and historical climate data, Ladau et al. (93) predicted that soil bacterial diversity will increase across the majority (about 75%) of the Tibetan Plateau and northern North America if bacterial communities equilibrate with existing climatic conditions. However, we still lack predictions of core microbiomes or keystone species at large spatial scales. In addition, the absolute quantification of microbiota abundance will become more competitive (94), because relative abundance cannot reflect the real microbial composition. In the future, with more detailed background information on microbes, soils, plants, and the climate of the studied sites, we may obtain high-resolution maps of soil microbes. Furthermore, scientists have begun to note the important role of microbial information in improving the resolution of these models (95, 96). The microbial environmental interactome (interactions between microbiomes and their surrounding environment) has shaped the planet's past and will continue to do so in the future (97). Therefore, the mathematical models used to predict the response and feedback of the ecosystem in the context of global climate change need to incorporate the microbiome data, including the spatial-temporal dynamics of the microbiome and microbial interactions within food webs. Although some efforts have been made to predict the future distributions of soil microbial communities, we still lack the ability to globally predict future soil biodiversity and ecosystem functions into the future.

FURTHER PERSPECTIVES

Soil microbial biogeography has become a research hot spot in the fields of soil biology and microbial ecology (Fig. 1) (6, 73, 98). Despite the recent advances in molecular techniques and the existence of global efforts, like the Earth Microbiome Project (98, 99), our minireview identified major challenges and research questions in the field of microbial biogeography. Our capacity to address some of these questions remains limited by methodological issues. For example, the differences in data collection and methods from different studies make integrative analysis difficult at regional or global scales. In addition, there are major gaps in current global and temporal sampling data (100) that limit our capacity to predict the distribution of soil microbes spatially and temporally. The present minireview identified some major research areas where studies are needed to move the field of microbial biogeography forward; there is a need for a stronger concept of microbial species, our capacity to generate projections of the soil microbiome toward future global change scenarios needs to be improved, the importance of the complexity within the soil food web should be



embraced, and the culture and isolation approaches that determine microbial functional profiles should not be neglected. Such knowledge may help us to cope with the challenges of future environmental changes and improve our ability to accurately predict microbial communities and their function in a changing world. We suggest the following challenges and research opportunities in future microbial biogeographic studies.

(i) A clear definition of microbial species is still lacking and is essential for biogeographic studies. Advances in high-throughput sequencing have led to the rapid development of microbial species definitions based on the species' genealogical, genomic, and phenotypic coherence (101); however, there is not a clear consensus about the concept of "microbial species." The microbial species definitions based on PCR (phylotype) result in *ad hoc* species groups, which limits our capacity to identify new species and their ecological preferences. In this way, further work based on non-PCR-based methods, high-throughput culturing and identification techniques, and faster microbial isolation and cultivation are needed to enlarge the reference databases (e.g., GenBank, Greengenes, and Silva) and fill the gaps of microbial classification (102). Recently, the genome taxonomy database (GTDB) was developed to provide more pragmatic and objective definitions of taxonomic levels based on sequence distance (103), which is now a primary starting point for gathering sequences to be used in phylogenetic analyses that lead to designations of species and other taxonomic levels.

(ii) The temporal distribution of microbial communities remains largely unknown. Information on changes in microbial communities over time (e.g., seasons, years, or much longer time scales) at the large spatial scale is currently a major knowledge gap, although large temporal variances of microbial community compositions have been observed (104). Based on the space-for-time substitution approach (e.g., soil chronosequence, succession, and restoration of ecosystems), it has been found that soil microbial communities may undergo predictable changes over time (5, 105–107). However, such studies are often challenging, because the resolution of DNA-sequencing approaches for temporal dynamics is limited by sequencing both dead and living organisms, and we lack information on microbial communities in globally distributed locations and across multiple years. Thus, for example, considering the existence of relic DNA in samples as a confounding factor is key in detecting fine-scale temporal patterns in microbial communities using DNA approaches (108, 109). Moreover, future global efforts, including existing global cooperative efforts like CLIMIFUN, NEON, LUCA, and NUTNET, should aim to monitor temporal variations in soil microorganisms and to set up additional sites in poorly studied regions from polar, tropical, and arid regions and in continents like Africa, Antarctica, and South America. Such accumulated samples and data over years will help us to better understand the change in microbial communities under future global-change scenarios.

(iii) Organisms should be engineered to support human development. Future advances in microbial biogeography associated with the use of synthetic biology, new approaches for microbial culturing, and multiple bio-omics (e.g., metatranscriptomics, metaproteomics, and metabolomics) may help us to harness the soil microbiome to promote crop production and health in a changing world. We are still far from knowing what functions are being conducted by every single microbial species and their contribution to terrestrial functions, yet synthetic biology approaches and engineering of microorganisms have been postulated to boost ecosystem restoration, rhizospheredriven crop yield, and pest control (110, 111), to fight global environmental change (112, 113), and even to aid the terraformation of other moons and planets to make them more similar to Earth (114, 115).

(iv) Researchers should make predictions for the soil microbiome into the future. Improving the prediction accuracy of models is critical for microbial mapping. First, soil samples must be collected from more types of habitats and as many locations as possible to enrich the database. Second, soil microbial communities are temporally dynamic; therefore, it is necessary to understand microbial variations at different time



scales. Third, not only the microbial diversity and community composition but also other information, such as microbial interactions (microbe-microbe, microbe-plant, microbe-host interactions), need to be considered and integrated into the model to improve our capacity to predict changes in key ecosystem functions (e.g., carbon storage) on a global scale. These accuracy-improved models may be further used to predict the temporal-spatial dynamics in soil biodiversity and ecosystem functions under changing environments, which will aid the conservation of soil biodiversity and the display of ecological functions under future climate change.

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REFERENCES

- Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, Costello EK, Fierer N, Peña AG, Goodrich JK, Gordon JI, Huttley GA, Kelley ST, Knights D, Koenig JE, Ley RE, Lozupone CA, McDonald D, Muegge BD, Pirrung M, Reeder J, Sevinsky JR, Turnbaugh PJ, Walters WA, Widmann J, Yatsunenko T, Zaneveld J, Knight R. 2010. QIIME allows analysis of high-throughput community sequencing data. Nat Methods 7:335–336. https://doi.org/10.1038/nmeth.f.303.
- Baas-Becking L. 1934. Geobiologie of inleiding tot de milieukunde. van Stockum and Zoon, The Hague, Nethetherlands.
- 3. Cermeno P, Falkowski PG. 2009. Controls on diatom biogeography in the ocean. Science 325:1539–1541. https://doi.org/10.1126/science .1174159.
- Hellweger FL, van Sebille E, Fredrick ND. 2014. Biogeographic patterns in ocean microbes emerge in a neutral agent-based model. Science 345:1346–1349. https://doi.org/10.1126/science.1254421.
- Tripathi BM, Stegen JC, Kim M, Dong K, Adams JM, Lee YK. 2018. Soil pH mediates the balance between stochastic and deterministic assembly of bacteria. ISME J 12:1072–1083. https://doi.org/10.1038/s41396-018 -0082-4.
- Bahram M, Hildebrand F, Forslund SK, Anderson JL, Soudzilovskaia NA, Bodegom PM, Bengtsson-Palme J, Anslan S, Coelho LP, Harend H, Huerta-Cepas J, Medema MH, Maltz MR, Mundra S, Olsson PA, Pent M, Põlme S, Sunagawa S, Ryberg M, Tedersoo L, Bork P. 2018. Structure and function of the global topsoil microbiome. Nature 560:233–237. https://doi.org/10.1038/s41586-018-0386-6.
- Filker S, Sommaruga R, Vila I, Stoeck T. 2016. Microbial eukaryote plankton communities of high-mountain lakes from three continents exhibit strong biogeographic patterns. Mol Ecol 25:2286–2301. https:// doi.org/10.1111/mec.13633.
- Liu L, Yang J, Yu Z, Wilkinson DM. 2015. The biogeography of abundant and rare bacterioplankton in the lakes and reservoirs of China. ISME J 9:2068–2077. https://doi.org/10.1038/ismej.2015.29.
- 9. MacArthur J. 1975. Ecology and evolution of communities. Belknap Press, Cambridge, MA.
- Pommier T, Canbäck B, Riemann L, Boström KH, Simu K, Lundberg P, Tunlid A, Hagström Å. 2007. Global patterns of diversity and community structure in marine bacterioplankton. Mol Ecol 16:867–880. https:// doi.org/10.1111/j.1365-294X.2006.03189.x.
- Fuhrman JA, Steele JA, Hewson I, Schwalbach MS, Brown MV, Green JL, Brown JH. 2008. A latitudinal diversity gradient in planktonic marine bacteria. Proc Natl Acad Sci U S A 105:7774–7778. https://doi.org/10 .1073/pnas.0803070105.
- Delgado-Baquerizo M, Maestre FT, Reich PB, Trivedi P, Osanai Y, Liu YR, Hamonts K, Jeffries TC, Singh BK. 2016. Carbon content and climate variability drive global soil bacterial diversity patterns. Ecol Monogr 86:373–380. https://doi.org/10.1002/ecm.1216.

- Fierer N, Jackson RB. 2006. The diversity and biogeography of soil bacterial communities. Proc Natl Acad Sci U S A 103:626–631. https:// doi.org/10.1073/pnas.0507535103.
- Chu H, Fierer N, Lauber CL, Caporaso JG, Knight R, Grogan P. 2010. Soil bacterial diversity in the Arctic is not fundamentally different from that found in other biomes. Environ Microbiol 12:2998–3006. https://doi .org/10.1111/j.1462-2920.2010.02277.x.
- Fierer N, McCain CM, Meir P, Zimmermann M, Rapp JM, Silman MR, Knight R. 2011. Microbes do not follow the elevational diversity patterns of plants and animals. Ecology 92:797–804. https://doi.org/10 .1890/10-1170.1.
- Singh D, Takahashi K, Adams JM. 2012. Elevational patterns in archaeal diversity on Mt. Fuji. PLoS One 7:e44494. https://doi.org/10.1371/ journal.pone.0044494.
- Singh D, Takahashi K, Kim M, Chun J, Adams JM. 2012. A hump-backed trend in bacterial diversity with elevation on Mount Fuji, Japan. Microb Ecol 63:429–437. https://doi.org/10.1007/s00248-011-9900-1.
- Shen C, Xiong J, Zhang H, Feng Y, Lin X, Li X, Liang W, Chu H. 2013. Soil pH drives the spatial distribution of bacterial communities along elevation on Changbai Mountain. Soil Biol Biochem 57:204–211. https:// doi.org/10.1016/j.soilbio.2012.07.013.
- Shen C, Liang W, Shi Y, Lin X, Zhang H, Wu X, Xie G, Chain P, Grogan P, Chu H. 2014. Contrasting elevational diversity patterns between eukaryotic soil microbes and plants. Ecology 95:3190–3202. https://doi .org/10.1890/14-0310.1.
- Miyamoto Y, Nakano T, Hattori M, Nara K. 2014. The mid-domain effect in ectomycorrhizal fungi: range overlap along an elevation gradient on Mount Fuji, Japan. ISME J 8:1739–1746. https://doi.org/10.1038/ismej .2014.34.
- Wang J, Meier S, Soininen J, Casamayor EO, Pan F, Tang X, Yang X, Zhang Y, Wu Q, Zhou J, Shen J. 2017. Regional and global elevational patterns of microbial species richness and evenness. Ecography (Cop) 40:393–402. https://doi.org/10.1111/ecog.02216.
- Nottingham AT, Fierer N, Turner BL, Whitaker J, Ostle NJ, McNamara NP, Bardgett RD, Leff JW, Salinas N, Silman MR, Kruuk LEB, Meir P. 2018. Microbes follow Humboldt: temperature drives plant and soil microbial diversity patterns from the Amazon to the Andes. Ecology 99: 2455–2466. https://doi.org/10.1002/ecy.2482.
- Delgado-Baquerizo M, Eldridge DJ. 2019. Cross-biome drivers of soil bacterial alpha diversity on a worldwide scale. Ecosystems 22: 1220–1231. https://doi.org/10.1007/s10021-018-0333-2.
- Fierer N. 2017. Embracing the unknown: disentangling the complexities of the soil microbiome. Nat Rev Microbiol 15:579–590. https://doi.org/ 10.1038/nrmicro.2017.87.
- Tedersoo L, Bahram M, Polme S, Koljalg U, Yorou NS, Wijesundera R, Ruiz LV, Vasco-Palacios AM, Thu PQ, Suija A, Smith ME, Sharp C,

mSystems*

Saluveer E, Saitta A, Rosas M, Riit T, Ratkowsky D, Pritsch K, Poldmaa K, Piepenbring M, Phosri C, Peterson M, Parts K, Partel K, Otsing E, Nouhra E, Njouonkou AL, Nilsson RH, Morgado LN, Mayor J, May TW, Majuakim L, Lodge DJ, Lee SS, Larsson K-H, Kohout P, Hosaka K, Hiiesalu I, Henkel TW, Harend H, Guo L-D, Greslebin A, Grelet G, Geml J, Gates G, Dunstan W, Dunk C, Drenkhan R, Dearnaley J, De Kesel A, Dang T, Chen X, Buegger F, Brearley FQ, Bonito G, Anslan S, Abell S, Abarenkov K. 2014. Global diversity and geography of soil fungi. Science 346: 1256688–1256688. https://doi.org/10.1126/science.1256688.

- Delgado-Baquerizo M, Bissett A, Eldridge DJ, Maestre FT, He J-Z, Wang J-T, Hamonts K, Liu Y-R, Singh BK, Fierer N. 2017. Palaeoclimate explains a unique proportion of the global variation in soil bacterial communities. Nat Ecol Evol 1:1339–1347. https://doi.org/10.1038/s41559-017 -0259-7.
- Geisen S, Briones MJI, Gan H, Behan-Pelletier VM, Friman V-P, de Groot GA, Hannula SE, Lindo Z, Philippot L, Tiunov AV, Wall DH. 2019. A methodological framework to embrace soil biodiversity. Soil Biol Biochem 136:107536. https://doi.org/10.1016/j.soilbio.2019.107536.
- Thakur MP, Geisen S. 2019. Trophic regulations of the soil Microbiome. Trends Microbiol 27:771–780. https://doi.org/10.1016/j.tim.2019.04 .008.
- Wang X, Liu J, Yu Z, Jin J, Liu X, Wang G. 2016. Novel groups and unique distribution of phage *phoH* genes in paddy waters in northeast China. Sci Rep 6:38428. https://doi.org/10.1038/srep38428.
- Liu J, Wang G, Wang Q, Liu J, Jin J, Liu X. 2012. Phylogenetic diversity and assemblage of major capsid genes (g23) of T4-type bacteriophages in paddy field soils during rice growth season in Northeast China. Soil Sci Plant Nutr 58:435–444. https://doi.org/10.1080/00380768.2012 .703610.
- Bates ST, Clemente JC, Flores GE, Walters WA, Parfrey LW, Knight R, Fierer N. 2013. Global biogeography of highly diverse protistan communities in soil. ISME J 7:652–659. https://doi.org/10.1038/ismej.2012 .147.
- 32. Liu J, Yu Z, Wang X, Jin J, Liu X, Wang G. 2016. The distribution characteristics of the major capsid gene (g23) of T4-type phages in paddy floodwater in Northeast China. Soil Sci Plant Nutr 62:133–139. https://doi.org/10.1080/00380768.2016.1163507.
- 33. de Vries FT, Manning P, Tallowin JRB, Mortimer SR, Pilgrim ES, Harrison KA, Hobbs PJ, Quirk H, Shipley B, Cornelissen JHC, Kattge J, Bardgett RD. 2012. Abiotic drivers and plant traits explain landscape-scale patterns in soil microbial communities. Ecol Lett 15:1230–1239. https://doi.org/10.1111/j.1461-0248.2012.01844.x.
- 34. Prober SM, Leff JW, Bates ST, Borer ET, Firn J, Harpole WS, Lind EM, Seabloom EW, Adler PB, Bakker JD, Cleland EE, DeCrappeo NM, DeLorenze E, Hagenah N, Hautier Y, Hofmockel KS, Kirkman KP, Knops JMH, La Pierre KJ, MacDougall AS, McCulley RL, Mitchell CE, Risch AC, Schuetz M, Stevens CJ, Williams RJ, Fierer N. 2015. Plant diversity predicts beta but not alpha diversity of soil microbes across grasslands worldwide. Ecol Lett 18:85–95. https://doi.org/10.1111/ele.12381.
- Yang T, Adams JM, Shi Y, He J-S, Jing X, Chen L, Tedersoo L, Chu H. 2017. Soil fungal diversity in natural grasslands of the Tibetan Plateau: associations with plant diversity and productivity. New Phytol 215: 756–765. https://doi.org/10.1111/nph.14606.
- Delgado-Baquerizo M, Fry EL, Eldridge DJ, de Vries FT, Manning P, Hamonts K, Kattge J, Boenisch G, Singh BK, Bardgett RD. 2018. Plant attributes explain the distribution of soil microbial communities in two contrasting regions of the globe. New Phytol 219:574–587. https://doi .org/10.1111/nph.15161.
- 37. Yang T, Tedersoo L, Soltis PS, Soltis DE, Gilbert JA, Sun M, Shi Y, Wang H, Li Y, Zhang J, Chen Z, Lin H, Zhao Y, Fu C, Chu H. 2019. Phylogenetic imprint of woody plants on the soil mycobiome in natural mountain forests of eastern China. ISME J 13:686–697. https://doi.org/10.1038/s41396-018-0303-x.
- Ramirez KS, Snoek LB, Koorem K, Geisen S, Bloem LJ, ten Hooven F, Kostenko O, Krigas N, Manrubia M, Caković D, van Raaij D, Tsiafouli MA, Vreš B, Čelik T, Weser C, Wilschut RA, van der Putten WH. 2019. Range-expansion effects on the belowground plant microbiome. Nat Ecol Evol 3:604–611. https://doi.org/10.1038/s41559-019-0828-z.
- Eldridge DJ, Delgado-Baquerizo M, Woodhouse JN, Neilan BA. 2016. Mammalian engineers drive soil microbial communities and ecosystem functions across a disturbance gradient. J Anim Ecol 85:1636–1646. https://doi.org/10.1111/1365-2656.12574.
- 40. Delgado-Baquerizo M, Eldridge DJ, Hamonts K, Singh BK. 2019. Ant

colonies promote the diversity of soil microbial communities. ISME J 13:1114–1118. https://doi.org/10.1038/s41396-018-0335-2.

- 41. Zinger L, Taberlet P, Schimann H, Bonin A, Boyer F, De Barba M, Gaucher P, Gielly L, Giguet-Covex C, Iribar A, Réjou-Méchain M, Rayé G, Rioux D, Schilling V, Tymen B, Viers J, Zouiten C, Thuiller W, Coissac E, Chave J. 2019. Body size determines soil community assembly in a tropical forest. Mol Ecol 28:528–543. https://doi.org/10.1111/mec .14919.
- 42. Graham EB, Knelman JE, Schindlbacher A, Siciliano S, Breulmann M, Yannarell A, Beman JM, Abell G, Philippot L, Prosser J, Foulquier A, Yuste JC, Glanville HC, Jones DL, Angel R, Salminen J, Newton RJ, Bürgmann H, Ingram LJ, Hamer U, Siljanen HMP, Peltoniemi K, Potthast K, Bañeras L, Hartmann M, Banerjee S, Yu R-Q, Nogaro G, Richter A, Koranda M, Castle SC, Goberna M, Song B, Chatterjee A, Nunes OC, Lopes AR, Cao Y, Kaisermann A, Hallin S, Strickland MS, Garcia-Pausas J, Barba J, Kang H, Isobe K, Papaspyrou S, Pastorelli R, Lagomarsino A, Lindström ES, Basiliko N, Nemergut DR. 2016. Microbes as engines of ecosystem function: when does community structure enhance predictions of ecosystem processes? Front Microbiol 7:214. https://doi.org/10 .3389/fmicb.2016.00214.
- Mougi A, Kondoh M. 2012. Diversity of interaction types and ecological community stability. Science 337:349–351. https://doi.org/10.1126/ science.1220529.
- 44. Weiss S, Van Treuren W, Lozupone C, Faust K, Friedman J, Deng Y, Xia LC, Xu ZZ, Ursell L, Alm EJ, Birmingham A, Cram JA, Fuhrman JA, Raes J, Sun F, Zhou J, Knight R. 2016. Correlation detection strategies in microbial data sets vary widely in sensitivity and precision. ISME J 10:1669–1681. https://doi.org/10.1038/ismej.2015.235.
- Röttjers L, Faust K. 2018. From hairballs to hypotheses—biological insights from microbial networks. FEMS Microbiol Rev 42:761–780. https://doi.org/10.1093/femsre/fuy030.
- Barberán A, Bates ST, Casamayor EO, Fierer N. 2012. Using network analysis to explore co-occurrence patterns in soil microbial communities. ISME J 6:343–351. https://doi.org/10.1038/ismej.2011.119.
- He D, Shen W, Eberwein J, Zhao Q, Ren L, Wu QL. 2017. Diversity and co-occurrence network of soil fungi are more responsive than those of bacteria to shifts in precipitation seasonality in a subtropical forest. Soil Biol Biochem 115:499–510. https://doi.org/10.1016/j.soilbio.2017 .09.023.
- de Vries FT, Griffiths RI, Bailey M, Craig H, Girlanda M, Gweon HS, Hallin S, Kaisermann A, Keith AM, Kretzschmar M, Lemanceau P, Lumini E, Mason KE, Oliver A, Ostle N, Prosser JI, Thion C, Thomson B, Bardgett RD. 2018. Soil bacterial networks are less stable under drought than fungal networks. Nat Commun 9:3033. https://doi.org/10.1038/s41467 -018-05516-7.
- Delgado-Baquerizo M, Reith F, Dennis PG, Hamonts K, Powell JR, Young A, Singh BK, Bissett A. 2018. Ecological drivers of soil microbial diversity and soil biological networks in the Southern Hemisphere. Ecology 99:583–596. https://doi.org/10.1002/ecy.2137.
- Banerjee S, Schlaeppi K, van der Heijden M. 2019. Reply to 'Can we predict microbial keystones?' Nat Rev Microbiol 17:194–194. https:// doi.org/10.1038/s41579-018-0133-x.
- 51. Röttjers L, Faust K. 2019. Can we predict keystones? Nat Rev Microbiol 17:193–193. https://doi.org/10.1038/s41579-018-0132-y.
- Thebault E, Fontaine C. 2010. Stability of ecological communities and the architecture of mutualistic and trophic networks. Science 329: 853–856. https://doi.org/10.1126/science.1188321.
- 53. Trøjelsgaard K, Olesen JM. 2013. Macroecology of pollination networks. Glob Ecol Biogeogr 22:149–162. https://doi.org/10.1111/j.1466-8238 .2012.00777.x.
- Lv X, Zhao K, Xue R, Liu Y, Xu J, Ma B. 2019. Strengthening insights in microbial ecological networks from theory to applications. mSystems 4:e00124-19. https://doi.org/10.1128/mSystems.00124-19.
- Philippot L, Spor A, Hénault C, Bru D, Bizouard F, Jones CM, Sarr A, Maron P-A. 2013. Loss in microbial diversity affects nitrogen cycling in soil. ISME J 7:1609–1619. https://doi.org/10.1038/ismej.2013.34.
- Wagg C, Bender SF, Widmer F, van der Heijden MGA. 2014. Soil biodiversity and soil community composition determine ecosystem multifunctionality. Proc Natl Acad Sci U S A 111:5266–5270. https://doi .org/10.1073/pnas.1320054111.
- Delgado-Baquerizo M, Maestre FT, Reich PB, Jeffries TC, Gaitan JJ, Encinar D, Berdugo M, Campbell CD, Singh BK. 2016. Microbial diversity drives multifunctionality in terrestrial ecosystems. Nat Commun 7:10541. https://doi.org/10.1038/ncomms10541.



- Schnitzer SA, Klironomos JN, Hillerislambers J, Kinkel LL, Reich PB, Xiao K, Rillig MC, Sikes BA, Callaway RM, Mangan SA, van Nes EH, Scheffer M. 2011. Soil microbes drive the classic plant diversity-productivity pattern. Ecology 92:296–303. https://doi.org/10.1890/10-0773.1.
- Kanchiswamy CN, Malnoy M, Maffei ME. 2015. Chemical diversity of microbial volatiles and their potential for plant growth and productivity. Front Plant Sci 6:151. https://doi.org/10.3389/fpls.2015.00151.
- Limmathurotsakul D, Golding N, Dance DAB, Messina JP, Pigott DM, Moyes CL, Rolim DB, Bertherat E, Day N, Peacock SJ, Hay SI. 2016. Predicted global distribution of Burkholderia pseudomallei and burden of melioidosis. Nat Microbiol 1:15008. https://doi.org/10.1038/ nmicrobiol.2015.8.
- Nelson MB, Martiny AC, Martiny J. 2016. Global biogeography of microbial nitrogen-cycling traits in soil. Proc Natl Acad Sci U S A 113: 8033–8040. https://doi.org/10.1073/pnas.1601070113.
- Fierer N, Leff JW, Adams BJ, Nielsen UN, Bates ST, Lauber CL, Owens S, Gilbert JA, Wall DH, Caporaso JG. 2012. Cross-biome metagenomic analyses of soil microbial communities and their functional attributes. Proc Natl Acad Sci U S A 109:21390–21395. https://doi.org/10.1073/ pnas.1215210110.
- Jing X, Sanders NJ, Shi Y, Chu H, Classen AT, Zhao K, Chen L, Shi Y, Jiang Y, He J-S. 2015. The links between ecosystem multifunctionality and above- and belowground biodiversity are mediated by climate. Nat Commun 6:8159. https://doi.org/10.1038/ncomms9159.
- Delgado-Baquerizo M, Trivedi P, Trivedi C, Eldridge DJ, Reich PB, Jeffries TC, Singh BK. 2017. Microbial richness and composition independently drive soil multifunctionality. Funct Ecol 31:2330–2343. https://doi.org/ 10.1111/1365-2435.12924.
- Delgado-Baquerizo M, Eldridge DJ, Ochoa V, Gozalo B, Singh BK, Maestre FT. 2017. Soil microbial communities drive the resistance of ecosystem multifunctionality to global change in drylands across the globe. Ecol Lett 20:1295–1305. https://doi.org/10.1111/ele.12826.
- Wagg C, Schlaeppi K, Banerjee S, Kuramae EE, van der Heijden MGA. 2019. Fungal-bacterial diversity and microbiome complexity predict ecosystem functioning. Nat Commun 10:4841. https://doi.org/10.1038/ s41467-019-12798-y.
- 67. Averill C, Hawkes CV. 2016. Ectomycorrhizal fungi slow soil carbon cycling. Ecol Lett 19:937–947. https://doi.org/10.1111/ele.12631.
- Wei Z, Yang T, Friman V-P, Xu Y, Shen Q, Jousset A. 2015. Trophic network architecture of root-associated bacterial communities determines pathogen invasion and plant health. Nat Commun 6:8413. https://doi.org/10.1038/ncomms9413.
- Martinez-Alonso E, Pena-Perez S, Serrano S, Garcia-Lopez E, Alcazar A, Cid C. 2019. Taxonomic and functional characterization of a microbial community from a volcanic englacial ecosystem in Deception Island, Antarctica. Sci Rep 9:12158. https://doi.org/10.1038/ s41598-019-47994-9.
- Jaswal R, Pathak A, Chauhan A. 2019. Metagenomic evaluation of bacterial and fungal assemblages enriched within diffusion chambers and microbial traps containing uraniferous soils. Microorganisms 7:324. https://doi.org/10.3390/microorganisms7090324.
- Swenson TL, Karaoz U, Swenson JM, Bowen BP, Northen TR. 2018. Linking soil biology and chemistry in biological soil crust using isolate exometabolomics. Nat Commun 9:19. https://doi.org/10.1038/s41467 -017-02356-9.
- 72. Bastida F, Crowther TW, Prieto I, Routh D, García C, Jehmlich N. 2018. Climate shapes the protein abundance of dominant soil bacteria. Sci Total Environ 640–641:18–21. https://doi.org/10.1016/j.scitotenv.2018 .05.288.
- Delgado-Baquerizo M, Oliverio AM, Brewer TE, Benavent-González A, Eldridge DJ, Bardgett RD, Maestre FT, Singh BK, Fierer N. 2018. A global atlas of the dominant bacteria found in soil. Science 359:320–325. https://doi.org/10.1126/science.aap9516.
- Egidi E, Delgado-Baquerizo M, Plett JM, Wang J, Eldridge DJ, Bardgett RD, Maestre FT, Singh BK. 2019. A few Ascomycota taxa dominate soil fungal communities worldwide. Nat Commun 10:2369. https://doi.org/ 10.1038/s41467-019-10373-z.
- 75. van den Hoogen J, Geisen S, Routh D, Ferris H, Traunspurger W, Wardle DA, de Goede RGM, Adams BJ, Ahmad W, Andriuzzi WS, Bardgett RD, Bonkowski M, Campos-Herrera R, Cares JE, Caruso T, de Brito Caixeta L, Chen X, Costa SR, Creamer R, Mauro da Cunha Castro J, Dam M, Djigal D, Escuer M, Griffiths BS, Gutiérrez C, Hohberg K, Kalinkina D, Kardol P, Kergunteuil A, Korthals G, Krashevska V, Kudrin AA, Li Q, Liang W, Magilton M, Marais M, Martín JAR, Matveeva E, Mayad EH, Mulder C,

Mullin P, Neilson R, Nguyen TAD, Nielsen UN, Okada H, Rius JEP, Pan K, Peneva V, Pellissier L, Carlos Pereira da Silva J, et al. 2019. Soil nematode abundance and functional group composition at a global scale. Nature 572:194–198. https://doi.org/10.1038/s41586-019-1418-6.

- Bastida F, Eldridge DL, Abades S, Alfaro F, Gallardo A, García-Velázquez L, García C, Hart S, Pérez C, Santos F, Trivedi P, Williams M, Delgado-Baquerizo M. 7 November 2019. Climatic vulnerabilities and ecological preferences of soil invertebrates across biomes. Mol Ecol https://doi .org/10.1111/mec.15299.
- 77. Phillips HRP, Guerra CA, Bartz MLC, Briones MJI, Brown G, Crowther TW, Ferlian O, Gongalsky KB, van den Hoogen J, Krebs J, Orgiazzi A, Routh D, Schwarz B, Bach EM, Bennett J, Brose U, Decaëns T, König-Ries B, Loreau M, Mathieu J, Mulder C, van der Putten WH, Ramirez KS, Rillig MC, Russell D, Rutgers M, Thakur MP, de Vries FT, Wall DH, Wardle DA, Arai M, Ayuke FO, Baker GH, Beauséjour R, Bedano JC, Birkhofer K, Blanchart E, Blossey B, Bolger T, Bradley RL, Callaham MA, Capowiez Y, Caulfield ME, Choi A, Crotty FV, Dávalos A, Cosin DJD, Dominguez A, Duhour AE, van Eekeren N, et al. 2019. Global distribution of earthworm diversity. Science 366:480–485. https://doi.org/10.1126/ science.aax4851.
- Steidinger BS, GFBI Consortium, Crowther TW, Liang J, Van Nuland ME, Werner GDA, Reich PB, Nabuurs GJ, De-Miguel S, Zhou M, Picard N, Herault B, Zhao X, Zhang C, Routh D, Peay KG. 2019. Climatic controls of decomposition drive the global biogeography of forest-tree symbioses. Nature 569:404–408. https://doi.org/10.1038/s41586-019-1128-0.
- Delgado-Baquerizo M. 2019. Obscure soil microbes and where to find them. ISME J 13:2120–2124. https://doi.org/10.1038/s41396-019 -0405-0.
- Ferrier S, Manion G, Elith J, Richardson K. 2007. Using generalized dissimilarity modelling to analyse and predict patterns of beta diversity in regional biodiversity assessment. Divers Distrib 13:252–264. https:// doi.org/10.1111/j.1472-4642.2007.00341.x.
- Bru D, Ramette A, Saby NPA, Dequiedt S, Ranjard L, Jolivet C, Arrouays D, Philippot L. 2011. Determinants of the distribution of nitrogencycling microbial communities at the landscape scale. ISME J 5:532–542. https://doi.org/10.1038/ismej.2010.130.
- Terrat S, Horrigue W, Dequietd S, Saby NPA, Lelièvre M, Nowak V, Tripied J, Régnier T, Jolivet C, Arrouays D, Wincker P, Cruaud C, Karimi B, Bispo A, Maron PA, Prévost-Bouré NC, Ranjard L. 2017. Mapping and predictive variations of soil bacterial richness across France. PLoS One 12:5–8. https://doi.org/10.1371/journal.pone.0186766.
- Karimi B, Terrat S, Dequiedt S, Saby NPA, Horrigue W, Lelièvre M, Nowak V, Jolivet C, Arrouays D, Wincker P, Cruaud C, Bispo A, Maron P-A, Bouré NCP, Ranjard L. 2018. Biogeography of soil bacteria and archaea across France. Sci Adv 4:eaat1808. https://doi.org/10.1126/sciadv.aat1808.
- Griffiths RI, Thomson BC, Plassart P, Gweon HS, Stone D, Creamer RE, Lemanceau P, Bailey MJ. 2016. Mapping and validating predictions of soil bacterial biodiversity using European and national scale datasets. Appl Soil Ecol 97:61–68. https://doi.org/10.1016/j.apsoil.2015.06.018.
- Jiao S, Xu Y, Zhang J, Lu Y. 2019. Environmental filtering drives distinct continental atlases of soil archaea between dryland and wetland agricultural ecosystems. Microbiome 7:15. https://doi.org/10.1186/s40168 -019-0630-9.
- Maestre FT, Delgado-Baquerizo M, Jeffries TC, Eldridge DJ, Ochoa V, Gozalo B, Quero JL, García-Gómez M, Gallardo A, Ulrich W, Bowker MA, Arredondo T, Barraza-Zepeda C, Bran D, Florentino A, Gaitán J, Gutiérrez JR, Huber-Sannwald E, Jankju M, Mau RL, Miriti M, Naseri K, Ospina A, Stavi I, Wang D, Woods NN, Yuan X, Zaady E, Singh BK. 2015. Increasing aridity reduces soil microbial diversity and abundance in global drylands. Proc Natl Acad Sci U S A 112:15684–15689. https:// doi.org/10.1073/pnas.1516684112.
- Ochoa-Hueso R, Collins SL, Delgado-Baquerizo M, Hamonts K, Pockman WT, Sinsabaugh RL, Smith MD, Knapp AK, Power SA. 2018. Drought consistently alters the composition of soil fungal and bacterial communities in grasslands from two continents. Glob Chang Biol 24: 2818–2827. https://doi.org/10.1111/gcb.14113.
- Oliverio AM, Bradford MA, Fierer N. 2017. Identifying the microbial taxa that consistently respond to soil warming across time and space. Glob Chang Biol 23:2117–2129. https://doi.org/10.1111/gcb.13557.
- Leff JW, Jones SE, Prober SM, Barberán A, Borer ET, Firn JL, Harpole WS, Hobbie SE, Hofmockel KS, Knops JMH, McCulley RL, La Pierre K, Risch AC, Seabloom EW, Schütz M, Steenbock C, Stevens CJ, Fierer N. 2015. Consistent responses of soil microbial communities to elevated nutri-



ent inputs in grasslands across the globe. Proc Natl Acad Sci U S A 112:10967–10972. https://doi.org/10.1073/pnas.1508382112.

- Parton W, Silver WL, Burke IC, Grassens L, Harmon ME, Currie WS, King JY, Adair EC, Brandt LA, Hart SC, Fasth B. 2007. Global-scale similarities in nitrogen release patterns during long-term decomposition. Science 315:361–364. https://doi.org/10.1126/science.1134853.
- See CR, Luke McCormack M, Hobbie SE, Flores-Moreno H, Silver WL, Kennedy PG. 2019. Global patterns in fine root decomposition: climate, chemistry, mycorrhizal association and woodiness. Ecol Lett 22: 946–953. https://doi.org/10.1111/ele.13248.
- Suseela V, Tharayil N. 2018. Decoupling the direct and indirect effects of climate on plant litter decomposition: accounting for stress-induced modifications in plant chemistry. Glob Chang Biol 24:1428–1451. https://doi.org/10.1111/gcb.13923.
- Ladau J, Shi Y, Jing X, He J-S, Chen L, Lin X, Fierer N, Gilbert JA, Pollard KS, Chu H. 2018. Existing climate change will lead to pronounced shifts in the diversity of soil prokaryotes. mSystems 3:e00167-18. https://doi .org/10.1128/mSystems.00167-18.
- Tkacz A, Hortala M, Poole PS. 2018. Absolute quantitation of microbiota abundance in environmental samples. Microbiome 6:110. https://doi .org/10.1186/s40168-018-0491-7.
- Perveen N, Barot S, Alvarez G, Klumpp K, Martin R, Rapaport A, Herfurth D, Louault F, Fontaine S. 2014. Priming effect and microbial diversity in ecosystem functioning and response to global change: a modeling approach using the SYMPHONY model. Glob Chang Biol 20:1174–1190. https://doi.org/10.1111/gcb.12493.
- 96. Bradford MA, Ciska GF, Bonis A, Bradford EM, Classen AT, Cornelissen JHC, Crowther TW, De Long JR, Freschet GT, Kardol P, Manrubia-Freixa M, Maynard DS, Newman GS, Logtestijn RSP, Viketoft M, Wardle DA, Wieder WR, Wood SA, Van Der Putten WH. 2017. A test of the hierarchical model of litter decomposition. Nat Ecol Evol 1:1836–1845. https://doi.org/10.1038/s41559-017-0367-4.
- Larsen P, Hamada Y, Gilbert J. 2012. Modeling microbial communities: current, developing, and future technologies for predicting microbial community interaction. J Biotechnol 160:17–24. https://doi.org/10 .1016/j.jbiotec.2012.03.009.
- 98. Thompson LR, Earth Microbiome Project Consortium, Sanders JG, Mc-Donald D, Amir A, Ladau J, Locey KJ, Prill RJ, Tripathi A, Gibbons SM, Ackermann G, Navas-Molina JA, Janssen S, Kopylova E, Vázquez-Baeza Y, González A, Morton JT, Mirarab S, Zech Xu Z, Jiang L, Haroon MF, Kanbar J, Zhu Q, Jin Song S, Kosciolek T, Bokulich NA, Lefler J, Brislawn CJ, Humphrey G, Owens SM, Hampton-Marcell J, Berg-Lyons D, McKenzie V, Fierer N, Fuhrman JA, Clauset A, Stevens RL, Shade A, Pollard KS, Goodwin KD, Jansson JK, Gilbert JA, Knight R. 2017. A communal catalogue reveals Earth's multiscale microbial diversity. Nature 551: 457–463. https://doi.org/10.1038/nature24621.
- Gilbert JA, Jansson JK, Knight R. 2018. Earth Microbiome Project and global systems biology. mSystems 3:e00217-17. https://doi.org/10.1128/ mSystems.00217-17.
- 100. Cameron EK, Martins IS, Lavelle P, Mathieu J, Tedersoo L, Gottschall F, Guerra CA, Hines J, Patoine G, Siebert J, Winter M, Cesarz S, Delgado-Baquerizo M, Ferlian O, Fierer N, Kreft H, Lovejoy TE, Montanarella L, Orgiazzi A, Pereira HM, Phillips HRP, Settele J, Wall DH, Eisenhauer N.

2018. Global gaps in soil biodiversity data. Nat Ecol Evol 2:1042–1043. https://doi.org/10.1038/s41559-018-0573-8.

- Achtman M, Wagner M. 2008. Microbial diversity and the genetic nature of microbial species. Nat Rev Microbiol 6:431–440. https://doi .org/10.1038/nrmicro1872.
- Pham VHT, Kim J. 2012. Cultivation of unculturable soil bacteria. Trends Biotechnol 30:475–484. https://doi.org/10.1016/j.tibtech.2012.05.007.
- 103. York A. 2018. Next-generation bacterial taxonomy. Nat Rev Microbiol 16:583–583. https://doi.org/10.1038/s41579-018-0083-3.
- 104. Shade A, Gregory Caporaso J, Handelsman J, Knight R, Fierer N. 2013. A meta-analysis of changes in bacterial and archaeal communities with time. ISME J 7:1493–1506. https://doi.org/10.1038/ismej.2013.54.
- 105. Delgado-Baquerizo M, Bardgett RD, Vitousek PM, Maestre FT, Williams MA, Eldridge DJ, Lambers H, Neuhauser S, Gallardo A, García-Velázquez L, Sala OE, Abades SR, Alfaro FD, Berhe AA, Bowker MA, Currier CM, Cutler NA, Hart SC, Hayes PE, Hseu Z-Y, Kirchmair M, Peña-Ramírez VM, Pérez CA, Reed SC, Santos F, Siebe C, Sullivan BW, Weber-Grullon L, Fierer N. 2019. Changes in belowground biodiversity during ecosystem development. Proc Natl Acad Sci U S A 116:6891–6896. https://doi.org/10.1073/pnas.1818400116.
- 106. Ortiz-Álvarez R, Fierer N, de los Ríos A, Casamayor EO, Barberán A. 2018. Consistent changes in the taxonomic structure and functional attributes of bacterial communities during primary succession. ISME J 12:1658–1667. https://doi.org/10.1038/s41396-018-0076-2.
- Nemergut DR, Anderson SP, Cleveland CC, Martin AP, Miller AE, Seimon A, Schmidt SK. 2007. Microbial community succession in an unvegetated, recently deglaciated soil. Microb Ecol 53:110–122. https://doi .org/10.1007/s00248-006-9144-7.
- Carini P, Marsden PJ, Leff JW, Morgan EE, Strickland MS, Fierer N. 2016. Relic DNA is abundant in soil and obscures estimates of soil microbial diversity. Nat Microbiol 2:16242. https://doi.org/10.1038/nmicrobiol .2016.242.
- 109. Carini P, Delgado-Baquerizo M, Hinckley E-L, Brewer TE, Rue G, Vanderburgh C, McKnight D, Fierer N. 2018. Unraveling the effects of spatial variability and relic DNA on the temporal dynamics of soil microbial communities. bioRxiv https://doi.org/10.1101/402438.
- 110. van Oppen MJH, Blackall LL. 2019. Coral microbiome dynamics, functions and design in a changing world. Nat Rev Microbiol 17:557–567. https://doi.org/10.1038/s41579-019-0223-4.
- Marian M, Shimizu M. 2019. Improving performance of microbial biocontrol agents against plant diseases. J Gen Plant Pathol 85:329–336. https://doi.org/10.1007/s10327-019-00866-6.
- Epstein HE, Smith HA, Torda G, Oppen MJ. 2019. Microbiome engineering: enhancing climate resilience in corals. Front Ecol Environ 17:100–108. https://doi.org/10.1002/fee.2001.
- Petipas RH, González JB, Palmer TM, Brody AK. 2017. Habitat-specific AMF symbioses enhance drought tolerance of a native Kenyan grass. Acta Oecologica 78:71–78. https://doi.org/10.1016/j.actao.2016.12.005.
- Solé RV, Montañez R, Duran-Nebreda S. 2015. Synthetic circuit designs for earth terraformation. Biol Direct 10:37. https://doi.org/10.1186/ s13062-015-0064-7.
- Solé RV, Montañez R, Duran-Nebreda S, Rodriguez-Amor D, Vidiella B, Sardanyés J. 2018. Population dynamics of synthetic terraformation motifs. R Soc Open Sci 5:180121. https://doi.org/10.1098/rsos.180121.



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