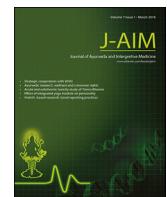




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# Journal of Ayurveda and Integrative Medicine

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## Review Article

# South African medicinal plants displaying angiotensin-converting enzyme inhibition: Potential use in the management of preeclampsia



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## ARTICLE INFO

### Article history:

Received 9 July 2021

Received in revised form

1 March 2022

Accepted 4 March 2022

Available online xxx

### Keywords:

Angiotensin-converting enzyme

Hypertension in pregnancy

Preeclampsia

Pregnancy

Phytotherapy

## ABSTRACT

In resource-limited settings, such as South Africa, hypertensive disorders of pregnancy such as preeclampsia, is the most common direct cause of maternal deaths. Current management strategies of preeclampsia primarily involve the use of pharmaceutical drugs, which are frequently associated with undesirable side-effects. Moreover, these drugs are often not easily accessible due to financial and economic constraints. Consequently, many patients rely on traditional medicine obtained from medicinal plants to manage health-related conditions.

Angiotensin-converting enzyme inhibitors are widely used drugs for the management of preeclampsia. This narrative review aims to highlight the use of indigenous medicinal plants from South Africa with Angiotensin-converting enzyme inhibitory activity whilst also evaluating their potential use in the treatment of hypertension in pregnancy. This information will influence traditional healers and sangomas in their patient management. Furthermore, the antihypertensive potential of these plants will be unraveled thus facilitating the development of new naturally occurring pharmaceutical products to reduce maternal and neonatal mortality and morbidity.

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## 1. Introduction

Globally, hypertensive disorders of pregnancy (HDP) such as chronic hypertension, gestational hypertension, preeclampsia, severe preeclampsia, eclampsia and the HELLP syndrome (haemolysis, elevated liver enzymes and low platelet levels) are a major cause of maternal and perinatal morbidity and mortality. In 2019, Rana et al. (2019) reported that preeclampsia (PE) accounts for >70,000 maternal and >500,000 foetal deaths each year [1]. PE affects up to 8% of pregnancies worldwide [2], with a higher prevalence in low and middle-income countries (LMICs) [3]. Complications associated with PE are common in resource-limited

settings such as South Africa (SA) [4]. Despite a decline in the triennium, HDP remains the second cause of maternal deaths, which emanate from patients attending antenatal clinics during the late stages of pregnancy, poor antenatal care such as failure to detect women at risk of PE, and inadequate emergency transport services to transfer women with complications associated with this disorder timeously [4].

Preeclampsia, a pregnancy-specific disorder, is defined as new-onset hypertension (systolic blood pressure  $\geq 140$  mmHg or diastolic blood pressure  $\geq 90$  mmHg) coupled with one or more of the following conditions: with/without proteinuria (urinary protein  $\geq 300$  mg per 24 h), maternal organ dysfunction, liver, and renal injury or foetal growth restriction; such clinical characteristics are usually detected at or after 20 weeks of gestation [5]. Clinical evidence suggests that hypertension is the most severe symptom influencing maternal and neonatal health in PE [6]. Whilst the exact aetiology of PE remains unknown, its pathogenesis is most likely

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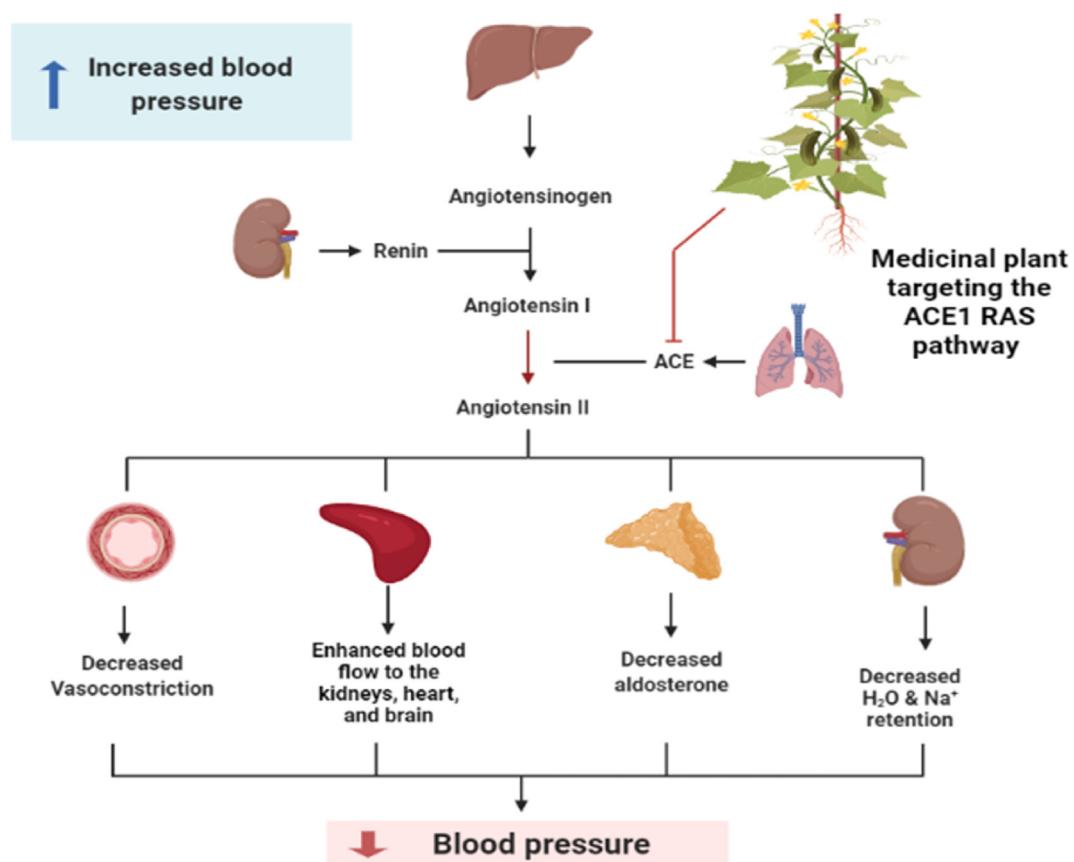
Peer review under responsibility of Transdisciplinary University, Bangalore.

dependent on the complex interaction among increased angiotensin II activity, endothelial dysfunction, neurovascular anomalies, and excessive vasoconstriction [7]. Therefore, the management of hypertension in PE involves pharmaceutical agents that target specific physiological mechanisms involved in blood pressure regulation, such as the renin-angiotensin-aldosterone system (RAAS) [8]. To-date, angiotensin-converting enzyme inhibitors (ACEI's) have proven to be valuable in the management of hypertensive disorders. ACEI's interfere with RAAS by inhibiting angiotensin II production and thereby stimulating blood vessel dilation that causes a decrease in blood pressure (Fig. 1) [9]. Notably, this inhibitory effect increases sodium and urine excretion, reduces resistance in renal blood vessels, increases venous capacity, whilst decreasing cardiac output [9].

## 2. Issues related to the current pharmaceutical management of hypertensive disorders during pregnancy

In addition to synthetic antihypertensive agents such as diuretics, beta-blockers, calcium channel blockers, ACEI's are recommended for the management of hypertension in pregnancy (Table 1). Nonetheless, the use of ACEI's is costly, thus inaccessible to patients from lower socioeconomic backgrounds [10]. Other limitations include a reduced efficacy with prolonged use, various adverse side effects, and teratogenic effects if used during the last two trimesters of pregnancy (Table 1) [5,10,11].

Clinically in South Africa, ACEI's and beta-blockers are the most commonly prescribed drugs in the management of HDP, especially PE [23]. Despite the frequent prescribing of ACEIs for the treatment of HDP, evidence suggests that synthetic ACEIs are contraindicated during the second and third trimesters as a result of suspected fetopathy [24]. However, data pertaining to the consequences observed during first-trimester exposure in pregnant women with chronic hypertension is poorly described, resulting in conflicting opinions regarding their safety [24–28]. For example, Cooper et al. (2006) suggests that the use of ACEIs may be associated with possible teratogenicity, since an increased risk of foetal cardiac valve and central nervous system defects was shown after first trimester ACEI exposure [26] in contrast to no reported teratogenic risk by others [29,30]. Likewise, a systematic analysis of published cases involving intrauterine exposure to ACEI's, highlights that most complications are less frequent in first trimester exposure compared to exposure during the second and third trimesters or throughout gestation [26,31,32]. This may be attributed to the possibility that Angiotensin II is responsible for foetal kidney development towards the end of pregnancy rather than early foetal development [32]. Moreover, prenatal renal development is dependent on a fully functional RAAS, hence it is possible that abnormalities arising after *in-utero* ACEI exposure may be due to the drug itself or underlying maternal ailments [31]. Antenatal screening of underlying complications prior to administration of ACEI's should be a perquisite in pregnant women predisposed to



**Fig. 1. Diagrammatic representation of the effect of medicinal plants on the angiotensin-converting enzyme I renin-angiotensin system (ACE1 RAS) pathway.** Renin is produced by the kidneys in response to low blood volume, low sodium, or high potassium levels. Renin's primary substrate is angiotensinogen, which is produced in the liver. Renin catalyzes the cleavage of circulating angiotensinogen, resulting in angiotensin I. Angiotensin II stimulates the secretion of aldosterone and is involved in sodium retention. The retention of water and sodium causes an increase in blood volume and thus blood pressure. ACEIs such as medicinal plants inhibit the action of ACE, reducing the conversion of angiotensin I to angiotensin II. Muscle contraction around blood vessels is reduced, successfully dilating vessels, and lowering blood pressure. Aldosterone levels drop, as does water/sodium reabsorption, lowering blood pressure. Image created with BioRender.com.

**Table 1**

Antihypertensive drugs for hypertension treatment during pregnancy.

Class of drugs	Candidate example	Side effects	References
Central $\alpha$ -agonist $\alpha$ -/ $\beta$ -blocker	Methyldopa Labetalol	Sedation and impaired sleep patterns Fatigue and bronchospasm Fetal bradycardia Neonatal hypoglycemia Decreased uteroplacental flow	[12,13] [14–16]
Calcium channel blockers Diuretics	Nifedipine Furosemide	Circulatory collapse and neuromuscular blockade Vascular volume contraction causes further elevations of blood pressure in PE women	[17] [18]
Vasodilator	Hydralazine	Hypotension Oliguria Fetal distress Lupus-like syndrome Peripheral neuropathy Renal dysgenesis Pulmonary hypoplasia Neonatal anuric renal failure Fetal death	[19]
ACEI's and angiotensin receptor blockers	Captopril/Atenolol		[20–22]

hypertension. Of note, babies delivered by pregnant women managed with captopril, a drug with a short elimination half-life, throughout pregnancy or towards the end of the pregnancy, revealed no neonatal complications in 95% of these babies [31]. Thus, during pregnancy, ACEIs should be discontinued early in the first trimester to prevent potential harmful effects associated with late pregnancy exposure [25]. Based on the conflicting evidence and side effects associated with the use of synthetic antihypertensive drugs, there is an urgent need for safer, more effective, and less expensive treatment alternatives with minimal or no side effects for the management of HDP. Modern medicine includes the use of several drugs that are derived from medicinal plants [33]. Medicinal plants are accessible in low-income environments [34] and represent a valuable source in the development of new therapeutic compounds.

### 2.1. Medicinal plants for the treatment of pregnancy-related hypertension

Phytotherapy is a global tradition that involves the use of plants to preserve the general well-being of an individual. Herbal therapy is utilized by 60% of the population of high-income countries and 80% of LMICs, including SA [34]. Over the last decade, there has been an exponential increase in the use of herbal alternative therapies across the world [35]. South Africa is home to 9% of the world's higher plants [36] and over 3000 of these plants are currently utilized in the treatment and management of numerous illnesses by several traditional healers [37]. It is worth noting that over 60–80% of the SA population from rural areas, make use of medicinal herbs for their primary healthcare needs [34]. Many rural communities prefer to use medicinal plants and their products over synthetic commercial drugs for reasons such as ease of accessibility and economic affordability [38]. Notwithstanding, traditional medicines are more acceptable from a cultural and spiritual standpoint, hence most ethnic groups are dependent on their healing capacity as their primary source of health care [39].

In SA, more than 100 plant species have been documented to treat hypertension traditionally [34]. However, only thirty-five are endemic to South Africa and evaluated for their *in vitro* ACE inhibitory activity [40,41]. Of the thirty-five, only twelve displayed strong ACE inhibitory potential *in vitro* (greater than 50% ACE inhibition) (Fig. 2). Despite their scientific value and benefits against a variety of cons/infections, the mechanism of action of only a few plants has been investigated in pre-clinical trials for hypotensive and antihypertensive efficacy. This narrative review highlights

medicinal plants with ACE inhibitory activity and their potential use in the treatment of hypertension associated with preeclampsia, in an attempt to reduce maternal and neonatal mortality and morbidity in low resource environments such as SA.

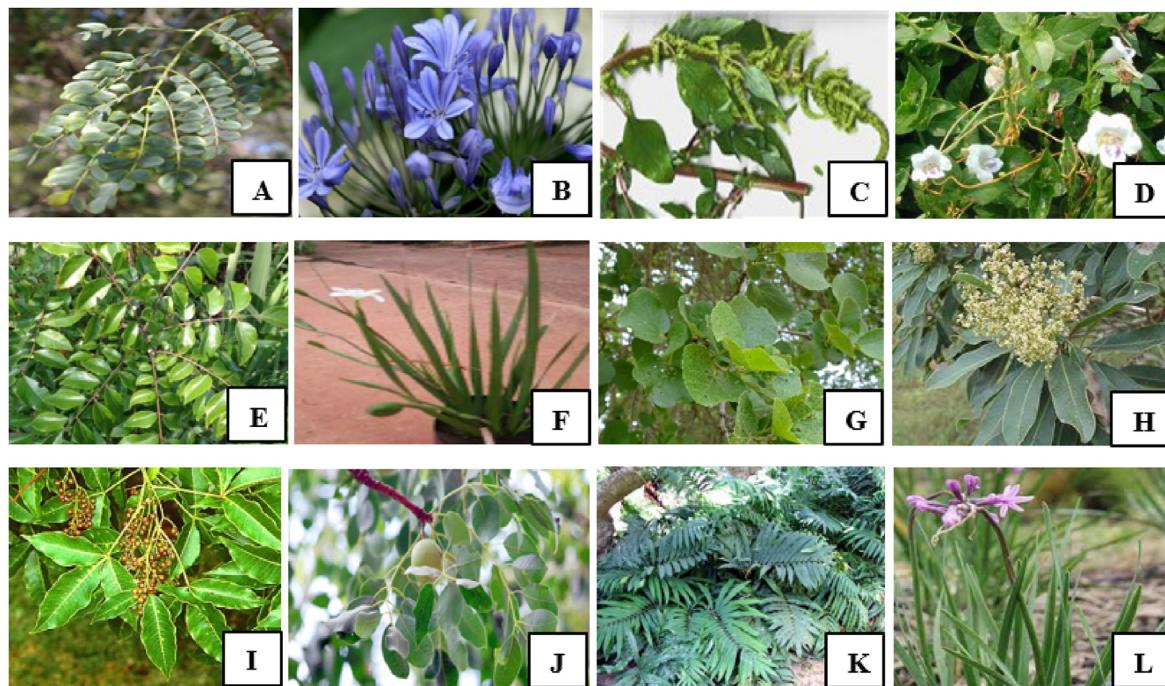
### 2.2. Medicinal plants targeting the ACE1 RAS pathway for the potential management of preeclampsia

Medicinal plants (together with their common names, location, traditional uses, and bioactive compounds) currently in use, that potentially lower blood pressure by modifying the ACE 1 RAS pathway is summarized in Table 2. Plants that are considered to have potential antihypertensive properties are required to inhibit the ACE enzyme (and the subsequent conversion of angiotensin I to angiotensin II) by more than 50%. These medicinal plants are widespread throughout SA. Different morphological parts of the plant are used for treatment, with the leaves mainly being used, which is in line with accepted protocols for plant conservation and sustainable use [34]. Most plant extracts isolated use polar solvents such as water, methanol, and ethanol and have high ACE inhibitory activity.

Phytochemical screening of 12 selected plants indicates secondary metabolites such as flavonoids, saponins, and alkaloids (Fig. 3). Additionally, the detection of saponins, tannins, flavonoids, and alkaloids in the plants that demonstrated antihypertensive effects proposes a link between these classes of compounds with ACE inhibition.

#### 2.2.1. *Adenopodia spicata*

The *in vitro* inhibition of ACE by *Adenopodia spicata* aqueous and ethanolic leaf extracts [40] were 97% and 72%, respectively but were not significant (8%) when using root extracts [40]. The phytochemical analysis confirmed flavonoids and saponins as the plant's main bioactive components [54]. While saponins isolated from *A. spicata* has not been researched for their antihypertensive effect, oral administration for five days of saponin isolated from the leaves of *Camellia sinensis* to spontaneously hypertensive rats showed a time-dependent decrease in blood pressure and mean blood pressure [55]. A single administration of saponin also showed a long-lasting hypotensive effect in these rats [55]. Likewise, partially purified soybean saponin also significantly reduced blood pressure in spontaneously hypertensive rats [56]. This data suggests that the saponin found in *A. spicata* may have antihypertensive potential, however, clinical trials are required to verify its therapeutic potential.



**Fig. 2. South African medicinal plants with promising ACE inhibitory activity and their traditional uses.** **A:** *Adenopodia spicata* (Chest or breast pain, syphilis, hypertension) **B:** *Agapanthus africanus* (Chest pains, coughs, heart disease) **C:** *Amaranthus dubius* (Kidney problems, anemia, fever, hemorrhage, stomach problems, hypertension) **D:** *Asystasia gangetica* (Asthma) **E:** *Clausena anisata* (Heart disease, tapeworms, fever, liver disease) **F:** *Dites iridioides* (Dysentery, hypertension) **G:** *Dombeya rotundifolia* (Heart problems, ulcers, stomach problems, fever, nausea, diarrhea) **H:** *Protorhus longifolia* (HBP, heartburn, internal bleeding, diarrhea, dysentery) **I:** *Rhus chirindensis* (Measles, cough, chest pain, syphilis, convulsions, epilepsy, HBP) **J:** *Sclerocarya birrea* (Dysentery, diarrhea, rheumatism, malaria, hemorrhoids) **K:** *Stangeria eriopus* (Headaches, internal parasites, HTN) **L:** *Tulbaghia violacea* (Sinus conditions, headaches, cough, colds, asthma, tuberculosis, intestinal worms and hypertension).

### 2.2.2. *Asystasia gangetica*

The *in vitro* inhibition of ACE by *Asystasia gangetica* aqueous and methanolic leaf extracts were 20% and 51%, respectively [41]. Furthermore, an *in-vivo* study on the effects of the aqueous leaf extract of *A. gangetica* (200 mg/kg) on blood pressure and heart rate of spontaneously hypertensive rats [57] significantly reduces the systolic, diastolic, and mean arterial blood pressure [57]. This reduction in systolic, diastolic, and mean arterial blood pressure produced by co-infusion with angiotensin I can be attributed to *A. gangetica* inhibiting the conversion of angiotensin I to angiotensin II, a potent vasoconstrictor [58].

### 2.2.3. *Clausena anisata*

*Clausena anisata* is a medicinal plant indigenous to Southern Africa [59]. The *in vitro* inhibition of ACE by *C. anisata* aqueous [40] and ethanolic leaf extracts were 54% and 1%, respectively [40]. An *in-vivo* study that administered bolus injections of the aqueous leaf extract of the plant (400 mg/kg/bw) to spontaneously hypertensive rats significantly reduced aortic blood pressure [59]. The report further demonstrated that the same amount of extract added daily to the drinking water of spontaneously hypertensive rats significantly decreases systolic, diastolic, and mean arterial blood pressure after 40 days of treatment [59]. The bioactive phytomolecules in the plant are carbozole alkaloids and coumarins [40,60], in which coumarins (Fig. 3) demonstrates antihypertensive properties [61]. *C. anisata* extracts were shown to significantly reduce the blood pressure of hypertensive rats, most likely by reducing the angiotensin II levels, which act via the ACE inhibitory mechanism [40]. This report delivers an essential basis for further studies into the isolation and characterization of active biomolecules that might be responsible for lowering blood pressure.

### 2.2.4. *Dites iridioides*

Infusions of *Dites iridioides* made from the inner part of its rhizomes are taken orally or in enemas, are utilized in childbirth and to treat hypertension [62]. The *in vitro*, antihypertensive impact of the leaf extracts of *D. iridioides* show 80% and 7% ACE inhibition using water and ethanol, respectively [40] in comparison to their roots which have low inhibition (13%) [40]. A study conducted on the cardiovascular effects of the leaf extract of *D. iridioides* (400 mg/kg/bw) in spontaneously hypertensive rats demonstrated significant decline of short- and long-term blood pressure (systolic, diastolic and mean arterial pressure parameters) within 20 min for 20 days, using bolus injections of the plant [63]. In addition, following the administration of the plant extract to the rats, an increase in plasma nitric oxide levels was noted indicating that the vasodilatory nitric oxide may be responsible for the decrease in aortic blood pressure [64]. Furthermore, a study that compared the blood pressure effects of *D. iridioides* with a combination of *D. iridioides* and perindopril (a known angiotensin-converting enzyme inhibitor) demonstrated a remarkable decrease in aortic blood pressure compared to both treatments alone [63].

### 2.2.5. *Sclerocarya birrea*

An *in vitro* study by Ojewole (2006) found that an aqueous extract of *Sclerocarya birrea* stem bark induced concentration-dependent relaxations of endothelium-intact rat isolated aortic rings precontracted with noradrenaline. Bolus intravenous administrations of the stem-bark extract (25–400 mg/kg) significantly decreases systemic arterial blood pressure and heart rate in anesthetized normotensive and hypertensive Dahl salt-sensitive rats [65]. Acute intravenous administration of *S. birrea* crude stem bark extract (120 mg/kg) to non-diabetic and streptozotocin-

**Table 2**

South African medicinal plants that have ACE inhibitory potential.

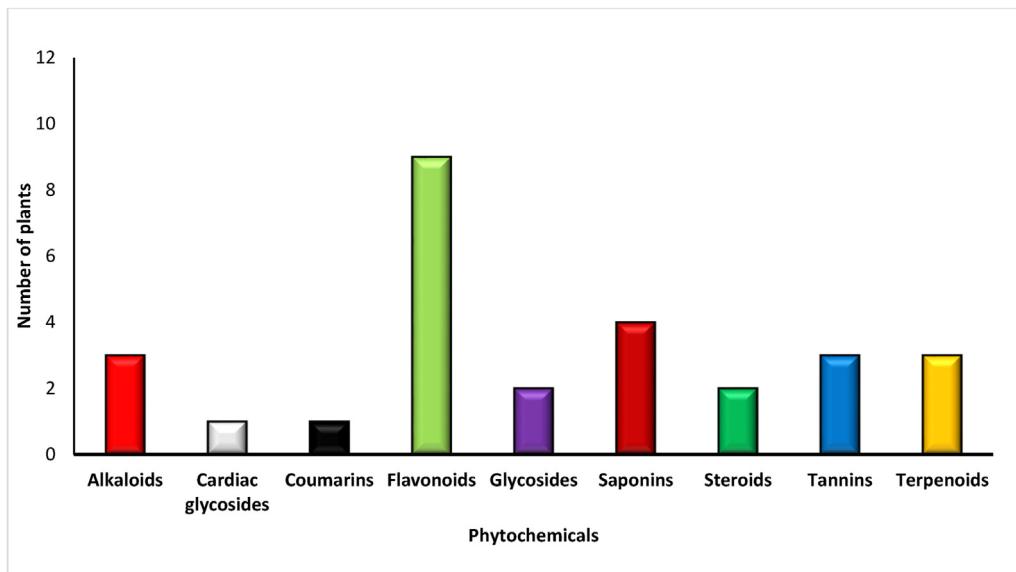
Plant Species (Family)	English Name/ (Traditional Name)	Traditional uses	Location	Phytochemicals	References
<i>Adenopodia spicata</i> (Fabaceae)	Spiny splinter bean (Ubobo)	Bark - chest or breast pain, syphilis, hypertension	Southern Africa	Flavonoids, Saponins	[40]
<i>Agapanthus africanus</i> (Amaryllidaceae)	African lily (Ubani)	Leaves and roots - chest pains, coughs, heart disease, ease labor	South Africa	Flavonoids, sitosterol, yuccagenin, agapanthagenin, spirostan sapogenins	[40]
<i>Amaranthus dubius</i> (Amaranthaceae)	Wild spinach (Imbuya)	Leaves - kidney problems, anemia, fever, hemorrhage, stomach problems, hypertension	Found worldwide	Flavonoids, Niacin, thiamine, riboflavin, ascorbic acid, hydrocyanic acid, oxalic acid	[41,42]
<i>Asystasia gangetica</i> (Acanthaceae)	Creeping foxglove (Isihobo)	Leaves – asthma	Tropics	Flavonoids, Alkaloids, terpenes, phenols, salidroside, apigenin, ajugol, megastigmane glucoside, benzyl-β-D-glucopyranoside, cardiac glycosides, tannins	[41,43]
<i>Clausena anisata</i> (Rutaceae)	Horsewood (Umnikambhiba)	Leaves and roots - heart disease, tapeworms, fever, liver disease	Africa	Not reported	[40,44]
<i>Dietes iridioides</i> (Iridaceae)	African iris (Isishuphe somfula)	Leaves, roots, and rhizomes - dysentery, hypertension	Sub-Saharan Africa	Flavonoids	[40,45]
<i>Dombeya rotundifolia</i> (Malvaceae)	Wild pear (iNhлизинонхulu)	Leaves and roots - heart problems, ulcers, stomach problems, fever, nausea, diarrhea	Southern Africa and northwards to central and eastern tropical Africa	Saponins, tannins and cardiac glycosides	[40,46]
<i>Protorhus longifolia</i> (Anacardiaceae)	Red beech (Uzintlwá)	Bark and leaves - HBP, heartburn, internal bleeding, diarrhea, dysentery	South Africa, Swaziland	Flavonoids, glycosides and sterols	[40,47]
<i>Rhus chirindensis</i> (Anacardiaceae)	Red currant (Umhlabamvudu)	All plant parts - measles, cough, chest pain, syphilis, convulsions, epilepsy, HBP	KwaZulu/Natal, Swaziland, Zimbabwe, and Mozambique	Flavonoids	[40,48]
<i>Sclerocarya birrea</i> (Anacardiaceae)	Marula (Ukanyi)	Bark, leaves, and stems - dysentery, diarrhea, rheumatism, malaria, hemorrhoids.	North-eastern South Africa and parts of eastern Botswana.	Polyphenols, tannins, flavonoids, alkaloids, anthocyanins, and saponosides coumarins, triterpenoids, and phytosterols ( $\beta$ -sitosterol) Quercetin, kaempferol, gallic acid, (-)-epicatechin 3-O-galloyl ester, (-)-epigallocatechin 3-O-galloyl ester	[40,49]
<i>Stangeria eriopus</i> (Zamiaceae)	Natal Grass Cycad (Umfigwani)	Root and leaves - headaches, internal parasites, HTN.	East coast of South Africa and southern Mozambique	Alkaloids, amino acids, biflavones, fatty acids, glycosides, polyphenols, saponins, and tannins	[40,50]
<i>Tulbaghia violacea</i> (Alliaceae)	Garlic (isihaqa)	Rhizome, bulb, leaves and roots- sinus conditions, headaches, cough, colds, asthma, tuberculosis, intestinal worms and hypertension	Eastern Cape, Limpopo and KwaZulu-Natal	Bioflavonoids, steroid saponins	[51–53]

treated diabetic rats resulted in momentary vasodepressive effects, with maximal activity occurring within 60 min of the extract's infusion. Long-term administration of the plant's stem-bark extract (120 mg/kg for 5 weeks) resulted in a significant reduction in mean arterial blood pressure. Compared to the control group, blood pressure was lower over the course of the 5-week study [66]. Furthermore, in a study conducted by Masoko et al. (2008), dichloromethane, hexane and acetone *S. birrea* stem, bark and leaves displayed strong antioxidant activity [67]. Polyphenols such as galloylate catechins contributed significantly to the antioxidant activity of *S. birrea* [68].

#### 2.2.6. *Tulbaghia violacea*

The ACE inhibitory potential of *Tulbaghia violacea* was demonstrated *in vitro* using aqueous (68%) and methanolic (71%) leaf

extracts [40,41,69]. The aqueous extracts of the leaf and bark also exhibited an ACE inhibitory potential of 72% and 49%, respectively, whilst the ethanolic leaf extracts inhibited the activity of ACE by 61% [40]. Analyses of the methanolic leaf extract tested at varying concentrations accentuated the reduction in systolic, diastolic, and mean arterial pressure of normotensive and spontaneously hypertensive rats in a dose-dependent manner, underpinning its antihypertensive impact. The reduction in blood pressure may be stimulated by the plant's actions on the ACE and  $\beta$ -adrenoceptors. Furthermore, *T. violacea* decreased systolic blood pressure in Dahl salt-sensitive rats by reducing renal angiotensin II type 1 receptor gene expression [69]. Antioxidant studies of the extracts reveal potent antioxidant activity with low IC<sub>50</sub> values [53,70]. In addition, a two-week co-treatment with the *T. violacea* extracts significantly decreased elevated thiobarbituric reacting substance (TBARS) and



**Fig. 3.** Distribution of reported secondary metabolites in the plants displaying ACE inhibitory potential.

reversed endothelial dysfunction and tissue antioxidant enzyme activity to near normal concentration [71]. The activity of serum markers of liver and kidney damage in extract-treated groups were significantly reduced, confirming this protective effect [71]. Treatment with the extract also decreased liver TBARS levels, improved liver superoxide dismutase, catalase, and glutathione peroxidase, and increased plasma nitric oxide concentrations in rats, supporting the antioxidant and hepatoprotective effects [72]. Additionally, the *in vitro* antioxidant activities of the plant extract validates its

use in preventing oxidative stress and, thereby, concomitant disorders such as hypertension [53].

### 2.3. Concerns regarding traditional medicines

The widespread accessibility and use of herbal medicines, potential herbal toxicity and herb–drug interactions are major global concerns; particularly the lack of scientific evidence with regards to efficacy and/or safety is worrying. Medicinal plants comprise a

**Table 3**  
Toxicity studies of some South African medicinal plants.

Plant	Acute/sub-chronic toxicity	Model of experimentation	Parts of plant/solvent used	Result	References
<i>Asystasia gangetica</i>	Acute toxicity	A single dose of 1000, 2000 and 5000 mg/kg of the extract were administered orally to male and female Wistar rats	Whole plant/methanolic extract	No mortality reported nor was there any sign of toxicity after 24 h and for 14 days thereafter.	[74]
<i>Clausena anisata</i>	Acute toxicity	A single dose of between 500 and 5000 mg/kg body weight was administered orally to male Swiss mice	Leaves/hexane extract	No mortality observed within 48 h. Physical signs observed (Decreased motor activity, respiration and feeding, closure of eyes)	[75]
			Leaves/Chloroform extract	Doses of 5000 and 2811 mg/kg produced 60% and 40% mortality, respectively, within 48 h. Oral LD <sub>50</sub> of the extract was calculated to be 4166.7 mg/kg.	[75]
<i>Sclerocarya birrea</i>	Acute toxicity	A single dose of 3000 mg/kg body weight was administered to male and female albino rats.	Kernel/aqueous extract	No sign of toxicity or mortality observed within 48 h.	[76]
	Sub-chronic toxicity	Animals were orally administered with doses of 1000, 2000, 3000 and 4000 mg/kg body weight of the extract once daily for 28 days.		Doses of 3000 and 4000 mg/kg/day revealed liver and kidney abnormalities	[76]
<i>Tulbaghia violacea</i>	Acute toxicity	A single dose of 5/kg body weight was administered orally to male and female Wistar rats	Rhizomes/methanolic extract	No mortality observed and no indication of toxicity, behavioural or physiological changes.	[77]
	Sub-chronic toxicity	Animals were orally administered with doses of 125, 250 and 500 mg/kg daily for 28 days		No mortality observed and no signs of toxicity reported	[77]

complex mixture of approximately 400 or more chemicals in comparison to synthetic drugs which are typically made up of a single chemical [73]. It is relatively simple to determine the activity and side effects of a single chemical, however, it is increasingly difficult to record the composite interactions and synergies occurring amongst the several chemicals found in a plant, or crude plant extract that is traditionally used. Toxicological issues linked with the use of traditional medicines are associated with serious adverse events including cardiovascular issues, liver toxicity or malfunction, hematologic, renal toxicity, and fatality (Table 3). The low frequency of adverse reports associated with traditional medicine in developing countries could be because consumers commonly esteem them as safe and thus assume their symptoms are unrelated to their use.

Therefore, it is imperative that more evidence-based studies demonstrating the efficacy of traditional medicine is conducted. Furthermore, medicinal plants should be phytochemically characterized to identify their bioactive compounds since other compounds present in crude extracts may cause unfavorable side effects. Albeit the phytochemical analysis should be complemented by studies on the mechanism of action and the toxicological profile of the medicinal plants. Additionally, there is a paucity of data on the toxicity of medicinal plants in SA during pregnancy. This warrants further toxicology studies that profile the potential use of medicinal plants and pre-clinical toxicological research during pregnancy. Since PE requires close monitoring and management across the gestational period, there is a need for chronic toxicity studies.

### 3. Conclusion and future prospective

Phytotherapy continues to create broader awareness and publicity due to its therapeutic properties and negligible side effects. We provide a summary of the effects of South African medicinal plants in the ACE1 RAS pathway, as a potential treatment of HDP, such as PE. *In vitro* screening of medicinal plants requires the support of *in vivo* studies that show clear evidence of its effectiveness to progress to the pre-clinical trial stage. Systematic pharmacokinetic and pharmacodynamic *in vivo* investigations are also required to evaluate the antihypertensive potential of documented medicinal plants for managing HDP. Additionally, research-based evidence on whether ACEI rich herbs are safer than synthetic ACEI's is lacking. Hence, large scale studies are required to evaluate the safety of ACEI rich herbs compared to synthetic ACEIs in pregnancy.

### Sources of funding

The study was supported by the National Research Foundation (grant no. 107236 and 122014).

### Conflict of interest

The authors declare no conflict of interest.

### Author contributions

Conceptualization: RR, SB, RM and NG; methodology and study design, SB, RM, NG; all authors contributed to the formal analysis and investigation; original draft preparation, RR; Review and editing, SB, RM, JM, TN, and NG. All authors have read and agreed to the final version of the manuscript.

### References

- [1] Rana S, Lemoine E, Granger JP, Karumanchi SA. Preeclampsia: pathophysiology, challenges, and perspectives. *Circ Res* 2019;124(7):1094–112.
- [2] Fasanya HO, Hsiao CJ, Armstrong-Sylvester KR, Beal SG. A critical review on the use of race in understanding racial disparities in preeclampsia. *J Appl Lab Med* 2021;6(1):247–56.
- [3] Organization WH. Maternal mortality ratio 2019. Available from: <https://www.who.int/news-room/fact-sheets/detail/maternal-mortality>.
- [4] Moodley J, Fawcett S, Pattinson R, editors. 21 years of confidential enquiries into maternal deaths in South Africa: reflections on maternal death assessments. *Obstetrics and Gynaecology Forum*. House Publications; 2020.
- [5] Brown MA, Magee LA, Kenny LC, Karumanchi SA, McCarthy FP, Saito S, et al. Hypertensive disorders of pregnancy: ISSHP classification, diagnosis, and management recommendations for international practice. *Hypertension* 2018;72(1):24–43.
- [6] Stamilio DM, Beckham AJ, Boggess KA, Jelovsek JE, Venkatesh KK. Risk factors for postpartum readmission for preeclampsia or hypertension before delivery discharge among low-risk women: a case-control study. *Am J Obstet Gynecol MFM* 2021;3(3):100317.
- [7] Ueki N, Takeda S, Koya D, Kanasaki K. The relevance of the Renin-Angiotensin system in the development of drugs to combat preeclampsia. *International journal of endocrinology* 2015;2015.
- [8] Brown CM, Garovic VD. Mechanisms and management of hypertension in pregnant women. *Curr Hypertens Rep* 2011;13(5):338.
- [9] Colafella KMM, Bovée DM, Danser AJ. The renin-angiotensin-aldosterone system and its therapeutic targets. *Exp Eye Res* 2019;186:107680.
- [10] Agrawal A, Wenger NK. Hypertension during pregnancy. *Curr Hypertens Rep* 2020;22(9):1–9.
- [11] Lindheimer MD, Taler SJ, Cunningham FG. Hypertension in pregnancy. *J Am Soc Hypertens* 2008;2(6):484–94.
- [12] Baekeland F, Lundwall L. Effects of methyldopa on sleep patterns in man. *Electroencephalogr Clin Neurophysiol* 1971;31(3):269–73.
- [13] Horwitz D, Pettinger WA, Orvis H, Thomas RE, Sjoerdsma A. Effects of methyldopa in fifty hypertensive patients. *Clin Pharmacol Therapeut* 1967;8(2):224–34.
- [14] Michelson EL, Fishman WH, Lewis JE, Edwards WT, Flanigan WJ, Bloomfield SS, et al. Multicenter clinical evaluation of long-term efficacy and safety of labetalol in treatment of hypertension. *Am J Med* 1983;75(4):68–80.
- [15] Vigil-De Gracia P, Lasso M, Ruiz E, Vega-Malek JC, de Mena FT, López JC. Severe hypertension in pregnancy: hydralazine or labetalol: a randomized clinical trial. *Eur J Obstet Gynecol Reprod Biol* 2006;128(1–2):157–62.
- [16] Heida KY, Zeeman GG, Van Veen TR, Hulzebos CV. Neonatal side effects of maternal labetalol treatment in severe preeclampsia. *Early Hum Dev* 2012;88(7):503–7.
- [17] Ben-Ami M, Giladi Y, Shalev E. The combination of magnesium sulphate and nifedipine: a cause of neuromuscular blockade. *Br J Obstet Gynaecol* 1994;101(3):262–3.
- [18] Pickkers P, Dormans TP, Russel FG, Hughes AD, Thien T, Schaper N, et al. Direct vascular effects of furosemide in humans. *Circulation* 1997;96(6):1847–52.
- [19] Magee LA, Cham C, Waterman EJ, Ohlsson A, Von Dadelszen P. Hydralazine for treatment of severe hypertension in pregnancy: meta-analysis. *BMJ* 2003;327(7421):955.
- [20] Guignard J. Persist anuria in a neonate: a side effect of captopril? *Int J Pediatr Nephrol* 1981;2:133.
- [21] Krct-Jais C, Plouin PF, Tchobroutsky C, Boutroy MJ. Angiotensin-converting enzyme inhibitors during pregnancy: a survey of 22 patients given captopril and nine given enalapril. *BJOG An Int J Obstet Gynaecol* 1988;95(4):420–2.
- [22] Briggs GG, Nageotte MP. Fatal fetal outcome with the combined use of valsartan and atenolol. *Ann Pharmacother* 2001;35(7–8):859–61.
- [23] Brown CM, Garovic VD. Drug treatment of hypertension in pregnancy. *Drugs* 2014;74(3):283–96.
- [24] Karthikeyan VJ, Ferner RE, Baghdadi S, Lane DA, Lip GY, Beevers DG. Are angiotensin-converting enzyme inhibitors and angiotensin receptor blockers safe in pregnancy: a report of ninety-one pregnancies. *J Hypertens* 2011;29(2):396–9.
- [25] Bateman BT, Patorno E, Desai RJ, Seely EW, Mogun H, Dejene SZ, et al. Angiotensin-converting enzyme inhibitors and the risk of congenital malformations. *Obstet Gynecol* 2017;129(1):174.
- [26] Cooper WO, Hernandez-Diaz S, Arbogast PG, Dudley JA, Dyer S, Gideon PS, et al. Major congenital malformations after first-trimester exposure to ACE inhibitors. *N Engl J Med* 2006;354(23):2443–51.
- [27] Diav-Citrin O, Shechtman S, Halberstadt Y, Finkel-Pekarsky V, Wajnberg R, Arnon J, et al. Pregnancy outcome after in utero exposure to angiotensin converting enzyme inhibitors or angiotensin receptor blockers. *Reprod Toxicol* 2011;31(4):540–5.
- [28] Porta M, Hainer J, Jansson S-O, Malm A, Bilous R, Chaturvedi N, et al. Exposure to candesartan during the first trimester of pregnancy in type 1 diabetes: experience from the placebo-controlled Diabetic REtinopathy Candesartan Trials. *Diabetologia* 2011;54(6):1298–303.
- [29] Lennestål R, Olausson PO, Källén B. Maternal use of antihypertensive drugs in early pregnancy and delivery outcome, notably the presence of congenital heart defects in the infants. *Eur J Clin Pharmacol* 2009;65(6):615–25.

- [30] Sullivan SD, Umans JG, Ratner R. Hypertension complicating diabetic pregnancies: pathophysiology, management, and controversies. *J Clin Hypertens* 2011;13(4):275–84.
- [31] Bullo M, Tschumi S, Bucher BS, Bianchetti MG, Simonetti GD. Pregnancy outcome following exposure to angiotensin-converting enzyme inhibitors or angiotensin receptor antagonists: a systematic review. *Hypertension* 2012;60(2):444–50.
- [32] Hanssens M, Keirse M, Vankelecom F, Van Assche FA. Fetal and neonatal effects of treatment with angiotensin-converting enzyme inhibitors in pregnancy. *Obstet Gynecol* 1991;78(1):128–35.
- [33] Dar RA, Shahnawaz M, Qazi PH. General overview of medicinal plants: a review. *J Phytopharmacol* 2017;6(6):349–51.
- [34] Balogun FO, Ashafa AOT. A review of plants used in South African Traditional Medicine for the management and treatment of hypertension. *Planta Med* 2019;85(4):312–34.
- [35] Romano B, Lucariello G, Capasso R. Topical Collection “pharmacology of medicinal plants”. Multidisciplinary Digital Publishing Institute; 2021.
- [36] Van Wyk B-E, Gericke N. People's plants: a guide to useful plants of Southern Africa. Briza publications; 2000.
- [37] Van Wyk B-E, Bv Oudtshoorn, Gericke N. Medicinal plants of South Africa. Briza; 1997.
- [38] Hosseinzadeh S, Jafarikukhdan A, Hosseini A, Armand R. The application of medicinal plants in traditional and modern medicine: a review of Thymus vulgaris. *Int J Clin Med* 2015;6(9):635.
- [39] Cock I, Selesho M, Van Vuuren S. A review of the traditional use of southern African medicinal plants for the treatment of selected parasite infections affecting humans. *J Ethnopharmacol* 2018;220:250–64.
- [40] Duncan AC, Jäger AK, van Staden J. Screening of Zulu medicinal plants for angiotensin converting enzyme (ACE) inhibitors. *J Ethnopharmacol* 1999;68(1–3):63–70.
- [41] Ramesar S, Baijnath H, Govender T, Mackraj I. Angiotensin I-converting enzyme inhibitor activity of nutritive plants in KwaZulu-Natal. *J Med Food* 2008;11(2):331–6.
- [42] Sarker U, Oba S. Nutraceuticals, antioxidant pigments, and phytochemicals in the leaves of Amaranthus spinosus and Amaranthus viridis weedy species. *Sci Rep* 2019;9(1):1–10.
- [43] Gopal T, Megha G, Chamundeeswari D, Reddy CU. Phytochemical and pharmacological studies on whole plant of Asystasia gangetica. *Indian J Res Pharm Biotechnol* 2013;1(3):365.
- [44] Senthilkumar A, Venkatesalu V. Phytochemical analysis and antibacterial activity of the essential oil of Clausena anisata (Willd.) Hook. f. ex Benth. *Int J Integr Biol* 2009;5(2):116–20.
- [45] Goldblatt P, Manning JC. Two new subspecies of Dietes (Iridaceae: iridoideae), Dietes iridooides subsp. angolensis from Angola and Dietes bicolor subsp. armeniaca from eastern South Africa, with notes and range extensions for Dietes butcheriana and Dietes iridooides. *Bothalia-African Biodivers Conserv* 2015;45(1):1–6.
- [46] Reid K, Jäger A, Van Staden J. Pharmacological and phytochemical properties of Dombeya rotundifolia. *South Afr J Bot* 2001;67(2):349–53.
- [47] Mhlongo NY, Naidoo KSB, Himakar RK, Cheriti A, Govender P. Phytochemical screening, antioxidant and antimicrobial efficacy of Protorhus longifolia (Bernh. Ex C. krauss) Engl. (Anacardiaceae) seed extracts. *Curr Trends Biotechnol Pharm* 2018;12(2):128–38.
- [48] Ojewole JA. Anticonvulsant effect of Rhus chirindensis (Baker F.) (Anacardiaceae) stem-bark aqueous extract in mice. *J Ethnopharmacol* 2008;117(1):130–5.
- [49] Russo D, Miglionico R, Carmosino M, Bisaccia F, Andrade PB, Valentão P, et al. A comparative study on phytochemical profiles and biological activities of Sclerocarya birrea (A. Rich.) Hochst leaf and bark extracts. *Int J Mol Sci* 2018;19(1):186.
- [50] Meurer-Grimes B, Stevenson DW. The biflavones of the Cycadales revisited: biflavones in Stangeria eriopus, Chigua restrepoi and 32 other species of Cycadales. *Biochem Systemat Ecol* 1994;22(6):595–603.
- [51] Davids D, Gibson D, Johnson Q. Ethnobotanical survey of medicinal plants used to manage high blood pressure and type 2 diabetes mellitus in Bitterfontein, Western Cape Province, South Africa. *J Ethnopharmacol* 2016;194:755–66.
- [52] Olorunnisola O, Bradley G, Afolayan A. Ethnobotanical information on plants used for the management of cardiovascular diseases in Nkonkobe Municipality, South Africa. *J Med Plants Res* 2011;5(17):4256–60.
- [53] Takaidza S, Mtunzi F, Pillay M. Analysis of the phytochemical contents and antioxidant activities of crude extracts from Tulbaghia species. *J Tradit Chin Med* 2018;38(2):272–9.
- [54] Breyer-Brandwijk MG. In: The medicinal and poisonous plants of southern and eastern africa being an account of their medicinal and other uses, chemical composition, pharmacological effects and toxicology in man and animal. The medicinal and poisonous plants of southern and eastern africa being an account of their medicinal and other uses, chemical composition, pharmacological effects and toxicology in man and animal. Edn 2 1962.
- [55] Sagesaka-Mitane Y, Sugiura T, Miwa Y, Yamaguchi K, Kyuki K. Effect of tea-leaf saponin on blood pressure of spontaneously hypertensive rats. *Yakugaku Zasshi: J Pharm Soc Jpn* 1996;116(5):388–95.
- [56] Takahashi S, Hori K, Shinbo M, Hiwatashi K, Gotoh T, Yamada S. Isolation of human renin inhibitor from soybean: soyasaponin I is the novel human renin inhibitor in soybean. *Biosci Biotechnol Biochem* 2008;0811051130.
- [57] Mugabo P, Raji IA. Effects of aqueous leaf extract of Asystasia gangetica on the blood pressure and heart rate in male spontaneously hypertensive Wistar rats. *BMC Compl Alternative Med* 2013;13(1):1–7.
- [58] Marchesi C, Paradis P, Schiffri EL. Role of the renin–angiotensin system in vascular inflammation. *Trends Pharmacol Sci* 2008;29(7):367–74.
- [59] Lechaba NMT, Schutte PJ, Hay L, Böhmer L, Govender MM. The effects of an aqueous leaf extract of Clausena anisata (Willd.) Hook. f. ex Benth. on blood pressure, urine output, angiotensin II levels and cardiac parameters in spontaneously hypertensive rats. *J Med Plants Res* 2016;10(28):425–34.
- [60] Chakraborty A, Chowdhury B, Bhattacharyya P. Clausenol and clausenine—two carbazole alkaloids from Clausena anisata. *Phytochemistry* 1995;40(1):295–8.
- [61] Gilane A, Shaheen F, Saeed S, Bibi S, Sadiq M, Faizi S. Hypotensive action of coumarin glycosides from Daucus carota. *Phytomedicine* 2000;7(5):423–6.
- [62] Pujol J. Natur africa: jean pujol. Natural Healers Foundation; 1990.
- [63] Moagi LE, Schutte P, Steinmann C. Cardiovascular effects of dietes iridooides in spontaneous hypertensive rats (SHR). University of Limpopo (Medunsa Campus); 2013.
- [64] Wilcox CS. Oxidative stress and nitric oxide deficiency in the kidney: a critical link to hypertension? *Am J Physiol Regul Integr Comp Physiol* 2005;289(4):R913–35.
- [65] Ojewole JA. Vasorelaxant and hypotensive effects of Sclerocarya birrea (A Rich) Hochst (Anacardiaceae) stem bark aqueous extract in rats: cardiovascular topic. *Cardiovasc J South Afr* 2006;17(3):117–23.
- [66] Gondwe M, Kamadyapa D, Tufts M, Chaturgoon A, Musabayane C. Sclerocarya birrea [(A. Rich.) Hochst.] [Anacardiaceae] stem-bark ethanolic extract (SBE) modulates blood glucose, glomerular filtration rate (GFR) and mean arterial blood pressure (MAP) of STZ-induced diabetic rats. *Phytomedicine* 2008;15(9):699–709.
- [67] Masoko P, Mmushu T, Mogashoa M, Mokgongo M, Mampuru L, Howard R. In vitro evaluation of the antifungal activity of Sclerocarya birrea extracts against pathogenic yeasts. *Afr J Biotechnol* 2008;7(20).
- [68] Braca A, Politi M, Sanogo R, Sanou H, Morelli I, Pizza C, et al. Chemical composition and antioxidant activity of phenolic compounds from wild and cultivated Sclerocarya birrea (Anacardiaceae) leaves. *J Agric Food Chem* 2003;51(23):6689–95.
- [69] Mackraj I, Ramesar S. ACE inhibitor activity of nutritive plants in Kwa-Zulu Natal. Wiley Online Library; 2007.
- [70] Olorunnisola O, Bradley G, Afolayan A. Antioxidant properties and cytotoxicity evaluation of methanolic extract of dried and fresh rhizomes of Tulbaghia violacea. *Afr J Pharm Pharmacol* 2011;5(22):2490–7.
- [71] Olorunnisola OS, Bradley G, Afolayan AJ. Protective effect of Tulbaghia violacea Harv. on aortic pathology, tissue antioxidant enzymes and liver damage in diet-induced atherosclerotic rats. *Int J Mol Sci* 2012;13(10):12747–60.
- [72] Moodley K, Joseph K, Naidoo Y, Islam S, Mackraj I. Antioxidant, antidiabetic and hypolipidemic effects of Tulbaghia violacea Harv.(wild garlic) rhizome methanolic extract in a diabetic rat model. *BMC Compl Alternative Med* 2015;15(1):408.
- [73] Fugh-Berman A. Herb-drug interactions. *Lancet* 2000;355(9198):134–8.
- [74] Eriamiatoe I, Edema M, Eriamiatoe T, Okpara G. Chemical constituents and pharmacological use of asystasia gangetica (Chinese violet) as an anti-ulcer plant. *J Chem Soc Niger* 2011;45(2).
- [75] Irungu BN, Mbabu MJ, Kiboi DM, Moindi E, Kinyua J, Mwirichia RK. In vivo antimalarial and acute toxicity properties of hexane and chloroform extracts from Clausena anisata (Willd.) Benth. 2012.
- [76] Muhammad S, Hassan L, Dangoggo S, Hassan S, Umar K, Aliyu R. Acute and subchronic toxicity studies of kernel extract of Sclerocarya birrea in rats. *Sci World J* 2011;6(3):11–4.
- [77] Olorunnisola O, Bradley G, Afolayan A. Acute and sub-chronic toxicity studies of methanolic extract of Tulbaghia violacea rhizomes in Wistar rats. *Afr J Biotechnol* 2012;11(83):14934–40.