



RESEARCH ARTICLE

Open Access

# Silymarin effect on amyloid- $\beta$ plaque accumulation and gene expression of APP in an Alzheimer's disease rat model

Parichehreh Yaghmaei<sup>1\*</sup>, Katia Azarfar<sup>1</sup>, Mehrooz Dezfulian<sup>2</sup> and Azadeh Ebrahim-Habibi<sup>3,4\*</sup>

## Abstract

**Background:** The deposition of amyloid peptides is associated with Alzheimer's disease (AD). These amyloid peptides are derived from the amyloid protein precursor (APP). Silymarin, a standardized extract of milk thistle, which is currently used in liver diseases, may be effective in the inhibition of amyloid formation. However, its effect has not been assessed on APP expression.

**Results:** In this study, first, the effect of silymarin was examined on the passive avoidance learning in a rat model of AD. This model was induced by the intracerebroventricular injection of A $\beta$  peptide (A $\beta$ <sub>1-42</sub>) in Wistar rats. Rats were treated with 70 and 140 mg/kg of the extract, once a day, for 4 weeks. Memory function that was evaluated in a shuttle-cage test, showed improvement upon administration of this extract. Brain amyloid plaques had also decreased upon administration of the extract. Furthermore, APP gene expression was compared in treated and untreated groups. The result showed that silymarin was able to suppress APP expression.

**Conclusion:** Our results are in accordance with the *in vitro* tests concerning the positive anti-amyloidogenic property of the main component of silymarin, namely silibinin. We suggest that the beneficial effect of silymarin in the AD model is related to its capacity to disaggregate amyloid plaques and to suppress APP expression. Considering the limited side effects of silymarin, this compound could be of use in AD therapy.

**Keywords:** Silymarin, RT-PCR, Passive avoidance learning, Alzheimer's disease, Beta amyloid

## Background

Alzheimer's disease (AD) is a progressive neurodegenerative disorder which has been characterized by the existence of extraneuronal aggregates of amyloid  $\beta$  (A $\beta$ ) peptide as well as intraneuronal deposits of hyperphosphorylated tau [1]. It is usually assumed that A $\beta$  (abeta) aggregation in the brain is the starting trigger of a pathological cascade which ultimately leads to synaptic dysfunction and loss, neuronal death, and eventually cognitive dysfunction [2]; the process includes also oxidative stress and inflammatory response, which play their roles in neuronal dysfunction [3,4]. It is suggested that the reactive microglia and astrocytes that

surround the A $\beta$  plaques in the AD brain may release reactive oxygen species and proinflammatory molecules [5].

Various isoforms of A $\beta$  peptide with lengths varying from 14 to 42 aminoacids have been characterized which are derived from "APP", the amyloid precursor protein (APP) [6,7]. However, it is generally accepted that the main dominant forms of A $\beta$  (40 and 42 aminoacids) are the toxic species involved in AD pathophysiology, as for example, it has been shown that the presence of A $\beta$  42 in cerebrospinal fluid could be a reliable predictor of AD progression [8].

The intracerebroventricular administration of A $\beta$  peptide into rodent brain has been used as a mean to simulate AD disease, since this injection could induce histological and biochemical changes as well as oxidative damage and inflammatory responses which result into memory deficits [9,10]. With this animal model, *in vivo* studies could be performed to test potential new candidates for AD therapy.

\* Correspondence: yaghmaei\_p@srbiau.ac.ir; aehabibi@sina.tums.ac.ir

<sup>1</sup>Department of Biology, Science and Research Branch, Islamic Azad University, Tehran, Iran

<sup>3</sup>Biosensor Research Center, Endocrinology and Metabolism Molecular-Cellular Sciences Institute, Tehran University of Medical Sciences, Tehran, Iran

Full list of author information is available at the end of the article

The flavonoid silibinin (or silybin) [(2*R*,3*R*)-3,5,7-trihydroxy-2-[(2*R*,3*R*)-3-(4-hydroxy-3-methoxyphenyl)-2-(hydroxy-methyl)-2,3-dihydrobenzo[*b*][1,4]dioxin-6-yl]chroman-4-one] is the main compound of the herb milk thistle (*Silybum marianum*) extract (silymarin) [11]. This compound possess anti-inflammatory and antioxidative effects [12]. It has also been reported that silymarin has protective effects against ethanol-induced brain injury [13], and neurotoxicity induced by lipopolysaccharide (LPS) [14]. In this study, we investigated the effect of silymarin on the memory impairment induced by A $\beta$ 1-42 injection in rats. We also examined its effect on changes in APP gene expression in the rats' brain.

## Materials and methods

### Animals

Male Wistar rats weighing 250  $\pm$  300 g were housed at six per cage (42  $\times$  26 cm), 23  $\pm$  0.5°C, under a 12/12-h light/dark cycle (lights on from 8:00 AM to 8:00 PM). Animals had free access to standard pellet food and water. Behavioral experiments were carried out in a sound-attenuated room, to which the rats were habituated for at least 1 hour. All experiments were performed in accordance with to the international guidelines set out in the *Guide for the Care and Use of Laboratory Animals* (Institute of Laboratory Animal Resources, 1996) and approved by the Research and Ethics Committee of Science and Research Branch, Azad University.

### Inducing Alzheimer's disease in animals and used compound

A $\beta$ 1-42 (Sigma, St Louis, MO, USA), was dissolved in PBS and incubated at 37°C for 7 days, after which bilateral surgery and injection of A $\beta$ 1-42 peptide was done in rats hippocampus with the help of a stereotaxic apparatus (SR-6 N Narishig, Japan) [15]. Ketamine and xylazine (Alfasan, Woerden-Holland) were used to anesthetize the rats. Injection was performed slowly into the CA1 region of the hippocampus at both sides of the brain as mentioned in the Paxinos atlas [16].

Silymarin tablets, containing ethyl acetate extracted Silymarin, were purchased from Goldaru Pharmaceutical Laboratory, (Isfahan, Iran) and suspended in a distilled water solution. Treatment group was administered two different dosages of the compound (70 and 140 mg/kg/day P.O.) for 4 weeks after the injection with A $\beta$ 1-42. A volume of 0.1 ml/10 g body weight was used.

Animals were divided into four groups with n = 6: the control group did not undergo surgery, the sham group underwent surgery (AD inducing) and was given distilled water 7 days after surgery, the experimental groups Exp1 and Exp2 received 70 and 140 mg/kg/day of the compound respectively (7 days after surgery).

## Testing the treatment efficacy

### a. Passive avoidance test

A shuttle-cage consisting of two compartments of equal size (26  $\times$  26 cm) separated by a sliding door (8  $\times$  8 cm) was used. The shock compartment was dark in contrast to the starting compartment. Each experiment started with a pre-training trial, where the rat was placed in the starting compartment for 5 seconds, after which the sliding door was raised, and the rat was allowed to stay in the dark compartment for 10 seconds. The rat was then put back in its cage and stayed there for 30 minutes after which it was again put into the shuttle box, and this time, after entering the dark compartment, a footshock (50 Hz, 1 mA, and 5 s) was delivered. The rat was then put back into cage and stayed there for 120 seconds. When put back in the shuttle box, if a latency (in the order of 120 seconds) is observed before entering the dark compartment, successful acquisition of passive avoidance is recorded. A similar procedure was used 24 hours after training sessions to make a retention test for evaluating long-term memory. Higher or lower latencies are taken as indicative of increase or decrease in memory retention [17].

### b. Histological studies of brain tissue

At the end of experiment animals were decapitated under anesthesia and their brain removed for histological assessments. First, the brains were fixed in 10% formalin and later processed for embedding with paraffin, after which serial sections in 6 mm of thickness were prepared. For staining of hippocampus cells, Thioflavin-S method which is detected by fluorescent microscopy was used [18].

### c. Assessing amyloid precursor protein (APP) expression

Semi-quantitative RT-PCR was performed to assess APP expression. RNA was extracted from the homogenized brain tissue of the rats by use of RNX plus kit (CinnaGen, Iran). Isolated RNA was reverse-transcribed using the following gene-specific primers: 5'-GGA TGC GGA GTT CGG ACA TG -3' (forward) and 5'-GTT CTG CAT CTG CTC AAA G -3' (reverse). GAPDH was used as housekeeping gene as a control [5'-GACATGCCGCCTGGAGAAAC -3' (forward) and 5'-AGCCCAGGATGCCCTTTAGT -3' (reverse)]. The difference in threshold cycles between APP gene and GAPDH gives the standardized expression level. PCR were performed using the One-Step SYBR PrimeScript RT-PCR kit (Cinagen, Tehran-Iran). The reaction profile consisted of a first round at 95°C for 3 min and then 40 cycles of denaturation at 95°C for 30 s, annealing at 57°C for 30 s, and extension at 72°C for 30 s, with a final extension reaction carried out at

72°C for 10 min. The RT-PCR products were loaded onto agarose gels and the resulting bands from electrophoresis were photographed with a UV transilluminator.

#### Statistical analyses

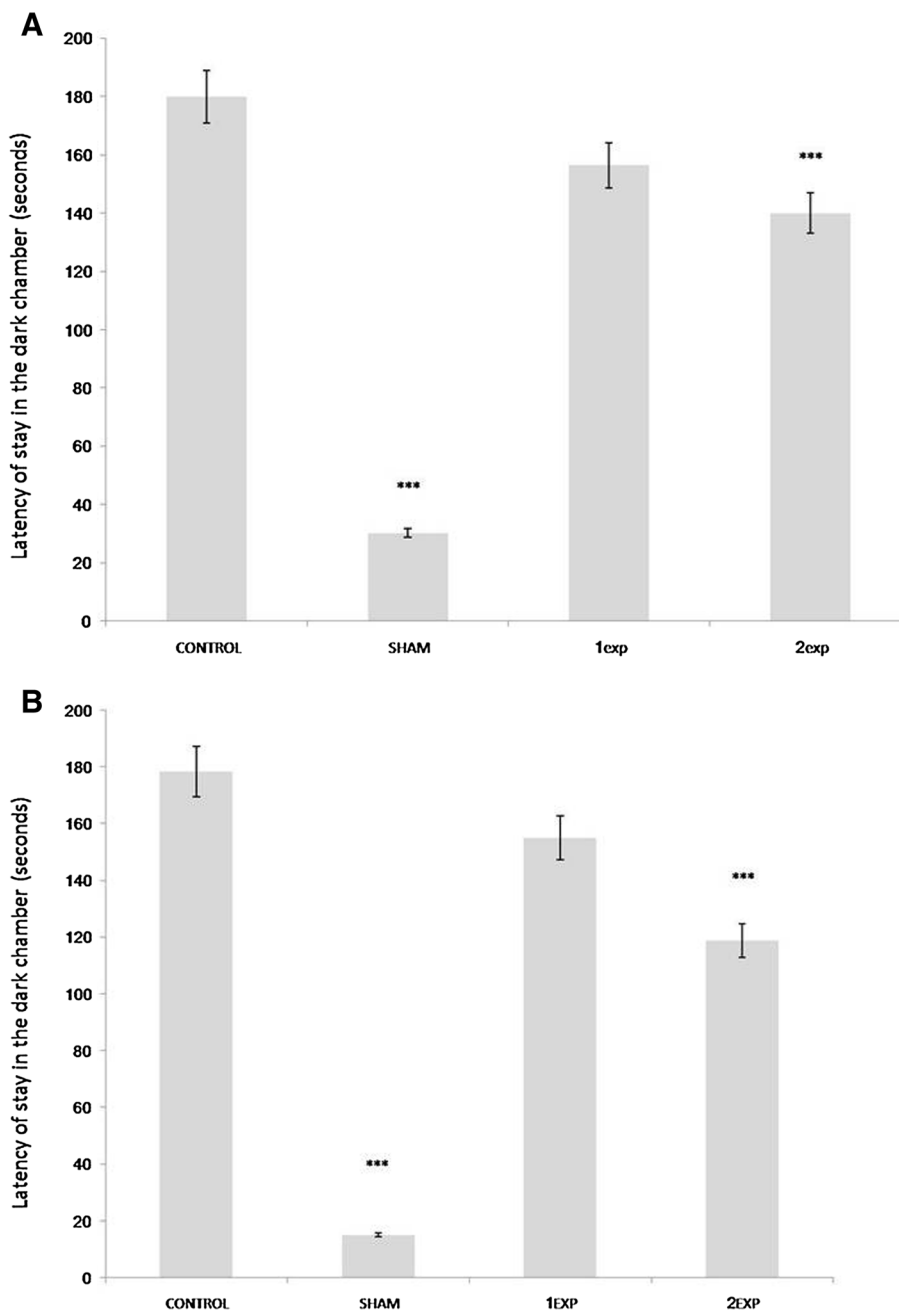
The results are expressed as the mean  $\pm$  S.E.M. Statistical significance was determined with the one-way ANOVA followed by Tukey's multiple comparisons test. A Pearson

correlation analysis was performed to elucidate the relationships.  $p < 0.05$  was taken as a significant level of difference.

#### Results

##### Passive avoidance test

Both experimental groups showed significantly longer step-through latency compared to control group ( $p < 0.05$ ) both in the training (Figure 1A) and test (Figure 1B) days.



**Figure 1** The mean latency to enter the dark chamber on the training (A) and test day (B). \*\*\* is indicative of ( $P < 0.001$ ). 1Exp group was treated with 70 mg/kg Silymarin and 2Exp group had received 140mg/kg Silymarin.

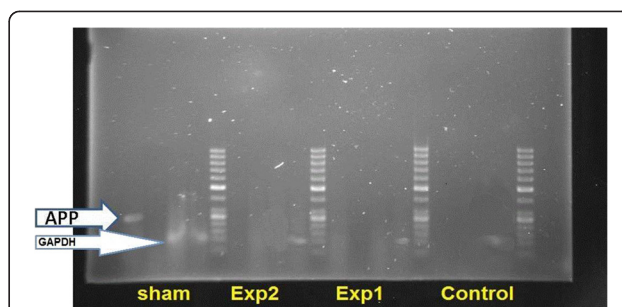
However, it is interesting to note that the 2Exp group, which had received a higher dose of silymarin showed slightly lower results in comparison with the 1Exp group. The 1Exp group has no significant difference in the step-through latency compared with the control group.

#### Histological studies of brain tissue

ThT staining was used to show the formation of plaques in the abeta-treated brain tissues. Usually, amyloid plaques could be detected in different areas of the brain, including the hippocampus and cortex (for an example of whole brain sections showing plaques distribution see Figure three A,B in reference # [19]). In the present study, both these areas were observed separately with regard to plaques concentration. The normal hippocampus and cortex tissues are showed in Figure 2A. Upon injection of abeta, plaques are formed in both tissues and observed as lighter areas (Figure 2B and C showing the cortex and hippocampus respectively). Treatment with silymarin is effective in diminishing the plaques amount, but here too, administration of 70 mg/Kg of the compound (Figure 2D and E) has a better effect than the higher used dose of 140 mg/Kg (Figure 2F and G).

#### Amyloid precursor protein (APP) expression

Results of electrophoresis are shown in Figure 3. When the housekeeping gene GAPDH is seen in all samples,

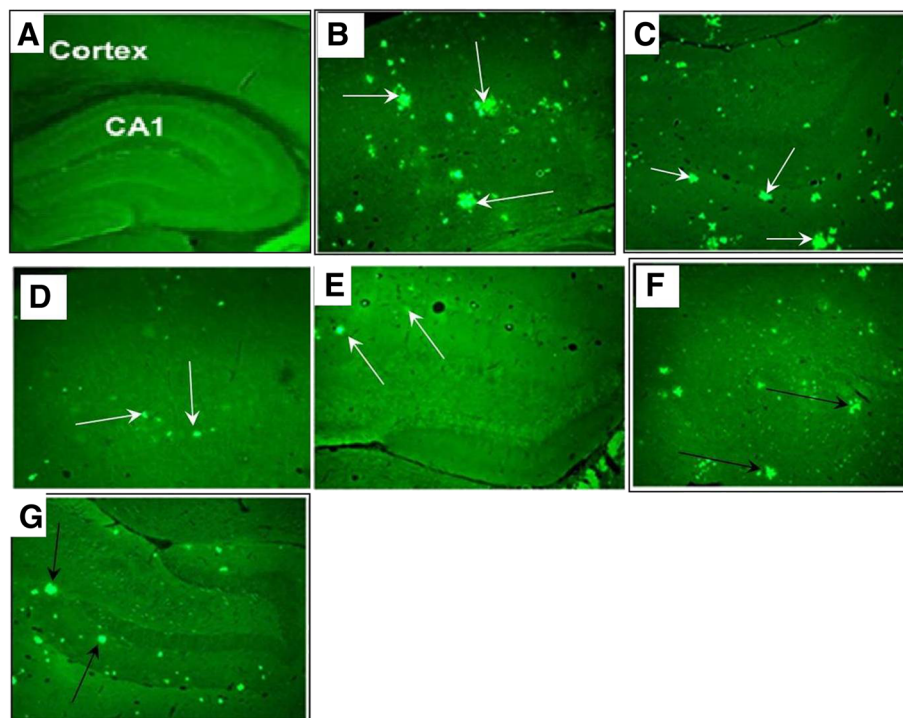


**Figure 3** RT-PCR results comparing gene expression of control, sham and experimental groups. The locations of the housekeeping gene and APP are indicated by arrows.

APP expression is detected in the sham group, which had only received the compound solvent, and is absent in both experimental groups.

#### Discussion

From the different theories that try to explain AD pathology, two have been more focused on: the cholinergic theory, which points to the decrease of acetyl choline as the main cause of the disease [20,21] and the amyloid hypothesis, which considers the aggregation of different derivatives of APP as having an important role in the pathophysiology of AD [22]. Therapeutic approaches may



**Figure 2** Brain slices from the frontal cortex & hippocampus with a magnification of 40. (A), control group, (B,C), sham group: arrows indicate positions where abeta plaques accumulation has occurred, (D,E) 1Ex group which was treated with 70 mg/kg silymarin and (F,G) 2Ex group which was treated with 140mg/kg silymarin. B,D, and E correspond to the cortex whereas C,E, and G correspond to the hippocampus.

include the design of acetyl cholinesterase inhibitors [23] as well as efforts toward finding A $\beta$  aggregation blockers [24], or compounds that would inhibit secretases, i.e. enzymes which cleave APP to A $\beta$  peptides of various length [25]. Some researchers have also tried to employ two or more therapeutic approaches simultaneously, or to find molecules that could affect more than one therapeutic target [26].

Abeta deposits are directly associated with AD. Various isoforms of abeta are derived from the precursor APP, including A $\beta$ 25–35 which is one of the most toxic species detected in the brain of AD patients [27,28]. At any rate, administration of abeta variants has led to cognitive impairment [9,29,30] which has also been observed in the present study. The hippocampus is known to be the part of the brain which is first affected by AD [31] and the result of this damage shows itself by cognitive impairment [32] that has been assessed here by the shuttle-box test.

Silymarin, the extract of *Silybum marianum* that contains flavonolignans, has been shown to possess multiple therapeutic properties, including protective effect against nerve damage and brain aging [33,34]. This extract may act via its antioxidant which could protect cells from oxidative stress damage [33], as it has been shown that silymarin effect against oxidative stress may be the result of an increase in reduced glutathione, ascorbic acid and superoxide dismutase levels [35]. Silymarin had also a neuroprotective effect in diabetic mice brains [36], and in mice treated with the brain damaging drug metamphatamine [37]. Previous studies have also reported the beneficial effect of silymarin on A $\beta$  plaques formation, and the effect was suggested to be related to microglial inflammation reduction and direct effect on A $\beta$  accumulation by blocking its aggregation in an Alzheimer's disease model of transgenic mice [19].

As a matter of fact, silibinin has the ability to inhibit the amyloid structure formation of various model or pathogenic proteins *in vitro*, including A $\beta$ <sub>1-42</sub> [38] human islet amyloid polypeptide [39], and human insulin and albumin [40]. This may be suggestive of a generic anti-amyloidogenic effect for this compound, as it has been suggested/observed for various other aromatic/polyphenolic molecules [41,42]. It should be pointed out that in the present study, what has been observed is a disaggregative effect of silymarin on A $\beta$  plaques, since the decrease of plaques was observed *after* they had been formed. This is an important fact, which is directly suggestive of a potential therapeutic effect for the compounds that are able to act on existing fibrils. The current view in the search for AD therapeutic compounds is preferably directed toward compounds that could preserve the native conformation of peptides that may form amyloid structure [43] or to focus on compounds that inhibit the formation

of intermediate structures in the course of amyloid formation [44,45]. However, it could make sense to include the compounds that have destabilizing effect on pre-formed fibrils into the spectra of potential AD therapies.

Flavonoids are able to cross the blood brain barrier [46], a fact that should also be true about silymarin, given its established neuroprotective effect [47]. Furthermore, this extract presents few side effects and drug interactions [48] which makes it a quite interesting potential drug for AD. Silymarin is already used as adjuvant therapy in liver insufficiency [49], which is another positive point for a potential drug candidate.

The beneficial effect of silymarin on the short term memory of rats is thus related with less abeta plaques, but also, as demonstrated in the present study, with a negative regulatory effect on the APP gene expression. Increased expression of APP has been detected in the ageing brain [50], and specifically related to AD pathophysiology [51]. Furthermore, the overexpression of APP that is present in the established transgenic mouse model of AD is definitely linked to the pathological characteristics of the model [52].

As far as we know, this should be a first report about the extract effect on APP, which is expanding our knowledge about the mechanism of action of silibinin in attenuating AD symptoms.

#### Competing interests

The authors declare that they have no competing interest.

#### Authors' contributions

PY has designed and supervised the project, analyzed the data and advised on writing the paper. This report is part of the results of KA MSc thesis project who performed experiments and wrote the first draft of the manuscript. MD has been an advisor to the thesis, supervised studies and analyzed data. A.E-H. advised on the project, analyzed data, and finalized the manuscript. All authors read and approved the final manuscript.

#### Author details

<sup>1</sup>Department of Biology, Science and Research Branch, Islamic Azad University, Tehran, Iran. <sup>2</sup>Department of Microbiology, College of Science, Islamic Azad University, Karaj Branch, Karaj, Iran. <sup>3</sup>Biosensor Research Center, Endocrinology and Metabolism Molecular-Cellular Sciences Institute, Tehran University of Medical Sciences, Tehran, Iran. <sup>4</sup>Endocrinology and Metabolism Research Center, Endocrinology and Metabolism Clinical Sciences Institute, Tehran University of Medical Sciences, North Kargar Avenue, 1411413137 Tehran, Iran.

Received: 18 July 2013 Accepted: 3 December 2013

Published: 24 January 2014

#### References

1. Blennow K, de Leon MJ, Zetterberg H: Alzheimer's disease. *Lancet* 2006, **368**(9533):387–403.
2. Walsh DM, Selkoe DJ: Deciphering the molecular basis of memory failure in Alzheimer's disease. *Neuron* 2004, **44**(1):181–193.
3. Butterfield DA, Reed T, Newman SF, Sultana R: Roles of amyloid beta-peptide-associated oxidative stress and brain protein modifications in the pathogenesis of Alzheimer's disease and mild cognitive impairment. *Free Radic Biol Med* 2007, **43**(5):658–677.
4. Farfara D, Lifshitz V, Frenkel D: Neuroprotective and neurotoxic properties of glial cells in the pathogenesis of Alzheimer's disease. *J Cell Mol Med* 2008, **12**(3):762–780.

5. Combs CK, Karlo JC, Kao S-C, Landreth GE: **Beta-amyloid stimulation of microglia and monocytes results in TNF alpha dependent expression of inducible nitric oxide synthase and neuronal apoptosis.** *J Neurosci* 2001, **21**(4):1179–1188.
6. Portelius E, Brinkmalm G, Tran AJ, Zetterberg H, Westman-Brinkmalm A, Blennow K: **Identification of novel APP/Abeta isoforms in human cerebrospinal fluid.** *Neurodegener Dis* 2009, **6**(3):87–94.
7. Portelius E, Mattsson N, Andreasson U, Blennow K, Zetterberg H: **Novel a isoforms in Alzheimers disease-their role in diagnosis and treatment.** *Curr Pharm Des* 2011, **17**(25):2594–2602.
8. van Harten AC, Visser PJ, Pijnenburg YA, Teunissen CE, Blankenstein MA, Scheltens P, van der Flier WM: **Cerebrospinal fluid Abeta42 is the best predictor of clinical progression in patients with subjective complaints.** *Alzheimers Dement* 2013, **9**(5):481–487.
9. Frautschy SA, Baird A, Cole GM: **Effects of injected Alzheimer beta-amyloid cores in rat brain.** *Proc Natl Acad Sci U S A* 1991, **88**(19):8362–8366.
10. Maurice T, Lockhart BP, Privat A: **Amnesia induced in mice by centrally administered beta-amyloid peptides involves cholinergic dysfunction.** *Brain Res* 1996, **706**(2):181–193.
11. Kvasnicka F, Biba B, Sevcik R, Voldrich M, Kratka J: **Analysis of the active components of silymarin.** *J Chromatogr A* 2003, **990**(1–2):239–245.
12. Kren V, Walterova D: **Silybin and silymarin—new effects and applications.** *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 2005, **149**(1):29–41.
13. la Grange L, Wang M, Watkins R, Ortiz D, Sanchez ME, Konst J, Lee C, Reyes E: **Protective effects of the flavonoid mixture, silymarin, on fetal rat brain and liver.** *J Ethnopharmacol* 1999, **65**(1):53–61.
14. Lu P, Mamiya T, Lu LL, Mouri A, Zou LB, Nagai T, Hiramatsu M, Ikejima T, Nabeshima T: **Silibinin prevents amyloid  $\beta$  peptide-induced memory impairment and oxidative stress in mice.** *Br J Pharmacol* 2009, **157**(7):1270–1277.
15. Cetin F, Dincer S: **The effect of intrahippocampal beta amyloid (1–42) peptide injection on oxidant and antioxidant status in rat brain.** *Ann N Y Acad Sci* 2007, **1100**:510–517.
16. Paxinos G, Watson C: *The rat brain in stereotaxic coordinates: hard cover edition.* The Netherlands: Academic Press, Elsevier; 2006.
17. Guaza C, Borrell J: **Prolonged ethanol consumption influences shuttle box and passive avoidance performance in rats.** *Physiol Behav* 1985, **34**(2):163–165.
18. Gandy S: **The role of cerebral amyloid beta accumulation in common forms of Alzheimer disease.** *J Clin Invest* 2005, **115**(5):1121–1129.
19. Murata N, Murakami K, Ozawa Y, Kinoshita N, Irie K, Shirasawa T, Shimizu T: **Silymarin attenuated the amyloid beta plaque burden and improved behavioral abnormalities in an Alzheimer's disease mouse model.** *Biosci Biotechnol Biochem* 2010, **74**(11):2299–2306.
20. Boller F, Forette F: **Alzheimer's disease and THA: a review of the cholinergic theory and of preliminary results.** *Biomed Pharmacother* 1989, **43**(7):487–491.
21. Robbins T, McAlonan G, Muir J, Everitt B: **Cognitive enhancers in theory and practice: studies of the cholinergic hypothesis of cognitive deficits in Alzheimer's disease.** *Behav Brain Res* 1997, **83**(1):15–23.
22. Selkoe DJ: **Toward a comprehensive theory for Alzheimer's disease. Hypothesis: Alzheimer's disease is caused by the cerebral accumulation and cytotoxicity of amyloid beta protein.** *Ann N Y Acad Sci* 2000, **924**(1):17–25.
23. Wilkinson DG, Francis PT, Schwam E, Payne-Parrish J: **Cholinesterase inhibitors used in the treatment of Alzheimer's disease.** *Drugs Aging* 2004, **21**(7):453–478.
24. Cheng B, Gong H, Xiao H, Petersen RB, Zheng L, Huang K: **Inhibiting toxic aggregation of amyloidogenic proteins: a therapeutic strategy for protein misfolding diseases.** *Biochimica et Biophysica Acta (BBA)—General Subjects* 2013, **1830**(10):4860–4871.
25. Citron M: **Beta-Secretase inhibition for the treatment of Alzheimer's disease—promise and challenge.** *Trends Pharmacol Sci* 2004, **25**(2):92–97.
26. Bajda M, Guziar N, Ignasik M, Malawska B: **Multi-Target-Directed ligands in Alzheimer's disease treatment.** *Curr Med Chem* 2011, **18**(32):4949–4975.
27. Pike CJ, Walencewicz-Wasserman AJ, Kosmoski J, Cribbs DH, Glabe CG, Cotman CW: **Structure-activity analyses of beta-amyloid peptides: contributions of the beta 25–35 region to aggregation and neurotoxicity.** *J Neurochem* 1995, **64**(1):253–265.
28. Kubo T, Nishimura S, Kumagai Y, Kaneko I: **In vivo conversion of racemized beta-amyloid ([D-Ser 26]A beta 1–40) to truncated and toxic fragments ([D-Ser 26]A beta 25–35/40) and fragment presence in the brains of Alzheimer's patients.** *J Neurosci Res* 2002, **70**(3):474–483.
29. Alkam T, Nitta A, Mizoguchi H, Itoh A, Nabeshima T: **A natural scavenger of peroxynitrites, rosmarinic acid, protects against impairment of memory induced by Abeta(25–35).** *Behav Brain Res* 2007, **180**(2):139–145.
30. Tsunekawa H, Noda Y, Mouri A, Yoneda F, Nabeshima T: **Synergistic effects of selegiline and donepezil on cognitive impairment induced by amyloid beta (25–35).** *Behav Brain Res* 2008, **190**(2):224–232.
31. Busche MA, Chen X, Henning HA, Reichwald J, Staufienbiel M, Sakmann B, Konnerth A: **Critical role of soluble amyloid beta for early hippocampal hyperactivity in a mouse model of Alzheimer's disease.** *Proc Natl Acad Sci* 2012, **109**(22):8740–8745.
32. Sabuncu MR, Desikan RS, Sepulcre J, Yeo BTT, Liu H, Schmansky NJ, Reuter M, Weiner MW, Buckner RL, Sperling RA: **The dynamics of cortical and hippocampal atrophy in Alzheimer disease.** *Arch Neurol* 2011, **68**(8):1040.
33. Raza SS, Khan MM, Ashafaq M, Ahmad A, Khuwaja G, Khan A, Siddiqui MS, Safhi MM, Islam F: **Silymarin protects neurons from oxidative stress associated damages in focal cerebral ischemia: a behavioral, biochemical and immunohistological study in Wistar rats.** *J Neural Sci* 2011, **309**(1):45–54.
34. Galhardi F, Mesquita K, Monserrat JM, Barros DM: **Effect of silymarin on biochemical parameters of oxidative stress in aged and young rat brain.** *Food Chem Toxicol* 2009, **47**(10):2655–2660.
35. Nencini C, Giorgi G, Micheli L: **Protective effect of silymarin on oxidative stress in rat brain.** *Phytomedicine* 2007, **14**(2):129–135.
36. Marrazzo G, Bosco P, la Delia F, Scapagnini G, di Giacomo C, Malaguarnera M, Galvano F, Nicolosi A, Li Volti G: **Neuroprotective effect of silibinin in diabetic mice.** *Neurosci Lett* 2011, **504**(3):252–256.
37. Lu P, Mamiya T, Lu L, Mouri A, Niwa M, Kim HC, Zou LB, Nagai T, Yamada K, Ikejima T, et al: **Silibinin attenuates cognitive deficits and decreases of dopamine and serotonin induced by repeated methamphetamine treatment.** *Behav Brain Res* 2010, **207**(2):387–393.
38. Yin F, Liu J, Ji X, Wang Y, Zidichouski J, Zhang J: **Silibinin: a novel inhibitor of Abeta aggregation.** *Neurochem Int* 2011, **58**(3):399–403.
39. Cheng B, Gong H, Li X, Sun Y, Zhang X, Chen H, Liu X, Zheng L, Huang K: **Silibinin inhibits the toxic aggregation of human islet amyloid polypeptide.** *Biochem Biophys Res Commun* 2012, **419**(3):495–499.
40. Chinisaz M, Ghasemi A, Larijani B, Ebrahim-Habibi A: **Amyloid formation and inhibition of an all-beta protein: a study on fungal polygalacturonase.** *J Mol Struct* 2014, **1059**:94–100.
41. Ono K, Yoshiike Y, Takashima A, Hasegawa K, Naiki H, Yamada M: **Potent anti-amyloidogenic and fibril-destabilizing effects of polyphenols in vitro: implications for the prevention and therapeutics of Alzheimer's disease.** *J Neurochem* 2003, **87**(1):172–181.
42. Choi Y-T, Jung C-H, Lee S-R, Bae J-H, Baek W-K, Suh M-H, Park J, Park C-W, Suh S-I: **The green tea polyphenol (–)-epigallocatechin gallate attenuates  $\beta$ -amyloid-induced neurotoxicity in cultured hippocampal neurons.** *Life Sci* 2001, **70**(5):603–614.
43. Johnson SM, Pettrassi HM, Palaninathan SK, Mohamedmohaideen NN, Purkey HE, Nichols C, Chiang KP, Walkup T, Sacchetti JC, Sharpless KB: **Bisaryloxime ethers as potent inhibitors of transthyretin amyloid fibril formation.** *J Med Chem* 2005, **48**(5):1576–1587.
44. Hafner-Bratkovič I, Gašperšič J, Šmid LM, Bresjanac M, Jerala R: **Curcumin binds to the  $\alpha$ -helical intermediate and to the amyloid form of prion protein—a new mechanism for the inhibition of PrPSc accumulation.** *J Neurochem* 2008, **104**(6):1553–1564.
45. Liu D, Xu Y, Feng Y, Liu H, Shen X, Chen K, Ma J, Jiang H: **Inhibitor discovery targeting the intermediate structure of  $\beta$ -amyloid peptide on the conformational transition pathway: implications in the aggregation mechanism of  $\beta$ -amyloid peptide.** *Biochemistry* 2006, **45**(36):10963–10972.
46. Youdim KA, Shukitt-Hale B, Joseph JA: **Flavonoids and the brain: interactions at the blood–brain barrier and their physiological effects on the central nervous system.** *Free Radic Biol Med* 2004, **37**(11):1683–1693.
47. Baluchnejadmojarada T, Roghanib M, Mafakheria M: **Neuroprotective effect of silymarin in 6-hydroxydopamine hemi-parkinsonian rat: involvement of estrogen receptors and oxidative stress.** *Neurosci Lett* 2010, **480**(3):206–210.
48. Jacobs BP, Dennehy C, Ramirez G, Sapp J, Lawrence VA: **Milk thistle for the treatment of liver disease: a systematic review and meta-analysis.** *Am J Med* 2002, **113**(6):506–515.
49. Feher J, Lengyel G: **Silymarin in the prevention and treatment of liver diseases and primary liver cancer.** *Curr Pharm Biotechnol* 2012, **13**(1):210–217.
50. Lukiw WJ: **Gene expression profiling in fetal, aged, and alzheimer hippocampus: a continuum of stress-related signaling.** *Neurochem Res* 2004, **29**(6):1287–1297.

51. Theuns J, Brouwers N, Engelborghs S, Sleegers K, Bogaerts V, Corsmit E, de Pooter T, van Duijn CM, de Deyn PP, van Broeckhoven C: **Promoter mutations that increase amyloid precursor-protein expression are associated with Alzheimer disease.** *Am J Hum Genet* 2006, **78**(6):936–946.
52. Howlett DR, Richardson JC: **The pathology of APP transgenic mice: a model of Alzheimer's disease or simply overexpression of APP?** *Histol Histopathol* 2009, **24**(1):83–100.

doi:10.1186/2008-2231-22-24

**Cite this article as:** Yaghmaei *et al.*: Silymarin effect on amyloid- $\beta$  plaque accumulation and gene expression of APP in an Alzheimer's disease rat model. *DARU Journal of Pharmaceutical Sciences* 2014 **22**:24.

**Submit your next manuscript to BioMed Central  
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

 **BioMed** Central