

Chinese Pharmaceutical Association Institute of Materia Medica, Chinese Academy of Medical Sciences

Acta Pharmaceutica Sinica B

www.elsevier.com/locate/apsb www.sciencedirect.com



REVIEW

Recent development on COX-2 inhibitors as promising anti-inflammatory agents: The past 10 years



Zhiran Ju^{a,d}, Menglan Li^a, Junde Xu^a, Daniel C. Howell^a, Zhiyun Li^a, Fen-Er Chen^{a,b,c,*}

^aInstitute of Pharmaceutical Science and Technology, Zhejiang University of Technology, Hangzhou 310014, China ^bEngineering Center of Catalysis and Synthesis for Chiral Molecules, Fudan University, Shanghai 200433, China ^cShanghai Engineering Center of Industrial Asymmetric Catalysis for Chiral Drugs, Shanghai 200433, China ^dCollaborative Innovation Center of Yangtze River Delta Region Green Pharmaceuticals, Zhejiang University of Technology, Hangzhou 310014, China

Received 16 November 2021; received in revised form 14 December 2021; accepted 30 December 2021

KEY WORDS

Inflammation; Cyclooxygenase; Prostaglandins; Adverse effects; COX-2 inhibitors **Abstract** Cyclooxygenases play a vital role in inflammation and are responsible for the production of prostaglandins. Two cyclooxygenases are described, the constitutive cyclooxygenase-1 and the inducible cyclooxygenase-2, for which the target inhibitors are the non-steroidal anti-inflammatory drugs (NSAIDs). Prostaglandins are a class of lipid compounds that mediate acute and chronic inflammation. NSAIDs are the most frequent choices for treatment of inflammation. Nevertheless, currently used anti-inflammatory drugs have become associated with a variety of adverse effects which lead to diminished output even market withdrawal. Recently, more studies have been carried out on searching novel selective COX-2 inhibitors with safety profiles. In this review, we highlight the various structural classes of organic and natural scaffolds with efficient COX-2 inhibitory activity reported during 2011–2021. It will be valuable for pharmaceutical scientists to read up on the current chemicals to pave the way for subsequent research.

© 2022 Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

*Corresponding author. Tel./fax: +86 571 88320242.

https://doi.org/10.1016/j.apsb.2022.01.002

E-mail address: rfchen@fudan.edu.cn (Fen-Er Chen).

Peer review under responsibility of Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences

^{2211-3835 © 2022} Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

Inflammation is an important part of the immune system's response against hostile world and has been linked to a variety of immunological diseases¹. Chemical agents, physical injuries, immunological reactions and Infection by pathogenic organisms usually cause acute or chronic inflammations². Non-steroidal anti-inflammatory drugs (NSAIDs) have long been known to alleviate inflammation. They treat a variety of diseases caused by inflammation, such as rheumatoid arthritis, acute fever as well as relieving common daily pains³. In the history of medicine development, the first therapeutic NSAID was aspirin, which has been used for more than 100 years since 1898. For more than a century, more typical NSAIDs, such as celecoxib, indomethacin, ibuprofen, and diclofenac (Fig. 1), have been latterly developed, and approved by the U.S. Food and Drug Administration (FDA) for clinical treatment. NSAIDs competitively inhibits the activity of cyclooxygenases (COXs) and thereby interferes with the bioconversion of the downstream inflammatory mediators. In 1971, Vane et al.^{4,5} first confirmed that the therapeutic target of NSAIDs is cyclooxygenase, further investigation found that the inhibition of cyclooxygenase directly resulted in termination of the biosynthesis of prostaglandins (PGs), which are crucial mediators of inflammation.

Four major bioactive PGs, prostaglandin E₂ (PGE₂), prostacyclin I₂ (PGI₂), prostaglandin D₂ (PGD₂), prostaglandin $F_{2\alpha}$ $(PGF_{2\alpha})$, along with a thromboxane A₂ (TXA₂) are generated during inflammation⁶. The formation of PGs was shown in Fig. 2, initially, arachidonic acid (AA) is released from the phospholipid by the catalysis of the phospholipase A₂ (PLA₂), and then a prostaglandin H₂ (PGH₂) is subsequently formed by the actions of both COX-1 and COX-2. These four PGs and TXA2 are eventually produced through the bioconversion of PGs and TXA₂ synthases in downstream mechanism. The level of prostaglandin production mainly depends on the expression of cyclooxygenases (COXs) in inflammatory tissues, especially cyclooxygenases-2⁷. PGs are hormone-like lipid compounds and are involved in many physiological reactions and play a key role in the generation of inflammatory responses⁸. In general, PGs exert their effects by mediating the body's responses to tissue injury or inflammation. Among them, PGE₂ is the dominated prostaglandin that induces typical symptoms of inflammation, such as pain, fever, tumor, and anaphylactic reaction⁹.

Two functional COXs are identified and defined as COX-1 and COX-2 according to their different structures and functions. COX-1 belongs to constitutive isoenzyme and extensively existed in most cells. Prostaglandins catalyzed by COX-1 have protective effects on gastrointestinal tract. COX-2 is an inducible enzyme which acts as the most important source of prostaglandins, so it is always regarded as a pathologic enzyme chiefly responsible for inflammation^{10–12}. NSAIDs exhibits anti-inflammatory effects by the non-selective/selective inhibition



Figure 2 A diagram of the biochemistry of prostanoids.

of cyclooxygenase (COX) activity and subsequently blocks the biosynthesis of prostaglandins in lesion sites $^{13-15}$. However, due to simultaneous inhibition of COX-1 and COX-2, non-selective NSAIDs not only achieve anti-inflammatory and analgesic purposes, but also cause serious adverse effects, such as digestive tract damage and platelet function disorder. While selective NSAIDs only inhibits COX-2 but does not affect the protective effect of prostaglandins catalyzed by COX-1 on gastrointestinal tract and platelet, thus greatly decreasing the risk of gastrointestinal side effects^{16,17}. The traditional anti-inflammatory mechanism of COX-2 inhibitor has been confirmed by recent studies. As depicted in Fig. 3, COX-2 inhibitors exert pharmacological activity through inhibition of the NF-kB pathway. Since COX-2 is responsible for producing reactive oxygen species (ROS), COX-2 inhibition causes a sharp drop in the amount of reactive oxygen species (ROS) in the upstream mechanism and keep NF- κ B in an inactive state of bondage to P-I κ B in the downstream, and thereby prohibit the production of proinflammatory cytokines, including NO, PGE2, IL-6, and TNF- α^{18} . Although selective COX-2 inhibitors are the most common choice of treatment for inflammatory diseases, they are often found to be associated with potential adverse effects of cardiovascular disorder, and a possible increased risk of heart attack, blood clots and stroke. Therefore, discovering new selective COX-2 inhibitors that can reduce such side effects appear more popular^{19,20}

To prevent undesirable outcomes, finding new NSAIDs with improved safety profiles remains the most effective approach to inflammation treatment. This paper reviews the classification and pharmacological action of new COX-2 inhibitors which have been



Figure 1 Structure formulas of clinically used COX-1 and COX-2 inhibitors.



Figure 3 A diagram of the putative anti-inflammatory mechanism of the COX-2 inhibitor in RAW264.7 cells.

reported in organic synthesis in the last ten years. In addition, new compounds from natural origins with potent COX-2 inhibitory and anti-inflammatory activity are also included, which are the promising COX-2 inhibitor for drug design and clinical use.

2. Chemistry and pharmacology of new synthetic COX-2 inhibitors

2.1. Compounds having a pyrazole ring

Pyrazole is a π -excess aromatic heterocycle, which has been recognized as a pharmacologically important active scaffold for organic synthesis, especially for new COX-2 inhibitor development. Clinical agents containing pyrazole fragments are celecoxib, antipyrine, aminopyrine, and metamizole²¹. During the past ten years, a number of pyrazole derivatives were reported and screened for their COX-2 inhibition and anti-inflammatory activity (Fig. 4). In 2014, a new compound **1** containing pyrazole fragment was synthesized by Bansal et al.²². Compound **1** showed excellent

selective inhibition of COX-2 [IC₅₀ = $0.31 \mu mol/L$, selectivity index (SI) > 222] and potential anti-inflammatory activity with ED₅₀ of 74.3 mg/kg in a carrageenan-induced rat paw edema model. The COXs inhibition activity of compound 1 was obtained by using a COX fluorescent inhibitor screening assay kit consisting of ovine COX-1 and human recombinant COX-2 enzymes [an enzyme immunoassay (EIA)]. Further investigation indicated that compound 1 presented suppression of acetic acid-induced writhes, and it showed better gastro-spasm profile compare to that of aspirin. In a molecular docking study, compound 1 showed higher selective binding affinity towards COX-2 than to COX-1. An important hydrogen bond between the oxygen of the nitro group and the hydrogen of Arg¹²⁰ was observed, which is important for the interaction with COX-2. El-Sayed et al.²³ reported the synthesis of a series of potentially useful 1,5-diphenyl pyrazoles. Compounds 2 and 3 displayed a considerable COX-2 activity and a good selectivity inhibitory (EIA) $(IC_{50} = 0.45 \text{ }\mu\text{mol/L}, SI = 111.1)$. Compounds 2 and 3 also presented high anti-inflammatory activity (ED₅₀ = 118 and 120 mg/kg) comparable with diclofenac (ED₅₀ = 114 mg/kg) in carrageenan-induced rat raw paw oedema assay. Notably, replacement of cycloalkanone moiety can significantly influence their activity. Meanwhile, molecular docking analysis indicated that compounds 2 and 3 bind into the active site of COX-2 in a similar manner to SC-558, a selective COX-2 inhibitor. Xu's group²⁴ reported a novel class of molecules, adopting a new pyrazole N-aryl sulfonate synthetic approach. This inspiration came from the selective COX-2 inhibitor-celecoxib, which has a typical sulfonamide fragment. In vitro EIA experiments indicated that compounds 4, 5, 6, and 8 have strong COX-2 inhibitory activity (Table 1). According to the selectivity index on COXs, compounds 4-8 displayed comparable selective COX-2 inhibition with that of celecoxib. Importantly, compounds 4, 5, 7, and 8 showed excellent in vivo anti-inflammatory activity (5, 7, and 8: % inhibition of auricular edemas = 27.0, 27.0 and 25.7, respectively; 4 and 7: % inhibition ratios of writhing = 50.7 and 48.5, separately, at the oral dose of 30 mg/kg, 8 mice/test group). El-Sayed et al.²⁵ continued to report some pyrazole derivatives



Figure 4 Chemical structures of compounds 1–16.

Table 1	In vitro	COX-1/COX-2	inhibition	$(IC_{50}, \mu$.mol/L)
and selectiv	vitv inde	x for compound	s 4-8, and	standard	l agent.

and serveen ny		ipoundo i o	, and standard agenti
Compd.	COX-1 ^a	COX-2 ^a	COX-2 selectivity ^b
4	0.5	0.0011	455
5	96.57	0.0092	10,479
6	>100.00	0.092	>1087
7	>100.00	0.53	>189
8	>100.00	< 0.01	-
Celecoxib	>100.00	0.056	295

 a The result (IC₅₀, µmol/L) is the mean of three determinations acquired using a COX fluorescent inhibitor screening assay kit (Cayman Chemical, MI, USA).

^bCOX-2 selectivity index (COX-1 IC₅₀/COX-2 IC₅₀).

based on the skeleton of SC-558 and celecoxib in 2012. Some of the newly synthesized compounds showed increased COX-2 inhibitory and anti-inflammatory activity. Example as compound 9, exhibiting excellent COX-2 inhibitory activity (EIA) with IC_{50} of 0.26 μ mol/L and selectivity index (SI = 192.3). Furthermore, the carrageenan-induced rat paw edema assay showed that compound 9 exerted equivalent anti-inflammatory activity with ED_{50} of 0.170 mmol/kg in comparison to the reference drug (diclofenac: $ED_{50} = 0.198$ and celecoxib: 0.185 mmol/kg, respectively). Per the docking result, the trifluoromethyl moiety of compound 9 inserts deep inside the COX-2 pocket and forming hydrogen bond with Gln¹⁹² and Arg⁵¹³, this result was consistent with COX-2 inhibition. A novel series of pyrazole derivatives that have unusual flexible fragments were reported by Gopi's group 26 , of these, compounds 10 and 11 exhibited moderate selective COX-2 inhibitory potency by using a chromogenic assay (IC₅₀ = 16.8and 14.3 μ mol/L, SI = 0.5100 and 0.4400, respectively). In docking calculations, an interaction between the ligands and Arg⁵¹³ was observed, which is required for the dependent inhibition of COX-2. In 2020, on the basis of the structure of celecoxib, several halogenated triarylpyrazoles were prepared by Abdellatif et al.²⁷. In vitro COX-2 inhibition assay indicated that three fluorinated compounds 12-14 exerted excellent efficacies (IC₅₀ = 0.049, 0.057, and 0.054 μ mol/L, respectively) close to that of celecoxib (IC₅₀ = $0.055 \text{ }\mu\text{mol/L}$) and showed better selective index (SI = 253.1, 201.8, and 214.8, respectively) than celecoxib (SI = 179.4). Moreover, compounds 12-14 exhibited close gastric profile (ulcer index (UI) = 1.25-2.5) to celecoxib (UI = 1.75). In this study, halogenated aryl ring was found to be crucial to affect activity and selectivity, and halogen atom fluoro derivatives showed better COX-2 selectivity than celecoxib.



Figure 5 Chemical structures of compounds 17–25.

Abdelall et al.²⁸ prepared a new series of 1,5-diaryl pyrazoles as both COX-2 and 15-lipoxygenase inhibitors. Compound 15 was more effective (ED₅₀ = $0.98 \mu mol/L$) on COX-2 inhibition than that of references celecoxib (ED₅₀ = $1.54 \text{ }\mu\text{mol/L}$) and meclofenamate sodium (ED₅₀ = 5.64 μ mol/L) in an EIA assay. Meanwhile, compound 15 showed good anti-inflammatory activity and selectivity index (SI = 4.89) in *in vivo* assay, which was almost identical to that of celecoxib (SI = 4.93). Moreover, the in vivo ulcerliability activity assay was explored in this study, compound 15 presented good ulcerous profile (UI = 2.78) and it was as safe as the reference celecoxib (UI = 2.9). In addition, the results suggested that presence of a (CF3) moiety in pyrazoles had no effect on COX-2 selectivity. Aiming to directly inhibit the production of PGE2 in serum samples of rats, some novel pyrazole derivatives were recently designed by Mohammed et al.²⁹. Of these, compound 16, which contains an acylamino linker, presented COX-2 inhibition with $IC_{50} = 1.76 \mu mol/L$ and a good selectivity index value of 11.1. Moreover, compound 16 showed potential anti-inflammatory activity (% edema inhibition = 81) and was less ulcerogenic than indomethacin in the in vivo ulcer liability assay. Unfortunately, the potency and selectivity of compound 16 cannot be compared with that of celecoxib, further structural modification is required to improve the activity.

2.2. Compounds having imidazole and imidazoline rings

The imidazole and imidazoline groups, as structurally similar pharmacophores, have been widely explored in the development of NSAIDs³⁰. From 2014 to 2021, several literatures have reported new COX-2 inhibitors containing imidazole and imidazoline moieties (Fig. 5). In 2014, Sampitak et al.³¹ designed and synthesized an active imidazoline analog 17. Compound 17 displayed prominent COX-2 inhibitory activity (IC₅₀ = $0.3 \ \mu mol/L$) comparable to clinically used celecoxib (IC₅₀ = $0.091 \mu mol/L$) upon in vitro evaluation. This study also proved that replacement of methylsulfonyl group by sulfonamide showed no pronounced suppressive effect on COX-2 inhibition. Four years later, Abdellatif et al.³² reported a number of 4-substituted-imidazoline analogs. Compounds 18-20 were more active towards COX-2 compare to celecoxib. Compounds 18, 20, and 21 were less ulcerogenic than clinical drugs including ibuprofen and celecoxib (Table 2). Structure-activity relationship study (SAR) revealed that multiple $-OCH_3$ (18) substituent on benzene ring has more favorable effect on the COX-2 inhibition and selectivity than other analogs. Some new substituted imidazoline-5-one derivatives 22, 23, and 24 were prepared by Metwally et al.³³ Compounds 22–24 showed similar anti-inflammatory activity (% inhibition of edema = 43.1, 41.8, and 49.0) compared to celecoxib (% inhibition of edema = 43.1%), which suggested that keeping the same sulfonamide (SO₂NH₂) moiety in new structures is crucial to maintain or increase the anti-inflammatory activity. Further study indicated that compounds 22-24 exhibited a high efficacy towards COX-2 inhibition (EIA) with IC50 of 0.090, 0.087, and 0.092 µmol/L, respectively. In addition, several substituted 1,5diarylimidazole derivatives having the thioalkyl group at position 2 were reported by Navidpour et al.³⁴ in 2014. Of these, compound 25 showed the moderate inhibition (EIA) $(IC_{50} = 14.2 \ \mu mol/L)$ of COX-2 and displayed less selectivity (SI = 3.1) than celecoxib (IC₅₀ = 0.544 µmol/L; SI = 19.4). The results suggested that compounds bearing thiomethyl group at position 2 have better activity as compared with thioethyl derivatives in this study.

Table 2 In vitro COX-1/COX-2 inhibition (IC_{50} , µmol/L), selectivity index, ulcerogenic evaluation for compounds **18–21**, and standard agents.

		U		
Compd.	COX-1 ^a	COX-2 ^a	COX-2 selectivity ^b	Ulcer index
18	4.52	0.42	10.76	1.22
19	6.74	0.62	10.87	3.02
20	4.52	0.52	8.69	2.60
21	7.86	0.86	9.14	2.61
Celecoxib	7.23	0.84	8.61	2.93
Ibuprofen	-	-	-	20.25

^aThe result (IC_{50} , µmol/L) is the mean of three determinations acquired using a COX fluorescent inhibitor screening assay kit (Cayman Chemical, MI, USA).

^bCOX-2 selectivity index (COX-1 IC₅₀/COX-2 IC₅₀).

2.3. Compounds having an indole ring

The indole moiety belongs to an important pharmacophore core for the synthesis of novel selective COX-2 inhibitors³⁵. To discover novel selective COX-2 inhibitors, Hayashi et al.36 designed a new acid-type compound 26 in 2012 (Fig. 6). Compound 26 maintained the basis structure of indomethacin and exerted potent selective COX-2 inhibition with IC₅₀ of 0.009 and 0.155 µmol/L in human cells and HWB cells, respectively. Moreover, compound 26 had good oral anti-inflammation efficacy and potent in vivo anti-oedematous effect. Meanwhile, a new N-1 and C-3 substituted indole derivative 27 was synthesized by Kaur's group³⁷ that showed selective COX-2 inhibitory activity (EIA) with IC₅₀ of 0.32 μ mol/L and SI of >312. According to the docking result, the phenyl CF₃ substituent attached to the C=N is located near the COX-2 active site and formed an important hydrogen bond to His⁹⁰, which is crucial for the COX-2 inhibition. Bhat's group³⁸ reported a new COX-2 inhibitor **28**, compound **28** not only inhibited COX-2 expression but also possessed desirable gastral safety profile. This work provided valuable information for exploring gastro-protective COX-2 inhibitors. Recently, Singh et al.39 reported several new compounds containing tosyl and dipeptide groups at N-1 and C-3 position, respectively, which were developed for COX-2 inhibitors. Of all the compounds,

compounds 29-31 showed similar in vivo anti-inflammatory activities to diclofenac. Moreover, an in vitro COX-2 selectivity assay showed that compounds 29 and 30 displayed competitive inhibition and selectivity of COX-2 (IC₅₀ = 0.006 and 0.099 μ mol/L; SI = 351 and 440, respectively). However, compound 31 showed excellent COX-2 inhibitory activity IC₅₀ of 0.54 µmol/L, but the selectivity (SI = 24) is poor. Additionally, Estevão et al.⁴⁰ synthesized a new indole derivative 32. In comparison with indomethacin, the methoxy at C-5 was replaced by a sulfonamide, the C-2 and C-3 positions were substituted by two 4-fluro benzyls, respectively. Compound 32 showed selective COX-2 inhibitory activity with $67 \pm 6\%$ (50 µmol/L) which is close to indomethacin (78 \pm 3%). More recently, Jung group⁴¹ designed and synthesized a novel N-1, C-3 substituted indole analog 33 that merge the structural motifs of anti-inflammatory ascidian metabolites, herdmanines. To form two vital hydrogen bonds with Tyr355 and Arg¹²⁰, the acid in indomethacin was replaced by a hydrazone moiety based on bioisosteric replacement drug design strategy. Compound 33 showed a considerable COX-2 inhibitory activity (7.59 μ mol/L) and selectivity (SI = 5.16) compared with diclofenac (IC₅₀ = 1.21 μ mol/L, SI = 15.18). The indole-containing analogues were mostly designed based on the structure of indomethacin. Consequently, replacement of the C-3 acetic acid moiety in indomethacin by various substitutes is an effective strategy to improves their activity and selectivity. In addition, the modification at N-1 and C-2 is also a reasonable option.

2.4. Compounds having a thiazole ring

Thiazole is a privileged pharmacophore in medicinal chemistry and bear high potential for the anti-inflammatory therapeutic option. In the past 10 years, three groups have reported several thiazole derivatives which were developed as selective COX-2 inhibitors⁴² (Fig. 7). Sağlık et al.⁴³ recently designed and synthesized novel derivatives bearing thiazolyl-hydrazine-methyl sulfonyl moiety as selective COX-2 inhibitors. Compound **34** demonstrated significant and selective COX-2 inhibition potency with an IC₅₀ value of 0.140 \pm 0.006 µmol/L and selectivity index of >714.28 comparable to nimesulide (IC₅₀ = 1.684 \pm 0.079 µmol/L) and celecoxib (IC₅₀ = 0.132 \pm 0.005 µmol/L) in *in vitro* COX-2 inhibition assay (EIA). Per the molecular docking results, compound **34** bounded in



Figure 6 Chemical structures of compounds 26–33.



Figure 7 Chemical structures of compounds 34–38.

a similar manner as celecoxib with COX-2 enzyme. Later, Abdel-Aziz et al.⁴⁴ synthesized a few novel anti-inflammatory EGFR inhibitors with cardiac and gastric safety profiles. Chemically, these compounds were formed with pyrimidine-5-carbonitrile hybrids with 2-amino-4-aryl-1,3-thiazole through an acetamide group linker. Compounds 35-37 displayed good and selective COX-2 inhibitions (EIA) (IC₅₀ = 1.17, 1.13, and $1.03 \mu mol/L$; SI = 5.78, 7.84, and 8.21, respectively) relative to celecoxib $(IC_{50} = 0.88 \ \mu mol/L, SI = 8.31)$. Further study indicated that compounds 35-37 exhibited anti-inflammatory activity (the percentage of edema inhibition) up to 90%, 94%, and 86% of meloxicam after 4 h interval and higher gastric safety profiles than meloxicam. Compounds 36 and 37 had a superior safety profile with an ulcer index of 2.70 and 2.40, respectively, compared to meloxicam (UI = 18). In addition, Hofmann's group⁴⁵ designed a new thiazole analogue, with 4-chloro- and 2-hydroxy-substituted compound 38, which displayed good and selective COX-2 inhibition with activity of 9.1 \pm 1.1% (COX-2 product formation) measured at a concentration of 10 µmol/L.

2.5. Compounds having a tetrazole ring

Since 2011, two groups have reported the synthesis and pharmacological studies of anti-inflammatory tetrazole derivatives (Fig. 8). Labib et al.⁴⁶ designed several tetrazole derivatives based on bio-isosteric replacement of SO₂NH₂ in celecoxib. Structurally, two classes of compounds were designed: isoxazoles (**39**, **40**) and pyrazoles (**41**–**44**). Compounds **39**–**44** are active and displayed potential *in vitro* COX-2 inhibitory activity in an EIA assay (IC₅₀ = 0.039–0.065 μ mol/L). Notably, compounds **40**, **42**, and **44** attained significant COX-2 selectivity index values which were as selective as celecoxib. Moreover, compounds **40** and **44** showed similar anti-inflammatory activity to celecoxib at different time intervals and were less ulcerogenic than celecoxib (Table 3). Downstream inflammatory factors were also detected, compounds 40 and 44 significantly decrease the production of PGE₂ (% inhibition = 81.042 and 82.724 in sequent) which is comparable to celecoxib (% inhibition = 79.666). The collected data indicated that the derivatives with methoxy are more active than those with hydrogen on the benzene ring. Al-Hourani et al.47 reported a tetrazole-containing compound 45, which exhibited potent COX-2 inhibition with IC_{50} value of 2.0 $\mu mol/L,$ but the SI value of compound 45 (SI = 210) was less than celecoxib (SI = 313). Five years later, this group⁴⁸ prepared more 1,5-diaryl-substituted tetrazoles by further modifications to the methylsulfonyl unit. The collected biological data showed that compounds 46 and 47 exhibited moderate COX-2 inhibitory activity (IC₅₀ = 24 and 38 μ mol/L, respectively) and selectivity (SI = 0.87 and 5.2, respectively); compound 48 displayed enhanced COX-2 inhibitory activity and selectivity towards COX-2 (EIA) (IC₅₀ = $3 \mu mol/L$, SI = > 67). The acquired results suggested that the presence of the methylsulfonyl unit, methylene spacer at C-1, and longer linker make the new derivatives more active towards COX-2 enzyme.

2.6. Compounds having an oxadiazole ring

Oxadiazole moiety has precedent for use as a bioisosteric substitute in drug design and synthesis. In the past ten years, three groups had reported the synthesis of COX-2 inhibitors containing oxadiazole group. As summarized in Fig. 9, El-Sayed et al.⁴⁹ designed a novel heterocyclic oxadiazoles **49**, which exhibited prominent COX-2 inhibitory activity (IC₅₀ = 0.041 µmol/L) and selectivity (SI = 89.72) comparably to celecoxib



Figure 8 Chemical structures of compounds 39–48.

Compd.	COX-1 ^a	COX-2 ^a	COX-2 selectivity ^b	Ulcer index	Rat paw edema (mm) (% edema inhibition, 6 h)
39	11.3	0.045	251.11	0.21 ± 0.02	7.27 ± 0.14
40	12.4	0.041	302.44	0.123 ± 0.01	5.10 ± 0.36
41	10.5	0.064	164.06	0.26 ± 0.01	$6.54 \pm 0.0.14$
42	12.8	0.043	297.67	0.21 ± 0.01	5.47 ± 0.23
43	10.9	0.065	167.69	0.55 ± 0.03	7.05 ± 0.35
44	12.4	0.039	317.95	0.11 ± 0.01	4.99 ± 0.19
Celecoxib	12.7	0.045	282.22	0.167 ± 0.01	5.21 ± 0.19
Indomethacin	0.10	0.080	1.25	0.88 ± 0.04	-

Table 3 In vitro COX-1/COX-2 inhibition (IC₅₀, μ mol/L), selectivity index, ulcerogenic evaluation and *in vivo* anti-inflammation activity (dose = 50 mg/kg) for compounds **39–44**, and standard agents.

-Not applicable.

^aThe result (IC₅₀, µmol/L) is the mean of three determinations acquired using a COX fluorescent inhibitor screening assay kit (Cayman Chemical, MI, USA).

^bCOX-2 selectivity index (COX-1 IC₅₀/COX-2 IC₅₀).

 $(IC_{50} = 0.049 \ \mu mol/L, SI = 308.16)$ by using an enzyme immunoassay. In 2020, Alfayomy et al.⁵⁰ reported two new selective COX-2 inhibitors 50 and 51, which belong to pyrimidine-5-carbonitrile hybrids with 1,3,4-oxadiazole scaffold. Compounds 50 and 51 showed significant and selectivity on COX-2 inhibition. Further investigation indicated that compounds 50 and 51 displayed good in vivo anti-inflammatory activity up to 89.5% inhibition at 4 h in carrageenan-induced rat paw edema assay. Moreover, compound 50 displayed superior safety profile than celecoxib. The results revealed that the pyrimidinyl substituent markedly affected the activity against COX-2. Grover et al.⁵ synthesized a new series of oxadiazole-comprising derivatives 52-55. Compounds 52-55 exhibited good and selective inhibition of COX-2 (EIA), but the efficacy and selectivity were less than reference drug celecoxib. Besides, compounds 53 and 55 had better in vivo anti-inflammatory activity than celecoxib (Table 4). The results confirmed that tert-butyl is an indispensable moiety to enhance COX-2 inhibitory activity and selectivity. This approach provides an alternative inspiration for new COX-2 inhibitor development.

2.7. Derivatives having fused heterocyclic fragments

Recently, fused heterocyclic rings have been flexibly used as crucial core for COX-2 inhibitors (Figs. 10 and 11). Szczukowski et al.⁵² produced a number of novel hybrid pyrrolo[3,4-d]pyridazinone derivatives bearing 4-aryl-1-(1-oxoethyl)piperazine pharmacophores. Compound 56 exerted no cytotoxicity and had significant selective COX-2 inhibition at lower concentrations. Structurally, the arylpiperazine pharmacophore is connected with 1,3,4oxadiazole ring via sulfur. The results indicated that elongating the linker part is important to enhance the anti-inflammatory activity. Khatri et al.⁵³ prepared several benzothiophene derivatives, and compounds 57, 58, 59, and 60 showed potent and selective COX-2 inhibition (EIA) (IC₅₀ = 0.33, 0.31, 0.67, and $1.40 \mu mol/L$ respectively, selectivity index: 48.8-183.8). Analysis of SAR indicated that various substitutions of benzyl were the major determinants for COX-2 inhibition, such as compounds 59 (4-SO₂NH₂) and **60** (-NHCOCH₃), which showed enhanced activity compared with 57 and 58. Moreover, compounds 57-60 showed considerable anti-inflammatory activity in vivo. Sun et al.⁵⁴ reported a series of novel selective inhibitors of enzyme COX-2. Compound 61 had potential anti-inflammatory activity with no cytotoxicity. Moreover, compound 61 showed selective inhibition towards COX-2 (IC₅₀ = $0.2 \,\mu$ mol/L) and COX-1 (IC₅₀ = $8.35 \,\mu$ mol/L) in an enzyme immuno assay (Bio-Swamp). Four bioactive benzoxazole analogs were prepared by Kaur et al.⁵⁵. Compounds 62-65showed significant COX-2 inhibitory activity and selectivity towards COX-2 over COX-1. Of all the compounds, compound 62 was the most active compound with excellent inhibition of COX-2 (EIA) (IC₅₀ = $0.04 \,\mu$ mol/L) and good selectivity (SI = 25.5). The in vivo assays results indicated that compounds 62-65 had significant anti-inflammatory activity (% inhibition = 84.09%, 68.18%, 79.54% and 72.72%, respectively), greater than reference drug ibuprofen (% inhibition = 65.90, dose = 60 mg/kg). More importantly, they demonstrated a more significant gastric tolerance than ibuprofen, the pharmacokinetic profile of compounds 62-65showed their available druggability. Later, a group of benzoxazolebenzamide analogs 66 was reported by the same group⁵⁶. Compound 66 exhibited potent and selective COX-2 inhibition (EIA) with IC_{50} of 0.14 μ mol/L as compared to celecoxib $(IC_{50} = 0.15 \ \mu mol/L)$. Compound **66** also exhibited *in vivo* antiinflammatory activity (79.54%) superior to ibuprofen (65.90%) (dose = 20 mg/kg); The ulcerogenic activity results indicated that compound 66 had significant more gastric tolerance than ibuprofen. The collected data revealed that electron withdrawing substitutions at ortho and para positions to phenyl ring aide in improving activity. Molecular docking results suggested that the benzoxazole ring is a crucial moiety to interact with Tyr355 and Arg120 of the COX-2 enzyme. All the experimental date demonstrated that compound 66 is a potential COX-2 inhibitor and valuable for further clinical investigation. Chen's group⁵⁷ reported a novel dihydropyrazole sulfonamide derivative 67, which exhibited remarkable and selective COX-2 inhibition (IC₅₀ = $0.33 \mu \text{mol/L}$), the potency almost



Figure 9 Chemical structures of compounds 49–55.

for compounds 52–55, and standard agents.							
Compd.	COX-1 ^a	COX-2 ^a	COX-2 selectivity ^b	Rat paw edema (% edema inhibition, 5 h)			
52	54.99	0.74	74.31	41.06 ± 2.64			
53	63.76	0.48	132.83	58.04 ± 1.30			
54	55.05	0.81	67.96	46.38 ± 2.47			
55	60.61	0.89	68.10	59.33 ± 2.19			
Celecoxib	37.98	0.10	379.80	49.81 ± 1.92			
Indomethacin	98.23	50.99	_	_			

Table 4 In vitro COX-1/COX-2 inhibition (IC₅₀, μ mol/L), selectivity index and *in vivo* anti-inflammation activity (dose = 150 μ mol/kg) for compounds **52–55**, and standard agents.

 a The result (IC₅₀, μ mol/L) is the mean of three determinations acquired using a COX fluorescent inhibitor screening assay kit (Cayman Chemical, MI, USA).

^bCOX-2 selectivity index (COX-1 IC₅₀/COX-2 IC₅₀).



Figure 10 Chemical structures of compounds 56–70.



Figure 11 Chemical structures of compounds 71–79.

Table 5 *In vitro* COX-1/COX-2 inhibition (IC₅₀, μmol/L), selectivity index, ulcerogenic evaluation for compounds **75–79**, and standard agent.

	0			
Compd.	COX-1 ^a	COX-2 ^a	COX-2 selectivity ^b	Ulcer index
75	276.44	4.320	63.99	0.603 ± 0.15
76	76.67	25.87	2.963	-
77	300.72	11.48	26.19	0.642 ± 0.25
78	33.58	7.750	4.332	-
79	225.68	21.87	10.31	1.991 ± 0.34
Celecoxib	>50	0.34	147.05	1.204 ± 0.06

^aThe result (IC_{50} , µmol/L) is the mean of three determinations acquired using a COX fluorescent inhibitor screening assay kit (Cayman Chemical, MI, USA).

^bCOX-2 selectivity index (COX-1 IC₅₀/COX-2 IC₅₀).

identical to that of celecoxib. Several new benzimidazole derivatives endowed with oxadiazole were described by Rathore et al.⁵⁸ in 2014. Compound **68** displayed reasonable COX-2 inhibition (EIA) (IC₅₀ = 8.2 μ mol/L) and selectivity (SI > 12.1). Moreover, compound 68 was much safer in terms of gastric toxicity with a severity index of 0.48, lower than that of indomethacin. The SAR studies revealed that the electron-withdrawing compounds showed better COX-2 inhibitory activity than those of the electronreleasing ones. An oxadiazole analog 69 was reported by Iyer et al.⁵⁹ in 2016 and exerted good COX-2 inhibition but associated with moderate COX-1 inhibition in vitro. The undesirable COX-2 selectivity indicated that compound 69 need further modification. In the same year, Nesaragi's group⁶⁰ prepared some novel coumarinyl-1,3,4-oxadiazolyl-2-mercaptobenzoxazoles. Of these, compounds 70-72 displayed moderate and selective COX-2 inhibitions (IC₅₀ = 23.71, 33.47 and 23.95 μ mol/L, respectively, SI = 33.95, 20.25 and 24.98, respectively). The results suggested that the activity is influenced by the bulkiness and lipophilicity of substituent on the benzene ring. Abdu-Allah et al.⁶¹ synthesized two novel 4-aminosalicylate based thiazolinone derivatives 73 and 74, both of which showed excellent COX-2 inhibitory efficacy $(IC_{50} = 44 \text{ and } 39 \text{ nmol/L})$ and selectivity indexes (SI = 66.82 and 68.46). Unfortunately, the selectivity indices of tested compounds (73 and 74) were lower than celecoxib but still higher than diclofenac sodium and indomethacin. Additionally, compounds 73 and 74 showed improved safety profiles than indomethacin. Analysis of the biological data revealed that the bulkiness of the substituent at heterocyclic ring enhanced the COX-2 inhibition activity. Murahari et al.⁶² introduced five active azomethine derivatives 75-79. Compounds 75-79 showed potent and selective COX-2 inhibition revealing that substitution with electron donors such as methoxyl and hydroxyl has unfavorable effect on anti-inflammatory activity. Meanwhile, compounds 75-79 were subjected to an ulcerogenic activity assay, and showed safer profiles with low ulcer indices when compared to the clinically used drug celecoxib (Table 5).

3. Chemistry and pharmacology of structurally modified COX-2 inhibitors

3.1. Derivatives of existing market drugs

Structural modification of existing drugs is an effective approach for drug development. The structures of bumetanide, celecoxib, indomethacin, and nimesulide have also been altered to develop new derivatives (Fig. 12). In 2020, Ibrahim et al.⁶³ have reported several novel benzenesulfonamide analogs which aim to developed as COX-2 inhibitors. Structurally, bumetanide was used as a precursor to synthesized new analogues. Of interest, the replacement of an acetic group by the bulky triazole moieties led to the potent COX-2 inhibitors, compounds 80 and 81. Compounds 80 and 81 exhibited excellent inhibition (Cayman's COX (ovine) Colorimetric Inhibitor Screening Assay) of COX-2 with IC₅₀ values of 0.28 and 0.17 μ mol/L, and a considerable selectivity index (SI = 71.93 and 115.82) in comparison to celecoxib (SI = 4.93). Further investigation indicated that compounds 80 and 81 showed good antiinflammatory activity and lower ulcerogenicity when administered orally. On the basis of the structure of indomethacin. Ikeda's group⁶ reported a fluorinated analog 82 of indomethacin, which bearing a lipophilic 3,3,3-trifluoroprop-1-enyl group at C-2 position. Compound 82 displayed greater COX-2 inhibitory activity and selectivity than indomethacin. Molecular docking results indicated that fluorine substituent of compound 82 contributed to a significant gain of the binding affinity for COX-2 by increasing van der Waals contacts. Aiming to discover novel selective COX-2 inhibitor. Chandna et al.⁶⁵ designed two series of celecoxib derivatives containing 1,5-diaryl fragments by bioisosteric replacement. The first series of celecoxib analogue were synthesized bearing a cyano group in place of sulfonamide moiety and then carbothioamide moiety was introduced and prepared the second series of analogues. Among these compounds, 83-86 exhibited potential selective COX-2 inhibitions $(IC_{50} = 7.07 - 19.22 \,\mu mol/L)$, but the activity of compounds 83-86 is weaker than that of celecoxib (Table 6). Nevertheless, compounds 83-86 showed potent in vivo anti-inflammatory activity which is comparable to indomethacin. Based on SAR study, the carbothioamide substituent compounds displayed better activity and selectivity than those of cyano substituent ones. In another study, Hassan et al.⁶⁶ reported a series of anti-inflammatory celecoxib analogs 87-91 by introducing a benzofuran moiety. It's worth noting that phenyl sulfonamide is an indispensable pharmacophore to maintain COX-2 selectivity. Accordingly, compounds 87-91 presented potent and selective COX-2 inhibitions with IC₅₀ values of 0.34-0.52 µmol/L. Meanwhile, changing the hydrogen in 87 into methyl in 89 led to minor decrease in COX-2 inhibition. The celecoxib analogue 91 with trifluoromethyl also had better COX-2 inhibition than fluoro analogue 90. Compounds 87–91 also possess better gastric safety profile and less gastric ulceration effect compared to clinical drug celecoxib (Table 7). Renard et al.⁶⁷ prepared a series of nimesulide analogs 92-94 in accordance with its favorable gastric and cardiovascular safety profile. Chemically, these derivatives were designed in which the nitrobenzene ring was replaced by pyridine nucleus based on isosteric replacement. The oxygen atom also has been replaced with nitrogen to construct a new linker between two aromatic rings. As a consequence, compounds 92-94 exhibited remarkable inhibitory activity associated to a COX-2/COX-1 selectivity ratio (7.46, 15.35, and 7.67, respectively; $IC_{50} = 0.26, 0.09$, and 0.30 µmol/L) similar or higher than that of celecoxib (ratio: 7.46, $IC_{50} = 0.35 \,\mu mol/L$) in a human whole blood model. The SAR study indicated that the various substitutions on the benzene ring are the main factor affecting their activity towards COXs.

3.2. Derivatives having fragments of natural products

Pharmaceutical chemists continuously design new chemical scaffolds inspired by reported natural products from 2011 to 2021, including examples that were displayed in Fig. 13. Ribeiro et al.⁶⁸ designed a series of cinnamic acid derivatives, and found three



Figure 12 Chemical structures of compounds 80–94.

active compounds 95-97 as new COX-2 inhibitors. Compounds 95-97 exhibited moderate inhibition of COX-2 (human whole blood assay) (IC₅₀ = 3.0, 2.4, and 1.09 μ mol/L; SI \geq 33, 10.0, and 3.9, respectively). The results confirmed that phenolic hydroxyl fragment is a potential pharmacological core for COXs inhibition. The data acquired also indicated that by introducing a couple of bulky hydrophobic groups may be a fruitful approach to increase the COX-2 selective inhibition. Takahash et al.⁶⁹ reported a new synthetic serotonin derivative: compound 98. Compound 98 showed weak inhibition on COX-2 (IC₅₀ = 42.5 μ mol/L) and considerable selectivity in serotonin derivatives testing assay. In this study, it was confirmed that extending amide linkage of 98 is crucial to increase COX-2 inhibitory activity. Rayar et al.⁷⁰ prepared a cyclocoumarol analog 99, which exhibited good inhibitory activity against PGE₂ production, and no inhibitory activity against the COX-1 was observed. Further study indicated that compound **99** showed considerable anti-inflammatory activity in a concentration-dependent manner.

4. Chemistry and pharmacology of potential COX-2 inhibitors from nature origin

Naturally occurring compounds have been reported to inhibit COX-2 enzyme, thereby possessing beneficial effects against inflammation. In the past ten years, a large number of natural compounds were identified as COX-2 inhibitors or exerting COX-2 inhibitory activity, examples as natural phenols, flavonoids, terpenoids, alkaloids, and other hybrids. The characteristics of their structural core scaffolds, COX-2 inhibitory activity, antiinflammatory effects, and structure–activity relationships are introduced as follows:

65–60 , and standard agents.					
Compd.	COX-1 ^a	COX-2 ^a	COX-2 selectivity ^b	Rat paw edema (% edema inhibition, 4 h)	
83	>30	19.22	>1.56	0.24 ± 0.05	
84	>30	7.07	>4.24	0.84 ± 0.01	
85	>30	9.07	>3.31	0.7 ± 0.07	
86	>30	17.43	>1.72	0.3 ± 0.01	
Celecoxib	>30	0.15	>200	-	
Indomethacin	0.18	>30	-	0.17 ± 0.03	

Table 6 In vitro COX-1/COX-2 inhibition (IC₅₀, μ mol/L), selectivity index and *in vivo* anti-inflammation activity for compounds **83–86**, and standard agents.

^aThe result (IC_{50} , μ mol/L) is the mean of three determinations acquired using a COX fluorescent inhibitor screening assay kit (Cayman Chemical, MI, USA).

^bCOX-2 selectivity index (COX-1 IC₅₀/COX-2 IC₅₀).

4.1. Phenols

Čulenová et al.⁷¹ investigated a phenolic compound **100** from Morus alba root bark. Compound 100 showed significant more in vitro COX-2 inhibition with the IC₅₀ value of 15.85 µmol/L than indomethacin (IC₅₀ = 27.04 μ mol/L, SI = 0.16), but the selectivity of COX-2 is low (SI = 0.47). Natu's group⁷² isolated an anti-inflammatory compound 101 from Alpinia officinarum Hance. Biological investigation indicated that compound 101 exhibited potent anti-inflammatory ability by the inhibition of the release and/or action of histamine, serotonin and kinin, and by COX-2 inhibition. Liu et al.⁷³ recently discovered two novel anti-inflammatory compounds 102 and 103 from Carissa spinarum. Compounds 102 and 103 were identified and showed good COX-2 inhibition by the COX-2 inhibition screening method (EIA) and the activity of compound 102 (IC₅₀) value = $0.3 \mu mol/L$) was comparable to indomethacin $(IC_{50} = 1.1 \ \mu mol/L)$. Nile et al.⁷⁴ investigated the antiinflammatory potency of three natural acids: ferulic acid (104), caffeicacid (105), and gallic acid (106). Compounds 104-106 showed potent COX-2 inhibitory activity (EIA) (IC₅₀ = 68.5, 62.5, and 65.2 µg/mL, respectively). Further investigation suggested that compounds 104-106 exerted anti-inflammatory effect through suppressing the activity of xanthine oxidase and COX-2 enzyme. Paulino et al.75 extracted and analyzed the phenols of propolis and grape pomace from Uruguayan species. Z-Fertaric acid 107 was identified and demonstrated good antiinflammatory activity, and COX-2 inhibitory activity. The potential pharmacological activity of curcumin 108 was investigated⁷⁶. Briefly, curcumin exhibited anti-inflammatory activity by significantly reducing the production of pro-inflammatory

 Table 7
 In vitro
 COX-1/COX-2
 inhibition
 (IC₅₀, μmol/L), selectivity index, ulcerogenic evaluation (rat 50 mg/kg) and for compounds 87–91, and standard agent.

-			U	
Compd.	COX-1 ^a	COX-2 ^a	COX-2 selectivity ^b	Ulcer index
87	>50	0.40	>6.67	13.82 ± 0.62
88	>50	0.52	>96.15	11.56 ± 0.54
89	>50	0.36	>138.90	10.50 ± 0.63
90	>50	0.46	>108.70	11.75 ± 0.63
91	>50	0.34	>147.06	10.50 ± 0.59
Celecoxib	>50	0.28	>178.57	16.12 ± 0.86

 a The result (IC₅₀, µmol/L) is the mean of three determinations acquired using a COX fluorescent inhibitor screening assay kit (Cayman Chemical, MI, USA).

^bCOX-2 selectivity index (COX-1 IC₅₀/COX-2 IC₅₀).

mediators including COX-2 and PGE₂ through NF- κ B pathway. A new homoegonol **109** was isolated from the extracts of Mamuyo (*Styrax ramirezii* Greenm)⁷⁷. **109** displayed antiinflammatory activity by nitric oxide reduction. In addition, **109** was able to decrease the LPS-induced transcription of inducible pro-inflammatory enzyme coding genes of COX-2. Cheng's group⁷⁸ reported the isolation and anti-inflammatory evaluation of two phenols, Periplanetol A (**110**) and Periplanetol B (**111**) (Fig. 14), from *Periplaneta americana*. **110** and **111** exhibited good COX-2 inhibition activity with IC₅₀ values of 0.768 and 0.617 µmol/L, but it is lower than that of celecoxib (IC₅₀ = 0.041 µmol/L).

4.2. Flavonoids

Honmore et al.⁷² discovered a flavonoid derivative 112, which exhibited selective COX-2 inhibition. The in vivo mice assay showed that 112 had potential anti-inflammatory activity in paw edema in comparison with diclofenac. Paulino et al.⁷⁵ extracted several flavonoids from propolis and grape pomace, flavonoid glycosylate (113), pinobanksin (114), and anthocyanin (115). 113-115 showed potent anti-inflammatory and selective COX-2 inhibitory activity (EIA) (SI = 1.82, 1.52, and 1.64, respectively). Hu et al.⁷⁹ identified a flavonoid, kaempferol-3-O-rutinoside 116, which showed VEGF-C-mediated anti-inflammation by interfering with VEGF-C-related signal transduction and interfered with the NF- κ B signaling pathway. Compound 116 exhibited high potency to trigger the receptor activation and inhibited the production of IL-6, TNF- α , and the expression of iNOS and COX-2. The anti-inflammatory effects of compound 116 was investigated in LPS-induced macrophages, and indicated that 116, as a natural compound, could be developed as an anti-inflammatory agent with good drug likeness. Nobiletin 117 (NOB) is a potential anti-inflammatory candidate. Xiao's group⁸⁰ recently reported their work on the anti-inflammatory potency of 117. 117 showed significant anti-inflammatory activity by inhibiting the expression of pro-inflammatory markers. Further investigation demonstrated that compound 117 sharply reduced the levels of iNOS and COX-2 protein in a concentration-dependent manner. Three new flavonoids 18-20 (Fig. 15) were found from the leaves of Myrica rubra sieb⁸¹, which can inhibit the expression levels of iNOS and COX-2 protein in a dose-dependent manner, and they also showed significant anti-inflammatory activity by inhibiting LPS-stimulated pro-inflammatory cytokines. Hanákova et al.⁸² isolated a new geranylated flavanone 121 from Paulownia tomentosa fruits. Compound 121 had moderate COX-2 inhibition activity (EIA) (IC₅₀ = 9.5 μ mol/L, SI = 2.8) and was more selective than ibuprofen $(IC_{50} = 4.2 \ \mu mol/L, SI = 1.5)$. Hosek et al.⁸³ reported a new



Figure 13 Chemical structures of compounds 95–99.

geranylated flavonoid, diplacone 122, which can significantly downregulated the expression of COX-2 in Western blot assay. Meanwhile, a new flavonoid **123** was purified from licorice residues⁸⁴, which displayed potent NO inhibitory effect (IC₅₀ = 9.89 μ mol/L) compared with minocycline (IC₅₀ = $33.20 \,\mu$ mol/L). Compound **123** also notably exhibited IL-1 β , IL-6, iNOS, and COX-2 inhibition. A new compound 124 was identified from Daphne genkwa Sieb and showed anti-inflammatory activity through NF- κ B signaling pathway⁸⁵, decreased expression levels of iNOS and COX-2 mRNA were observed. A large family of flavonoids, comprising compounds **125–132** (Fig. 16), was isolated from lotus plumule⁸⁶. Compounds 125–132 displayed significant anti-inflammatory activity by inhibiting the production of pro-inflammatory cytokines. Further study demonstrated that compounds 125-132 were considered as potential COX-2 ligands by computer modeling calculations. An et al.⁸⁷ analyzed the anti-inflammatory effect of Saxifragin (133) which is founded abundantly in plants, especially in Saxifrage stolonifera. Compound 133 showed outstanding anti-inflammatory activity and decreased the production of PGE₂ through suppressing the level of protein expression of COX-2. Puerarin (134) is a flavonoid derivative and possesses antipyretic and sedation activity. Pharmacological experiments indicated that puerarin exerted anti-inflammatory activity via the ERK/Nrf2/ARE pathway by inhibiting the production of pro-inflammatory markers including iNOS and COX-288, it has the

potential to be a COX-2 inhibitor. Toyama's group⁸⁹ reported their work on the anti-inflammatory activity of 8-C-rhamnosyl apigenin 135, which demonstrated selective COX-2 inhibitory activity with an IC50 value of 28.6 µmol/L and potent in vivo anti-inflammatory activity. Waller et al.⁹⁰ evaluated the COX-2 inhibitory activity of four flavonoids 136-139 from the bulbs of the Southern African Ledebouria socialis. Compounds 136-139 showed good activity towards COX-2. Notably, compounds 136 and 137 had reasonable and selective COX-2 inhibitory activity (EIA) (IC₅₀ = 1.12 and $2.87 \mu mol/$ L, respectively). Kim et al.⁹¹ evaluated the anti-inflammatory activity of a new chalcone 140 from Alpinia species. Compound 140 demonstrated inhibition of COX-2 expression and NF-kB activation in a luciferase transcriptional assay. Recently, Zhou et al.⁹² detected the anti-inflammatory therapeutic effects of a natural chalcone, isobavachalcone 141, which showed strong iNOS, COX-2, and NF-κB p65 inhibitory activity and attenuated the production levels of proinflammatory cytokine PGE₂. This compound is also a potential lead for further modifications and pharmacological evaluation. In addition, the methanol extract of Boerhaavia diffusa roots was investigated and led to the discovery of two active rotenoids 142 and 143^{93} , both of which showed moderate COX-2 inhibition with IC₅₀ value of 31.4 and 25.5 µmol/L, respectively. However, both compounds 142 and 143 exerted less COX-2 selectivity indices of 1.09 and 0.79.



Figure 14 Chemical structures of compounds 100–111.



Figure 15 Chemical structures of compounds 112–123.

4.3. Terpenoids

Bauer's group⁹⁴ reported two new terpenoid derivatives: **144** and **145** from *Hypericum cistifolium*. Compounds **144** and **145** exhibited antiinflammatory activity by inhibiting COXs activities (EIA). However, both of compounds **144** and **145** displayed relative low COX-1 and COX-2 inhibition. Zhang et al.⁹⁵ isolated and evaluated the ethyl acetate fraction of the ethanol extract from *Mallotus conspurcatus* croizat, and found two new terpenoids **146** and **147**, both of which demonstrated marked suppression of the secretion of PGE₂ and the expression of TNF- α , iNOS, NF- κ B, and COX-2 proteins. Three new sesquiterpenoids **148**, **149**, and **150** were isolated from the rhizomes and roots of *Nardostachys jatamansi* (Fig. 15)⁹⁶, which inhibit the expression of pro-inflammatory mediators COX-2 protein and cytokine PGE₂. An's group⁹⁷ reported a novel triterpenoid **151** from *Rosa rugosa* root, which potently inhibited the expression of COX-2 protein and also suppressed the production of PGE₂. Five new antiinflammatory sesquiterpenes **152–156** (Fig. 17) were found from the leaves of *Artemisia lavandulaefolia*⁹⁸, the biological evaluation results showed that compounds **152–156** had weak COX-2 inhibitory activity with IC₅₀ values of 43.29–236.33 µmol/L. In 2020, Choo group⁹⁹ investigated the potential activity of a sesquiterpene



Figure 16 Chemical structures of compounds 124–143.



Figure 17 Chemical structures of compounds 144-160.

lactone, costunolide **157**. Compound **157** decreased the expression level of COX-2 protein and have good anti-inflammatory activity through NF-κB signaling pathway. Gao's group¹⁰⁰ recently isolated several new cyathane diterpenoids from the bird's nest fungus *Cyathus africanus*. Of the new compounds, compound **158** exhibited the most active COX-2 and iNOS inhibitory effects. Kim's group¹⁰¹ reported two anti-inflammatory compounds, elatoside (**159**) and kalopanax-saponin F (**160**), which were first isolated from *Aralia elata*. Compounds **159** and **160** suppressed the NF-κB activation induced by TNF-α with IC₅₀ values of 4.1 and 9.5 µmol/L, respectively. Compounds **159** and **160** also showed inhibitory activity towards COX-2 in a dose-dependent manner.

4.4. Alkaloids

More recently, a large family of quinolizidine alkaloids were purified from the seeds of *Sophora alopecuroides* (Fig. 18)¹⁰². Among them, a new anti-inflammatory alkaloid **161** exhibited higher NO inhibition with IC₅₀ values of 29.19 µmol/L than matrine (IC₅₀ = 38.90 µmol/L). Compound **161** also showed antiinflammatory activity through decreasing the protein levels of COX-2. Feng et al.¹⁰³ had investigated the COX-2 inhibitory activity of berberine hydrochloride **162**, which showed inhibitory activity of the overexpressed COX-2 through PPAR- γ pathway.



Figure 18 Chemical structures of compounds 161–166.

Kang et al.¹⁰⁴ reported a new alkaloid 163 extracted from Amaryllidaceae, which exhibited anti-inflammatory activity via the P38 and STATs signal pathways. The expression levels of iNOS and COX-2 protein inhibited by 163 were observed, but didn't suppress the transcription of the COX-2 gene, suggesting that 163 may serve as a COX-2 inhibitor. Mohan et al.¹⁰⁵ investigated and assessed a major carbazole alkaloid girinimbine 164 presents in curry leaves, which exhibited potential antiinflammatory activity. Compound 164 demonstrated suppressing effect on COX-2 enzyme, but no effect on COX-1 in an EIA assay. The result indicated that girinimbine 164 significantly inhibited COX-2 enzyme (% inhibition = 52.5; Dose = $25 \mu g/mL$). Rui's group¹⁰⁶ reported an isoquinoline alkaloid (coptisine, **165**) from Coptidis rhizome. Similarly, compound 165 showed antiinflammatory activity via the inhibition of NF-KB pathway. In particular, compound 165 effectively blocked the production of PGE_2 through COX-2 inhibition. Lee's group¹⁰⁷ evaluated and assessed the anti-platelet activity of a major alkaloid of black pepper and long pepper: piperine 166, which showed antiinflammatory activity by regulating the AA-metabolizing enzymes. In the downstream mechanism, compound 166 decreased the production of PGE₂ and PGD₂ via COX-2 inhibition.

4.5. Others

In addition to the typical classes of natural products as introduced above, hybrid natural compounds (Fig. 19) were also discovered and showed COX-2 inhibitory and anti-inflammatory activity. Lin et al.¹⁰⁸ isolated and identified a novel quinone **167** from soft coral *Sinularia flexibilis*. The expression of COX-2 protein was significantly inhibited by compound **167** at 20 µmol/L with no cytotoxicity. Choi et al.¹⁰⁹ examined the anti-inflammatory activity of several naturally occurring anthraquinone derivatives, which were isolated from the Rhubarb Rhizome. The results indicated that compound **168** was the most potent of the compounds in inhibiting the protein expression of COX-2. Liu's group¹¹⁰ reported a new phenyl compound **169**, which was identified from a mangrove plant derived fungus *Botryosphaeria* sp. Compound **169** exhibited remarkable COX-2 inhibitory activity (IC₅₀ = 1.12 µmol/L). A few



Figure 19 Chemical structures of compounds 167–176.

phenylpropanoid derivatives **170–172** were purified from Chinese Olive by He's group¹¹¹. Western blot analyses were performed in this study and found that they significantly and dose-dependently reduced the expression level of COX-2 protein. Shen's group¹¹² isolated a new phenylpropanoid (+)-episesaminone **173** from *Cinnamomum camphora*, which prominently suppressed the expression levels of COX-2 protein. Additionally, three phenylpropanoids **174–176** were isolated from *Lilium* Asiatic hybrids flowers by Baek's group¹¹³. At a concentration of 50 µg/mL, compounds **174–176** can effectively decreased COX-2 expressions.

5. Conclusions and future perspectives

COX-2 is a bio-functional enzyme that catalyzes the biosynthesis of PGs during inflammation, and has become a significant therapeutic target when searching for anti-inflammatory drugs. Since 2011, more efforts have been focused on mining new chemical scaffolds as COX-2 inhibitors. The main emphasis of this review was on the potent COX-2 inhibitory and anti-inflammatory activity of various structural families of compounds, which have been reported within the last decade. With respect to the SAR, pyrazole analogs showed the most potent and selective inhibition of COX-2. Derivatives having fragments of natural products only showed moderate COX-2 inhibition and thereby demand more structural modification to improve their activity. Moreover, derivatives having indole, oxadiazole, thiazole, and tetrazole pharmacological cores also displayed acceptable COX-2 inhibitory activity. His⁹⁰, Arg¹²⁰ and Arg⁵¹³ were found to be most important amino acids for the inhibition and selectivity of COX-2. Meanwhile, extensive in vitro and in vivo pharmacological tests were performed and aim to discover new selective COX-2 inhibitors with safety profiles. In addition, a lot of natural compounds with good COX-2 inhibitory and anti-inflammatory activity were included herein. Natural products described in this review may provide inspiration for pharmaceutical chemists and also could serve as a foundation for novel COX-2 inhibitor design to avoid undesirable adverse effects.

Selective inhibition of COX-2 is a major feature of the new generation of NSAIDs, there are several prospects need to be considered for the development of next-generation of NSAIDs.

- 1) The new COX-2 inhibitors must be able to reduce stomach irritation and the risk of peptic ulcers.
- In some respects, COX-1/COX-2 balanced inhibitors maybe a new direction for the development of NSAIDs, when the serious adverse effects of either non-selective or selective inhibitors are considered.

3) Majority of derivatives require the presence of aryl group as the basic scaffold for COX-2 inhibitors and thereby the solubility of the new compounds need to be further considered in clinical use. Introducing hydrophilic groups into the structures may be helpful to address this problem.

The above discussion will undoubtedly attract more interest in the coming years. Regarding the future research on COX-2 inhibitor, we strongly feel that utilizing the traditional medicinal chemistry approach seems to be insufficient for current clinical needs, combing genetic engineering, enzyme engineering, and computer science may be a fruitful way to confront future challenges.

Acknowledgments

The author would like to thank Dr. K. Kaliyaperumal, Dr. M. Fredimoses, and Dr. B. Sachin for their carful revisions of this work.

Author contributions

Zhiran Ju and Fen-Er Chen generated the manuscript draft. Menglan Li, Junde Xu, Daniel C. Howell, and Zhiyun Li edited and revised the manuscript. All authors reviewed and approved the final version of the manuscript.

Conflicts of interest

The authors have no conflicts of interest to declare.

References

- 1. Nathan C, Ding A. Nonresolving inflammation. *Cell* 2010;140: 871–82.
- Lichtenberger LM, Wang ZM, Romero JJ, Ulloa C, Perez JC, Giraud MN, et al. Non-steroidal anti-inflammatory drugs (NSAIDs) associate with zwitterionic phospholipids: insight into the mechanism and reversal of NSAID-induced gastrointestinal injury. *Nat Med* 1995;1:154–8.
- **3.** Bindu S, Mazumder S, Bandyopadhyay U. Non-steroidal antiinflammatory drugs (NSAIDs) and organ damage: a current perspective. *Biochem Pharmacol* 2020:114147.
- Vane JR, Botting RM. Mechanism of action of nonsteroidal antiinflammatory drugs. Am J Med 1998;10:2S-8S.
- Vane JR, Botting RM. Mechanism of action of anti-inflammatory drugs. Scand J Rheumatol 1996;25(sup102):9–21.

- 6. Ricciotti E, FitzGerald GA. Prostaglandins and inflammation. Arterioscler Thromb Vasc Biol 2011;31:986–1000.
- 7. Morteau O. Prostaglandins and inflammation: the cyclooxygenase controversy. *Arch Immunol Ther Exp (Warsz)* 2000;**48**:473–80.
- Rossi A, Kapahi P, Natoli G, Takahashi T, Chen Y, Karin M, et al. Anti-inflammatory cyclopentenone prostaglandins are direct inhibitors of IκB kinase. *Nature* 2000;403:103–8.
- Funk CD. Prostaglandins and leukotrienes: advances in eicosanoid biology. *Science* 2001;294:1871–5.
- Smith WL, Dewitt DL. Prostaglandin endoperoxide H synthases-1 and -2. Adv Immunol 1996;62:167–215.
- Xie W, Robertson DL, Simmons DL. Mitogen-inducible prostaglandin G/H synthase: a new target for nonsteroidal antiinflammatory drugs. *Drug Dev Res* 1992;25:249–65.
- Nantel F, Denis D, Gordon R, Northey A, Cirino C, Chan CC. Distribution and regulation of cyclooxygenase-2 in carrageenan-induced inflammation. *Br J Pharmacol* 1999;**128**:853–9.
- Gupta AK, Gupta RA, Soni LK, Kaskhedikar SG. Exploration of physicochemical properties and molecular modelling studies of 2sulfonyl-phenyl-3-phenyl-indole analogs as cyclooxygenase-2 inhibitors. *Eur J Med Chem* 2008;43:1297–303.
- 14. Tanaka A, Araki H, Komoike Y, Hase S, Takeuchi K. Inhibition of both COX-1 and COX-2 is required for development of gastric damage in response to nonsteroidal antiinflammatory drugs. J Physiol Paris 2001;95:21–7.
- Ahlström MM, Ridderström M, Zamora I, Luthman K. CYP2C9 structure metabolism relationships: optimizing the metabolic stability of COX-2 inhibitors. *J Med Chem* 2007;50:4444–52.
- Wallace JL, McKnight W, Reuter BK, Vergnolle N. NSAID-induced gastric damage in rats: requirement for inhibition of both cyclooxygenase 1 and 2. *Gastroenterology* 2000;119:706–14.
- 17. Lamie PF, Ali WAM, Bazgier V, Rarova L. Novel N-substituted indole Schiff bases as dual inhibitors of cyclooxygenase-2 and 5lipoxygenase enzymes: synthesis, biological activities *in vitro* and docking study. *Eur J Med Chem* 2016;**123**:803–13.
- Ramalho TC, Rocha MVJ, da Cunha EFF, Freitas MP. The search for new COX-2 inhibitors: a review of 2002–2008 patents. *Expert Opin Ther Pat* 2009;50:1193–228.
- Meade EA, Smith WL, Dewitt DL. Differential inhibition of prostaglandin endoperoxide synthase (cyclooxygenase) isozymes by aspirin and other non-steroidal anti-inflammatory drugs. *J Biol Chem* 1993;268:6610–4.
- Charlier C, Michaux C. Dual inhibition of cyclooxygenase-2 (COX-2) and 5-lipoxygenase (5-LOX) as a new strategy to provide safer non-steroidal anti-inflammatory drugs. *Eur J Med Chem* 2003;38: 645–59.
- Somakala K, Amir M. Synthesis, characterization and pharmacological evaluation of pyrazolyl urea derivatives as potential antiinflammatory agents. *Acta Pharm Sin B* 2017;7:230–40.
- 22. Bansal S, Bala M, Suthar SK, Choudhary S, Bhattacharya S, Bhardwaj V, et al. Design and synthesis of novel 2-phenyl-5-(1,3-diphenyl-1*H*-pyrazol-4-yl)-1,3,4-oxadiazoles as selective COX-2 inhibitors with potent anti-inflammatory activity. *Eur J Med Chem* 2014;**80**:167–74.
- 23. El-Sayed MA, Abdel-Aziz NI, Abdel-Aziz AA, El-Azab AS, Asiri YA, Eltahir KE. Design, synthesis, and biological evaluation of substituted hydrazone and pyrazole derivatives as selective COX-2 inhibitors: molecular docking study. *Bioorg Med Chem* 2011;19:3416–24.
- 24. Yao HY, Guo Q, Wang MR, Wang R, Xu ZQ. Discovery of pyrazole *N*-aryl sulfonate: a novel and highly potent cyclooxygenase-2 (COX-2) selective inhibitors. *Bioorg Med Chem* 2021;46:116344.
- El-Sayed MAA, Abdel-Aziz NI, Abdel-Aziz AAM, El-Azab AS, ElTahir KEH. Synthesis, biological evaluation and molecular modeling study of pyrazole and pyrazoline derivatives as selective COX-2 inhibitors and anti-inflammatory agents, Part 2. *Bioorg Med Chem* 2012;20:3306–16.
- Tewari AK, Singh VP, Yadav P, Gupta G, Singh A, Goel RK, et al. Synthesis, biological evaluation and molecular modeling study of

pyrazole derivatives as selective COX-2 inhibitors and antiinflammatory agents. *Bioorg Chem* 2014;**56**:8–15.

- 27. Abdellatif KRA, Abdelall EKA, Labib MB, Fadaly WAA, Zidan TH. Synthesis of novel halogenated triarylpyrazoles as selective COX-2 inhibitors: anti-inflammatory activity, histopatholgical profile and in-silico studies. *Bioorg Chem* 2020;**105**:104418.
- Abdelall EK, Kamel GM. Synthesis of new thiazolo-Celecoxib analogues as dual cyclooxygenase-2/15-lipoxygenase inhibitors: determination of regio-specific different pyrazole cyclization by 2D NMR. *Eur J Med Chem* 2016;118:250–8.
- Mohammed KO, Nissan YM. Synthesis, molecular docking, and biological evaluation of some novel hydrazones and pyrazole derivatives as anti-inflammatory agents. *Chem Biol Drug Des* 2014;84: 473–88.
- **30.** Tyagi R. Imidazoline and its derivatives: an overview. *J Oleo Sci* 2007;**56**:211–22.
- Sarnpitak P, Mujumdar P, Morisseau C, Hwang SH, Hammock B, Iurchenko V, et al. Potent, orally available, selective COX-2 inhibitors based on 2-imidazoline core. *Eur J Med Chem* 2014;84: 160–72.
- Abdellatif KRA, Fadaly WAA. New 1,2-diaryl-4-substituted-benzylidene-5-4H-imidazolone derivatives: design, synthesis and biological evaluation as potential anti-inflammatory and analgesic agents. *Bioorg Chem* 2017;**72**:123–9.
- 33. Metwally NH, Mohamed MS. New imidazolone derivatives comprising a benzoate or sulfonamide moiety as anti-inflammatory and antibacterial inhibitors: design, synthesis, selective COX-2, DHFR and molecular-modeling study. *Bioorg Chem* 2020;99: 103438.
- Navidpour L, Amini M, Miri R, Firuzi O, Tavakkoli M, Shafiee A. Synthetic approaches towards the sulfonamide substituted-1,5-diarylimidazole-2-thiones as selective cyclooxygense-2 inhibitors. J *Heterocycl Chem* 2014;51:71–9.
- Liu H, Du DM. Recent advances in the synthesis of 2-imidazolines and their applications in homogeneous catalysis. *Adv Synth Catal* 2009;351:489–519.
- 36. Hayashi S, Ueno N, Murase A, Nakagawa Y, Takada J. Novel acidtype cyclooxygenase-2 inhibitors: design, synthesis, and structure– activity relationship for anti-inflammatory drug. *Eur J Med Chem* 2012;50:179–95.
- 37. Kaur J, Bhardwaj A, Huang Z, Knaus EE. N-1 and C-3 substituted indole Schiff bases as selective COX-2 inhibitors: synthesis and biological evaluation. *Bioorg Med Chem Lett* 2012;22:2154–9.
- Bhat MA, Al-Omar MA, Raish M, Ansari MA, Abuelizz HA, Bakheit AH, et al. Indole derivatives as cyclooxygenase inhibitors: synthesis, biological evaluation and docking studies. *Molecules* 2018;23:1250.
- 39. Singh P, Prasher P, Dhillon P, Bhatti R. Indole based peptidomimetics as anti-inflammatory and anti-hyperalgesic agents: dual inhibition of 5-LOX and COX-2 enzymes. *Eur J Med Chem* 2015;97: 104–23.
- 40. Estevão MS, Carvalho LC, Freitas M, Gomes A, Viegas A, Manso J, et al. Indole based cyclooxygenase inhibitors: synthesis, biological evaluation, docking and NMR screening. *Eur J Med Chem* 2012;54: 823–33.
- 41. Ju ZR, Su M, Hong J, La Kim E, Moon HR, Chung HY, et al. Design of balanced COX inhibitors based on anti-inflammatory and/or COX-2 inhibitory ascidian metabolites. *Eur J Med Chem* 2019;**180**:86–98.
- Vazzana I, Terranova E, Mattioli F, Sparatore F. Aromatic Schiff bases and 2,3-disubstituted-1,3-thiazolidin-4-one derivatives as antiinflammatory agents. *Arkivoc* 2004;5:364–74.
- 43. Sağlık BN, Osmaniye D, Levent S, Çevik UA, Çavusoğlu BK, Ozkay Y, et al. Design, synthesis and biological assessment of new selective COX-2 inhibitors including methyl sulfonyl moiety. *Eur J Med Chem* 2021;209:112918.
- 44. Abdel-Aziz SA, Taher ES, Lan P, Asaad GF, Gomaa HAM, El-Koussi NA, et al. Design, synthesis, and biological evaluation of new

pyrimidine-5-carbonitrile derivatives bearing 1,3-thiazole moiety as novel anti-inflammatory EGFR inhibitors with cardiac safety profile. *Bioorg Chem* 2021;**111**:104890.

- **45.** Rodl CB, Vogt D, Kretschmer SB, Ihlefeld K, Barzen S, Bruggerhoff A, et al. Multi-dimensional target profiling of *N*,4-diaryl-1,3-thiazole-2-amines as potent inhibitors of eicosanoid metabolism. *Eur J Med Chem* 2014;**84**:302–11.
- **46.** Labib MB, Fayez AM, El-Nahass ES, Awadallah M, Halim PA. Novel tetrazole-based selective COX-2 inhibitors: design, synthesis, anti-inflammatory activity, evaluation of PGE2, TNF- α , IL-6 and histopathological study. *Bioorg Chem* 2020;**104**:104308.
- 47. Al-Hourani BJ, Sharma SK, Mane JY, Tuszynski J, Baracos V, Kniess T, et al. Synthesis and evaluation of 1,5-diaryl-substituted tetrazoles as novel selective cyclooxygenase-2 (COX-2) inhibitors. *Bioorg Med Chem Lett* 2011;21:1823–6.
- 48. Al-Hourani BJ, Al-Awaida W, Matalka KZ, El-Barghouthi MI, Alsoubani F, Wuest F. Structure–activity relationship of novel series of 1,5-disubstituted tetrazoles as cyclooxygenase-2 inhibitors: design, synthesis, bioassay screening and molecular docking studies. *Bioorg Med Chem Lett* 2016;26:4757–62.
- 49. El-Sayed NA, Nour MS, Salem MA, Arafa RK. New oxadiazoles with selective-COX-2 and EGFR dual inhibitory activity: design, synthesis, cytotoxicity evaluation and *in silico* studies. *Eur J Med Chem* 2019;183:111693.
- 50. Alfayomy AM, Abdel-Aziz SA, Marzouk AA, Shaykoon MSA, Narumi A, Konno H, et al. Design and synthesis of pyrimidine-5carbonitrile hybrids as COX-2 inhibitors: anti-inflammatory activity, ulcerogenic liability, histopathological and docking studies. *Bioorg Chem* 2021;108:104555.
- Grover J, Bhatt N, Kumar V, Patel NK, Gondaliya BJ, Elizabeth Sobhia M, et al. 2,5-Diaryl-1,3,4-oxadiazoles as selective COX-2 inhibitors and anti-inflammatory agents. *RSC Adv* 2015;5:45535–44.
- 52. Ł Szczukowski, Krzyżak E, Zborowska A, Zajac P, Potyrak K, Peregrym K, et al. Design, synthesis and comprehensive investigations of pyrrolo [3,4-d] pyridazinone-based 1,3,4-oxadiazole as new class of selective COX-2 inhibitors. *Int J Mol Sci* 2020;21: 9623.
- 53. Khatri CK, Indalkar KS, Patil CR, Goyal SN, Chaturbhuj GU. Novel 2-phenyl-4,5,6,7-tetrahydro[b]benzothiophene analogues as selective COX-2 inhibitors: design, synthesis, anti-inflammatory evaluation, and molecular docking studies. *Bioorg Med Chem Lett* 2017;27: 1721–6.
- 54. Sun JA, Wang S, Sheng GH, Lian ZM, Liu HY, Zhu HL. Synthesis of phenylpiperazine derivatives of 1,4-benzodioxan as selective COX-2 inhibitors and anti-inflammatory agents. *Bioorg Med Chem* 2016;24: 5626–32.
- 55. Kaur A, Pathak DP, Sharma V, Wakode S. Synthesis, biological evaluation and docking study of a new series of di-substituted benzoxazole derivatives as selective COX-2 inhibitors and antiinflammatory agents. *Bioorg Med Chem* 2018;26:891–902.
- 56. Kaur A, Pathak DP, Sharma V, Narasimhan B, Sharma P, Mathur R, et al. Synthesis, biological evaluation and docking study of *N*-(2-(3,4,5-trimethoxybenzyl)benzoxazole-5-yl) benzamide derivatives as selective COX-2 inhibitor and anti-inflammatory agents. *Bioorg Chem* 2018;81:191–202.
- 57. Chen Z, Wang ZC, Yan XQ, Wang PF, Lu XY, Chen LW, et al. Design, synthesis, biological evaluation and molecular modeling of dihydropyrazole sulfonamide derivatives as potential COX-1/COX-2 inhibitors. *Bioorg Med Chem Lett* 2015;25:1947–51.
- Rathore A, Rahman MU, Siddiqui AA, Ali A, Shaharyar M. Design and synthesis of benzimidazole analogs endowed with oxadiazole as selective COX-2 inhibitor. *Arch Pharm* 2014;347:923–35.
- **59.** Iyer VB, Gurupadayya B, Koganti VS, Inturi B, Chandan RS. Design, synthesis and biological evaluation of 1,3,4-oxadiazoles as promising anti-inflammatory agents. *Med Chem Res* 2016;**26**: 190–204.
- **60.** Nesaragi AR, Kamble RR, Dixit S, Kodasi B, Hoolageri SR, Bayannavar PK, et al. Green synthesis of therapeutically active 1,3,4-

oxadiazoles as antioxidants, selective COX-2 inhibitors and their *in silico* studies. *Bioorg Med Chem Lett* 2021;**43**:128112.

- Abdu-Allah HHM, Abdelmoez AAB, Tarazi H, El-Shorbagi AA, El-Awady R. Conjugation of 4-aminosalicylate with thiazolinones afforded non-cytotoxic potent *in vitro* and *in vivo* anti-inflammatory hybrids. *Bioorg Chem* 2020;94:103378.
- Murahari M, Mahajan V, Neeladri S, Kumar MS, Mayur YC. Ligand based design and synthesis of pyrazole based derivatives as selective COX-2 inhibitors. *Bioorg Chem* 2019;86:583–97.
- 63. Ibrahim TS, Salem IM, Mostafa SM, El-Sabbagh OI, ElKhamisi MKM, Hegazy L, et al. Design, synthesis, and pharmacological evaluation of novel and selective COX-2 inhibitors based on bumetanide scaffold. *Bioorg Chem* 2020;100:103878.
- 64. Ikeda A, Funakoshi E, Araki M, Ma B, Karuo Y, Tarui A, et al. Structural modification of indomethacin toward selective inhibition of COX-2 with a significant increase in van der Waals contributions. *Bioorg Med Chem* 2019;27:1789–94.
- 65. Chandna N, Kumar S, Kaushik P, Kaushik D, Roy SK, Gupta GK, et al. Synthesis of novel celecoxib analogues by bioisosteric replacement of sulfonamide as potent anti-inflammatory agents and cyclooxygenase inhibitors. *Bioorg Med Chem* 2013;21:4581–90.
- 66. Hassan GS, Abou-Seri SM, Kamel G, Ali MM. Celecoxib analogs bearing benzofuran moiety as cyclooxygenase-2 inhibitors: design, synthesis and evaluation as potential anti-inflammatory agents. *Eur J Med Chem* 2014;76:482–93.
- 67. Renard JF, Lecomte F, Hubert P, de Leval X, Pirotte B. N-(3-Arylaminopyridin-4-yl)alkanesulfonamides as pyridine analogs of nimesulide: cyclooxygenases inhibition, anti-inflammatory studies and insight on metabolism. *Eur J Med Chem* 2014;74:12–22.
- **68.** Ribeiro D, Proenca C, Varela C, Janela J, Tavares da Silva EJ, Fernandes E, et al. New phenolic cinnamic acid derivatives as selective COX-2 inhibitors. Design, synthesis, biological activity and structure-activity relationships. *Bioorg Chem* 2019;**91**:103179.
- Takahashi T, Miyazawa M. N-Caffeoyl serotonin as selective COX-2 inhibitor. *Bioorg Med Chem Lett* 2012;22:2494–6.
- Rayar AM, Lagarde N, Martin F, Blanchard F, Liagre B, Ferroud C, et al. New selective cyclooxygenase-2 inhibitors from cyclocoumarol: synthesis, characterization, biological evaluation and molecular modeling. *Eur J Med Chem* 2018;**146**:577–87.
- Čulenová M, Sychrová A, Hassan STS, Berchová-Bímová K, Svobodová P, Helclová A, et al. Multiple *in vitro* biological effects of phenolic compounds from *Morus alba* root bark. *J Ethnopharmacol* 2020;248:112296.
- 72. Honmore VS, Kandhare AD, Kadam PP, Khedkar VM, Sarkar D, Bodhankar SL, et al. Isolates of *Alpinia officinarum* Hance as COX-2 inhibitors: evidence from anti-inflammatory, antioxidant and molecular docking studies. *Int Immunopharm* 2016;**33**:8–17.
- **73.** Liu Y, Zhang YL, Muema FW, Kimutai F, Chen GL, Guo MQ. Phenolic compounds from *Carissa spinarum* are characterized by their antioxidant, anti-inflammatory and hepatoprotective activities. *Antioxidants* 2021;**10**:652.
- 74. Nile SH, Ko EY, Kim DH, Keum YS. Screening of ferulic acid related compounds as inhibitors of xanthine oxidase and cyclooxygenase-2 with anti-inflammatory activity. *Rev Bras Farmacogn* 2016;26:50–5.
- Paulino M, Alvareda E, Iribarne F, Miranda P, Espinosa V, Aguilera S, et al. Toward the understanding of the molecular basis for the inhibition of COX-1 and COX-2 by phenolic compounds present in Uruguayan propolis and grape pomace. *J Biomol Struct Dyn* 2016; 34:2643–57.
- 76. Dai CS, Ciccotosto GD, Cappai R, Tang S, Li D, Xie S, et al. Curcumin attenuates colistin-induced neurotoxicity in N2a cells *via* antiinflammatory activity, suppression of oxidative stress, and apoptosis. *Mol Neurobiol* 2018;55:421–34.
- Timmers MA, Guerrero-Medina JL, Esposito D, Grace MH, Paredes-Lopez O, Garcia-Saucedo PA, et al. Characterization of phenolic compounds and antioxidant and anti-inflammatory activities from Mamuyo (*Styrax ramirezii* Greenm.) Fruit. *J Agric Food Chem* 2015; 63:10459–65.

- Bai HF, Li YP, Qin FY, Yan YM, Wang SM, Zhang HX, et al. Periplanetols A-F, phenolic compounds from *Periplaneta americana* with potent COX-2 inhibitory activity. *Fitoterapia* 2020;**143**:104589.
- 79. Hu WH, Dai DK, Zheng BZ, Duan R, Chan GK, Dong TT, et al. The binding of kaempferol-3-O-rutinoside to vascular endothelial growth factor potentiates anti-inflammatory efficiencies in lipopolysaccharidetreated mouse macrophage RAW264.7 cells. *Phytomedicine* 2021;80: 153400.
- 80. Wu X, Song M, Rakariyatham K, Zheng J, Wang M, Xu F, et al. Inhibitory effects of 4'-demethylnobiletin, a metabolite of nobiletin, on 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced inflammation in mouse ears. J Agric Food Chem 2015;63:10921–7.
- Kim HH, Kim DH, Kim MH, Oh MH, Kim SR, Park KJ, et al. Flavonoid constituents in the leaves of *Myrica rubra* sieb. et zucc. with anti-inflammatory activity. *Arch Pharmacal Res* 2013;36:1533–40.
- 82. Hanákova Z, Hosek J, Kutil Z, Temml V, Landa P, Vanek T, et al. Anti-inflammatory activity of natural geranylated flavonoids: cyclooxygenase and lipoxygenase inhibitory properties and proteomic analysis. J Nat Prod 2017;80:999–1006.
- 83. Hosek J, Toniolo A, Neuwirth O, Bolego C. Prenylated and geranylated flavonoids increase production of reactive oxygen species in mouse macrophages but inhibit the inflammatory response. *J Nat Prod* 2013;**76**:1586–91.
- 84. Bai M, Yao GD, Ren Q, Li Q, Liu QB, Zhang Y, et al. Triterpenoid saponins and flavonoids from licorice residues with antiinflammatory activity. *Ind Crop Prod* 2018;125:50–8.
- Sun YW, Bao YG, Yu H, Chen QJ, Lu F, Zhai S, et al. Anti-rheumatoid arthritis effects of flavonoids from Daphne genkwa. *Int Immunopharm* 2020;83:106384.
- Chen GL, Fan MX, Wu JL, Li N, Guo MQ. Antioxidant and antiinflammatory properties of flavonoids from lotus plumule. *Food Chem* 2019;277:706–12.
- 87. Cheon SY, Chung KS, Jeon E, Nugroho A, Park HJ, An HJ. Antiinflammatory activity of Saxifragin *via* inhibition of NF-κB involves caspase-1 activation. *J Nat Prod* 2015;**78**:1579–85.
- Ma JQ, Ding J, Xiao ZH, Liu CM. Puerarin ameliorates carbon tetrachloride-induced oxidative DNA damage and inflammation in mouse kidney through ERK/Nrf2/ARE pathway. *Food Chem Toxicol* 2014;71:264–71.
- 89. Tamayose CI, Romoff P, Toyama DO, Gaeta HH, Costa CRC, Belchor MN, et al. Non-clinical studies for evaluation of 8-Crhamnosyl apigenin purified from *Peperomia obtusifolia* against acute edema. *Int J Molecul Sci* 2017;18:1972.
- 90. Waller CP, Thumser AE, Langat MK, Crouch NR, Mulholland DA. COX-2 inhibitory activity of homoisoflavanones and xanthones from the bulbs of the Southern African *Ledebouria socialis* and *Ledebouria ovatifolia* (Hyacinthaceae: Hyacinthoideae). *Phytochemistry* 2013;95:284–90.
- Kim AY, Shim HJ, Kim SY, Heo S, Youn HS. Differential regulation of MyD88- and TRIF-dependent signaling pathways of Toll-like receptors by cardamonin. *Int Immunopharm* 2018;64:1–9.
- 92. Zhou YS, Zhong BL, Min XJ, Hou Y, Lin LG, Wu Q, et al. Therapeutic potential of isobavachalcone, a natural flavonoid, in murine experimental colitis by inhibiting NF-κB p65. *Phytother Res* 2021; 35:5861–70.
- Bairwa K, Singh IN, Roy SK, Grover J, Srivastava A, Jachak SM. Rotenoids from *Boerhaavia diffusa* as potential anti-inflammatory agents. J Nat Prod 2013;76:1393-8.
- 94. Crockett SL, Kunert O, Pferschy-Wenzig EM, Jacob M, Schuehly W, Bauer R. Phloroglucinol and terpenoid derivatives from *Hypericum cistifolium* and *H. galioides* (*Hypericaceae*). *Front Plant Sci* 2016;**7**:961.
- 95. Zhang YJ, Huang XS, Chen HC, Zhou DX, Yang ZM, Wang K, et al. Discovery of anti-inflammatory terpenoids from *Mallotus con-spurcatus* croizat. *J Ethnopharmacol* 2019;231:170–8.
- 96. Yoon CS, Kim DC, Park JS, Kim KW, Kim YC, Oh H. Isolation of novel sesquiterpeniods and anti-neuroinflammatory metabolites from *Nardostachys jatamansi. Molecules* 2018;23:2367.

- 97. An HJ, Kim IT, Park HJ, Kim HM, Choi JH, Lee KT. Tormentic acid, a triterpenoid saponin, isolated from *Rosa rugosa*, inhibited LPSinduced iNOS, COX-2, and TNF-alpha expression through inactivation of the nuclear factor-κB pathway in RAW 264.7 macrophages. *Int Immunopharm* 2011;11:504–10.
- Lv JL, Li Z, Guo LM. Sesquiterpene lactones with COX-2 inhibition activity from Artemisia lavandulaefolia. Chem Biodivers 2018;15: E1700548.
- 99. Nan L, Nam HH, Choo BK. Costunolide inhibits inflammation in LPSinduced RAW264.7 cells and ameliorates gastric acid reflux-induced esophageal injury in rat model. *Appl Biol Chem* 2020;63:1–9.
- 100. Yin X, Wei J, Wang WW, Gao YQ, Stadler M, Kou RW, et al. New cyathane diterpenoids with neurotrophic and anti-neuroinflammatory activity from the bird's nest fungus *Cyathus africanus*. *Fitoterapia* 2019;**134**:201–9.
- 101. Nhiem NX, Lim HY, Kiem PV, Minh CV, Thu VK, Tai BH, et al. Oleanane-type triterpene saponins from the bark of *Aralia elata* and their NF-κB inhibition and PPAR activation signal pathway. *Bioorg Med Chem Lett* 2011;21:6143–7.
- 102. Li JC, Dai WF, Liu D, Zhang ZJ, Jiang MY, Rao KR, et al. Quinolizidine alkaloids from *Sophora alopecuroides* with anti-inflammatory and anti-tumor properties. *Bioorg Chem* 2021;110:104781.
- 103. Feng AW, Gao W, Zhou GR, Yu R, Li N, Huang XL, et al. Berberine ameliorates COX-2 expression in rat small intestinal mucosa partially through PPARgamma pathway during acute endotoxemia. *Int Immunopharm* 2012;12:182–8.
- 104. Kang JJ, Zhang YS, Cao X, Fan J, Li GL, Wang Q, et al. Lycorine inhibits lipopolysaccharide-induced iNOS and COX-2 up-regulation in RAW264.7 cells through suppressing P38 and STATs activation and increases the survival rate of mice after LPS challenge. *Int Immunopharm* 2012;12:249–56.
- 105. Mohan S, Hobani YH, Shaheen E, Abou-Elhamd AS, Abdelhaleem A, Alhazmi HA, et al. Girinimbine from curry leaves promotes gastro protection against ethanol induced peptic ulcers and improves healing *via* regulation of anti-inflammatory and antioxidant mechanisms. *Food Funct* 2020;11:3493–505.
- 106. Zhou K, Hu L, Liao WJ, Yin DF, Rui F. Coptisine prevented IL-betainduced expression of inflammatory mediators in chondrocytes. *Inflammation* 2016;**39**:1558–65.
- 107. Son DJ, Akiba S, Hong JT, Yun YP, Hwang SY, Park YH, et al. Piperine inhibits the activities of platelet cytosolic phospholipase A2 and thromboxane A2 synthase without affecting cyclooxygenase-1 activity: different mechanisms of action are involved in the inhibition of platelet aggregation and macrophage inflammatory response. *Nutrients* 2014;6:3336–52.
- 108. Lin YF, Kuo CY, Wen ZH, Lin YY, Wang WH, Su JH, et al. Flexibilisquinone, a new anti-inflammatory quinone from the cultured soft coral Sinularia flexibilis. *Molecules* 2013;18:8160–7.
- **109.** Choi RJ, Ngoc TM, Bae K, Cho HJ, Kim DD, Chun J, et al. Antiinflammatory properties of anthraquinones and their relationship with the regulation of P-glycoprotein function and expression. *Eur J Pharm Sci* 2013;**48**:272–81.
- 110. Ju ZR, Qin XC, Lin XP, Wang JF, Kaliyaperumal K, Tian YQ, et al. New phenyl derivatives from endophytic fungus *Botryosphaeria* sp. SCSIO KcF6 derived of mangrove plant *Kandelia candel*. *Nat Prod Res* 2016;**30**:192–8.
- 111. Zhang SJ, Huang YY, Li Y, Wang YH, He XJ. Anti-neuroinflammatory and antioxidant phenylpropanoids from Chinese olive. *Food Chem* 2019;286:421-7.
- 112. Li YR, Fu CS, Yang WJ, Wang XL, Feng D, Wang XN, et al. Investigation of constituents from *Cinnamomum camphora* (L.) J. Presl and evaluation of their anti-inflammatory properties in lipopolysaccharide-stimulated RAW 264.7 macrophages. *J Ethnopharmacol* 2018;221:37–47.
- 113. Thi NN, Song HS, Oh EJ, Lee YG, Ko JH, Kwon JE, et al. Phenylpropanoids from *Lilium* Asiatic hybrid flowers and their antiinflammatory activities. *Appl biol chem* 2017;**60**:527–33.