



cis–trans-Amide isomerism of the 3,4-dehydroproline residue, the ‘unpuckered’ proline

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Abstract

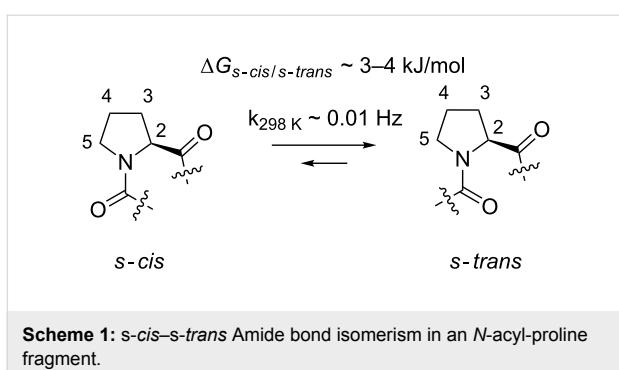
Proline (Pro) is an outstanding amino acid in various biochemical and physicochemical perspectives, especially when considering the *cis–trans* isomerism of the peptidyl-Pro amide bond. Elucidation of the roles of Pro in chemical or biological systems and engineering of its features can be addressed with various Pro analogues. Here we report an experimental work investigating the basic physicochemical properties of two Pro analogues which possess a 3,4-double bond: 3,4-dehydroproline and 4-trifluoromethyl-3,4-dehydroproline. Both indicate a flat pyrrolidine ring in their crystal structures, in agreement with previous theoretical calculations. In solution, the peptide mimics exhibit an almost unchanged equilibrium of the *trans/cis* ratios compared to that of Pro and 4-trifluoromethylproline derivatives. Finally we demonstrate that the 3,4-double bond in the investigated structures leads to an increase of the amide rotational barriers, presumably due to an interplay with the transition state.

Introduction

The sole genetically encoded secondary amino acid proline (Pro, **1**) is known for its unique properties in biological systems. In particular, Pro residues are often found in the *s-cis* peptidyl-Pro conformation, due to the low energy difference between the *s-trans* and the *s-cis* conformational states (ca. 3–4 kJ/mol) [1,2]. In addition, the high energy barrier of the *s-cis*–*s-trans* isomerization (84–89 kJ/mol) stabilizes the amide conformers kinetically (Scheme 1) [3]. By comparison of the amide rotational rates of peptidyl-Pro with the ones of the closest Pro

structural analogues, azetidine-2-carboxylic acid (norproline) and pipecolic acid (homoproline) [4], it appears that the high isomerization barrier is a feature associated with the 5-membered pyrrolidine ring of Pro [5].

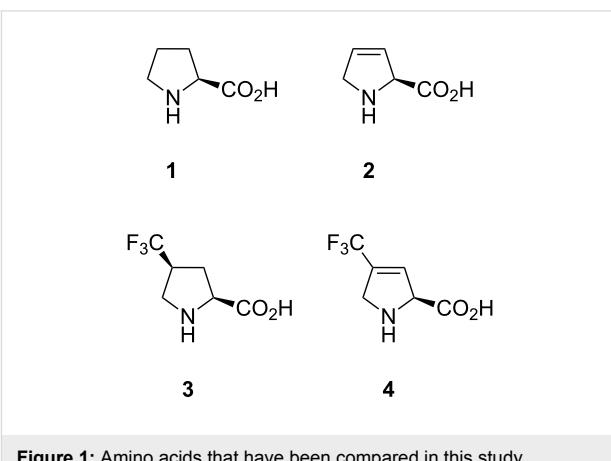
The pyrrolidine ring of Pro can be found in several conformations, designated as the *exo*- and *endo*-puckers, as well as in twisted forms [6,7]. Various ring substituents can significantly shift the equilibrium towards a high preference for one particu-



lar conformation. For instance, 3-[8,9] and 4-ring [10–15] substituents have been characterized to shift the pucker equilibrium, and as such adversely affect the *s*-*trans*/*s*-*cis* equilibrium preferences of the amide bond. Conversely, substitutions in the ring positions 2 [16–18] and 5 [19–21] shift the amide equilibrium towards higher contents of *s*-*trans* and *s*-*cis* forms, respectively, due to the steric reasons. However, it has also been reported that *N*-acylated pyroglutamic acid exhibits almost exclusively the *s*-*trans* amide conformation despite being formally a 5-substituted Pro [22]. The conformational preferences of the heterocyclic analogues of proline [23–25] and, in particular, pseudoprolines [26–28] have also been characterized.

3,4-Dehydroproline (Dhp, **2**, Figure 1) has been reported to exhibit a rather flat ring structure, as the result of theoretical analysis of Ac-Dhp-NHMe models [29,30]. It has also been demonstrated that Dhp inhibits collagen biosynthesis and suppresses the hydroxylation of proline [31–33]. Recently we found, in a comparative study of proline analogues, that Dhp is a translationally active amino acid, which, when compared to proline, exhibited lower rates of translation [34]. In order to further understand the role and potential of Dhp, this amino acid requires parametrization of its basic physicochemical features. Previously, in the literature the experimental characterization of Dhp has not been properly discussed. Herein we report the NMR and crystallographic characterization of Dhp (**2**) and 4-tri-

fluoromethyl-3,4-dehydroproline (TfmDhp, **4**) in simple models. Data on Pro (**1**) and (4*S*)-trifluoromethylproline (**3**) is used for comparison.



Results and Discussion

Firstly, we determined the pK_a of the ammonium group in the free amino acids (Table 1). Overall the values demonstrate a 0.9 pK_a reduction upon introduction of the 3,4-double bond, and a 2.2 pK_a reduction for the 4-CF₃-group. Thus, both modifications lead to electron depletion of the ring system. In addition, the larger effect upon incorporation of the CF₃-group indicates that both analogues **3** and **4** possess a similar orientation of the side-chain substituent with respect to the amine group. Previously we speculated that the CF₃-group in **3** should adopt an equatorial conformation, and thus stabilize the *exo*-pucker, based upon considerations of the vicinal *J*-couplings (in Ac-TfmPro-OMe, **7**) [35]. Considering that the CF₃-substituent should be located within the plane of the 3-pyrroline ring in **4**, this, indeed, has an orientation close to the equatorial CF₃-group placement in **3**, and is not axial.

Next, we determined the pK_a of the carboxyl groups in *N*-acetyl amino acids for two rotameric forms separately (Table 1). All

Table 1: pK_a Values determined for the amino acids **1–4** and their *N*-acetyl derivatives.

Xaa	ammonium group pK_a^a in Xaa	carboxyl group pK_a^a in Ac-Xaa			ΔpK_a
		<i>s</i> - <i>trans</i>	<i>s</i> - <i>cis</i>		
1 , Pro	10.68	3.55	2.85		0.70
2 , Dhp	9.78	3.03	2.37		0.66
3 , TfmPro	8.46	3.21	2.57		0.64
4 , TfmDhp	7.60	2.65	1.99		0.66

^aIn aqueous medium at 298 K, standard error for the amino group ±0.10, for the carboxyl group ± 0.05. $\Delta pK_a = pK_a$ (*s*-*trans*) – pK_a (*s*-*cis*).

four compounds exhibited similar ΔpK_a values [36,37]. Though, absolute acidity was depressed with both the 3,4-double bond and 4-CF₃ substitutions, the former had a stronger impact. Thus it is evident that the 3,4-double bond significantly increases the electrophilicity of the carbonyl group of the amino acid residue.

The effect on the *s-trans/s-cis* equilibrium was revealed upon NMR investigations of the conventional methyl esters of the *N*-acetyl amino acids (Ac-Xaa-OMe) [38]. The equilibrium $K_{s\text{-}trans/s\text{-}cis}$ constants in the model compounds were found to be: **5** – 4.97 ± 0.07 , **6** – 5.45 ± 0.09 , **7** – 4.31 ± 0.05 and **8** – 4.82 ± 0.03 (50 mM, D₂O, 296 K). In terms of the free energy the 3,4-double bond increased the relative stability of the *s-trans* conformer by 0.2–0.3 kJ/mol, whereas the 4-CF₃-group demonstrated an opposite effect of about 0.3 kJ/mol (standard error ± 0.1 kJ/mol). Despite both effects being rather marginal, this indicates that the increase of the electrophilicity of the terminal carbonyl groups (as seen previously in Ac-Xaa acidity) does not have a significant impact on the intramolecular interaction between the two carbonyl groups (as seen from $K_{s\text{-}trans/s\text{-}cis}$ values). Similarly, Jenkins et al. reported on bicyclic proline analogues and demonstrated that axially oriented electron withdrawing substituents (4-fluoro and 4-hydroxy groups) maintained the $K_{s\text{-}trans/s\text{-}cis}$ equilibrium values of the parent unsubstituted structure [39].

We also analyzed X-ray crystal structures of the Ac-Xaa-OMe models. All four compounds crystallized in the *s-trans* conformation: **5** and **7** as racemates [40], **6** and **8** as single enantiomers (Figure 2). In addition, *rac*-Boc-TfmDhp (**9**) crystals were subjected for analysis as well. The latter crystal structure demonstrated a pseudo *s-cis* conformation due to the helical hydrogen bond structure established between the free carboxylic and the carbamate groups as C(=O)–O–H…O=C(–Ot-Bu)–N. The same has been recently re-

ported in particular in the cases of *N*-Boc-2-methylproline [41] and *N*-Boc 4,5-difluoromethanoproline [42].

In the crystal structure the ring conformations were found to be: twisted *endo*-pucker for **5**, *exo*-pucker for **7**, and compounds **6**, **8** and **9** exhibited reasonably flat pyrrolidine rings, that is in agreement with previous computational works. (Alternative ‘flattened’ proline analogues – 4,5-methanoprolines have also been characterized, see [43,44].) For **6** and **8** the ϕ -angles were found to be -69 and -67° respectively, which are typical values for a Pro residue. Importantly, both structures did not indicate any pyramidalization around the amide nitrogen atom. The N–C=O→C=O(OMe) angle was 94 – 97° , which is below the optimal Bürgi–Dunitz trajectory angle [45,46]. The ϕ -angles in **6** and **8** found in the crystals and the $K_{s\text{-}trans/s\text{-}cis}$ values found in solution both indicate close conformational similarities between Dhp and Pro fragments.

Finally, we identified the amide rotation rates and corresponding activation barriers by EXSY NMR (D₂O, 310 K). In order to establish a proper reference system we correlated the observed activation energies with the pK_a of the ammonium groups in the corresponding free amino acids (Scheme 2). The resulting correlation indicates a remarkable offset in the activation energy in the 3,4-double bond containing residues (Figure 3). This effect was not only observed in water, but also persisted in organic solvents (MeOD, DMSO, CDCl₃, see Supporting Information File 1). The activation energies are thus generally increased by the presence of the double bond by ca. 2.7 kJ/mol, that corresponds to about a factor of 1/3 of the rotational rates.

Considering that no nitrogen pyramidalization has been observed in the crystal structures, this would indicate a destabilization of the transition state by the 3,4-double bond in Dhp

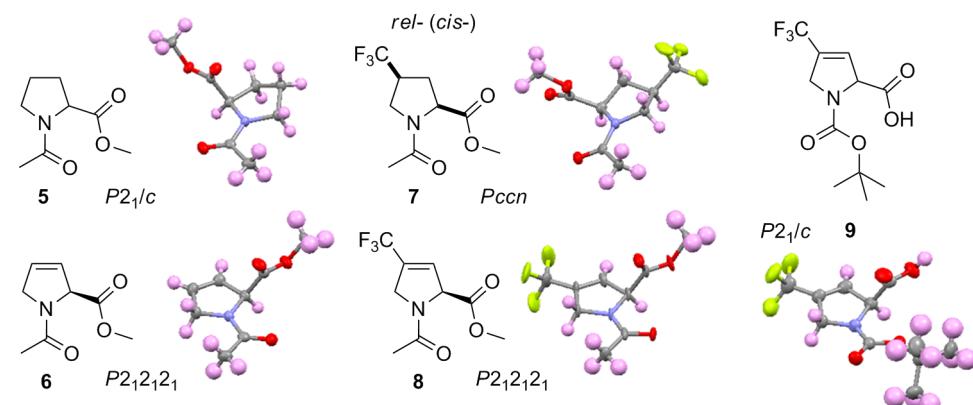
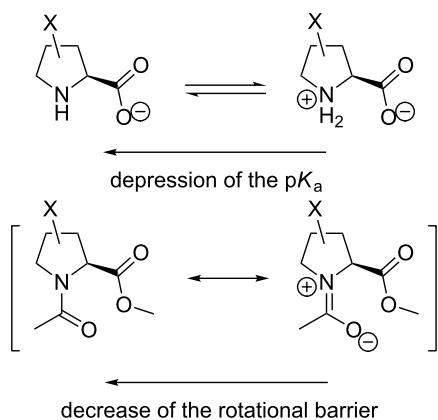


Figure 2: X-ray crystal structures of compounds **5**–**9**. Carbon—grey, nitrogen—blue, oxygen—red, fluorine—yellow, hydrogen—purple.



Scheme 2: The relationship between the pK_a of the ammonium function in the amino acid and the amide rotational barrier in proline analogues. The substituents that impose a pK_a depression effect should also decrease the content of the resonance structure with the separate charges in the ground state of the corresponding amide, that leads to a lowering of the rotational barrier.

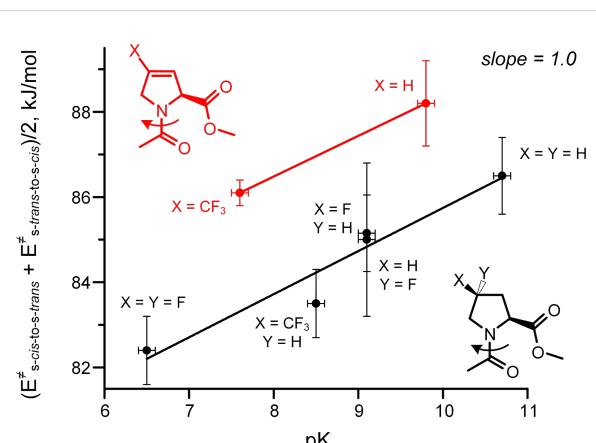


Figure 3: The double bond between C³ and C⁴ atoms in 3,4-dehydroproline residues induces an increase in the amide rotational barriers in Ac-Xaa-OMe.

and TfmDhp. Indeed, it is well known, that the peptidyl-Pro amide bond rotation proceeds via the *syn/exo* transition state, where the oxygen atom of the amide group moves under the pyrrolidine ring, approaching to C³ and C⁴ atoms [47]. Repulsion between the oxygen lone pairs and the double bond in the Dhp residue could cause the experimentally evident increase in the rotation barriers.

Conclusion

In summary, we performed the experimental characterization of proline analogues with a 3,4-double bond: Dhp and TfmDhp. Our results confirmed ‘flattening’ of the proline ring by the double bond, in agreement with what has been previously suggested by theoretical studies. Both, the 3,4-double bond and the

4-CF₃-group impose electron-withdrawing effects on the functional groups of the amino acids. Though, the carboxyl function is influenced more strongly by the double bond, whereas the amino group is more affected by the structurally proximal 4-CF₃-substituent. Conversely, the backbone conformational properties and the s-*trans*/s-*cis* energy differences remain nearly non-affected in both cases. Finally, the 3,4-double bond was found to increase the barrier of the amide rotation presumably due to the repulsive effect between the amide oxygen and the double bond in the *syn/exo* transition state. Thus 3,4-dehydroproline can be considered as a potential structural ‘freezer’ for polypeptide structures.

Supporting Information

The crystal structures are deposited in Cambridge Structural Database under the following IDs: **5**-CCDC1443104, **6**- CCDC1443105, **8**- CCDC1443103, **9**-CCDC1443102. The crystal structure of **7** have already been discussed in [35] and the deposit number was CCDC1042476. The structure files can be retrieved free of charge at <http://www.ccdc.cam.ac.uk>.

Supporting Information File 1

Experimental procedures, values for the amide rotational barriers in different solvents, copies of the NMR spectra and ellipsoid diagrams of the X-ray crystal structures. [<http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-12-57-S1.pdf>]

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References

- Owens, N. W.; Braun, C.; O'Neil, J. D.; Marat, K.; Schweizer, F. *J. Am. Chem. Soc.* **2007**, *129*, 11670–11671. doi:10.1021/ja073488d
- Renner, C.; Alefelder, S.; Bae, J. H.; Budisa, N.; Huber, R.; Moroder, L. *Angew. Chem., Int. Ed.* **2001**, *40*, 923–925. doi:10.1002/1521-3773(20010302)40:5<923::AID-ANIE923>3.0.CO;2-923
- Lu, K. P.; Finn, G.; Lee, T. H.; Nicholson, L. K. *Nat. Chem. Biol.* **2007**, *3*, 619–629. doi:10.1038/nchembio.2007.35
- Kern, D.; Schutkowski, M.; Drakenberg, T. *J. Am. Chem. Soc.* **1997**, *119*, 8403–8408. doi:10.1021/ja970606w
- Kleinpeter, E. *J. Mol. Struct.* **1996**, *380*, 139–156. doi:10.1016/0022-2860(95)09188-2
- Kang, Y. K.; Choi, H. Y. *Biophys. Chem.* **2004**, *111*, 135–142. doi:10.1016/j.bpc.2004.05.006

7. Braga, C. B.; Ducati, L. C.; Tormena, C. F.; Rittner, R. *J. Phys. Chem. A* **2014**, *118*, 1748–1758. doi:10.1021/jp5007632
8. Kim, W.; Hardcastle, K. I.; Conticello, V. P. *Angew. Chem., Int. Ed.* **2006**, *45*, 8141–8145. doi:10.1002/anie.200603227
9. Thomas, C. A.; Talaty, E. R.; Bann, J. G. *Chem. Commun.* **2009**, 3366–3368. doi:10.1039/b821952d
10. Eberhardt, E. S.; Panasik, N., Jr.; Raines, R. T. *J. Am. Chem. Soc.* **1996**, *118*, 12261–12266. doi:10.1021/ja9623119
11. Hodges, J. A.; Raines, R. T. *J. Am. Chem. Soc.* **2005**, *127*, 15923–15932. doi:10.1021/ja054674r
12. Shoulders, M. D.; Hodges, J. A.; Raines, R. T. *J. Am. Chem. Soc.* **2006**, *128*, 8112–8113. doi:10.1021/ja061793d
13. Kümin, M.; Sonntag, L.-S.; Wennemers, H. *J. Am. Chem. Soc.* **2007**, *129*, 466–467. doi:10.1021/ja067148o
14. Erdmann, R. S.; Wennemers, H. *Angew. Chem., Int. Ed.* **2011**, *50*, 6835–6838. doi:10.1002/anie.201008118
15. Siebler, C.; Erdmann, R. S.; Wennemers, H. *Angew. Chem., Int. Ed.* **2014**, *53*, 10340–10344. doi:10.1002/anie.201404935
16. Delaney, N. G.; Madison, V. *J. Am. Chem. Soc.* **1982**, *104*, 6635–6641. doi:10.1021/ja00388a027
17. De Poli, M.; Moretto, A.; Crisma, M.; Peggion, C.; Formaggio, F.; Kaptein, B.; Broxterman, Q. B.; Toniolo, C. *Chem. – Eur. J.* **2009**, *15*, 8015–8025. doi:10.1002/chem.200900688
18. Torbeev, V.; Ebert, M.-O.; Dolenc, J.; Hilvert, D. *J. Am. Chem. Soc.* **2015**, *137*, 2524–2535. doi:10.1021/ja510109p
19. Beausoleil, E.; Lubell, W. D. *Biopolymers* **2000**, *53*, 249–256. doi:10.1002/(SICI)1097-0282(200003)53:3<249::AID-BIP4>3.0.CO;2-J
20. Lummis, S. C. R.; Beene, D. L.; Lee, L. W.; Lester, H. A.; Broadhurst, W.; Dougherty, D. A. *Nature* **2005**, *438*, 248–252. doi:10.1038/nature04130
21. Melis, C.; Bussi, G.; Lummis, S. C. R.; Molteni, C. *J. Phys. Chem. B* **2009**, *113*, 12148–12153. doi:10.1021/jp9046962
22. Galardy, R. E.; Alger, J. R.; Liakopoulou-Kyriakides, M. *Int. J. Pept. Protein Res.* **1982**, *19*, 123–132. doi:10.1111/j.1399-3011.1982.tb02599.x
23. Shireman, B. T.; Miller, M. J.; Jonas, M.; Wiest, O. *J. Org. Chem.* **2001**, *66*, 6046–6056. doi:10.1021/jo102841
24. Humbert-Voss, E.; Arrault, A.; Jamart-Grégoire, B. *Tetrahedron* **2014**, *70*, 363–370. doi:10.1016/j.tet.2013.11.049
25. Duttagupta, I.; Misra, D.; Bhunya, S.; Paul, A.; Sinha, S. *J. Org. Chem.* **2015**, *80*, 10585–10604. doi:10.1021/acs.joc.5b01668
26. Dumy, P.; Keller, M.; Ryan, D. E.; Rohwedder, B.; Wöhr, T.; Mutter, M. *J. Am. Chem. Soc.* **1997**, *119*, 918–925. doi:10.1021/ja962780a
27. Chaume, G.; Barbeau, O.; Lesot, P.; Brigaut, T. *J. Org. Chem.* **2010**, *75*, 4135–4145. doi:10.1021/jo100518t
28. Feytens, D.; Chaume, G.; Chassaing, G.; Lavielle, S.; Brigaut, T.; Byun, B. J.; Kang, Y. K.; Miclet, E. *J. Phys. Chem. B* **2012**, *116*, 4069–4079. doi:10.1021/jp300284u
29. Flores-Ortega, A.; Casanovas, J.; Zanuy, D.; Nussinov, R.; Alemán, C. *J. Phys. Chem. B* **2007**, *111*, 5475–5482. doi:10.1021/jp0712001
30. Kang, Y. K.; Park, H. S. *Pept. Sci.* **2009**, *92*, 387–398. doi:10.1002/bip.21203
31. Salvador, R. A.; Tsai, I.; Marcel, R. J.; Felix, A. M.; Kerwar, S. S. *Arch. Biochem. Biophys.* **1976**, *174*, 381–392. doi:10.1016/0003-9861(76)90366-0
32. Kerwar, S. S.; Felix, A. M. *J. Biol. Chem.* **1976**, *251*, 503–509.
33. Cooper, J. B.; Varner, J. E. *Plant Physiol.* **1983**, *73*, 324–328. doi:10.1104/pp.73.2.324
34. Doerfel, L.; Wohlgemuth, I.; Kubyshkin, V.; Starosta, A. L.; Wilson, D. N.; Budisa, N.; Rodnina, M. V. *J. Am. Chem. Soc.* **2015**, *137*, 12997–13006. doi:10.1021/jacs.5b07427
35. Kubyshkin, V.; Afonin, S.; Kara, S.; Budisa, N.; Mykhailiuk, P. K.; Ulrich, A. S. *Org. Biomol. Chem.* **2015**, *13*, 3171–3181. doi:10.1039/C5OB00034C
36. Bedford, G. R.; Sadler, P. J. *Biochim. Biophys. Acta* **1974**, *343*, 656–662. doi:10.1016/0304-4165(74)90286-4
37. Evans, C. A.; Rabenstein, D. L. *J. Am. Chem. Soc.* **1974**, *96*, 7312–7317. doi:10.1021/ja00830a023
38. Shoulders, M. D.; Raines, R. T. *Annu. Rev. Biochem.* **2009**, *78*, 929–958. doi:10.1146/annurev.biochem.77.032207.120833
39. Jenkins, C. L.; Lin, G.; Duo, J.; Rapolu, D.; Guzei, I. A.; Raines, R. T.; Krow, G. R. *J. Org. Chem.* **2004**, *69*, 8565–8573. doi:10.1021/jo049242y
40. Panasik, N. J.; Eberhardt, E. S.; Edison, A. S.; Powell, D. R.; Raines, R. T. *Int. J. Pept. Protein Res.* **1994**, *44*, 262–269. doi:10.1111/j.1399-3011.1994.tb00169.x
41. Torbeev, V. Y.; Fumi, E.; Ebert, M.-O.; Schweizer, W. B.; Hilvert, D. *Helv. Chim. Acta* **2012**, *95*, 2411–2420. doi:10.1002/hlca.201200483
42. Kubyshkin, V. S.; Mykhailiuk, P. K.; Afonin, S.; Ulrich, A. S.; Komarov, I. V. *Org. Lett.* **2012**, *14*, 5254–5257. doi:10.1021/o1302412a
43. Hanessian, S.; Reinhold, U.; Gentile, G. *Angew. Chem., Int. Ed. Engl.* **1997**, *36*, 1881–1884. doi:10.1021/anie.199718811
44. Berger, G.; Vilchis-Reyes, M.; Hanessian, S. *Angew. Chem., Int. Ed.* **2015**, *54*, 13268–13272. doi:10.1002/anie.201506208
45. Bartlett, G. J.; Choudhary, A.; Raines, R. T.; Woolfson, D. N. *Nat. Chem. Biol.* **2010**, *6*, 615–620. doi:10.1038/nchembio.406
46. Choudhary, A.; Gandra, D.; Krow, G. R.; Raines, R. T. *J. Am. Chem. Soc.* **2009**, *131*, 7244–7246. doi:10.1021/ja901188y
47. Fischer, S.; Dunbrack, R. L., Jr.; Karplus, M. *J. Am. Chem. Soc.* **1994**, *116*, 11931–11937. doi:10.1021/ja00105a036

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