





## Five Complete Salmonella enterica Serotype Reading Genomes Recovered from Patients in the United States

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**ABSTRACT** Between 2018 and 2019, *Salmonella enterica* serotype Reading caused a large, multistate outbreak linked to contact with raw turkey products in the United States. Here, we provide five *Salmonella* Reading reference genomes collected from US patients between 2016 and 2018.

Salmonella enterica serotype Reading is uncommonly associated with human illness, but caused a multistate outbreak linked to contact with raw turkey products from 2018 to 2019 in the US (1). Previous phylogenetic analyses identified three Hadar clades. Clade 1 contained the "emergent" 2018 to 2019 outbreak-associated subclade, which was genetically distinct from a "contemporary" subclade of previously circulating Reading strains, partially due to the acquisition of mobile genetic elements (MGE) (2). Clade 2 primarily contained human isolates, and its pangenome was considerably smaller than those of the other clades, but further analysis to understand these differences was not performed (2). Closed sequences of Reading are required for pangenome exploration and to further understand the novel "emergent" subclade, but only 11 are currently available (3, 4), and none are from US patients. Here, we generated five complete clinical Reading sequences from Clades 1 and 2 to serve as references.

Five Reading isolates from human illnesses were chosen for long-read sequencing to represent human-associated Reading diversity before and during the outbreak: two "contemporary" Clade 1 isolates (2018), one "emergent" Clade 1 isolate (2017), and two Clade 2 isolates collected before the outbreak (2016 to 2017). Isolates originated from clinical diagnostic or public health laboratories (PHL) as part of the CDC's national passive Salmonella surveillance (https://www.cdc.gov/nationalsurveillance/salmonella-surveillance.html); thus, isolation methods vary by site (5). Serotype was confirmed in silico using SeqSero2 v0.1 (6). Genomic DNA was extracted (Wizard Genomic DNA purification kit, modified manufacturer's protocol, Promega, WI, USA) from cultures incubated on tryptic soy agar-sheep blood overnight (37°C). Libraries were prepared (Rapid Barcoding kit SQK-RBK004; manufacturer's protocol, Oxford Nanopore Technologies [ONT], Oxford, United Kingdom) and sequenced for 72 h on a GridlON sequencing platform (R9.4.1 flowcells; ONT). Reads were base-called using Guppy v4.2.2 and filtered for quality using MinKNOW (ONT). Hybrid assemblies were generated, polished, circularized, and rotated using Unicycler v0.4.8 (conservative option) (7); the corresponding Illumina short reads (previously generated at PHL through PulseNet, https://www.cdc.gov/pulsenet/) were accessed through NCBI's Short Read Archive (https:// www.ncbi.nlm.nih.gov/sra). Assemblies were quality controlled using QUAST v5.0.2 (8) and blastn v2.9.0 (9). Resistance determinants and plasmid replicons were detected using an internal workflow that employs the ResFinder database (downloaded 30JUL2020; 90% identity,

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**TABLE 1** Summary information for five Salmonella enterica serotype Reading genomes isolated from humans in the US<sup>a</sup>

	NCBI accession numbers	ımbers			Short-read		Long-read												
Strain no.	BioSample	BioSample Short read SRA GenBank		Long read SRA	Mean read length	No. of reads	Contig N <sub>so</sub> (bp)	N <sub>50</sub> (bp)	Mean read No. of length (bp) reads	No. of reads	No. of contigs	GC content (%)	Total size (bp)	Collection yr	Clade	TS	Resistance determinants	Plasmid replicon(s)	PTU
PNUSAS060563	NUSAS060563 SAMN10601908 SRR8327737	SRR8327737	CP093147, CP093148	SRR18753113	241	835,744	4,678,719	10,579	5,144.1	208,391	2	52.19	4,680,815	2018	Clade 1 contemporary	412 None		ColpVC	PTU-E11
PNUSAS039653	PNUSAS039653 SAMN09199050 SRR7155940		CP093129, CP093130, CP093131	SRR18753114 245	245	1,043,912	4,681,904 11,665		5,454	201,613	m	52.06	4,742,074	2018	Clade 1 contemporary	412	None	Incl2(delta), ColpVC	PTU-12, PTU-E11
PNUSAS014950	PNUSAS014950 SAMN07193435 SRR5659649	SRR5659649	CP093132, CP093133	SRR18753116	241	556,070	4,647,387	11,386	4,975.5	148,565	2	52.20	4,657,771	2017	Clade 1 emergent 412	412	2 bla <sub>TEM-1C</sub>	ColpHAD28, Col440II	No PTU assigned <sup>©</sup>
PNUSAS003019	PNUSAS003019 SAMN05437761 SRR3979113	SRR3979113	CP093134	SRR18753117	240	1,431,150	4,543,956	12,425	5,968.3	284,196	-	52.19	4,543,956	2016	2	93	None	None	None
PNUSAS020177	NUSAS020177 SAMN07436240	SRR19599933	CP094293	SRR18753115	255	4,123,842	4,544,675	13,652	6,436.9	146,777	1	52.19	4,544,696	2017	2	93	None	None	None

<sup>o</sup>ST, sequence type; PTU, plasmid taxonomic unit.

<sup>o</sup>Whole-genome core single nucleotide polymorphism (SNP)-based phylogenetic dade as defined by Miller et al. (2).

<sup>o</sup>No PTU assigned, but query is part of a sHSBM cluster of size 2 using COPLA (https://castillo.dicom.unican.es/copla/[10]).

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50% coverage) and the PointFinder scheme for *Salmonella* spp. (downloaded 30 August 2019), and an in-house database adapted from PlasmidFinder (90% identity, 60% coverage; https://cge.food.dtu.dk/services/PlasmidFinder/), all implemented in staramr v.0.4.0 (https://github.com/phac-nml/staramr). Plasmid taxonomic units (PTUs) were identified using COPLA (10). Sequence types (ST) were determined using staramr (multilocus sequence typing [MLST] software [https://github.com/tseemann/mlst] and the PubMLST database [11]). The default parameters were used for all software unless otherwise specified.

Consistent with the previous analysis (2), Clade 1 genomes were larger than Clade 2 genomes by at least  $\sim$ 110 kb, due to the presence of plasmids and MGE (Table 1). Of note, PNUSAS014950 contained a  $\sim$ 10-kb resistance plasmid (replicons ColpHAD28 and Col440II, PTU not assigned; Table 1) that was previously found to be significantly more common in the "emergent" 2018 to 2019 outbreak-associated subclade (2). This plasmid was first seen in Reading in 2014 and may be of particular interest for further investigation (2).

**Data availability.** The sequences discussed here have been deposited in GenBank and SRA under the accession and BioSample numbers listed in Table 1.

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