

Research progress on V delta 1⁺ T cells and their effect on pathogen infection

Yuxia Li¹, Yanfei Liu², Xiaoxiao Bu², Yuanyuan Qin² and Yanyan Zhang³

¹ School of Basic Medical Sciences, Shandong Second Medical University, Key Laboratory of Immune Microenvironment and Inflammatory Disease Research in Universities of Shandong Province, Weifang, Shandong, China

² School of Basic Medical Sciences, Shandong Second Medical University, Weifang, Shandong, China

³ Department of Rheumatology and Immunology, Weifang Second People's Hospital, Weifang, Shandong, China

ABSTRACT

The ongoing high occurrence of harmful infectious diseases significantly threatens human health. Existing methods used to control such diseases primarily involve targeting the pathogens, usually neglecting the vital role of host factors in disease advancement. Gamma delta ($\gamma\delta$) T cells act as a bridge between innate and adaptive immunity, playing a crucial role in combating pathogen invasion. Among these $\gamma\delta$ T cell subsets, which are categorized based on T cell receptor delta variable expression patterns, V delta (δ) 1⁺ T cells possess unique recognition abilities and regulatory characteristics and actively engage in various immune responses. The differentiation, development, and immune reactivity of V δ 1⁺ T cells are closely associated with the initial and progressive stages of infectious diseases. This article provides an overview of the classification, distribution, differentiation, and development of V δ 1⁺ T cells and their mechanisms in combating pathogenic infections, offering new insights for disease diagnosis and treatment.

Subjects Cell Biology, Microbiology, Molecular Biology, Immunology, Infectious Diseases

Keywords V δ 1⁺ T cells, $\gamma\delta$ T cells, Pathogen infection, Development, Cytokine, Mechanism

INTRODUCTION

With the continuous emergence of new and unidentified pathogens and the recurrence of existing pathogens, the incidence of infectious diseases has shown an increasing trend. For example, the spread of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), which caused the rapid infection of millions of people (*Senevirathne et al., 2024*), and the outbreak of non-epidemic Monkeypox virus in 2022 is regarded as a new global threat (*Li et al., 2022; Yi et al., 2024*). Other plaguing pathogens, such as *Mycobacterium tuberculosis* (*Mtb*), the human immunodeficiency virus (HIV), and *Plasmodium falciparum*, still have not been completely eradicated. This poses a significant challenge to public healthcare and economic infrastructure. These pathogens enter the body through various means, such as the respiratory, reproductive, and skin pathways. For example, SARS-CoV-2 primarily spreads among people through respiratory secretions, including those from coughing, sneezing, and even droplets produced when talking (*Zaidi & Singh, 2024*). Therefore, the

Submitted 26 July 2024

Accepted 24 September 2024

Published 30 October 2024

Corresponding authors

Yuxia Li, lyx_16ncmc@163.com

Yanyan Zhang,

17865367335@163.com

Academic editor

Vladimir Uversky

Additional Information and
Declarations can be found on
page 11

DOI 10.7717/peerj.18313

© Copyright

2024 Li et al.

Distributed under

Creative Commons CC-BY 4.0

OPEN ACCESS

treatment, prevention, and control of infectious diseases have become the focus and basis of current studies. Similarly, regulating the immune function of the human body to resist pathogen invasion is a recent topic of discussion.

The human immune system responds to microbial infections using various mechanisms to provide protection. As a distinct subclass of T lymphocytes, gamma delta ($\gamma\delta$) T cells serve as a frontline defense in human immune system response. With advancements in studies, emerging evidence has shown that $\gamma\delta$ T cell functionality extends beyond their innate immune response and is crucial in initiating and orchestrating adaptive immunity. For instance, some $\gamma\delta$ T cell subsets exhibit professional antigen-presenting cell capabilities (Yang *et al.*, 2022), which are vital for developing and maintaining adaptive immunity and can receive and transmit signals (Perko *et al.*, 2015; Chan *et al.*, 2022). Unlike alpha beta ($\alpha\beta$) T cells, the proportion of $\gamma\delta$ T cells in the circulating blood is relatively low, accounting for less than 5% of the total T cells in circulation; however, a high proportion of these cells are present in tissues, including the intestinal tissue (approximately 40%) and the skin (10–30%) (Hu *et al.*, 2023). Based on the expression patterns of the T cell receptor delta variable, $\gamma\delta$ T cells are primarily categorized into three subtypes, namely $V\delta 1^+$ T, $V\delta 2^+$ T, and $V\delta 3^+$ T cells (Zhao *et al.*, 2023). Owing to the limited proportion of $V\delta 1^+$ T cells in circulating blood, most studies have primarily focused on $V\delta 2^+$ T cells, which are the primary source of $\gamma\delta$ T cells in circulating blood. However, emerging observations indicate that $V\delta 1^+$ T cells also have a crucial impact on disease development (Chan *et al.*, 2024). Therefore, a comprehensive understanding of the traits of $V\delta 1^+$ T cells and an in-depth exploration of their role and mechanism in diseases resulting from pathogen infection can provide a clearer study orientation for immunologists, microbiologists, and clinical researchers, providing new insights for disease treatment. Based on this, we wrote this review which focuses on the classification, distribution, differentiation, and development of $V\delta 1^+$ T cells and the mechanism underlying their role in infectious diseases.

SURVEY METHODOLOGY

Our review focuses on $V\delta 1^+$ T cells and their effect on pathogen infection. We conducted a literature search using the Web of Science and PubMed databases. As $V\delta 1^+$ T cells constitute a subgroup of $\gamma\delta$ T cells, initially, when summarizing the characteristics of $V\delta 1^+$ T cells, we used search keywords such as “gamma delta T cell” or “V delta 1 T cell”. Subsequently, when delineating the relationship between $V\delta 1^+$ T cell and pathogen infection, in addition to “gamma delta T cell” and “V delta 1 T cell”, the keywords we searched also encompassed “bacteria” or “fungi” or “virus” or “parasite.” Studies related to pathogen infection involving $V\delta 1^+$ T cells were screened from these four categories of microorganisms. Further, we searched for “V delta 1 T cell” with “*Mycobacterium tuberculosis* (MTB)” or “human immunodeficiency virus (HIV)” or “Coronavirus Disease 2019 (COVID-19)” or “*Plasmodium*” or “*cytomegalovirus* (CMV)” or “Epstein-Barr Virus (EBV)”. Similarly, some related terms were also considered, such as “TB”, “tuberculosis”, and “SARS-CoV-2”. The articles were checked for compliance with the requirements of this review, excluding irrelevant articles (*e.g.* changings in $V\delta 1^+$ T cells are not caused by

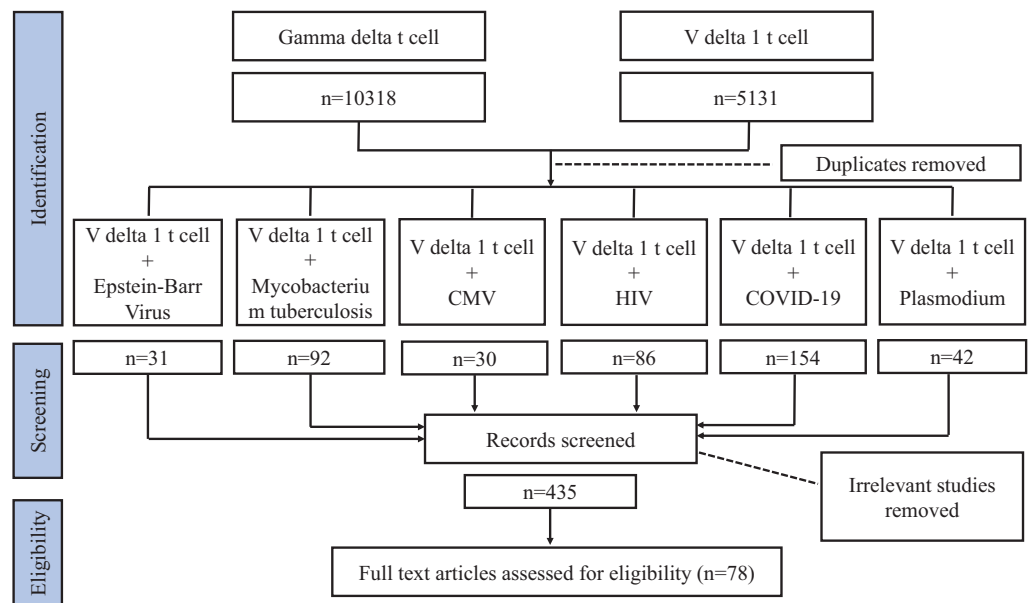


Figure 1 Flowchart article selection.

Full-size  DOI: 10.7717/peerj.18313/fig-1

pathogenic infections) and articles with older publication years where citations and conclusions were duplicated, and we screened for the results of studies on human specimens and focused on recognized articles with relatively high impact (Fig. 1). We incorporated 78 articles in this review.

Classification and distribution of V δ 1⁺ T cells

$\gamma\delta$ T cells are a crucial subgroup of T lymphocytes with distinct characteristics and functions that exhibit features of innate and adaptive immunity (Wang et al., 2024a). Several methods are currently used to classify $\gamma\delta$ T cells, with the most common being based on the composition of the antigen receptor chains on cell surfaces. The antigen recognition receptor on $\gamma\delta$ T cells comprises γ and δ chains. Based on their different γ chains, $\gamma\delta$ T cells can be classified into six different cell subpopulations, namely, V γ 2⁺, V γ 3⁺, V γ 4⁺, V γ 5⁺, V γ 8⁺, and V γ 9⁺; based on their different δ chains, $\gamma\delta$ T cells are categorized into eight cell subpopulations (V δ 1 through V δ 8) (Dart et al., 2023; Gray et al., 2024). The V δ 2⁺ subset primarily pairs with V γ 9⁺ T-cell receptors (TCRs), while the V δ 1⁺ subset exhibits flexible pairing with various V γ chains (Hu et al., 2023). V δ 1⁺ T cells are more distributed (Fig. 2) but in a smaller proportion of peripheral blood and primarily in tissues, including the intestinal epithelium, liver, spleen, and skin (Schadeck et al., 2023).

Human $\gamma\delta$ T cells, excluding V γ 9V δ 2⁺ T cells, are categorized as “non-V γ 9V δ 2⁺ T cells” (Papadopoulou, Sanchez Sanchez & Vermijlen, 2020). A common feature shared by these non-V γ 9V δ 2⁺ T cells is the adaptive characteristics of their TCR repertoires (Davey et al., 2017). V δ 1⁺ T cells are an essential part of non-V γ 9V δ 2⁺ T cells, with diverse and private repertoires (Davey et al., 2017; Gray et al., 2024), including circulating and tissue-resident repertoires (Hunter et al., 2018). In adult intestinal tissues, for example, their private repertoires contain many N additions (Holtmeier et al., 1997). One study evaluated the

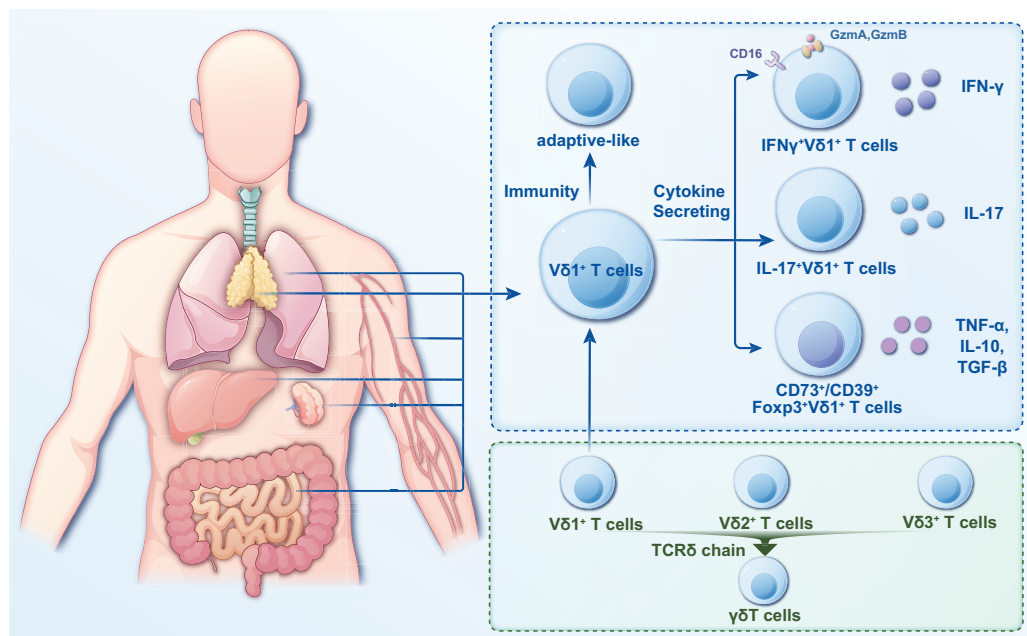


Figure 2 Classification and distribution characteristics of $V\delta 1^+$ T cells. $V\delta 1^+$ T cells are widely distributed and primarily in tissues such as intestinal epithelium, liver, spleen, lung and skin. They are categorized into two distinct classes based on their immunity and cytokine secretion. In the study of infectious diseases, the pro- or anti-infective functions can be more readily described based on the functional or cytokine secretion approach. [Full-size !\[\]\(5f471a71b78d7676bc356df190b88ab4_img.jpg\) DOI: 10.7717/peerj.18313/fig-2](https://doi.org/10.7717/peerj.18313/fig-2)

function of $V\delta 1^+$ T cells and $V\gamma 9V\delta 2^+$ T cells at the single-cell level and showed that $V\delta 1$ is more similar to natural killer (NK) cells, whereas $V\delta 2$ is similar to $CD8 \alpha\beta$ T cells (Pizzolato *et al.*, 2019). Unlike $V\gamma 9V\delta 2^+$ T cells, $V\delta 1^+$ T cells do not recognize phosphorylated antigens; however, they can recognize lipid antigens of the CD1 family, Annexin A2, and MHC class I chain-related protein A and B (Kang, Kim & Lee, 2023; Lien *et al.*, 2024). Furthermore, during pathogenic infections, $V\delta 1^+$ T cells increase the expression of natural killer cell receptors and CD16 (FcγRIII). This boosts antibody-dependent cellular cytotoxicity, aiding in recognizing and eliminating pathogens (Field *et al.*, 2024).

Based on cytokine secretion, $\gamma\delta$ T cells can be grouped into $\gamma\delta T1$, $\gamma\delta T2$, $\gamma\delta T17$, and $\gamma\delta Treg$ cells, and the associated cytokines are interferon- γ (IFN- γ), interleukin (IL)-4, IL-17A, and transforming growth factor- β , respectively (Das *et al.*, 2024). An increase in the amount of IFN- γ -producing $V\delta 1$ T cells was found in intrahepatic lymphocytes from hepatitis C virus (HCV) infected patients with high liver inflammation which could aggravate hepatic necroinflammation (Agrati *et al.*, 2001). These cells, identified as IFN- γ -producing $V\delta 1$ T cells, exhibit distinct cytotoxic markers (Hu *et al.*, 2023). Moreover, a study that involved using single-cell RNA sequencing to inspect the functional phenotypes of tumor-infiltrating $\gamma\delta$ T cells in patients with hepatocellular carcinoma revealed that these cells, primarily $V\delta 1^+$ T cells, express abundant IFNG and limited IL17A, suggesting that $V\delta 1^+$ T cells in tumor tissues exert cytotoxic effects through IFN- γ secretion (Carbone, Vaccher & Glohini, 2022). Increasing evidence indicates that $\gamma\delta$ T cells play a regulatory

role in cancer or inflammatory diseases (Yao *et al.*, 2022; Das *et al.*, 2024). For instance, in patients with breast cancer, CD73⁺Vδ1⁺ T cells constitute most of the tumor-infiltrating cell population and effectively hinder the development and functionality of dendritic cells, similar to the cytokine production by helper CD4⁺ and effector CD8⁺ T cells (Wang, Lim & Tan, 2023). These findings suggest that Vδ1⁺ T cells possess significant functional adaptability, with their function heavily influenced by the surrounding environment.

Differentiation and development of Vδ1⁺ T cells

The differentiation and development of Vδ1⁺ T cells are intricate processes governed by numerous factors. The thymus serves as a pivotal organ for T-cell development, where immature T cells progress through various differentiation stages to mature into functional T cells. γδT cells originate from T-cell precursors originating from bone marrow hematopoietic stem cells. In humans, the development of γδT cells requires TCR and sustained Notch signaling, particularly Jagged2/Notch3 signaling (Tubero Euzebio Alves *et al.*, 2024). Differentiation of human γδT cells involves the Notch-independent double-negative (DN) pathway and the Notch-dependent double-positive (DP) pathway. These yield a TCR γδ⁺ population containing DN, DP, and single-positive (SP) in the postnatal thymus (Van de Walle *et al.*, 2009; Van Coppennolle *et al.*, 2012). This indicates that a fraction of γδT cells express CD8 or CD4 molecules on their surfaces. CD8⁺γδT cells are primarily formed from Vδ1⁺ T cells in some pathogen infections (Roy Chowdhury *et al.*, 2023), and their gene transcription levels are highly consistent with those of CD8⁺αβT cells (McMurray *et al.*, 2022). This suggests that SP γδT cells undergo a developmental process that is similar to that of αβ T cells. Throughout pregnancy, γδT cell subsets shift dynamically, marked by increased Vδ1⁺ T cell production, which dominates the population in cord blood and the thymus of children, resulting in a higher count of Vδ1⁺ T cells than Vδ2⁺ T cells at birth (Boehme, Roels & Taghon, 2022). The frequencies of Vδ1⁺ T cells and Vδ2⁺ T cells changed with increasing age. It has been shown that the frequency of Vδ1 cells in the blood decreased significantly at the age of 12, whereas blood Vδ2 cells increased to adult levels; in addition, Vδ1 cells in the lungs increased to adult levels (Gray *et al.*, 2024). The diversity of distribution and tissue specificity of Vδ1⁺ T cells at different times suggests that they can play critical roles at different times.

The migration of γδT cells from the thymus to peripheral tissues involves various regulatory mechanisms, with the diversity of chemokine receptors on γδT cells playing a crucial role in migration. For example, C-C chemokine receptor six (CCR6) and C-X-C motif chemokine receptor six (CXCR6) on the surface of γδT cells facilitate their migration into mucosal tissues (Moriyama *et al.*, 2023; Das *et al.*, 2024). Vδ1⁺ T cells, a subgroup of γδT cells, express CCR2 in addition to CCR6 and CXCR6 and show migratory response to C-C motif ligand 2 (Deng *et al.*, 2023). The expression of these chemokine receptors undergoes significant changes in response to different physiological conditions. For example, the inflammatory chemokine receptors CCR2, CCR5, and CCR6 are expressed in the resting state, whereas the lymphatic chemokine receptor CCR7 is activated (Brandes *et al.*, 2003). This indicates that Vδ1⁺ T cells display tissue-specific homing mediated by chemokine receptors, and in scenarios such as tumor development, infections, and other

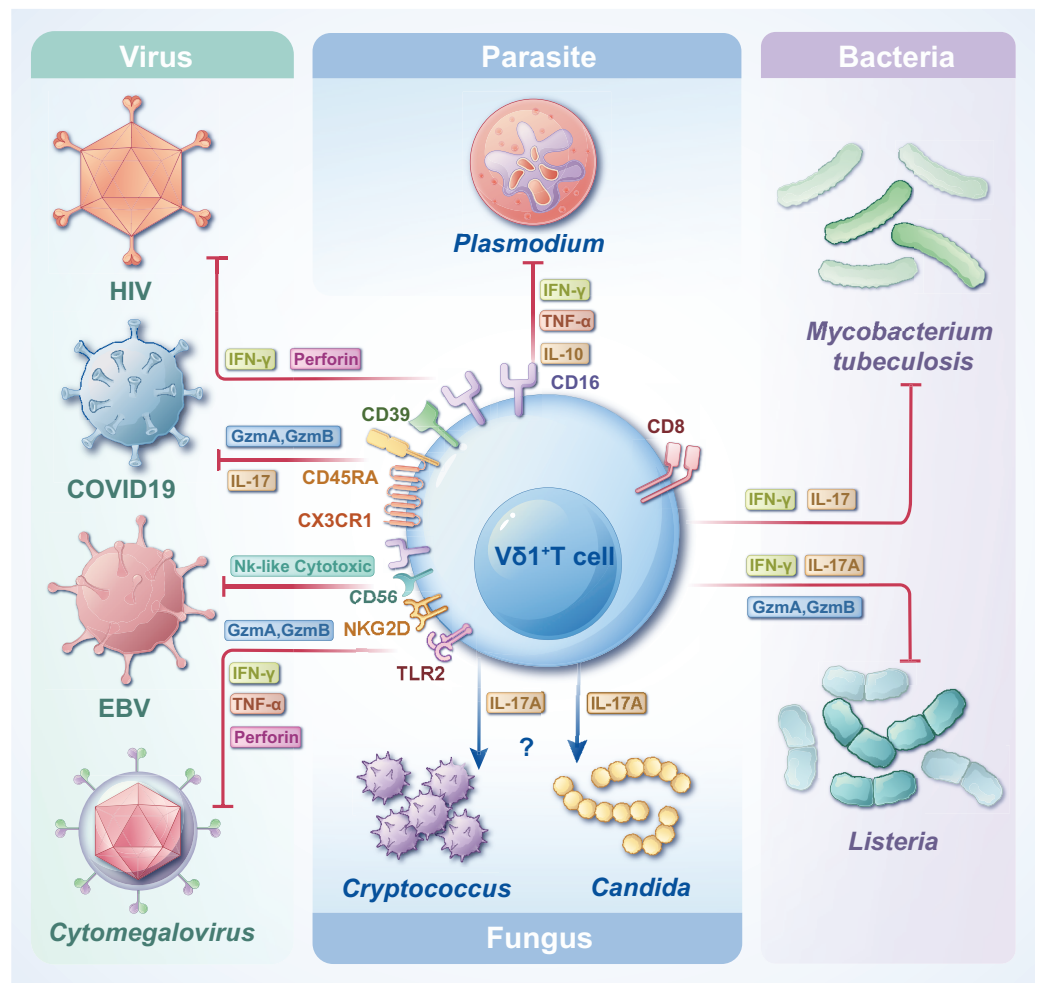


Figure 3 The roles of $V\delta 1^+$ T cells in infections with various pathogens. $V\delta 1^+$ T cells, when encountering diverse pathogens, express distinct CD molecules and secrete diverse types of cytokines, thereby attaining cytotoxicity and/or immunoregulatory effects. Facing different pathogen infections, $V\delta 1^+$ T cells exert different immune mechanisms, which need to be further clarified.

Full-size DOI: 10.7717/peerj.18313/fig-3

diseases, $V\delta 1^+$ T cells exhibit distinct selective targeting in response to alterations in their microenvironment.

$V\delta 1^+$ T cells and pathogen infection

$\gamma\delta$ T cells are crucial in combating infectious diseases. Upon activation, $\gamma\delta$ T cells exhibit cytotoxic capabilities, eliminating target cells by releasing cytotoxic substances such as perforin and granzyme, inducing apoptosis. $V\delta 1^+$ T cells, functioning as adaptive immune cells, undergo rapid clonal expansion in response to antigenic stimuli from tumors or infectious agents (Chen *et al.*, 2022). Despite their small proportion in the circulating blood, the significance of $V\delta 1^+$ T cells in the pathogenesis of infectious diseases is notable. One study analyzed the transcriptional profiles of $V\delta 1^+$ T cells and revealed that $V\delta 1^+$ T_{naive} and $V\delta 1^+$ $T_{effector}$ were highly congruent with $CD8^+$ T_{naive} and $CD8^+$ $T_{effector}$, respectively, at the cellular, molecular level (including an increase in cytotoxicity and

cytokine production) and the gene transcription level (McMurray et al., 2022). Another study on *Mycobacterium leprae* showed that resident $\gamma\delta$ T cells in the skin of patients with leprosy can protect against the disease by producing IL-17 in response to IL-23 stimulation, and most of these cells are $V\delta 1^+$ T cells. This could provide a potential target for disease treatment (Liu et al., 2022a). The subsequent sections delineate the contributions of $V\delta 1^+$ T cells to various types of pathogen infections and their distinct effector functions (Fig. 3). In most cases, $V\delta 1^+$ T cells can trigger apoptosis in pathogen-infected target cells by secreting perforin and granzymes. Furthermore, they can release various cytokines, including IFN- γ and IL-17, to actively regulate immune responses and facilitate the elimination of pathogens.

V $\delta 1^+$ T cells and Plasmodium falciparum infection

Plasmodium falciparum, a prominent malaria parasite affecting humans, imposes significant burdens on social and economic aspects of life. As a distinct subset of T cells, $\gamma\delta$ T cells are pivotal in combatting *Plasmodium* infections. Numerous studies have documented a notable and sustained expansion of $\gamma\delta$ T cells in individuals suffering from *Plasmodium* infection, revealing an increased abundance and frequency of $V\delta 1^+$ T cells (McMurray et al., 2022; Nana et al., 2024). Particularly in malaria-endemic regions, reactive $\gamma\delta$ T cells in patients, spanning children to adults frequently exposed to *P. falciparum*, predominantly comprise $V\delta 1^+$ T cells. Some studies have suggested that $V\delta 1^+$ T cells can help the body to generate naturally acquired immunity against *P. falciparum*, which is further enhanced when $V\delta 1^+$ T cells are induced to mature by the pathogens (Hviid, Smith-Togobo & Willcox, 2019; León-Lara et al., 2022). These cells exhibit a markedly heightened cytotoxic effector phenotype characterized by $CD16^+CD94^{neg}V\delta 1^+$ T cells. They demonstrate a versatile cytokine production profile, encompassing IFN- γ , tumor necrosis factor- α , and/or IL-10, with high polyclonality (León-Lara et al., 2022). Notably, recurrent *P. falciparum* infections trigger pronounced clonal expansion within the $V\delta 1^+$ T-cell population, where naïve $V\delta 1^+$ T cells gradually transition into cytotoxic $V\delta 1^+$ T cells, exerting their cytotoxic effects (von Borstel et al., 2021). These findings could be sufficient to draw our attention to $V\delta 1^+$ T cells, and the focus of studies could be shifted from the relationship between $\gamma\delta$ T cells and *P. falciparum* to $V\delta 1^+$ T cells with adaptive immune functions.

V $\delta 1^+$ T cells and Mtb infection

Tuberculosis (TB), caused by *Mtb* infection, is a highly contagious airborne disease. According to the World Health Organization's Global Tuberculosis Report 2023, an estimated 10.6 million individuals worldwide are expected to be infected with TB by 2022. Notably, China alone accounts for approximately 748,000 new TB cases, with approximately 30,000 patients diagnosed with multidrug-resistant TB, and a TB mortality rate of 2.0 per 100,000 individuals (Geneva: World Health Organization, 2023). These data underscore the urgent need to curtail the spread and mutation of *Mtb*. Previous investigations have revealed that *Mtb* infection triggers the proliferation of $\gamma\delta$ T cells in peripheral blood (Roy Chowdhury et al., 2023). Notably, the predominant $\gamma\delta$ T cell subset

in the lung tissue of patients in the acute phase is $V\delta 1^+$ T cells (Ogongo *et al.*, 2020), indicating their pivotal role in frontline defense against *Mtb* infection. Studies have unveiled substantial polymorphism in the $\gamma\delta$ TCR repertoire using high-throughput immune repertoire sequencing technology (Xia *et al.*, 2023), offering novel insights into the protective mechanisms of $\gamma\delta$ T cells against *Mtb*. One study used high-throughput sequencing to screen CDR3 δ sequences that can specifically bind to *Mtb* proteins through the $V\delta 1$ gene sequence. Results demonstrated that stimulation with *Mtb* proteins triggers significant proliferation and secretion of cytotoxic factors from $V\delta 1^+$ T cells *in vitro* (Li *et al.*, 2019). Additionally, another study revealed a distinct subset of $\gamma\delta$ T cells, NK-like $CD8^+$ $\gamma\delta$ T cells, which expand during chronic *Mtb* infection, primarily comprising $V\delta 1^+$ T cells (Roy Chowdhury *et al.*, 2023). Consequently, exploring the mechanism underlying the $\gamma\delta$ T cell response to *Mtb* infection is essential, particularly regarding the function of diverse subsets of peripheral blood $\gamma\delta$ T cells and the functional diversity of these subsets, especially $V\delta 1^+$ T cells, at the local infection site.

V $\delta 1^+$ T cells and HIV infection

HIV is a retrovirus that specifically targets $CD4^+$ T cells, progressively impairing cellular and adaptive immune responses. Studies have extensively demonstrated the pivotal role of $\gamma\delta$ T cells in combating HIV infection. A recent study involved using a model of bone marrow-liver-thymus humanized mice to evaluate the quantity and functionality of $\gamma\delta$ T cells in peripheral blood post-HIV infection (Biradar *et al.*, 2022). These findings are similar to those of previous clinical observations, revealing an increase in $V\delta 1^+$ T cells and a decrease in $V\delta 2^+$ T cells, altering the $V\delta 1/V\delta 2$ ratio, offering a novel perspective for Acquired Immunodeficiency Syndrome. Clinical intervention has increasingly focused on antiretroviral therapy (ART) (Suleman *et al.*, 2024). Studies indicate that post-ART treatment, patients manifest robust activation and degranulation of $V\delta 1^+$ T cells, which exhibit a mature NK-like phenotype enriched with CD16 (Pihl *et al.*, 2024; Field *et al.*, 2024). In instances where HIV infection precipitates tumors, $V\delta 1^+$ T cells also adopt a cytotoxic phenotype, exerting their cytotoxic effects *via* IFN- γ secretion (Carbone, Vaccher & Gloghini, 2022). Analysis of HIV-infected patients revealed a notable increase in $CD39^+V\delta 1^+$ T cells, indicating exhaustion and terminal differentiation, which is independent of the patient's disease state (Kolbe *et al.*, 2022). Consequently, $V\delta 1^+$ T cell character in HIV infection is multifaceted. Although they can identify HIV-infected cells and exert antiviral activity, HIV has evolved mechanisms to evade the immune response, causing impaired $V\delta 1^+$ T-cell function. Despite these challenges, comprehending the dynamics of $V\delta 1^+$ T-cell responses in the viral infection environment is crucial for devising strategies to bolster their antiviral functions.

V $\delta 1^+$ T cells in patients with COVID-19

COVID-19 is caused by SARS-CoV-2. Numerous studies have revealed the effect of COVID-19 on the quantity and functionality of circulating $\gamma\delta$ T cells (Lin *et al.*, 2024). Investigations have revealed that individuals with acute COVID-19 exhibit a substantial increase in the ratio of effector $V\delta 1^+$ T cells ($CD27^{neg}CD45RA^+CX3CR1^+$) in their

circulating blood. Furthermore, the fraction of cytolytic $V\delta 1^+$ T cells among $GzmA^+GzmB^+$ T cells is markedly elevated, with no such distinction observed for $V\delta 2^+$ T cells (*von Borstel et al., 2023*). Additionally, primate studies underscored a rapidly activating and expanding profile of circulating blood $V\delta 1^+$ T cells following SARS-CoV-2 infection. In the initial phases of infection, a correlated relationship was observed between the proportion of $V\delta 1^+$ T cells and the viral burden in bronchoalveolar lavage fluid (BALF). Moreover, $V\delta 1^+$ T cells in the circulating blood and BALF exhibited a predisposition towards IL-17 secretion, indicating their dual role in viral suppression and proinflammatory responses (*Fears et al., 2022*). Similarly, as previously mentioned, the prevalence of $V\delta 1^+$ T cells was higher in the cord blood and the thymus of children who possessed a naïve $V\delta 1^+$ T-cell pool and consequently exhibited a lower susceptibility to COVID-19 infection (*Zimmermann & Curtis, 2021*). This reveals a potential close association between $V\delta 1^+$ T cells and the resilience of children to SARS-CoV-2 infection, underscoring the need for further comprehensive studies on the mechanisms underlying the effect of $V\delta 1^+$ T cells on SARS-CoV-2 infection.

V $\delta 1^+$ T cells and CMV infection

CMV is an opportunistic β -herpesvirus that is widely distributed and infects most of the world's population from early childhood (*Cannon, Schmid & Hyde, 2010*). Studies have shown that the seroprevalence of CMV is approximately 40–90%, and infection in healthy individuals is usually latent and asymptomatic (*Seale et al., 2006; Griffiths & Reeves, 2021*). However, seropositive individuals have a higher frequency of the $V\delta 1^+$ subpopulation than CMV seronegative individuals (*Pitard et al., 2008*). When the organism is immunocompromised, CMV is reactivated persistently (*Chan et al., 2024*), a common viral infection complication after allogeneic haematopoietic cell transplantation (*Liu et al., 2022b*). In hematopoietic stem cell transplantation recipients, CMV reactivation can drive the immune reconstitution of $V\delta 1^+$ T cells (*de Witte et al., 2018*) and the concentration of $V\delta 1^+$ T cells was inversely related to CMV reactivation (*Bian et al., 2018*). Studies have confirmed that $V\delta 1^+\gamma\delta$ T cells are activated and expanded following CMV infection, in transplantation, neonatal infection, or common variable immunodeficiency (*Vermijlen et al., 2010; Tuengel et al., 2021; Griffiths & Reeves, 2021*). In addition, CMV infection causes a significant incidence of $V\delta 1^+$ cells with a terminally differentiated (including high cytotoxic potential) phenotype at the age of 2 years, manifested as $CD27^-CD45RA^+$, $CD57^+$, granzyme B⁺, and $CD16^+$ (*Rovito et al., 2017*). It has been shown that native and *in-vitro*-amplified $V\delta 1^+$ T cells can directly and independently recognize CMV and CMV-infected cells and inhibit CMV duplication *in vitro*, which can be achieved through the TCR $\gamma\delta$ and toll-like receptor two and natural killer cell group (NKG) 2D receptor pathways (*Liu et al., 2022b*). The percentage of $NKG2C^+V\delta 1^+$ T cells is significantly higher, and Granzyme A is strongly expressed in CMV-infected infants and adults compared with that in CMV-uninfected individuals (*Tuengel et al., 2021*), and Granzyme B is also highly expressed in CMV-infected older adults (*Verschoor et al., 2022*). The CMV antigens that cause clonal proliferation of $V\delta 1^+$ T cells are unknown. Some studies have claimed that $V\delta 1^+$ T cells are specific for endothelial protein C receptor (*Willcox et al.,*

2012), and others have revealed that V δ 1⁺ T cells recognize human leucocyte antigen-DR in a peptide-independent manner (Deseke *et al.*, 2022). These results indicate the significant influence of V δ 1⁺ T cells in combating CMV infection, and also provide new strategies and directions for the therapy of CMV-infectious diseases.

DISCUSSION

V δ 1⁺ T cells represent a unique subgroup of $\gamma\delta$ T cells (10–30% of circulating $\gamma\delta$ T cells) (Yokobori *et al.*, 2009). Regarding infection, V δ 1⁺ T cells play vital roles in controlling and eliminating pathogens. Their direct cytotoxicity, cytokine production, and modulation of immune responses significantly contribute to overall immune defense. Their participation in immune responses against infectious diseases underscores their pivotal role in host defense mechanisms. In addition to the aforementioned pathogenic infections, V δ 1⁺ T cells also undergo clonal expansion and exhibit cytotoxic phenotypes in other infections, including viral infections such as Epstein-Barr virus (EBV) (Hirai *et al.*, 2023), bacterial infections such as *Listeria* (Wang *et al.*, 2023), and fungal infections such as *Candida* (Wang *et al.*, 2024b) and *Cryptococcus* (Sato *et al.*, 2020). These results confirm that the broad pathogen-recognition capacity of V δ 1⁺ T cells underscores their versatility in combating infections. Moreover, the transcriptional profiles of V δ 1⁺ T cells are reportedly differentially expressed in various diseases, such as autoimmune diseases (Devan *et al.*, 2024) and malignancy (Ye *et al.*, 2020; Schadeck *et al.*, 2023). Therefore, further investigation into V δ 1⁺ T cells is required for increased attention. For example, enhancing the quantity or concentration of V δ 1⁺ T cells with some type of effective expansion protocol or immunostimulant-targeting V δ 1⁺ T cells, and observing their positive or negative effect on pathogens is essential. This approach will help clarify the direction of the study and disease treatment.

Despite considerable efforts in understanding V δ 1⁺ T cell biology, numerous questions and challenges persist. Further investigations are required to unravel the factors governing the commitment of thymocytes to the V δ 1 lineage and the mechanisms influencing V δ 1⁺ T-cell differentiation. Additionally, gaining clearer insights into the interactions between V δ 1⁺ T cells and diverse pathogens will facilitate the development of targeted therapeutic strategies that can be used to harness the potential of these cells. Some studies have focused on allogeneic V δ 1 CART cell therapy, however, based on the available reports, they have been primarily focused on tumor treatment (Makkouk *et al.*, 2021; Ferry *et al.*, 2022; Sánchez Martínez *et al.*, 2022), while no reports have been published on such therapy for infectious diseases. Therefore, applying the chimeric T cell technique to V δ 1⁺ T cells may help optimize effectiveness. However, this application is currently making slow progress in the clinic, which may be due to some technical challenges that need further exploration.

CONCLUSIONS

The attributes of V δ 1⁺ T cells in response to pathogen infection, particularly their cytotoxicity and immunomodulatory functions, indicate that they possess significant

potential applications in immunotherapy. $V\delta 1^+$ T cells are crucial components in the intricate network of immune responses, furnishing an additional layer of defense layer against infectious diseases. Therefore, a deeper comprehension of the development and biological function of $V\delta 1^+$ T cells is anticipated to facilitate the understanding of immunity and foster innovative approaches for preventing and treating infectious diseases. The investigation of $V\delta 1^+$ T cells is an ongoing process, and their distinctive characteristics are expected to be leveraged to develop $V\delta 1^+$ T cell therapy for the treatment of clinical diseases. The further exploration of $V\delta 1^+$ T cells is expected to offer more possibilities for the future development of cellular immunotherapy.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was supported by the Natural Science Foundation of Shandong Province (ZR2020QH001) and the Science and Technology Development Plan of Shandong Medical and Health Science (No. 202302070979). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

Natural Science Foundation of Shandong Province: ZR2020QH001.

Science and Technology Development Plan of Shandong Medical and Health Science: 202302070979.

Competing Interests

The authors declare that they have no competing interests.

Author Contributions

- Yuxia Li conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Yanfei Liu performed the experiments, prepared figures and/or tables, and approved the final draft.
- Xiaoxiao Bu analyzed the data, prepared figures and/or tables, and approved the final draft.
- Yuanyuan Qin analyzed the data, prepared figures and/or tables, and approved the final draft.
- Yanyan Zhang conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

This is a literature review.

REFERENCES

- Agrati C, D'Offizi G, Narciso P, Abrignani S, Ippolito G, Colizzi V, Poccia F. 2001. Vdelta1 T lymphocytes expressing a Th1 phenotype are the major gammadelta T cell subset infiltrating the liver of HCV-infected persons. *Molecular Medicine* 7(1):11–19 DOI 10.1007/BF03401834.
- Bian Z, Xu LP, Fu Q, Huo M, Liu L, Zhao X, Huang XJ, Liu J. 2018. Homeostatic $\gamma\delta$ T cell contents are preserved by granulocyte colony-stimulating factor priming and correlate with the early recovery of $\gamma\delta$ T cell subsets after haploidentical hematopoietic stem cell transplantation. *Biology of Blood and Marrow Transplantation* 24(2):252–259 DOI 10.1016/j.bbmt.2017.10.027.
- Biradar S, Agarwal Y, Lotze MT, Bility MT, Mailliard RB. 2022. The BLT humanized mouse model as a tool for studying human gamma Delta T cell-HIV interactions in vivo. *Frontiers in Immunology* 13:881607 DOI 10.3389/fimmu.2022.881607.
- Boehme L, Roels J, Taghon T. 2022. Development of $\gamma\delta$ T cells in the thymus—a human perspective. *Seminars in Immunology* 61–64:101662 DOI 10.1016/j.smim.2022.101662.
- Brandes M, Willimann K, Lang AB, Nam KH, Jin C, Brenner MB, Morita CT, Moser B. 2003. Flexible migration program regulates gamma Delta T-cell involvement in humoral immunity. *Blood* 102(10):3693–3701 DOI 10.1182/blood-2003-04-1016.
- Cannon MJ, Schmid DS, Hyde TB. 2010. Review of cytomegalovirus seroprevalence and demographic characteristics associated with infection. *Reviews in Medical Virology* 20(4):202–213 DOI 10.1002/rmv.655.
- Carbone A, Vaccher E, Gloghini A. 2022. Hematologic cancers in individuals infected by HIV. *Blood* 139(7):995–1012 DOI 10.1182/blood.2020005469.
- Chan KF, Duarte JDG, Ostrouska S, Behren A. 2022. $\gamma\delta$ T cells in the tumor microenvironment-interactions with other immune cells. *Frontiers in Immunology* 13:894315 DOI 10.3389/fimmu.2022.894315.
- Chan S, Morgan B, Yong MK, Margetts M, Farchione AJ, Lucas EC, Godsell J, Giang NA, Slade CA, von Borstel A, Bryant VL, Howson LJ. 2024. Cytomegalovirus drives V δ 1+ $\gamma\delta$ T cell expansion and clonality in common variable immunodeficiency. *Nature Communications* 15(1):4286 DOI 10.1038/s41467-024-48527-3.
- Chen D, Guo Y, Jiang J, Wu P, Zhang T, Wei Q, Huang J, Wu D. 2022. $\gamma\delta$ T cell exhaustion: opportunities for intervention. *Journal of Leukocyte Biology* 112(6):1669–1676 DOI 10.1002/JLB.5MR0722-777R.
- Dart RJ, Zlatareva I, Vantourout P, Theodoridis E, Amar A, Kannambath S, East P, Recaldin T, Mansfield JC, Lamb CA, Parkes M, Irving PM, Prescott NJ, Hayday AC. 2023. Conserved $\gamma\delta$ T cell selection by BTNL proteins limits the progression of human inflammatory bowel disease. *Science* 381(6663):eadh0301 DOI 10.1126/science.adh0301.
- Das D, Arava S, Khandpur S, Santosh KV, Akhtar S, Sharma A. 2024. Dominance and improved survivability of human $\gamma\delta$ T17 cell subset aggravates the immunopathogenesis of pemphigus vulgaris. *Immunologic Research* 72(1):72–81 DOI 10.1007/s12026-023-09413-0.
- Davey MS, Willcox CR, Joyce SP, Ladell K, Kasatskaya SA, McLaren JE, Hunter S, Salim M, Mohammed F, Price DA, Chudakov DM, Willcox BE. 2017. Clonal selection in the human V δ 1 T cell repertoire indicates $\gamma\delta$ TCR-dependent adaptive immune surveillance. *Nature Communications* 8(1):14760 DOI 10.1038/ncomms14760.
- de Witte MA, Sarhan D, Davis Z, Felices M, Vallera DA, Hinderlie P, Curtsinger J, Cooley S, Wagner J, Kuball J, Miller JS. 2018. Early reconstitution of NK and $\gamma\delta$ T cells and its implication for the design of post-transplant immunotherapy. *Biology of Blood and Marrow Transplantation* 24(6):1152–1162 DOI 10.1016/j.bbmt.2018.02.023.

- Deng S, Zhou F, Wang F, Jiang Y, Tang J, Hu X, Luo L, Jin Y, Huang L, Sun D, Xiao G, Feng J, Li X. 2023. C5a enhances V δ 1 T cells recruitment via the CCL2-CCR2 axis in IgA nephropathy. *International Immunopharmacology* 125(25):111065 DOI 10.1016/j.intimp.2023.111065.
- Deseke M, Rampoldi F, Sandroock I, Borst E, Böning H, Ssebyatika GL, Jürgens C, Plückerbaum N, Beck M, Hassan A, Tan L, Demera A, Janssen A, Steinberger P, Koenecke C, Viejo-Borbolla A, Messerle M, Krey T, Prinz I. 2022. A CMV-induced adaptive human V δ 1⁺ $\gamma\delta$ T cell clone recognizes HLA-DR. *Journal of Experimental Medicine* 219:e20212525 DOI 10.1084/jem.20212525.
- Devan J, Nosi V, Spagnuolo J, Chancellor A, Beshirova A, Loureiro JP, Vacchini A, Hendrik Niess J, Calogero R, Mori L, De Libero G, Hruz P. 2024. Surface protein and functional analyses identify CD4⁺CD39⁺ TCR $\alpha\beta$ ⁺ and activated TCR V δ 1⁺ cells with distinct pro-inflammatory functions in Crohn's disease lesions. *Clinical and Experimental Immunology* 215(1):79–93 DOI 10.1093/cei/uxad098.
- Fears AC, Walker EM, Chirichella N, Slisarenko N, Merino KM, Golden N, Picou B, Spencer S, Russell-Lodrigue KE, Doyle-Meyers LA, Blair RV, Beddingfield BJ, Maness NJ, Roy CJ, Rout N. 2022. The dynamics of $\gamma\delta$ T cell responses in nonhuman primates during SARS-CoV-2 infection. *Communications Biology* 5:1380 DOI 10.1038/s42003-022-04310-y.
- Ferry GM, Agbuduwe C, Forrester M, Dunlop S, Chester K, Fisher J, Anderson J, Barisa M. 2022. A simple and robust single-step method for CAR-V δ 1 $\gamma\delta$ T cell expansion and transduction for cancer immunotherapy. *Frontiers in Immunology* 13:863155 DOI 10.3389/fimmu.2022.863155.
- Field KR, Wragg KM, Kent SJ, Lee WS, Juno JA. 2024. $\gamma\delta$ T cells mediate robust anti-HIV functions during antiretroviral therapy regardless of immune checkpoint expression. *Clinical and Translational Immunology* 13(2):e1486 DOI 10.1002/cti2.1486.
- Gray JI, Caron DP, Wells SB, Guyer R, Szabo P, Rainbow D, Ergen C, Rybkina K, Bradley MC, Matsumoto R, Pethe K, Kubota M, Teichmann S, Jones J, Yosef N, Atkinson M, Brusko M, Brusko TM, Connors TJ, Sims PA, Farber DL. 2024. Human $\gamma\delta$ T cells in diverse tissues exhibit site-specific maturation dynamics across the life span. *Science Immunology* 9(96):eadn3954 DOI 10.1126/sciimmunol.adn3954.
- Griffiths P, Reeves M. 2021. Pathogenesis of human cytomegalovirus in the immunocompromised host. *Nature Reviews. Microbiology* 19(12):759–773 DOI 10.1038/s41579-021-00582-z.
- Hirai Y, Iwatsuki K, Takahashi T, Miyake T, Nakagawa Y, Tanimoto S, Kawakami Y, Morizane S. 2023. Coexpression of natural killer cell antigens by T-cell large granular lymphocytes in hydroa vacciniforme lymphoproliferative disorder and the involvement of V δ 1 + epithelial-type $\gamma\delta$ T cells. *International Journal of Hematology* 118(1):54–64 DOI 10.1007/s12185-023-03599-7.
- Holtmeier W, Rowell DL, Nyberg A, Kagnoff MF. 1997. Distinct delta T cell receptor repertoires in monozygotic twins concordant for coeliac disease. *Clinical and Experimental Immunology* 107(1):148–157 DOI 10.1046/j.1365-2249.1997.d01-887.x.
- Hu Y, Hu Q, Li Y, Lu L, Xiang Z, Yin Z, Kabelitz D, Wu Y. 2023. $\gamma\delta$ T cells: origin and fate, subsets, diseases and immunotherapy. *Signal Transduction and Targeted Therapy* 8:434 DOI 10.1038/s41392-023-01653-8.
- Hunter S, Willcox CR, Davey MS, Kasatskaya SA, Jeffery HC, Chudakov DM, Oo YH, Willcox BE. 2018. Human liver infiltrating $\gamma\delta$ T cells are composed of clonally expanded circulating and tissue-resident populations. *Journal of Hepatology* 69(3):654–665 DOI 10.1016/j.jhep.2018.05.007.

- Hviid L, Smith-Togobo C, Willcox BE. 2019. Human V δ 1+ T cells in the immune response to Plasmodium falciparum infection. *Frontiers in Immunology* 10:259 DOI 10.3389/fimmu.2019.00259.
- Kang I, Kim Y, Lee HK. 2023. Double-edged sword: $\gamma\delta$ T cells in mucosal homeostasis and disease. *Experimental and Molecular Medicine* 55(9):1895–1904 DOI 10.1038/s12276-023-00985-3.
- Kolbe K, Wittner M, Hartjen P, Hufner AD, Degen O, Ackermann C, Cords L, Stellbrink HJ, Haag F, Schulze Zur Wiesch J. 2022. Inversed ratio of CD39/CD73 expression on $\gamma\delta$ T cells in HIV versus healthy controls correlates with immune activation and disease progression. *Frontiers in Immunology* 13:867167 DOI 10.3389/fimmu.2022.867167.
- León-Lara X, Yang T, Fichtner AS, Bruni E, von Kaisenberg C, Eiz-Vesper B, Dodoo D, Adu B, Ravens S. 2022. Evidence for an adult-like type 1-immunity phenotype of V δ 1, V δ 2 and V δ 3 T cells in Ghanaian children with repeated exposure to malaria. *Frontiers in Immunology* 13:807765 DOI 10.3389/fimmu.2022.807765.
- Li Y, Wang X, Teng D, Chen H, Wang M, Wang J, Zhang J, He W. 2019. Identification of the ligands of TCR $\gamma\delta$ by screening the immune repertoire of $\gamma\delta$ T cells from patients with tuberculosis. *Frontiers in Immunology* 10:2282 DOI 10.3389/fimmu.2019.02282.
- Li H, Zhang H, Ding K, Wang XH, Sun GY, Liu ZX, Luo Y. 2022. The evolving epidemiology of monkeypox virus. *Cytokine and Growth Factor Reviews* 68:1–12 DOI 10.1016/j.cytogfr.2022.10.002.
- Lien SC, Ly D, Yang SYC, Wang BX, Clouthier DL, St Paul M, Gadalla R, Noamani B, Garcia-Batres CR, Boross-Harmer S, Bedard PL, Pugh TJ, Spreafico A, Hirano N, Razak ARA, Ohashi PS. 2024. Tumor reactive $\gamma\delta$ T cells contribute to a complete response to PD-1 blockade in a Merkel cell carcinoma patient. *Nature Communications* 15(1):1094 DOI 10.1038/s41467-024-45449-y.
- Lin J, Bai S, He L, Yang Y, Li X, Luo L, Wang Y, Chen YY, Qin J, Zhong Y. 2024. Cytotoxic lymphocyte-monocyte complex reflects the dynamics of coronavirus disease 2019 systemic immune response. *The Journal of Infectious Diseases* 230(1):5–14 DOI 10.1093/infdis/jiae048.
- Liu Y, Shi C, Ma S, Ma Y, Lu X, Zhu J, Yang D. 2022a. The protective role of tissue-resident interleukin 17A-producing gamma Delta T cells in Mycobacterium leprae infection. *Frontiers in Immunology* 13:961405 DOI 10.3389/fimmu.2022.961405.
- Liu R, Wu N, Gao H, Liang S, Yue K, Dong T, Dong X, Xu LP, Wang Y, Zhang XH, Liu J, Huang XJ. 2022b. Distinct activities of V δ 1+ T-cells upon different cytomegalovirus reactivation status after haematopoietic transplantation. *Immunology* 167(3):368–383 DOI 10.1111/imm.13542.
- Makkouk A, Yang XC, Barca T, Lucas A, Turkoz M, Wong JTS, Nishimoto KP, Brodey MM, Tabrizid M, Gundurao SRY, Bai L, Bhat A, An Z, Abbot S, Satpayev D, Aftab BT, Herrman M. 2021. Off-the-shelf V δ 1 gamma Delta T cells engineered with glypican-3 (GPC-3)-specific chimeric antigen receptor (CAR) and soluble IL-15 display robust antitumor efficacy against hepatocellular carcinoma. *Journal for ImmunoTherapy of Cancer* 9(12):e003441 DOI 10.1136/jitc-2021-003441.
- McMurray JL, von Borstel A, Taher TE, Syrimi E, Taylor GS, Sharif M, Rossjohn J, Remmerswaal EBM, Bemelman FJ, Vieira Braga FA, Chen X, Teichmann SA, Mohammed F, Berry AA, Lyke KE, Williamson KC, Stubbington MJT, Davey MS, Willcox CR, Willcox BE. 2022. Transcriptional profiling of human V δ 1 T cells reveals a pathogen-driven adaptive differentiation program. *Cell Reports* 39(8):110858 DOI 10.1016/j.celrep.2022.110858.
- Moriyama N, Saito M, Ono Y, Yamashita K, Aoi T, Kotani J. 2023. Increased interleukin-17-producing $\gamma\delta$ T cells in the brain exacerbate the pathogenesis of sepsis-associated

- encephalopathy and sepsis-induced anxiety in mice. *Journal of Clinical Medicine* **12**(13):4309 DOI [10.3390/jcm12134309](https://doi.org/10.3390/jcm12134309).
- Nana CMM, Tchakounté BDK, Bitye BMZ, Fogang B, Zangue BKT, Seumko'o RMN, Nana BC, Leke RGF, Djontu JC, Argüello RJ, Ayong L, Megnekou R. 2024.** Phenotypic changes of $\gamma\delta$ T cells in *Plasmodium falciparum* placental malaria and pregnancy outcomes in women at delivery in Cameroon. *Frontiers in Immunology* **15**:1385380 DOI [10.3389/fimmu.2024.1385380](https://doi.org/10.3389/fimmu.2024.1385380).
- Ogongo P, Steyn AJ, Karim F, Dullabh KJ, Awala I, Madansein R, Leslie A, Behar SM. 2020.** Differential skewing of donor-unrestricted and $\gamma\delta$ T cell repertoires in tuberculosis-infected human lungs. *The Journal of Clinical Investigation* **130**(1):214–230 DOI [10.1172/JCI130711](https://doi.org/10.1172/JCI130711).
- Papadopoulou M, Sanchez Sanchez G, Vermijlen D. 2020.** Innate and adaptive $\gamma\delta$ T cells: how, when, and why. *Immunological Reviews* **298**(1):99–116 DOI [10.1111/imr.12926](https://doi.org/10.1111/imr.12926).
- Perko R, Kang G, Sunkara A, Leung W, Thomas PG, Dallas MH. 2015.** Gamma Delta T cell reconstitution is associated with fewer infections and improved event-free survival after hematopoietic stem cell transplantation for pediatric leukemia. *Biology of Blood and Marrow Transplantation* **21**(1):130–136 DOI [10.1016/j.bbmt.2014.09.027](https://doi.org/10.1016/j.bbmt.2014.09.027).
- Pihl RMF, Smith-Mahoney EL, Olson A, Yuen RR, Asundi A, Lin N, Belkina AC, Snyder-Cappione JE. 2024.** V δ 1 effector and V δ 2 $\gamma\delta$ T-Cell subsets shift in frequency and are linked to plasma inflammatory markers during antiretroviral therapy-suppressed HIV infection. *The Journal of Infectious Diseases* **229**(5):1317–1327 DOI [10.1093/infdis/jiae091](https://doi.org/10.1093/infdis/jiae091).
- Pitard V, Roumanes D, Lafarge X, Couzi L, Garrigue I, Lafon ME, Merville P, Moreau JF, Déchanet-Merville J. 2008.** Long-term expansion of effector/memory Vdelta2-gammadelta T cells is a specific blood signature of CMV infection. *Blood* **112**(4):1317–1324 DOI [10.1182/blood-2008-01-136713](https://doi.org/10.1182/blood-2008-01-136713).
- Pizzolato G, Kaminski H, Tosolini M, Franchini DM, Pont F, Martins F, Valle C, Labourdette D, Cadot S, Quillet-Mary A, Poupot M, Laurent C, Ysebaert L, Meraviglia S, Dieli F, Merville P, Milpied P, Déchanet-Merville J, Fournié JJ. 2019.** Single-cell RNA sequencing unveils the shared and the distinct cytotoxic hallmarks of human TCRV δ 1 and TCRV δ 2 $\gamma\delta$ T lymphocytes. *Proceedings of the National Academy of Sciences of the United States of America* **116**(24):11906–11915 DOI [10.1073/pnas.1818488116](https://doi.org/10.1073/pnas.1818488116).
- Rovito R, Korndewal MJ, van Zelm MC, Ziagkos D, Wessels E, van der Burg M, Kroes ACM, Langerak AW, Vossen ACTM. 2017.** T and B cell markers in dried blood spots of neonates with congenital cytomegalovirus infection: B cell numbers at birth are associated with long-term outcomes. *Journal of Immunology* **198**(1):102–109 DOI [10.4049/jimmunol.1601182](https://doi.org/10.4049/jimmunol.1601182).
- Roy Chowdhury R, Valainis JR, Dubey M, von Boehmer L, Sola E, Wilhelmy J, Guo J, Kask O, Ohanyan M, Sun M, Huang H, Huang X, Nguyen PK, Scriba TJ, Davis MM, Bendall SC, Chien YH. 2023.** NK-like CD8+ $\gamma\delta$ T cells are expanded in persistent Mycobacterium tuberculosis infection. *Science Immunology* **8**(81):eade3525 DOI [10.1126/sciimmunol.ade3525](https://doi.org/10.1126/sciimmunol.ade3525).
- Sánchez Martínez D, Tirado N, Mensurado S, Martínez-Moreno A, Romecín P, Gutiérrez Agüera F, Correia DV, Silva-Santos B, Menéndez P. 2022.** Generation and proof-of-concept for allogeneic CD123 CAR-Delta One T (DOT) cells in acute myeloid leukemia. *Journal for ImmunoTherapy of Cancer* **10**(9):e005400 DOI [10.1136/jitc-2022-005400](https://doi.org/10.1136/jitc-2022-005400).
- Sato K, Yamamoto H, Nomura T, Kasamatsu J, Miyasaka T, Tanno D, Matsumoto I, Kagesawa T, Miyahara A, Zong T, Oniyama A, Kawamura K, Yokoyama R, Kitai Y, Ishizuka S, Kanno E, Tanno H, Suda H, Morita M, Yamamoto M, Iwakura Y, Ishii K, Kawakami K. 2020.** Production of IL-17A at innate immune phase leads to decreased Th1 immune response and attenuated host defense against infection with *Cryptococcus deneoformans*. *Journal of Immunology* **205**(3):686–698 DOI [10.4049/jimmunol.1901238](https://doi.org/10.4049/jimmunol.1901238).

- Schadeck J, Oberg HH, Peipp M, Hedemann N, Schamel WW, Bauerschlag D, Wesch D. 2023. Vdelta1 T cells are more resistant than Vdelta2 T cells to the immunosuppressive properties of galectin-3. *Frontiers in Immunology* 14:1286097 DOI 10.3389/fimmu.2023.1286097.
- Seale H, MacIntyre CR, Gidding HF, Backhouse JL, Dwyer DE, Gilbert L. 2006. National serosurvey of cytomegalovirus in Australia. *Clinical and Vaccine Immunology CVI* 13:1181–1184 DOI 10.1128/CVI.00203-06.
- Senevirathne TH, Wekking D, Swain JWR, Solinas C, De Silva P. 2024. COVID-19: from emerging variants to vaccination. *Cytokine and Growth Factor Reviews* 76(7798):127–141 DOI 10.1016/j.cytogfr.2023.11.005.
- Suleman M, Khan SU, Hussain T, Khan MU, Shamsul Hassan S, Majid M, Khan SU, Shehzad Khan M, Shan Ahmad RU, Arif M, Ahmad Z, Crovella S, Anthony S. 2024. Cardiovascular challenges in the era of antiretroviral therapy for AIDS/HIV: a comprehensive review of research advancements, pathophysiological insights, and future directions. *Current Problems in Cardiology* 49(3):102353 DOI 10.1016/j.cpcardiol.2023.102353.
- Tubero Euzebio Alves V, Bruno MEC, Mukherjee S, Wang L, Danaher RJ, Su L, Starr ME, Gonzalez OA. 2024. Sex-related effect of aging in gingival gamma-delta T cells. *Journal of Dental Research* 103(1):62–70 DOI 10.1177/00220345231205210.
- Tuengel J, Ranchal S, Maslova A, Aulakh G, Papadopoulou M, Drissler S, Cai B, Mohsenzadeh-Green C, Soudeyns H, Mostafavi S, van den Elzen P, Vermijlen D, Cook L, Gantt S. 2021. Characterization of adaptive-like $\gamma\delta$ T cells in Ugandan infants during primary cytomegalovirus infection. *Viruses* 13(10):1987 DOI 10.3390/v13101987.
- Van Coppennolle S, Vanhee S, Verstichel G, Snauwaert S, van der Spek A, Velghe I, Sinnesael M, Heemskerk MH, Taghon T, Leclercq G, Plum J, Langerak AW, Kerre T, Vandekerckhove B. 2012. Notch induces human T-cell receptor $\gamma\delta^+$ thymocytes to differentiate along a parallel, highly proliferative and bipotent CD4 CD8 double-positive pathway. *Leukemia* 26(1):127–138 DOI 10.1038/leu.2011.324.
- Van de Walle I, De Smet G, De Smedt M, Vandekerckhove B, Leclercq G, Plum J, Taghon T. 2009. An early decrease in Notch activation is required for human TCR-alphabeta lineage differentiation at the expense of TCR-gammadelta T cells. *Blood* 113(13):2988–2998 DOI 10.1182/blood-2008-06-164871.
- Vermijlen D, Brouwer M, Donner C, Liesnard C, Tackoen M, Van Rysselberge M, Twité N, Goldman M, Marchant A, Willems F. 2010. Human cytomegalovirus elicits fetal gammadelta T cell responses in utero. *Journal of Experimental Medicine* 207(4):807–821 DOI 10.1084/jem.20090348.
- Verschoor CP, Picard E, Andrew MK, Haynes L, Loeb M, Pawelec G, Kuchel GA. 2022. NK- and T-cell granzyme B and K expression correlates with age, CMV infection and influenza vaccine-induced antibody titres in older adults. *Frontiers in Aging* 3:1098200 DOI 10.3389/fragi.2022.1098200.
- von Borstel A, Nguyen TH, Rowntree LC, Ashhurst TM, Allen LF, Howson LJ, Holmes NE, Smibert OC, Trubiano JA, Gordon CL, Cheng AC, Kent SJ, Rossjohn J, Kedzierska K, Davey MS. 2023. Circulating effector $\gamma\delta$ T cell populations are associated with acute coronavirus disease 19 in unvaccinated individuals. *Immunology and Cell Biology* 101(4):321–332 DOI 10.1111/imcb.12623.
- von Borstel A, Chevour P, Arsovski D, Krol JMM, Howson LJ, Berry AA, Day CL, Ogongo P, Ernst JD, Nomicos EYH, Boddey JA, Giles EM, Rossjohn J, Traore B, Lyke KE, Williamson KC, Crompton PD, Davey MS. 2021. Repeated Plasmodium falciparum infection

- in humans drives the clonal expansion of an adaptive $\gamma\delta$ T cell repertoire. *Science Translational Medicine* **13**(622):eabe7430 DOI [10.1126/scitranslmed.abe7430](https://doi.org/10.1126/scitranslmed.abe7430).
- Wang Y, Hu Y, Liu Y, Shi C, Yu L, Lu N, Zhang C. 2023.** Liver-resident CD44hiCD27⁻ $\gamma\delta$ T cells help to protect against *Listeria monocytogenes* infection. *Cellular and Molecular Gastroenterology and Hepatology* **16**(6):923–941 DOI [10.1016/j.jcmgh.2023.08.008](https://doi.org/10.1016/j.jcmgh.2023.08.008).
- Wang CQ, Lim PY, Tan AHM. 2023.** Gamma/delta T cells as cellular vehicles for anti-tumor immunity. *Frontiers in Immunology* **14**:1282758 DOI [10.3389/fimmu.2023.1282758](https://doi.org/10.3389/fimmu.2023.1282758).
- Wang X, Wu H, Fang C, Li Z. 2024a.** Insights into innate immune cell evasion by *Chlamydia trachomatis*. *Frontiers in Immunology* **15**:1289644 DOI [10.3389/fimmu.2024.1289644](https://doi.org/10.3389/fimmu.2024.1289644).
- Wang X, Wu S, Li L, Yan Z. 2024b.** *Candida albicans* overgrowth disrupts the gut microbiota in mice bearing oral cancer. *Mycology* **15**(1):57–69 DOI [10.1080/21501203.2023.2256761](https://doi.org/10.1080/21501203.2023.2256761).
- Willcox CR, Pitard V, Netzer S, Couzi L, Salim M, Silberzahn T, Moreau JF, Hayday AC, Willcox BE, Déchanet-Merville J. 2012.** Cytomegalovirus and tumor stress surveillance by binding of a human $\gamma\delta$ T cell antigen receptor to endothelial protein C receptor. *Nature Immunology* **13**(9):872–879 DOI [10.1038/ni.2394](https://doi.org/10.1038/ni.2394).
- World Health Organization. 2023.** *Global Tuberculosis Report 2023*. Geneva: World Health Organization. Available at <https://www.who.int/teams/global-tuberculosis-programme/tb-reports/global-tuberculosis-report-2023>.
- Xia M, Blazevic A, Fiore-Gartland A, Hoft DF. 2023.** Impact of BCG vaccination on the repertoire of human $\gamma\delta$ T cell receptors. *Frontiers in Immunology* **14**:1100490 DOI [10.3389/fimmu.2023.1100490](https://doi.org/10.3389/fimmu.2023.1100490).
- Yang XW, Li H, Feng T, Zhang W, Song XR, Ma CY, Nie M, Wang L, Tan X, Kang Y, Liao X. 2022.** Impairment of antigen-presenting function of peripheral $\gamma\delta$ T cells in patients with sepsis. *Clinical and Experimental Immunology* **207**(1):104–112 DOI [10.1093/cei/uxab029](https://doi.org/10.1093/cei/uxab029).
- Yao YE, Qin CC, Yang CM, Huang TX. 2022.** $\gamma\delta$ T17/ $\gamma\delta$ Treg cell subsets: a new paradigm for asthma treatment. *The Journal of Asthma* **59**(10):2028–2038 DOI [10.1080/02770903.2021.1980585](https://doi.org/10.1080/02770903.2021.1980585).
- Ye W, Kong X, Zhang W, Weng Z, Wu X. 2020.** The roles of $\gamma\delta$ T cells in hematopoietic stem cell transplantation. *Cell Transplantation* **29**(9):963689720966980 DOI [10.1177/0963689720966980](https://doi.org/10.1177/0963689720966980).
- Yi XM, Lei YL, Li M, Zhong L, Li S. 2024.** The monkeypox virus-host interplays. *Cell Insight* **3**(5):100185 DOI [10.1016/j.cellin.2024.100185](https://doi.org/10.1016/j.cellin.2024.100185).
- Yokobori N, Schierloh P, Geffner L, Balboa L, Romero M, Musella R, Castagnino J, De Stéfano G, Alemán M, de la Barrera S, Abbate E, Sasiain MC. 2009.** CD3 expression distinguishes two gammadeltaT cell receptor subsets with different phenotype and effector function in tuberculous pleurisy. *Clinical and Experimental Immunology* **157**(3):385–394 DOI [10.1111/j.1365-2249.2009.03974.x](https://doi.org/10.1111/j.1365-2249.2009.03974.x).
- Zaidi AK, Singh RB. 2024.** Epidemiology of COVID-19. *Progress in Molecular Biology and Translational Science* **202**(3):25–38 DOI [10.1016/bs.pmbts.2023.09.002](https://doi.org/10.1016/bs.pmbts.2023.09.002).
- Zhao Y, Zhu R, Wang Y, Wang K. 2023.** Classification and function of $\gamma\delta$ T cells and its research progress in anti-glioblastoma. *Discover Oncology* **14**(1):150 DOI [10.1007/s12672-023-00770-8](https://doi.org/10.1007/s12672-023-00770-8).
- Zimmermann P, Curtis N. 2021.** Why is COVID-19 less severe in children? A review of the proposed mechanisms underlying the age-related difference in severity of SARS-CoV-2 infections. *Archives of Disease in Childhood* **106**(5):429–439 DOI [10.1136/archdischild-2020-320338](https://doi.org/10.1136/archdischild-2020-320338).