i:S



https:/doi.org/10.1093/ckj/sfac152 Advance Access Publication Date: 9 June 2022 Editorial Comment

EDITORIAL COMMENT

Gain-of-function TLR7 and loss-of-function A20 gene variants identify a novel pathway for Mendelian lupus and lupus nephritis

Priscila Villalvazo ^{1,2,3}, Sol Carriazo^{1,2,3}, Jorge Rojas-Rivera^{1,2}, Adrián M. Ramos^{1,2}, Alberto Ortiz ^{1,2,3,*} and Maria Vanessa Perez-Gomez ^{1,2,3,*}

¹Department of Nephrology and Hypertension, Instituto de Investigación Sanitaria, Fundacion Jimenez Diaz, Universidad Autónoma de Madrid, Madrid, Spain, ²RICORS2040, Madrid, Spain and ³Departamento de Medicina, Facultad de Medicina, Universidad Autónoma de Madrid, Madrid, Spain

*Share senior authorship Correspondence to: Maria Vanessa Perez-Gomez; E-mail: mvanessa@fjd.es

ABSTRACT

Systemic lupus erythematosus (SLE) is a chronic and inflammatory autoimmune disease of unknown origin that may cause kidney disease, i.e. lupus nephritis (LN). Within a wider trend towards an expanding field of genetic causes of kidney disease, two recent reports have emphasized the role of Mendelian autoimmune disorders in causing LN both in children and in young adults. Loss-of-function (LOF) variants of tumor necrosis factor alpha–induced protein 3 (TNFAIP3) and gain of function (GOF) variants of Toll-like receptor 7 (TLR7) cause SLE and LN, respectively. Interestingly, both genes regulate the same signaling route, as A20, the protein encoded by TNFAIP3, inhibits nuclear factor κ B (NF- κ B) activation while TLR7 promoted NF- κ B activation. Moreover, TNFAIP3 and TLR7 variants are relatively frequent, potentially contributing to polygenic risk for LN. Finally, they both may be expressed by kidney cells, potentially contributing to the severity of kidney injury in persons who have already developed autoimmunity. The fact that both genes regulate the same pathway may lead to novel therapeutic approaches targeting the shared molecular pathway.

Keywords: A20, HA20, inherited kidney disease, lupus nephritis, systemic lupus erythematosus, TLR7

INTRODUCTION

Systemic lupus erythematosus (SLE) is a chronic and inflammatory autoimmune disease of unknown cause, characterized by the loss of immune tolerance to nuclear self-antigens, B-cell hyperreactivity and the production of autoantibodies and inflammatory cytokines, resulting in damage to several tissues and organs and in increased morbidity and mortality [1, 2]. Lupus nephritis (LN) is one of the most common severe manifestations of SLE, as up to 60% of SLE patients develop LN, mainly people with juvenile-onset SLE. The incidence and severity of LN vary according to the geographical area, race/ethnicity, sex/age and applied diagnostic criteria [3, 4]. LN is an immune complex glomerulonephritis characterized by the development of proteinuria, hematuria, leukocyturia and/or reduced kidney function. The course is characterized by relapses and remissions. LN may be the only initial manifestation of SLE [5]. Classically, evidence of LN was one of the potential

Received: 20.5.2022; Editorial decision: 21.5.2022

[©] The Author(s) 2022. Published by Oxford University Press on behalf of the ERA. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (https://creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

diagnostic criteria for SLE, including persistent proteinuria (>0.5 g/24 hours) or the presence of urinary casts in the 1982 American College of Rheumatology SLE classification criteria [6] or proteinuria >0.5 g/24 hours or per gram of urinary creatinine or red blood cell casts in urinary sediment in the 2012 systemic lupus erythematosus international collaborating clinics criteria [7]. Kidney biopsy confirms the diagnosis of LN, assesses severity and helps to predict outcomes and determine treatment. The 2003 International Society of Nephrology/Renal Pathology Society (ISN/RPS) LN classification and its 2018 update [8, 9] establish six histologic classes and activity and chronicity parameters of severity [3, 5, 10, 11]. The role of the kidney biopsy was recently highlighted in the 2019 European League Against RheumatismAmerican College of Rheumatology classification criteria: a positive antinuclear antibody with proteinuria >0.5 g/24 hours (or equivalent) and the presence of LN on kidney biopsy according to the 2003 ISN/RPS classification are sufficient to diagnose SLE [12].

THE EXPANDING FIELD OF GENETIC CAUSES OF KIDNEY DISEASE

Inherited kidney diseases (IKDs) are more common than previously thought. They account for at least 10% of adult CKD cases [13, 14]. Prior to the increasing availability of genetic testing, many of these patients were incorrectly classified as having hypertensive nephropathy, CKD of unknown cause or assigned a different cause of CKD. Contributing to the invisibility of IKDs, major registries only report autosomal dominant polycystic kidney disease separately as a cause of CKD, while all other causes of IKD are grouped under 'other' or, if not diagnosed, in any other category. However, when all IKDs are grouped and reported together, we get a different perspective: IKDs are the third leading cause of kidney failure in Catalonia and the fourth in Madrid, Spain [15]. This may still be an underestimation, given the low uptake of genetic diagnostic tests even when they are freely available [16]. A genetic basis for CKD was also identified for one of the most common causes of CKD, so-called hypertensive nephropathy, which in African Americans is usually an IKD APOL1 variant nephropathy [17]. Although there is no specific treatment available for most IKDs, a correct diagnosis may prevent unnecessary invasive procedures and treatments and, as IKD can be directly attributed to the dysfunction of the responsible gene, this may lead to the design and development of specific therapies [15]. Recent reports have also provided a genetic basis for some forms of LN [18].

GENETIC CAUSES OF LUPUS

SLE is recognized as a polygenic autoimmune disease. The strong genetic component in SLE is estimated to be 66% of heritability in twin studies [19]. In recent decades, genome-wide association studies (GWASs) have identified >100 SLE susceptibility loci [20]. The proportion of phenotypic variances explained by variants in human leukocyte antigen (HLA) is 2.6% [21] and non-HLA 38% [22]. Additionally, >30 genes causing monogenic forms of SLE or SLE-like syndromes have been reported [23– 25] (Table 1). Among them, deficiencies of complement factors such as C1q, C4A, B and C2 confer a high disease susceptibility [26]. Approximately 90% of people with C1q deficiency develop a lupus-like phenotype [27]. TREX1 variants are also associated to monogenic diseases, such as familial chilblain lupus 1, a cuta-

| Table 1. Examples of genes whose variants are associated with hu- |
|---|
| man SLE and impact of the gene variants on the activity of the pro- |
| tein product (modified from Brown et al. [25]) |

| SLE predisposition resulting from functional deficiency in protein product | SLE predisposition resulting from excess activity of protein product |
|--|--|
| TNFAIP3 [18] C1QA C1QB C1QC C1R C2 CFB C4A C4B DNASE1 TREX1 PRKCD DNASE1L3 ACP5 SOCS1 NCKAP1L C1S C3 SAMHD1 ADAR1 RNASEH2B | TLR7 [25] TMEM173 TNFSF6 IFIH1 STAT4 |

neous form of SLE, and Aicardi–Goutieres syndrome, an inflammatory encephalopathy that shares features with SLE [26].

The type I interferon (IFN) system also plays a major role in SLE pathogenesis [28]. GWASs have reported associations with type I IFN–induced genes, and several monogenic lupus or lupus-like diseases are associated with interferonopathy (e.g. IFIH1, TNFAIP3, RNASEH2A, RNASEH2B, IRF7). IFN- α -induced genes are overexpressed in the peripheral blood of 60–80% of patients with SLE [29, 30].

The HLA region is a strong predictor of genetic risk, predominately HLA class II {e.g. HLA-DR2 [hazard ratio (HR) 1.2] and HLA-DR3 [HR 2.4]} loci related to T-cell-dependent antibody responses [31–33]. Other predisposing genes involve those encoding lymphocyte signaling molecules that regulate the activation or suppression of T- or B-cell activity or survival, such as PTPN22, OX40L and PD1 [34–37].

Lastly, in recent years, evidence suggests the role of genetic factors in both disease susceptibility and on different disease phenotypes [38]. In this regard, *ITGAM* and *FCGR2A* variants have been associated with susceptibility to skin involvement, while *ITGAM*, *HLDR2* and *STAT4* are associated with kidney disease [39].

Loss-of-function (LOF) variants of tumor necrosis factor alpha-induced protein 3 (TNFAIP3) and gain of function (GOF) variants of Toll-like receptor 7 (TLR7) were recently reported to underlie LN in children and adults in the Clinical Kidney Journal (CKJ) and Nature, respectively [18, 25].

A20, HA20 AND LN

A20 is a negative regulator of inflammation encoded by the TNFAIP3 gene [40]. A20 inhibits nuclear factor (NF)- κ B signaling and restricts the interferon regulatory factor (IRF) pathway and autophagy [41] (Figure 1). Haploinsufficiency of

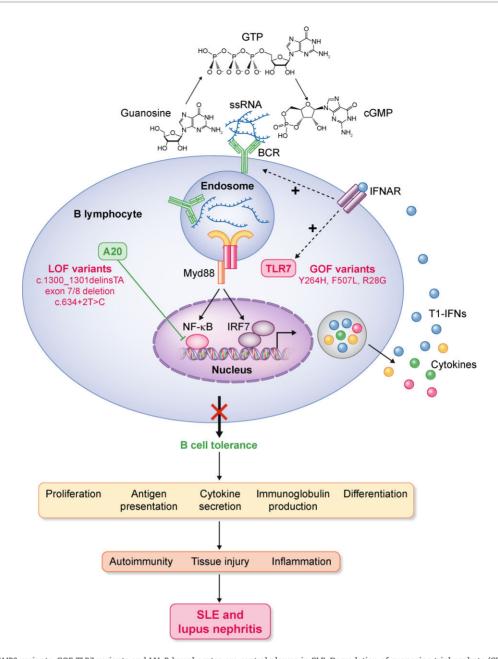


FIGURE 1: LOF TNFAIP3 variants, GOF TLR7 variants and LN. B lymphocytes are central players in SLE. Degradation of guanosine triphosphate (GTP) from processed ssRNA yields guanosine and 2¢,3¢-cyclophosphate guanosine monophosphate (cGMP) that are recognized by the BCR and ssRNA is endocytosed complexed with BCR and delivered to endosomes, thus allowing ssRNA interaction with TLR7. TLR7 then recruits the adaptor Myd88 and signals to activate NF-κB and IRF7, leading to synthesis and secretion of cytokines (yellow, green, red dots) and type I IFN (T1-INF, blue dots). The TNFAIP3 gene encodes the A20 protein, a negative regulator of NF-κB (and of NF-κB association with IRF7), contributing to B-cell tolerance [58]. GOF variants of TLR7 break B-cell tolerance, leading to SLE, as exemplified by Y264H, F507L and R28G [25]. TLR7 GOF variants may disrupt B-cell tolerance resulting in increased proliferation of autoimmune B cells, antigen presentation to autoreactive T cells, differentiation to plasma cells and cytokine production [59, 60]. Cytokines contribute to inflammation, while T1-INF endocrine or paracrine (from activated DCs [61]) loops decisively contribute to loss of tolerance. The introduction of the TLR7-Y264H variant in mice resulted in SLE characterized by autoimmunity, tissue injury and inflammation, including LN, and this was prevented by Myd88 deficiency [25]. Also recently, LOF mutations in TNFAIP3 decisively contribute to loss of tolerance and LN [18]. BCR: B-cell receptor; IFNAR: interferon-α/β receptor; ssRNA: single-stranded RNA.

A20 (HA20; Online Mendelian Inheritance in Man 616744) is an autosomal dominant monogenic disease caused by heterozygous LOF TNFAIP3 variants and is characterized by early onset systemic inflammation in multiple organs [40]. Although phenotypes may vary according to specific TNFAIP3 variants [42], the major phenotype is Behçet-like symptoms. In a literature review of clinical manifestations, the most common symptoms were oral ulcers (70%), recurrent fever (42%), gastrointestinal ulcers (40%), skin lesions (38%), genital ulcers (36%), musculoskeletal disorders (34%) and autoimmune thyroid disorder (19%). Ocular involvement, vasculitis, atrophic gastritis, kidney or liver injury, recurrent respiratory tract infection, interstitial lung disease or dental anomaly were found in <10% of patients [42]. The diverse clinical manifestations may result from variable penetrance as well as from the interaction with other genes and the environment.

| Gene | HGVS consequence | VEP annotation | Clinical significance | Allele frequency | Range of allele frequency |
|---------|-----------------------|----------------|---|------------------|---------------------------|
| TNFAIP3 | c.805+28A>C | Intron | Benign | 6.09E-01 | 0.17–0.85 |
| | c.296–15_296–13delCCT | Intron | Benign | 6.05E-01 | 0.17-0.86 |
| | p.Phe127Cys | Missense | Benign/Likely benign | 6.17E-02 | 0.015-0.36 |
| | c.805+26C>T | Intron | Benign | 4.41E-02 | 0.001-0.10 |
| | p.Asn102Ser | Missense | Benign | 1.24E-02 | 0.000-0.06 |
| | c.2089–42G>A | Intron | C C | 6.53E-03 | 0.000-0.01 |
| | c.487–8C>G | Splice region | Benign | 6.11E-03 | 0.000-0.05 |
| | p.Thr647Pro | Missense | Conflicting interpretations of pathogenicity | 1.81E-03 | 0.000-0.006 |
| | p.Ala125Val | Missense | Conflicting interpretations of pathogenicity | 1.65E-03 | 0.000-0.02 |
| TLR7EG | p.Gln11Leu | Missense | | 1.79E-01 | 0.000-0.27 |
| | p.Val219Ile | Missense | Benign | 4.59E-03 | 0.000-0.03 |
| | p.Ala448Val | Missense | Benign | 3.01E-03 | 0.000-0.005 |
| | p.Val222Asp | Missense | Benign | 2.45E-03 | 0.000-0.005 |

| Tabl | e 2. Alle | le distri | bution fo | r gnomAD | variants f | for TNFAIP3 | and TLR7 |
|------|-----------|-----------|-----------|----------|------------|-------------|----------|
|------|-----------|-----------|-----------|----------|------------|-------------|----------|

HGVS: Human Genome Variation Society; VEP: variant effect predictor.

Gene variants with an allele frequency >1 in 1000 are shown. Gene changes are shown in or within 75 base pairs of a coding exon [56, 57]. The range of allele frequency refers to allele frequencies for different ethnicities.

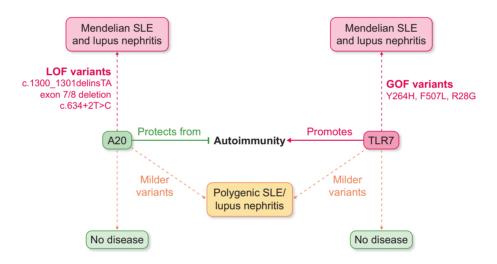


FIGURE 2: TNFAIP3 and TLR7 gene variants and SLE/LN. TNFAIP3 encodes A20, an inhibitor of the pro-inflammatory transcription factor NF-kB that is activated by TLR7. This means that A20 and TLR7 can be traced to the same intracellular signaling pathway and have the potential for clinically relevant interactions. Several severe LOF TNFAIP3 variants cause Mendelian SLE/LN, as do several GOF TLR7 gene variants. These variants with a severe impact on function are generally associated with severe, early onset disease, but this may represent the tip of the iceberg, as milder variants may exist that cause late-onset disease that has not yet being characterized, just as familial hypocholesteremia was initially identified in patients having coronary artery disease in childhood. Additionally, even milder gene variants, with the potential to be present in genetic databases from the general population (Table 1) and that are considered benign when isolated, may contribute to polygenic risk for more classical forms of SLE/LN, especially when associated with risk variants of the other gene.

In this issue of CKJ, Zhang et al. [18]. report that HA20 is a cause of biopsy-proven LN with both early and late onset in males and females in three families with different TNFAIP3 variants.

A male patient had late-onset (age 29 years) SLE with multiorgan involvement: alopecia, arthralgia, nephrotic proteinuria, thrombocytopenia, hypocomplementemia and positive autoantibodies. Additionally, atopy-like clinical manifestations and high immunoglobulin E levels were present. A novel heterozygous variant c.634+2T>C in the TNFAIP3 gene affected messenger RNA (mRNA) splicing and created a frameshift mutation that removed both the Ovarian TUmor (OTU) domain and all Zinc Finger (ZnF) domains. Family members with the same genetic variant had milder involvement, including oral ulcers with or without duodenal ulcers, skin rashes, anemia and allergic history, illustrating the clinical variability even within the same family [43]. The other two patients were girls with an early onset (3 years old). One had recurrent fever, autoimmune hemolytic anemia, hepatosplenomegaly, lymphadenopathy, acute cutaneous lupus, serositis, cardiovascular compromise, mild growth retardation and kidney injury. A deletion of exons 7 and 8 in the TNFAIP3 gene resulted in loss of both OUT and ZnF domains. The other girl also had recurrent fever, autoimmune hemolytic anemia, hepatosplenomegaly, lymphadenopathy and kidney

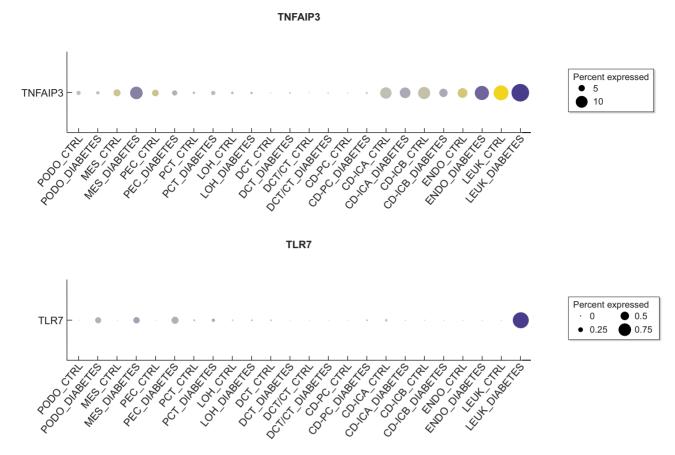


FIGURE 3: TNFAIP3 and TLR7 mRNA expression in human control and diabetic kidney tissue. Images from Humphreys Laboratory [62] with permission (Wilson et al. [63] and Wu et al. [64]).

injury, and additionally had oral ulcers and musculoskeletal involvement. A c.1300_1301delinsTA (p.A434*) TNFAIP3 variant disrupted the ZnF coding region. This report adds TNFAIP3 variants to the list of causes of Mendelian LNs. Although mainly found in children, this report illustrates that monogenic LNs may be found in adults.

TLR7

TLR7 is an intracellular receptor mainly expressed in B cells and plasmacytoid dendritic cells (pDCs) that recognizes pathogenassociated molecular patterns [44]. Overexpression of TLR7 results in more severe autoimmune responses and greater incidence of lupus-like disease [45, 46]. TLR7 is encoded by the X chromosome. Although female cells randomly inactivate one of the two X chromosomes, 15-23% of X-linked human genes escape X chromosome inactivation and both alleles are simultaneously expressed. Female B cells with biallelically expressed TLR7 have an increased susceptibility to TLR7-dependent autoimmune syndromes [47]. In this regard, female pDCs produce more IFN- α than male pDCs upon stimulation with synthetic ligands or single-stranded RNA (ssRNA) that selectively activate TLR7 [48, 49]. Furthermore, increased IFN- α is related to exogenous and endogenous estrogens [47]. Indeed, a TLR7dependent, IFN-independent immune activation has been proposed to be sufficient to accelerate SLE [50]. Additionally, TLR7 stimulation activates the proinflammatory transcription factor NF-KB [51] that links TLR7 and A20 on opposite sides of the same

pathway. TLR7 gene dosage also contributed to accelerating autoimmunity when a cluster of at least 16 X-linked genes is duplicated and translocated to the Y chromosome in mice (Yaachromosome, Y-linked autoimmune accelerator) [45]. Finally, in a Mexican population, an increased copy number of TLR7 was associated with an increased risk of pediatric SLE [52].

Brown et al. [25] have described female patients with SLE and GOF TLR7 variants. A 7-year-old patient had a de novo TLR7 p.Tyr264His (Y264H) missense variant. Whole exome sequencing of additional patients with SLE identified TLR7 p.Arg28Gly (R28G) in a young female with mucosal and hematological involvement and TLR7 p.Phe507Leu (F507L) in a pediatric patient with optic neuritis. To explore the impact of TLR7 gene variants, Brown et al [25]. overexpressed TLR7^{Y264H}, TLR7^{R28G} and TLR7^{F507L} in cultured RAW264.7 macrophages and found that these variants caused NF-KB activation. Moreover, they demonstrated that TLR7^{Y264H} could cause SLE, as CRISPR-Cas9 editing into C57BL/6 mice caused splenomegaly, decreased survival, development of antinuclear antibodies, thrombocytopenia and proliferative glomerulonephritis with mesangial electron-dense deposits and increased mesangial cellularity in male or female mice carrying one or two alleles. Lymphoid cells infiltrated the liver, salivary glands and pancreas and mice displayed increased levels of IFN- γ , interleukin-6 (IL-6), IL-10 and TNF [53]. These findings are consistent with previous studies [50, 54] and confirm the role of excess TLR7 activity in the pathogenesis of SLE, including LN. This has clear therapeutic implications. In this regard, an intravenous Toll-like receptor inhibitory peptide 1 (IP1)

decreased albuminuria, kidney inflammation and mRNA expression downstream of TLR7 or TLR9 in MRL/lpr mice with SLE [54]. However, preclinical results of therapeutic interventions may be more solid if they are confirmed at multiple sites [55].

PATHOPHYSIOLOGICAL AND CLINICAL IMPACT

The unstoppable advance of kidney genetics is now expanding into immune-mediated kidney disease. TNFAIP3 and TLR7 should be added to the list of genes to be assessed in the evaluation of patients with LN. Interestingly, both genes regulate the same signaling route (Figure 1) and gene variants are relatively frequent (Table 2). Although most of the more common gene variants are labeled benign, this means that they are not associated with Mendelian inherited disease, but they might contribute to polygenic risk scores, and this should be explored (Figure 2). Furthermore, the identification of individual contributors to the pathogenesis of SLE and LN will allow the development of new targeted therapies. Moreover, although studies on their role in SLE have focused on the driving events of LN (i.e. autoimmunity), both genes are also expressed by kidney parenchymal cells. Thus a potential role in kidney injury, independent from the presence of autoimmunity, including a specific role in LN once autoimmunity develops, should be explored. In this regard, both TNFAIP3 and TLR7 may be differentially expressed in kidney parenchymal cells in the course of kidney injury (Figure 3).

FUNDING

FIS/Fondos FEDER (PI18/01366, PI18/01133, PI19/00588, PI19/00815, DTS18/00032), ERA-PerMed-JTC2018 (KIDNEY ATTACK AC18/00064 and PERSTIGAN AC18/00071, ISCIII-RETIC REDinREN RD016/0009), Sociedad Española de Nefrología, FRIAT, Comunidad de Madrid en Biomedicina B2017/BMD-3686 CIFRA2-CM, Instituto de Salud Carlos III (ISCIII) RICORS program to RICORS2040 (RD21/0005/0001) and SPACKDc PMP21/00109, FEDER funds.

CONFLICT OF INTEREST STATEMENT

A.O. has received grants from Sanofi and consultancy or speaker fees or travel support from Advicciene, Astellas, AstraZeneca, Amicus, Amgen, Fresenius Medical Care, GlaxoSmithKline, Bayer, Sanofi-Genzyme, Menarini, Mundipharma, Kyowa Kirin, Alexion, Freeline, Idorsia, Chiesi, Otsuka, Novo Nordisk and Vifor Fresenius Medical Care Renal Pharma and is Director of the Catedra Mundipharma-UAM of diabetic kidney disease and the Catedra AstraZeneca-UAM of chronic kidney disease and electrolytes. A.O. is Emeritus Editor-in-Chief of CKJ.

(See related article by Zhang et al. Novel loss-of-function mutations in TNFAIP3 gene in patients with lupus nephritis. Clin *Kidney J* (2022) 15: 2027–2038.)

REFERENCES

- Tsokos GC. Systemic lupus erythematosus. N Engl J Med 2011; 365: 2110–2121
- Cervera R, Khamashta MA, Font J et al. Morbidity and mortality in systemic lupus erythematosus during a 10-year period: a comparison of early and late manifestations in a cohort of 1,000 patients. *Medicine* 2003; 82: 299–308

- 3. Anders HJ, Saxena R, Zhao MH et al. Lupus nephritis. Nat Rev Dis Primers 2020; 6: 7
- Croca SC, Rodrigues T, Isenberg DA. Assessment of a lupus nephritis cohort over a 30-year period. Rheumatology 2011; 50: 1424–1430
- Moroni G, Vercelloni PG, Quaglini S et al. Changing patterns in clinical-histological presentation and renal outcome over the last five decades in a cohort of 499 patients with lupus nephritis. Ann Rheum Dis 2018; 77: 1318–1325
- Tan EM, Cohen AS, Fries JF et al. The 1982 revised criteria for the classification of systemic lupus erythematosus. Arthritis Rheum 1982; 25: 1271–1277
- Petri M, Orbai AM, Alarcón GS et al. Derivation and validation of the systemic lupus international collaborating clinics classification criteria for systemic lupus erythematosus. Arthritis Rheum 2012; 64: 2677–2686
- Weening JJ, D'Agati VD, Schwartz MM et al. The classification of glomerulonephritis in systemic lupus erythematosus revisited. J Am Soc Nephrol 2004; 15: 241–250; erratum: J Am Soc Nephrol 2004; 15: 835–83.
- Bajema IM, Wilhelmus S, Alpers CE et al. Revision of the International Society of Nephrology/Renal Pathology Society classification for lupus nephritis: clarification of definitions, and modified national institutes of health activity and chronicity indices. Kidney Int 2018; 93: 789–796
- Parikh SV, Almaani S, Brodsky S et al. Update on lupus nephritis: core curriculum 2020. Am J Kidney Dis 2020; 76: 265–281
- Kidney Disease: Improving Global Outcomes Glomerular Diseases Work Group. KDIGO 2021 clinical practice guideline for the management of glomerular diseases. *Kidney Int* 2021; 100(4 Suppl): S1–S276
- Aringer M, Costenbader K, Daikh D et al. 2019 European League Against Rheumatism/American College of Rheumatology classification criteria for systemic lupus erythematosus. Arthritis Rheum 2019; 71: 1400–1412
- Schrezenmeier E, Budde K, Bergmann C. Diagnostic utility of exome sequencing for kidney disease. N Engl J Med 2019; 380: 2078
- Carriazo S, Ortiz A, Perez-Gomez MV. Diagnostic utility of exome sequencing for kidney disease. N Engl J Med 2019; 380: 2078
- Torra R, Furlano M, Ortiz A et al. Genetic kidney diseases as an underrecognized cause of chronic kidney disease: the key role of international registry reports. Clin Kidney J 2021; 14: 1879–1885
- Doreille A, Villié P, Mesnard L. National survey on genetic test prescription in French adult nephrologists: a call for simplification and education. Clin Kidney J 2022; 15: 1213– 1215
- Carriazo S, Vanessa Perez-Gomez M, Ortiz A. Hypertensive nephropathy: a major roadblock hindering the advance of precision nephrology. Clin Kidney J 2020; 13: 504–509
- Zhang C, Han X, Sun L et al. Novel loss-of-function mutations in TNFAIP3 gene in patients with lupus nephritis. Clin Kidney J 2022; https://doi.org/10.1093/ckj/sfac130
- Alarcón-Segovia D, Alarcón-Riquelme ME, Cardiel MH et al. Familial aggregation of systemic lupus erythematosus, rheumatoid arthritis, and other autoimmune diseases in 1,177 lupus patients from the GLADEL cohort. Arthritis Rheum 2005; 52: 1138–1147
- Guerra SG, Vyse TJ, Cunninghame Graham DS. The genetics of lupus: a functional perspective. Arthritis Res Ther 2012; 14: 211

- 21. Molineros JE, Looger LL, Kim K et al. Amino acid signatures of HLA class-I and II molecules are strongly associated with SLE susceptibility and autoantibody production in Eastern Asians. PLoS Genet 2019; **15**: e1008092
- 22. Morris DL, Sheng Y, Zhang Y et al. Genome-wide association meta-analysis in Chinese and European individuals identifies ten new loci associated with systemic lupus erythematosus. Nat Genet 2016; **48**: 940–946
- Rullo OJ, Tsao BP. Recent insights into the genetic basis of systemic lupus erythematosus. Ann Rheum Dis 2013; 72 (Suppl 2): ii56-ii61
- Alperin JM, Ortiz-Fernández L, Sawalha AH. Monogenic lupus: a developing paradigm of disease. Front Immunol 2018; 9: 2496
- Brown GJ, Cañete PF, Wang H et al. TLR7 gain-of-function genetic variation causes human lupus. Nature 2022; 605: 349– 356
- 26. Demirkaya E, Sahin S, Romano M et al. New horizons in the genetic etiology of systemic lupus erythematosus and lupus-like disease: monogenic lupus and beyond. *J Clin Med* 2020; **9**: 712
- Lood C, Gullstrand B, Truedsson L et al. C1q inhibits immune complex-induced interferon-alpha production in plasmacytoid dendritic cells: a novel link between C1q deficiency and systemic lupus erythematosus pathogenesis. Arthritis Rheum 2009; 60: 3081–3090
- Hagberg N, Rönnblom L. Systemic lupus erythematosus—a disease with a dysregulated type i interferon system. Scand J Immunol 2015; 82: 199–207
- Kariuki SN, Kirou KA, MacDermott EJ et al. Cutting edge: autoimmune disease risk variant of STAT4 confers increased sensitivity to IFN-α in lupus patients in vivo. J Immunol 2009; 182: 34–38
- Banchereau R, Hong S, Cantarel B et al. Personalized immunomonitoring uncovers molecular networks that stratify lupus patients. Cell 2016; 165: 1548–1550
- Fernando MMA, Stevens CR, Walsh EC et al. Defining the role of the MHC in autoimmunity: a review and pooled analysis. PLos Genet 2008; 4: e1000024
- Barcellos LF, May SL, Ramsay PP et al. High-density SNP screening of the major histocompatibility complex in systemic lupus erythematosus demonstrates strong evidence for independent susceptibility regions. PLoS Genet 2009; 5: e1000696
- Rioux JD, Goyette P, Vyse TJ et al. Mapping of multiple susceptibility variants within the MHC region for 7 immunemediated diseases. Proc Natl Acad Sci USA 2009; 106: 18680– 18685
- Berghöfer B, Frommer T, Haley G et al. TLR7 ligands induce higher IFN-α production in females. J Immunol 2006; 177: 2088–2096
- Abelson AK, Delgado-Vega AM, Kozyrev SV et al. STAT4 associates with systemic lupus erythematosus through two independent effects that correlate with gene expression and act additively with IRF5 to increase risk. Ann Rheum Dis 2009; 68: 1746–1753
- 36. Aitman TJ, Dong R, Vyse TJ et al. Copy number polymorphism in Fcgr3 predisposes to glomerulonephritis in rats and humans. Nature 2006; **439**: 851–855
- 37. Yang Y, Chung EK, Yee LW et al. Gene copy-number variation and associated polymorphisms of complement component C4 in human systemic lupus erythematosus (SLE): low copy number is a risk factor for and high copy number is a protective factor against SLE susceptibility in European Americans. Am J Hum Genet 2007; 80: 1037–1054

- Kariuki SN, Ghodke-Puranik Y, Dorschner JM et al. Genetic analysis of the pathogenic molecular sub-phenotype interferon-alpha identifies multiple novel loci involved in systemic lupus erythematosus. *Genes Immun* 2015; 16: 15–23
- Ceccarelli F, Perricone C, Borgiani P et al. Genetic factors in systemic lupus erythematosus: contribution to disease phenotype. J Immunol Res 2015; 2015: 745647
- Martens A, van Loo G. A20 at the crossroads of cell death, inflammation, and autoimmunity. Cold Spring Harb Perspect Biol 2020; 12: a036418
- 41. Catrysse L, Vereecke L, Beyaert R et al. A20 in inflammation and autoimmunity. *Trends Immunol* 2014; **35**: 22–31
- Chen Y, Ye Z, Chen L et al. Association of clinical phenotypes in haploinsufficiency A20 (HA20) with disrupted domains of A20. Front Immunol 2020; 11: 574992
- Yu MP, Xu XS, Zhou Q et al. Haploinsufficiency of A20 (HA20): updates on the genetics, phenotype, pathogenesis and treatment. World J Pediatr 2020; 16: 575–584
- O'Neill LAJ, Hennessy EJ, Parker AE. Targeting Toll-like receptors: emerging therapeutics? Nat Rev Drug Discov 2010; 9: 293–307
- Deane JA, Pisitkun P, Barrett RS et al. Control of Toll-like receptor 7 expression is essential to restrict autoimmunity and dendritic cell proliferation. *Immunity* 2007; 27: 801–810
- Pisitkun P, Deane JA, Difilippantonio MJ et al. Autoreactive B cell responses to RNA-related antigens due to TLR7 gene duplication. Science 2006; 312: 1669–1672
- Souyris M, Mejía JE, Chaumeil J et al. Female predisposition to TLR7-driven autoimmunity: gene dosage and the escape from X chromosome inactivation. Semin Immunopathol 2019; 41: 153–164
- Meier A, Chang JJ, Chan ES et al. Sex differences in the TLRmediated response of pDCs to HIV-1 are associated with higher immune activation in infected women. Nat Med 2009; 15: 955–959
- Berghöfer B, Frommer T, Haley G et al. TLR7 ligands induce higher IFN-α production in females. J Immunol 2006; 177: 2088–2096
- Wolf SJ, Theros J, Reed TJ et al. TLR7-mediated lupus nephritis is independent of type I IFN signaling. J Immunol 2018; 201: 393–405
- Nanda SK, Lopez-Pelaez M, Arthur JSC et al. Suppression of IRAK1 or IRAK4 catalytic activity, but not type 1 IFN signaling, prevents lupus nephritis in mice expressing a ubiquitin binding–defective mutant of ABIN1. J Immunol 2016; 197: 4266–4273
- 52. García-Ortiz H, Velázquez-Cruz R, Espinosa-Rosales F et al. Association of TLR7 copy number variation with susceptibility to childhood-onset systemic lupus erythematosus in Mexican population. Ann Rheum Dis 2010; 69: 1861–1865
- Brown GJ, Cañete PF, Wang H et al. TLR7 gain-of-function genetic variation causes human lupus. Nature 2022; 605: 349– 356
- 54. Baek WY, Lee SM, Lee SW et al. Intravenous administration of Toll-like receptor inhibitory peptide 1 is effective for the treatment of systemic lupus erythematosus in a Mus musculus model. J Rheum Dis 2021; 28: 133–142
- 55. Lei Y, Sehnert B, Voll RE et al. A multicenter blinded preclinical randomized controlled trial on jak1/2 inhibition in MRL/MpJ-Fas^{lpr} mice with proliferative lupus nephritis predicts low effect size. *Kidney Int* 2021; 99: 1331–1341
- 56. TNFAIP3 | gnomAD v2.1.1 | gnomAD. https://gnomad. broadinstitute.org/gene/ENSG00000118503?dataset= gnomad_r2_1 (13 May 2022, date last accessed)

- TLR7 | unknown | gnomAD. https://gnomad.broadinstitute. org/gene/ENSG00000196664?dataset=gnomad_r2_ (13 May 2022, date last accessed)
- 58. Das T, Chen Z, Hendriks RW, Kool M. A20/tumor necrosis factor α -induced protein 3 in immune cells controls development of autoinflammation and autoimmunity: lessons from mouse models. Front Immunol 2018; **9**: 104
- Rawlings DJ, Schwartz MA, Jackson SW et al. Integration of B cell responses through Toll-like receptors and antigen receptors. Nat Rev Immunol 2012; 12: 282–294
- Meyer-Bahlburg A, Rawlings DJ. B cell autonomous TLR signaling and autoimmunity. Autoimmun Rev 2008; 7: 313–316
- Saitoh SI, Abe F, Kanno A et al. TLR7 mediated viral recognition results in focal type I interferon secretion by dendritic cells. Nat Commun 2017; 8: 1592
- 62. KIT | results page. http://humphreyslab.com/SingleCell/ displaycharts.php (13 May 2022, date last accessed)
- 63. Wilson PC, Wu H, Kirita Y et al. The single-cell transcriptomic landscape of early human diabetic nephropathy. Proc Natl Acad Sci USA 2019; **116**: 19619–19625
- 64. Wu H, Malone AF, Donnelly EL et al. Single-cell transcriptomics of a human kidney allograft biopsy specimen defines a diverse inflammatory response. J Am Soc Nephrol 2018; **29**: 2069–2080