



Identification of Prognostic Biomarkers for Multiple Solid Tumors Using a Human Villi Development Model

Botao Zhang^{1,2}, Yuanjing Wang³, Hongxia Li⁴, Lin Feng^{2*}, Wenbin Li^{1*} and Shujun Cheng^{2*}

¹ Department of Neuro-oncology, Neurosurgery Center, Beijing Tiantan Hospital, Capital Medical University, Beijing, China, ² State Key Laboratory of Molecular Oncology, Department of Etiology and Carcinogenesis, National Cancer Center/National Clinical Research Center for Cancer/Cancer Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China, ³ Department of Gynecological Oncology, Beijing Obstetrics and Gynecology Hospital, Capital Medical University, Beijing, China, ⁴ Department of Obstetrics and Gynecology, Beijing Shijitan Hospital, Capital Medical University, Beijing, China

The processes of embryonic development that rely on epithelial-mesenchymal transition (EMT) for the implantation of trophoblast cells are co-opted by tumors, reflecting their inherent uncontrolled characteristics and leading to invasion and metastasis. Although tumorigenesis and embryogenesis have similar EMT characteristics, trophoblasts have been shown to exhibit "physiological metastasis" or be "pseudo-malignant," resulting in different outcomes. The gene co-expression network is the basis of embryonic development and tumorigenesis. We hypothesize that if the gene co-expression network in tumors is "off-track" from that in villi, it is more likely to develop into malignant tumors and have a worse prognosis, and we proposed the "off-track theory" for the first time. In this study, we examined gene co-expression networks in villi and multiple solid tumors. Through network functional enrichment analyses, we found that most tumors and villi exhibited a significantly enriched EMT, but the genes that performed this function were not identical. Then, we identified the "off-track genes" in the EMT-related gene interaction network using the "off-track theory," and through survival analysis, we discovered that the risk score of "off-track genes" was associated with poor survival of cancer patients. Our study indicated that villi development is a reliable and strictly regulated model that can illuminate the trajectory of human cancer development and that the gene co-expression networks in tumor development are "off-track" from those in villi. These "off-track genes" may have a substantial impact on tumor development and could reveal novel prognostic biomarkers.

Keywords: villi, solid tumor, gene co-expression networks, off-track, prognosis

OPEN ACCESS

Edited by:

Luisa Lanfrancone, European Institute of Oncology (IEO), Italy

Reviewed by:

Jürgen Pollheimer, Medical University of Vienna, Austria Daniele Vergara, University of Salento, Italy

*Correspondence:

Lin Feng fenglin@cicams.ac.cn Wenbin Li liwenbin@ccmu.edu.cn Shujun Cheng chengshj@cicams.ac.cn

Specialty section:

This article was submitted to Molecular and Cellular Oncology, a section of the journal Frontiers in Cell and Developmental Biology **Received:** 17 February 2020

Accepted: 25 May 2020 Published: 23 June 2020

Citation:

Zhang B, Wang Y, Li H, Feng L, Li W and Cheng S (2020) Identification of Prognostic Biomarkers for Multiple Solid Tumors Using a Human Villi Development Model. Front. Cell Dev. Biol. 8:492. doi: 10.3389/fcell.2020.00492

INTRODUCTION

Cancer is a complex disease with various pathologic properties (Machnik et al., 2019), and cancer incidence and mortality have been increasing, making cancer the leading cause of death and a major public health problem worldwide (Chen et al., 2016; Siegel et al., 2018). Although multiple forms of treatment exist for cancers, such as surgery, chemotherapy, radiotherapy, targeted therapy, and so on, the prognosis of patients with cancer is still poor, and cancer still poses a serious threat to human health and affects the quality of life of patients (Kovar et al., 2016; Gigic et al., 2018). In addition, there is a strong heterogeneity in tumor tissues, and the genome is extremely unstable (Burrell et al., 2013; Troiani et al., 2014). Therefore, a model similar to the biological behavior and molecular characteristics of tumor cells but without heterogeneity is urgently needed to study tumors.

Tumorigenesis and placental villi development have many similar biological behaviors, especially during the early stage of pregnancy (Strickland and Richards, 1992; Holtan et al., 2009; Perry et al., 2009). The epithelial-mesenchymal transition (EMT) plays an important role in embryonic development and cancer progression (Chaffer et al., 2016; Simeone et al., 2019). The EMT is a fundamental cell biological process that plays a critical role in embryogenesis, which was reactivated during cancer progression and enhances the metastatic phenotype. Implantation of trophoblast cells of the placenta into the decidual and muscular layers of the uterus is the sine qua non of a successful pregnancy, and trophoblast cells promote implantation through EMT, which results in loss of cell-tocell contact inhibition (Holtan et al., 2009). EMT of tumor cells in cancer progression possibly generates the different cell types of the tumor stroma, such as cancer-associated myofibroblasts. EMT contributes to new tumor cell properties required for invasiveness and vascular intravasation during metastasis (Moustakas and Heldin, 2010).

The processes of implantation during placental development that rely on EMT are co-opted by tumors, reflecting their inherent uncontrolled characteristic, which leads to invasion and metastasis (Hay, 2005; Huber et al., 2005; Jean Paul and Sleeman, 2006). However, tumorigenesis and placental development have different outcomes. Placental development is a physiological process; the placenta is an important organ used to nourish the fetus during pregnancy, and the EMT process is strictly regulated. However, tumorigenesis is a pathological process, and the EMT process is not regulated by the host. Disequilibrium is the key feature of cancer occurrence and development (Hanahan and Weinberg, 2011; Macklin et al., 2017). In addition, the gene regulatory network induces the expression of other target genes via the protein products of differentially expressed genes, which is the basis of many biological processes, such as embryonic development and tumorigenesis (Singh et al., 2018). During the process of placental development, a strong correlation exists between different genes, suggesting that all biological functions could be modulated through the precise regulation of gene networks (An et al., 2015). However, during the process of tumor progression, close gene regulation relationships are abolished, and the correlations between genes responsible for normal

placental development are disrupted. Based on this principle, we proposed the "off-track theory" for the first time. Such "offtrack genes" are likely to promote the progression of cancer and may also predict the prognosis of cancer patients. Therefore, we deemed that in placental villi development and tumorigenesis, the gene regulatory networks are altered, resulting in gene coexpression disorders and different outcomes. In summary, when the gene co-expression network in tumors is "off-track" from that in villi development, cells are more likely to develop into malignant tumors, or have a worse prognosis.

In this study, we utilized RNA sequencing data combined with RNA transcriptome data of multiple solid tumors from The Cancer Genome Atlas (TCGA) to construct gene coexpression networks for chorionic villi and multiple solid tumors, respectively. Then, according to the "off-track theory," we identified "off-track genes" from the EMT-related gene coexpression networks and then screened "off-track genes" that were highly expressed in cancers. Afterward, we performed a survival analysis to assess whether these "off-track genes" could predict the prognosis of cancer patients.

Our research indicated that placental villi development is a reliable and strictly regulated model. The villi development model could produce new insights for the understanding of tumors and provide new methods for cancer research. Additionally, through the different co-expression relationships between physiological and pathological states, we found new biomarkers which may be helpful for the treatment and posttreatment monitoring of cancers.

MATERIALS AND METHODS

Patients, Tissue Samples, and TCGA RNA Sequence Data Curation

Tissue samples of developing villi were obtained from Beijing Shijitan Hospital between March 2015 and August 2016. The samples included 15 chorionic villus samples at 6 to 10 weeks of gestation (3 samples at each time point) and 6 leaf chorionic samples from postpartum placental tissue representing the mature placenta. The detailed information regarding those samples is presented in **Supplementary Table S1**. The inclusion

TABLE 1 | The sample numbers of these 11 types of tumors.

| Tumor | Stage I/WHC | Stage II/WHC | Stage III/WH | Ottalge IV/WH | 10 N A | Normal |
|--------|-------------|--------------|--------------|---------------|---------------|--------|
| BLCA | 2 | 130 | 140 | 134 | 2 | 19 |
| BRCA | 181 | 620 | 250 | 20 | 22 | 112 |
| COAD | 44 | 108 | 79 | 39 | 9 | 41 |
| EAC | 12 | 23 | 29 | 5 | 20 | 11 |
| ESCC | 7 | 55 | 27 | 4 | 2 | 11 |
| GBMLGG | 0 | 515 | 0 | 152 | 0 | 5 |
| HNSC | 27 | 74 | 81 | 266 | 72 | 44 |
| LUAD | 269 | 120 | 80 | 25 | 8 | 59 |
| LUSC | 244 | 162 | 84 | 7 | 4 | 51 |
| READ | 12 | 26 | 33 | 13 | 9 | 10 |
| STAD | 52 | 121 | 165 | 37 | 13 | 35 |

criteria for the pregnant women have been described previously (Zhang et al., 2017). All donors signed informed consent forms. This study was reviewed and approved by the Ethics Committee of the National Cancer Center/Cancer Hospital, Chinese Academy of Medical Sciences, and Peking Union Medical College.

For TCGA datasets, RNA sequencing data of bladder urothelial carcinoma (BLCA), breast invasive carcinoma (BRCA), colon adenocarcinoma (COAD), rectum adenocarcinoma (READ), esophageal adenocarcinoma (EAC), lung adenocarcinoma (LUAD), stomach adenocarcinoma (STAD), glioma (GBMLGG), head and neck squamous cell carcinoma (HNSC), esophageal squamous cell carcinoma (ESCC), and lung squamous cell carcinoma (LUSC) and the corresponding clinical information were downloaded through the R package "TCGA2STAT." Detailed information on these 11 types of tumors is shown in **Table 1**.

We also collected datasets from the Gene Expression Omnibus (GEO) and Chinese Glioma Genome Atlas (CGGA) for validation, including 3 independent sets of COAD (GSE14333, GSE12945, and GSE17536), 3 independent sets of LUAD (GSE30219, GSE13213, and GSE68465), 3 independent sets



| Tissue type | <i>p</i> -value |
|-------------|-----------------|
| villi.early | 0.00000 |
| BLCA | 0.00000 |
| BRCA | 0.00000 |
| COAD | 0.00272 |
| ECA | 0.01548 |
| ESCC | 0.00001 |
| GBMLGG | 0.00083 |
| HNSC | 0.00002 |
| LUAD | 0.00000 |
| LUSC | 0.26326 |
| READ | 0.00000 |
| STAD | 0.00000 |

of GBMLGG (GSE109857, GSE16011, and CGGA), and their corresponding clinical information. All of the datasets were downloaded and analyzed directly.

RNA-Seq and Analysis

Total RNA was isolated from frozen chorionic villus and mature placenta tissues with TRIzol reagent (Thermo Fisher, United States) according to the manufacturer's instructions. A complementary DNA library was prepared, and sequencing was performed according to the Illumina standard protocol by Beijing Novel Bioinformatics Co., Ltd.¹ Raw reads from the RNAseq libraries were trimmed to remove the adaptor sequence, reads with adaptor contaminants, and low-quality reads. The indexes of the reference genome were built using Salmon, and the pairedend clean reads were aligned to the reference genome using UCSC version hg19. The transcripts per million (TPM) value of each gene was calculated based on the gene read counts mapped to that gene. The raw sequence data of villus tissues reported in this paper have been deposited in the Genome Sequence Archive (Genomics, Proteomics & Bioinformatics 2017) in the Beijing Institute of Genomics (BIG) Data Center (Nucleic Acids Res 2018), BIG, Chinese Academy of Sciences, under the accession number HRA000050.

¹https://en.novogene.com/

FIGURE 2 UpSet plot of the intersection of EMT-related genes between villi and cancers. The transverse bar graph at the bottom left shows the number of EMT-related genes in each set. The black points in the dot matrix at the bottom left and in the bar graph at the top indicate genes included in the correlation set. The bars above represent the number of genes corresponding to each intersection.

Construction of Gene Co-expression Networks and Enrichment Analysis

For chorionic villus and multiple cancer datasets, we used the "MatrixEQTL" package in R to compute all pairwise associations of gene expression levels and to estimate false discovery rates (FDRs; Shabalin, 2012). To increase the computational speed, FDRs were only reported for interactions achieving a raw *p*-value less than 0.001, and in subsequent analyses the self-interactions were removed from the co-expression networks.

For chorionic villus and multiple cancer gene co-expression interactions, we selected the 10,000 most significant coexpression relationships to build the networks and perform enrichment analysis. We used the "iGraph" package to compute each node's degree (Netzer et al., 2011) and the "RedeR" package to visualize the networks (Castro, 2012). Functional enrichment analyses of networks were performed with the Spatial Analysis of Network Associations (SANTA) package using custom gene sets (Cornish and Markowetz, 2014), including 50 hallmark gene sets from the Molecular Signatures Database (MSigDB).

Identification of "Off-Track Genes"

Villi development and cancer progression exhibit many similar biological behaviors but have distinct outcomes. For the first time,

we proposed the "off-track theory" to explore the genes that are out of balance in cancers.

First, genes related to EMT were assessed in villi and tumors. We supposed that Pearson's correlation coefficient of a gene-pair (gene A, B) in the villi is x, and that of the gene-pair in cancer is y. Then, we projected this onto a coordinate axis as a point (x, y). When the correlation coefficients were equal, the point was projected on the line y = x in the coordinate axis. When a point (x, y) deviated from the line y = x, this indicated that the gene interaction was more "off-track" in cancer progression than in the normal physiological process, resulting in a larger contribution to the synergistic disorder. We defined genes with a distance from the point (x, y) to the line y = x greater than 1 as "off-track genes."

To further identify genes related to the prognosis of cancer patients among "off-track genes," we used Student's *t*-test to screen for genes that are highly expressed in cancers.

Survival Analysis

To explore the genes that could predict the prognosis of cancer patients, a risk factor score was calculated to assess the survival of patients. In brief, we used a univariate Cox regression analysis to evaluate the association between survival time and the expression levels of those genes. A mathematical formula was constructed to

predict survival (Li et al., 2016), and the risk score of each patient was calculated as follows:

$$Risk_Score = \sum_{i=1}^{n} \beta_i * expression (genes)$$

 β_i represents the regression coefficient of each gene expression value. All patients in the datasets were thus assigned to high-risk and low-risk groups using the median risk score as the cut-off point. The Kaplan-Meier method was used to estimate the overall survival (OS) time for the two subgroups, and differences in survival time were analyzed using the log-rank test (R package "survival").

Statistical Analysis

All statistical analyses in this study were performed with R software. The UpSet plot was generated by the R package "UpSetR." All statistical tests were two-sided, and p < 0.05 was considered significant.

RESULTS

Computation of Gene Co-expression Relationships in Villi and Tumors and Functional Enrichment Analyses

For each gene in villi and tumors, we used the Matrix eQTL engine function in the "MatrixEQTL" package to calculate the correlation between each row of the two matrices by linear regression, in other words the associations between pairwise gene expression levels, and tested its significance with t statistics. To enable quantitative comparisons of gene co-expression relationships in villi and cancer, we standardized the number of edges in each network by constructing networks from the most significant 10,000 interactions in villi and tumors.

To visualize the gene co-expression networks, we used the "RedeR" package and limited the analysis to genes with a network degree greater than ten (**Figure 1**). To systematically test for network functional enrichment and to directly compare network functional enrichment in villi and tumors, we used the SANTA method to test the association between a query gene set and a network, enabling the functional annotation of networks. By determining the association between hallmark gene sets and networks in villi and cancers, these results demonstrated that the villi and cancer co-expression networks were enriched for EMT gene sets, and the *p*-values of EMT enrichment in villi and 11 cancers are shown in **Table 2**.

Identification of "Off-Track" and Highly Expressed Genes in Cancers

The developing placental villi and cancers networks were both enriched in EMT; in other words, they undergo a similar EMT process, but the genes that perform this function are not identical (**Figure 2**). For example, 49 genes were involved in the EMT in villi development, 38 genes were involved in BRCA, and 16 EMT-related genes were involved in both processes, accounting for 32.7% and 42.1% of the EMT genes involved in villi and BRCA, respectively. Then, according to the "off-track theory," we identified the "off-track genes" in multiple cancers (**Figure 3**).

To further screen genes to assess the prognosis of cancer patients, we used Student's *t*-test to identify differentially expressed genes in cancers. The genes that were significantly upregulated and downregulated in multiple cancers are shown in **Table 3**.

Prognostic Significance of "Off-Track Genes" in Cancer Patients

To determine whether these "off-track genes" were related to the prognosis of cancer patients, we used Kaplan-Meier curves for the analysis. A log-rank test confirmed that the risk score of "off-track genes" was significantly related to the OS of cancer patients, and a higher risk score suggested a poor prognosis (BLCA, n = 408, p = 0.008; BRCA, n = 1093, p = 0.052;

TABLE 3 | High and low expression of off-track genes in each tumor.

| Tumor | High-expression genes | Low-expression genes |
|--------|--|--|
| BLCA | MMP14, COL5A2, CTHRC1, VCAN | ACTA2, DCN, FERMT2, FMOD, ITGA5, LUM, MMP2, MYL9, TAGLN, TPM1 |
| BRCA | CDH11, COL12A1, COL1A1, COL1A2, COL3A1, COL5A2, COL8A2, CTHRC1, LUM, MMP14 | DCN, ECM2, FERMT2 |
| COAD | CALU, COL1A1, COL1A2, COL3A1, COL4A1, COL4A2, COL5A2, CTHRC1, LUM, MMP14, MMP2, PLOD1, VCAN | FERMT2, TPM1, EMP3, VIM, ACTA2, DCN, IGFBP4, MGP, MYL9, TAGLN, SNAI2, PMP22, FBLN2 |
| READ | CALU, COL1A1, COL1A2, COL3A1, COL4A1, COL5A2, CTHRC1, MMP14, MMP2, VCAN | ACTA2, CALD1, DCN, FBLN2, FERMT2, MGP, MYL9, PMP22, TAGLN, TPM1, VIM |
| EAC | COL1A1, COL3A1, COL4A1, COL5A2, CTHRC1, MMP14 | NA |
| ESCC | COL1A1, COL1A2, COL3A1, COL5A2, CTHRC1, LUM, MMP14, VCAN | NA |
| GBMLGG | ACTA2, COL1A1, COL1A2, COL3A1, COL5A2, COL6A2, ITGB1, LUM, MMP14, PLOD1, SNAI2, TAGLN, VIM | NA |
| HNSC | MMP2, COL1A1, COL1A2, COL3A1, COL5A2, COL6A2, CTHRC1, LUM, MMP14, PCOLCE, VCAN | TPM1 |
| LUAD | COL1A1, COL1A2, COL3A1, COL5A2, LUM, MMP2 | ACTA2, DCN |
| PRAD | CTHRC1 | ACTA2, COL4A1, COL4A2, COL6A2, DCN, EMP3, FERMT2, IGFBP4, ITGA5, LUM, MATN2, MMP14, MMP2, MYL9, PMP22, TAGLN, TPM1, VIM |
| STAD | COL1A2, COL3A1, COL5A2, CTHRC1, LUM, MMP14, MMP2 | CALD1, FERMT2, TPM1, ACTA2, TAGLN, SFRP1, MYL9, MYLK, DCN, FLNA |

FIGURE 4 The representative survival curve and forest plot of "off-track gene" risk scores for multiple cancers. (**A–B**) The survival curves and the gene expression intensity in the low risk group as well as in the high risk group of LUAD patients in TCGA datasets. *P < 0.05, **P < 0.01, ***P < 0.01, and no stars for genes whose *p*-value is > 0.05. Box plots compare the difference of gene expression between risk groups using a *t*-test. (**C–D**) The survival curves and the gene expression intensity in the low risk group as well as in the high risk group of STAD patients in TCGA datasets. *P < 0.05, **P < 0.01, ***P < 0.01, ***P < 0.001, and no stars for genes whose *p*-value is > 0.05. Box plots compare the difference of gene expression between risk groups using a *t*-test. (**C–D**) The survival curves and the gene expression intensity in the low risk group as well as in the high risk group of STAD patients in TCGA datasets. *P < 0.05, **P < 0.01, ***P < 0.001, and no stars for genes whose *p*-value is > 0.05. Box plots compare the difference of gene expression between risk groups using a *t*-test. (**E**) Forest plot of "off-track genes" using data from multiple cancer cohorts in TCGA using a univariate cox repression model. The forest plot shows the HR, 95% confidence interval, and *p*-value for each type of cancer.

COAD, n = 279, p = 0.012; EAC, n = 89, p = 0.016; ESCC, n = 95, p = 0.184; GBMLGG, n = 421, p = 1.70e-29; HNSC, n = 520, p = 0.004; LUAD, n = 502, p = 4.28e-04; READ, n = 93, p = 0.039; and STAD, n = 388, p = 0.026). These survival curves and differences in their gene expression of representative respiratory and digestive tract tumors are shown in Figures 4A-D. To further assess the impact of "off-track genes" on cancer patients' survival, we performed univariate and multivariate Cox regressions; the results demonstrated that high risk scores for these genes increased the risk of death in cancer patients, and these off-track genes were independent prognostic factors in cancers except BLCA. In addition, high risk scores for these genes in ESCC were not related to patient survival. The results of the Cox proportional hazards regression analysis of clinical characteristics (including age, gender, and tumor stages) are presented in Supplementary Tables S2-S11, and the results of univariate Cox regression of the "off-track gene" risk scores are shown in Figure 4E.

We also used certain independent cancer datasets to validate the prognostic value of these "off-track genes." The survival analysis results revealed that the risk scores of "off-track genes" were significantly correlated with cancer patient survival, such as in the COAD datasets (GSE14333, n = 196, p = 0.017; GSE12945, n = 29, p = 0.012; and GSE17536, n = 177, p = 0.006), LUAD datasets (GSE30219, n = 85, p = 0.020; GSE13213, n = 117, p = 0.002; and GSE68465, n = 443, p = 0.012), and GBMLGG datasets (GSE109857, n = 225, p = 1.167e-15; GSE16011, n = 284, p = 4.320e-13; and CGGA, n = 325, p = 6.430e-28). These survival curves are shown in **Figure 5**.

DISCUSSION

Increasing evidences support the atavistic theory of cancer, that is, the biological origin of cancer can be found in the early transitional phase from unicellularity to multicellularity (Bussey et al., 2017; Trigos et al., 2017). Tumorigenesis and placental villi development exhibit many similar biological behaviors, but their outcomes are different. Gene co-expression networks are the basis of embryonic development and tumorigenesis (Singh et al., 2017), and the gene regulatory network in tumors becomes disrupted (Bussey et al., 2017).

The EMT during placental development is a normal physiological process that involves a strong correlation between

FIGURE 5 | Survival curves for cancers. (A) The survival curve of COAD patients in GSE14333, GSE12945, and GSE17536. (B) The survival curve of LUAD patients in GSE30219, GSE13213, and GSE68465. (C) The survival curve of GBMLGG patients in GSE109857, GSE16011, and CGGA.

genes, suggesting that all biological functions are under the tightly precise regulation of gene networks. However, during the process of tumor progression, the close relationship of genes is disrupted and becomes "off-track" from that in normal placental development. Therefore, we proposed the "off-track theory" for the first time. Such "off-track genes" are likely to promote the progression of cancer and may also predict the prognosis of cancer patients.

Different gene co-expression networks lead to different biological outcomes, and this finding has received increased

attention from researchers. However, methods to measure changes in the network have not been well explored. In this study, we focused on biological processes involved in the EMT, considering a recent paper "Guidelines and definitions for research on epithelial-mesenchymal transition" (Yang et al., 2020), which authors put forward Consensus Statement about EMT and suggest use cellular properties together with a set of molecular markers to define the EMT status. We used 200 EMT related markers from the MSigDB and identified "offtrack genes" in cancer networks using the "off-track theory." First, we used Pearson's correlation analysis to calculate the correlation coefficient of pairs of genes that are involved in the EMT in villi and cancer development. The correlation coefficient of genes in biological networks can reflect the degree of biological connection (Teschendorff et al., 2010; Bockmayr et al., 2013). Then, Pearson's correlation coefficients of the EMT-related gene pairs in villi and cancer were projected on a coordinate axis to identify the gene pairs with a distance from the line y = x greater than 1. Based on the results, we identified "off-track genes" in multiple cancers and deemed that these genes were associated with tumor progression or the prognosis.

To further evaluate the relationship between "off-track genes" and the prognosis of patients, we performed a survival analysis. A high risk score of these genes was significantly associated with poor prognosis in all forms of adenocarcinoma that we analyzed as well as GBMLGG and HNSC, but it was not associated with the prognosis for LUSC and ESCC. There are several reasons that might explain the different prognostic values. First, endodermderived organ tumors are more closely related to embryonic development. The endoderm generates the digestive tube and its accessory organs, the respiratory and pulmonary epithelium, the thymus and the bladder; in addition, nerve cells also originate from the endoderm (Solnica-Krezel, 2005). Second, LUSC has been reported to possibly be derived from basal cells (Sutherland and Berns, 2010; Terry et al., 2010); it exhibits a stepwise progression from normal bronchial epithelium to squamous metaplasia to LUSC, and it has a different development process than LUAD (Nishisaka et al., 2000). Third, ESCC is caused by abnormal hyperplasia of esophageal squamous epithelium, which does not appear during embryonic development (di Pietro et al., 2018). Fourth, HNSC was the only type of squamous cell carcinoma associated with a significantly poor prognosis in this study; however, stage IV patients accounted for 59% of the total HNSC patients, which may influence the survival results.

The results of this study requires further verification. First, although we confirmed that the co-expression network is altered in the cancerous process compared with normal physiological processes and identified the "off-track genes," the network relationships are very complex, and it is difficult to determine a standard for measuring the changes in network relationships. Further exploration is urgently needed. Second, these identified off-track EMT related genes not validated in tumors in vitro and in vivo. This requires a lot of work, and it is hard to verify every gene in every cancer, further studies are planned. Third, the transcriptome represents only one level of the biological process and cannot fully reflect the function of the entire biological system. These results should be combined with other "omics" data, such as proteomics and methylation data, to conduct an integrated analysis of multiomics data to provide more comprehensive information on the biological system.

The placental villi development model may be a useful tool for exploring tumorigenesis. Development models are widely used to study the complex mechanisms of tumorigenesis, and many important signaling pathways and molecular markers have been found (Kho et al., 2004; Liu et al., 2006; Kaiser et al., 2007). Our study indicate that villus development is a reliable and strictly regulated model that can illuminate the trajectory of human cancer development, and gene co-expression in disease states is "off-track" from that in the normal state. The "off-track genes" may have a substantial impact on tumor development and reveal novel prognostic biomarkers to assist in cancer treatment and evaluate the therapeutic effect.

DATA AVAILABILITY STATEMENT

The datasets generated for this study can be found in the Genome Sequence Archive in the Beijing Institute of Genomics (BIG) Data Center, BIG, Chinese Academy of Sciences, under the accession number HRA000050.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Cancer Institute and Hospital, Chinese Academy of Medical Sciences. The patients/participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

AUTHOR CONTRIBUTIONS

SC and WL conceived and designed the study. BZ extracted RNA, performed the data analysis, and wrote the manuscript. HL and YW collected villi tissues. LF provided advice on the study. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by the CAMS Innovation Fund for Medical Sciences (CIFMS2016-I2M-3-005).

ACKNOWLEDGMENTS

We are thankful to our colleagues who provided their expertise that greatly assisted this research work.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fcell.2020.00492/ full#supplementary-material

TABLE S1 | The detailed information of villi samples.

TABLE S2 | Cox proportional hazards regression analysis of OS in BLCA.

TABLE S3 | Cox proportional hazards regression analysis of OS in BRCA.

- TABLE S4 | Cox pro portional hazards regression analysis of OS in COAD.
- TABLE S5 | Cox proportional hazards regression analysis of OS in EAC.
- TABLE S6 | Cox proportional hazards regression analysis of OS in ESCC.
- TABLE S7 | Cox proportional hazards regression analysis of OS in GBMLGG.

REFERENCES

- An, N., Shi, X., Zhang, Y., Lv, N., Feng, L., Di, X., et al. (2015). Discovery of a novel immune gene signature with profound prognostic value in colorectal cancer: a model of cooperativity disorientation created in the process from development to cancer. *PLos One* 10:e0137171. doi: 10.1371/journal.pone.0137171
- Bockmayr, M., Klauschen, F., Györffy, B., Denkert, C., and Budczies, J. (2013). New network topology approaches reveal differential correlation patterns; in breast cancer. *BMC Syst. Biol.* 7:78. doi: 10.1186/1752-0509-7-78
- Burrell, R. A., Nicholas, M. G., Jiri, B., and Charles, S. (2013). The causes and consequences of genetic heterogeneity in cancer evolution. *Nature* 501, 338– 345. doi: 10.1038/nature12625
- Bussey, K. J., Cisneros, L. H., Lineweaver, C. H., and Davies, P. C. W. (2017). Ancestral gene regulatory networks drive cancer. *Proc. Natl. Acad. Sci. U.S.A.* 114:201706990.
- Castro, M. A. (2012). RedeR: R/Bioconductor package for representing modular structures, nested networks and multiple levels of hierarchical associations. *Genome Biol.* 13:R29.
- Chaffer, C. L., San Juan, B. P., Lim, E., and Weinberg, R. A. (2016). EMT, cell plasticity and metastasis. *Cancer Metastasis Rev.* 35, 645–654. doi: 10.1007/ s10555-016-9648-7
- Chen, W., Zheng, R., Baade, P. D., Zhang, S., Zeng, H., Bray, F., et al. (2016). Cancer statistics in China, 2015. *CA Cancer J. Clin.* 66, 115–132. doi: 10.3322/caac. 21338
- Cornish, A. J., and Markowetz, F. (2014). SANTA: quantifying the functional content of molecular networks. *PLoS Comput. Biol.* 10:e1003808. doi: 10.1371/ journal.pcbi.1003808
- di Pietro, M., Canto, M. I., and Fitzgerald, R. C. (2018). Endoscopic management of early adenocarcinoma and squamous cell carcinoma of the esophagus: screening. *Diagnosis Ther. Gastroenterol.* 154, 421–436. doi: 10.1053/j.gastro. 2017.07.041
- Gigic, B., Boeing, H., Toth, R., Bohm, J., Habermann, N., Scherer, D., et al. (2018). Associations between dietary patterns and longitudinal quality of life changes in colorectal cancer patients: the colocare study. *Nutr. Cancer* 70, 51–60. doi: 10.1080/01635581.2018.1397707
- Hanahan, D., and Weinberg, R. A. (2011). Hallmarks of cancer: the next generation. *Cell* 144, 646–674. doi: 10.1016/j.cell.2011.02.013
- Hay, E. D. (2005). The mesenchymal cell, its role in the embryo, and the remarkable signaling mechanisms that create it. *Dev. Dyn.* 233, 706–720. doi: 10.1002/dvdy. 20345
- Holtan, S. G., Creedon, D. J., Paul, H., and Markovic, S. N. (2009). Cancer and pregnancy: parallels in growth, invasion, and immune modulation and implications for cancer therapeutic agents. *Mayo Clin. Proc.* 84, 985–1000. doi: 10.4065/84.11.985
- Huber, M. A., Kraut, N., and Beug, H. (2005). Molecular requirements for epithelial–mesenchymal transition during tumor progression. *Curr. Opin. Cell Biol.* 17, 548–558. doi: 10.1016/j.ceb.2005.08.001
- Jean Paul, T., and Sleeman, J. P. (2006). Complex networks orchestrate epithelialmesenchymal transitions. *Nat. Rev. Mol. Cell Biol.* 7, 131–142. doi: 10.1038/ nrm1835
- Kaiser, S., Park, Y. K., Franklin, J. L., Halberg, R. B., Yu, M., Jessen, W. J., et al. (2007). Transcriptional recapitulation and subversion of embryonic colon development by mouse colon tumor models and human colon cancer. *Genome Biol.* 8:R131.
- Kho, A. T., Qing, Z., Zhaohui, C., Butte, A. J., Kim, J. Y. H., Pomeroy, S. L., et al. (2004). Conserved mechanisms across development and tumorigenesis revealed by a mouse development perspective of human cancers. *Genes Dev.* 18, 629–640. doi: 10.1101/gad.1182504
- Kovar, H., Amatruda, J., Brunet, E., Burdach, S., Cidre-Aranaz, F., Alava, E. D., et al. (2016). The second European interdisciplinary Ewing sarcoma research

- TABLE S8 | Cox proportional hazards regression analysis of OS in HNSC.
- **TABLE S9** | Cox proportional hazards regression analysis of OS in LUAD.
- TABLE S10 | Cox proportional hazards regression analysis of OS in READ.
- TABLE S11 | Cox proportional hazards regression analysis of OS in STAD.

summit – A joint effort to deconstructing the multiple layers of a complex disease. *Oncotarget* 7, 8613–8624. doi: 10.18632/oncotarget.6937

- Li, Y., Wang, Z., Wang, Y., Zhao, Z., Zhang, J., Lu, J., et al. (2016). Identification and characterization of lncRNA mediated transcriptional dysregulation dictates lncRNA roles in glioblastoma. *Oncotarget* 7, 45027–45041. doi: 10.18632/ oncotarget.7801
- Liu, H., Kho, A. T., Kohane, I. S., and Sun, Y. (2006). Predicting survival within the lung cancer histopathological hierarchy using a multi-scale genomic model of development. *PLoS Med.* 3:e232. doi: 10.1371/journal.pmed.0030232
- Machnik, M., Cylwa, R., Kielczewski, K., Biecek, P., Liloglou, T., Mackiewicz, A., et al. (2019). The expression signature of cancer-associated KRAB-ZNF factors identified in TCGA pan-cancer transcriptomic data. *Mol. Oncol.* 13, 701–724. doi: 10.1002/1878-0261.12407
- Macklin, P. S., Mcauliffe, J., Pugh, C. W., and Yamamoto, A. (2017). Hypoxia and HIF pathway in cancer and the placenta. *Placenta* 56:S0143400417301959.
- Moustakas, A., and Heldin, C. (2010). Signaling networks guiding epithelialmesenchymal transitions during embryogenesis and cancer progression. *Cancer Sci.* 98, 1512–1520. doi: 10.1111/j.1349-7006.2007.00550.x
- Netzer, M., Weinberger, K. M., Handler, M., Seger, M., Fang, X., Kugler, K. G., et al. (2011). Profiling the human response to physical exercise: a computational strategy for the identification and kinetic analysis of metabolic biomarkers. *J. Clin. Bioinformat.* 1:34. doi: 10.1186/2043-9113-1-34
- Nishisaka, T., Takeshima, Y., and Inai, K. (2000). Evaluation of p53 gene mutation and loss of heterozygosity of 3p, 9p and 17p in precancerous lesions of 29 lung cancer patients. *Hiroshima J. Med. Sci.* 49, 109–116.
- Perry, J. K., Lins, R. J., Lobie, P. E., and Mitchell, M. D. (2009). Regulation of invasive growth: similar epigenetic mechanisms underpin tumour progression and implantation in human pregnancy. *Clin. Sci.* 118, 451–457. doi: 10.1042/ cs20090503
- Shabalin, A. A. (2012). Matrix eQTL: ultra fast eQTL analysis via large matrix operations. *Bioinformatics* 28, 1353–1358. doi: 10.1093/bioinformatics/bts163
- Siegel, R. L., Miller, K. D., and Jemal, A. (2018). Cancer statistics, 2018. CA: Cancer J. Clin. 68, 7–30. doi: 10.3322/caac.21442
- Simeone, P., Trerotola, M., Franck, J., Cardon, T., Marchisio, M., Fournier, I., et al. (2019). The multiverse nature of epithelial to mesenchymal transition. *Semin. Cancer Biol.* 58, 1–10.
- Singh, A. J., Ramsey, S. A., Filtz, T. M., and Kioussi, C. (2017). Differential gene regulatory networks in development and disease. *Cell. Mol. Life Sci.* 75, 1–13.
- Singh, A. J., Ramsey, S. A., Filtz, T. M., and Kioussi, C. (2018). Differential gene regulatory networks in development and disease. *Cell Mol Life Sci.* 75, 1013–1025. doi: 10.1007/s00018-017-2679-6
- Solnica-Krezel, L. (2005). Conserved patterns of cell movements during vertebrate gastrulation. Curr. Biol. 15, R213–R228.
- Strickland, S., and Richards, W. G. (1992). Invasion of the trophoblasts. Cell 71, 355–357. doi: 10.1016/0092-8674(92)90503-5
- Sutherland, K. D., and Berns, A. (2010). Cell of origin of lung cancer. *Mol. Oncol.* 4, 397–403.
- Terry, J., Leung, S., Laskin, J., Leslie, K. O., Gown, A. M., and Ionescu, D. N. (2010). Optimal immunohistochemical markers for distinguishing lung adenocarcinomas from squamous cell carcinomas in small tumor samples. *Am. J. Surgical Pathol.* 34, 1805–1811.
- Teschendorff, A. E., Gomez, S., Arenas, A., El-Ashry, D., Schmidt, M., Gehrmann, M., et al. (2010). Improved prognostic classification of breast cancer defined by antagonistic activation patterns of immune response pathway modules. *BMC Cancer* 10:604. doi: 10.1186/1471-2407-10-604
- Trigos, A. S., Pearson, R. B., Papenfuss, A. T., and Goode, D. L. (2017). Altered interactions between unicellular and multicellular genes drive hallmarks of transformation in a diverse range of solid tumors. *Proc. Natl. Acad. Sci. U.S.A.* 114, 6406–6411. doi: 10.1073/pnas.1617743114

- Troiani, T., Martinelli, E., Napolitano, S., Morgillo, F., Belli, G., Cioffi, L., et al. (2014). Molecular aspects of resistance to biological and non-biological drugs and strategies to overcome resistance in colorectal cancer. *Curr. Med. Chem.* 21, 1639–1653. doi: 10.2174/09298673113209990224
- Yang, J., Antin, P., Berx, G., Blanpain, C., Brabletz, T., Bronner, M., et al. (2020). Guidelines and definitions for research on epithelial-mesenchymal transition. *Nat. Rev. Mol. Cell Biol.* 21, 341–352.
- Zhang, B., Lin, F., Honglin, G., Hongxia, L., Yuanjing, W., Kaitai, Z., et al. (2017). Villi-specific gene expression reveals novel prognostic biomarkers in multiple human cancers. J. Cancer 8, 2793–2801. doi: 10.7150/jca.19787

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Zhang, Wang, Li, Feng, Li and Cheng, This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.