

Available online at http://www.toxmut.or.kr

Effects of DHU001, a Mixed Herbal Formula on Acute Inflammation in Mice

Young-Doo Back¹, Hyeung-Sik Lee¹ and Sae-Kwang Ku²

¹Department of Herbal Biotechnology, ²Department Anatomy and Histology, College of Oriental Medicine, Daegu Haany University, Gyeongsan 712-715, Korea

(Received June 11, 2008; Revised August 13, 2008; Accepted August 16, 2008)

The effects of DHU001, a mixed herbal formula consisted of 7 types aqueous extracts for treating respiratory disorders were observed on xylene-induced acute inflammation. The xylene was topically applied 60 min after administration of 500, 250 and 125 mg/kg of DHU001, and all animals were sacrificed 2 hrs after xylene application. The changes on ear weights, histolopathological analyses of ear were evaluated and compared to those of indomethacin and dexamethasone (15 mg/kg treated) - Both of drugs are well-known by anti-inflammatory agents. Xylene application resulted in marked increases in induced ear weights as compared with intact control ear. Severe vasodilation, edematous changes of ear skin and increase in the thickness of the ear tissues, neutrophil infiltration as acute inflammation were detected in xylene-treated control ears at histopathological observation. However, these xylene-induced acute inflammatory changes were dose-dependently decreased by oral treatment of DHU001. Therefore, it is concluded that DHU001 has favorable anti-inflammatory effects on xylene-applicated acute ear inflamed mice.

Key words: Mixed herbal formula, DHU001, Acute inflammation, Histology, Mouse

INTRODUCTION

Inflammation is an essential protective process preserving the integrity of organisms against physical, chemical and infective insults. However, the inflammatory response to several insults frequently leads to erroneous damage to normal tissues (Habashy et al., 2005). Physical damage, chemical substances, microorganisms and other agents are all possible causes of acute inflammation. The inflammatory responses to such insults consist of changes in blood flow, increased permeability of blood vessels and the subsequent escape of cells from the blood into the tissues. The changes are essentially the same regardless of the cause or its location. Acute inflammation is short-lived, typically lasting only a few days. If the inflammation is longer lasting, however, it is referred to as chronic inflammation. The xylene-induced acute inflammatory mouse ear model has generally been used as one of the classic methods for detecting the efficacy of antiinflammatory agents. In this model, the anti-inflammatory effect of a drug is based on observations of ear weight and histopathology (Hosseinzadeh *et al.*, 2003; Kou *et al.*, 2005). As control anti-inflammatory drugs, indomethacin are cyclo-oxygenase inhibitors and dexamethasone is a well-known glucocorticoid; these drugs are the most widely used as controls in the development of the new anti-inflammatory drugs (Hosseinzadeh and Younsei 2002; Ramprasath *et al.*, 2006).

DHU001 is a mixed herbal formula consisted of 7 types aqueous extracts; Ficis fructus, Liriopis tuber, Platycodi radix, Schisandrae fructus, Glycyrrhizae radix, Zingiberis rhizome and Menthae herba and being developed for respiratory disorders. Among 7 types of herbal components of DHU001, Liriopis tuber (Park and Geon, 2003), Platycodi radix (Kim *et al.*, 2004), Schisandrae fructus (Narimanian *et al.*, 2005; Rhyu *et al.*, 2006), Glycyrrhizae radix (Sun and Pan, 2006), Zingiberis rhizome (Aimbire *et al.*, 2007; Ghayur *et al.*, 2008) and Menthae herba (Shin, 2003) has been used for treating various respiratory symptoms. In addition, anti-inflam-

Correspondence to: Sae-Kwang Ku, Department of Anatomy and Histology, College of Oriental Medicine, Daegu Haany University, 290, Yugok-dong, Gyeongsan-si, Gyeongsangbukdo 712-715, Korea E-mail: gucci200@hanmail.net

matory effects of Platycodi radix (Ahn *et al.*, 2005; Jang *et al.*, 2006; Kim *et al.*, 2006), Glycyrrhizae radix (Park *et al.*, 2005), Zingiberis rhizome (Ojewole, 2006; Aimbire *et al.*, 2007; Minghetti *et al.*, 2007). In the present study, the effects of DHU001 on the xylene-induced acute inflammation were investigated.

MATERIALS AND METHODS

Animals and husbandry. Sixty-three male ICR mice (6-week-old upon receipt; SLC, Japan) were used after acclimatization for 7 days. Animals were housed five or four per polycarbonate cage in a temperature ($20 \sim 25^{\circ}$ C)- and humidity ($40 \sim 45^{\circ}$)-controlled room with a 12 h : 12 h light:dark cycle. Feed (Samyang, Korea) and water were supplied *ad libitum*. All animals were fasted overnight before dosing and sacrifice (about 18 h with *ad libitum* access to water).

Test articles, grouping and dosing. The herbal compositions of DHU001 were listed in Table 1. Each herbal component was purchased from Cho-Heung Pharmaceutical Ind. Co. (Daegu, Korea) after confirmation of the morphology under microscopy. Approximated amounts of each herbal component was mixed (317.5 g) and boiled in 21 of distilled water for 2 hours and than filtrated. The filtrate was decompressed using a rotary vacuum evaporator (Lab. Camp, Daejeon, Korea) and lyophilized in a programmable freeze-dryer (IIShin Lab, Daejeon, Korea). Total acquired lyophilized extracts (DHU001) were 34.93 g (yield 11%). Powders of extracts were stored in a desiccator to protect against light and moisture. Indomethacin (Fluka, Switzerland) and dexamethasone (Sigma, USA) were used as reference drugs in this study. The animals were distributed into seven groups with 9 mice per group: intact control, xylene-treated control, 500, 250 and 125 mg/kg DHU001 dosing groups, 15 mg/kg indomethacin and dexamethasone dosing groups. DHU001 was orally administered

Table 1. Herbal composition of DHU001 used in this study

once, and indomethacin and dexamethasone were administered intraperitoneally once. DHU001 was dissolved in distilled water, and indomethacin and dexamethasone were dissolved or suspended in saline.

Induction of acute inflammation. 1 h after dosing with test substances, 0.03 ml of xylene (Merck, Germany) was topically applied to the anterior surface of the right ear. The left ear was considered a control.

Ear weight measurement. Two hours after xylene application, all animals were sacrificed and both ears were removed. Circular sections were taken using a cork borer with a 7-mm diameter and weighed as previously described (Kou *et al.*, 2005). The weight of the induced ear was regarded as the absolute weight. The differences between the intact ear and induced ear were calculated as follows:

Differences in weights (g) = xylene-treated ear weight - intact ear weight

Histology and histomorphometry. Following removal, induced ears were trimmed crosswise to include ear cartilage and fixed in 10% neutral buffered formalin, then embedded in paraffin, sectioned (3~4 µm) and stained with Hematoxylin & Eosin stain. The histological profiles of the ears were observed compared to that of intact and/or xylene-treated control. The thickness of the induced ear was calculated using automated image analysis (DMI-300; DMI, Korea) under 50 × magnification (Nikon, Japan). The thicknesses from epidermis to dermis of induced ear skin (anterior skin thickness; um) and anterior to posterior ear (full ear thickness, µm) were detected as histomorphometry on crossly trimmed ear specimens in regular corresponding regions in all induced ear samples. In addition, infiltrated neutrophil numbers were also calculated as cells/ 1 mm² of histological filed.

Herbs	Scientific name	Amounts (g) 140	
Ficis fructus	Ficus carica Linn.		
Liriopis tuber	Liriope spicata Lour.	45	
Platycodi radix	Platycodon grandiflorum Jacq.	60	
Schisandrae fructus	Schisandra chinensis Baill	22.5	
Glycyrrhizae radix	Glycyrrhiza uralensis Fisch	15	
Zingiberis rhizoma recens	Zingiberis rhizoma recens Zingiber officinale Roscoe		
Menthae Herba	Mentha arvensis Linne var piperascens	20	
Total	7 types	317.5	

All herbs were purchase from from Cho-Heung Pharmaceutical Ind. Co. (Daegu, Korea) and yield 11% aqueous extracts were acquired.

Statistical analyses. Multiple comparison tests of the different dose groups were carried out. The variance homogeneity was examined using the Levene test. If the Levene test indicated no significant deviations from the variance homogeneity, obtained data was analyzed using a one way ANOVA test followed by a least-significant difference multi-comparison test to determine which pairs of group comparisons were significantly different. A non-parametric comparison test, Kruskal-Wallis H test was performed in the case of significant deviations from the variance homogeneity was observed at Levene test. When a significant difference was observed in the Kruskal-Wallis H test, the Mann-Whitney U-Wilcoxon Rank Sum W test was used to determine the specific pairs of group comparisons that were significantly different. The SPSS statistical software (Release 6.1.3, SPSS Inc., USA) was used and a p value <0.05 was considered significant.

RESULTS

Changes in ear weights. Significant (p < 0.01) increases in the weight of the induced ear were detected in xylene-treated control as compared with intact control and, accordingly, the differences between intact and induced ear were also significantly (p < 0.01) increased. However, the induced ear weights and the differences between intact and induced ear of all dosing groups were significantly (p < 0.01 or p < 0.05) decreased compared to that of xylene-treated control (Table 2).

Histological analyses of induced ear. The classic histological profile of acute inflammation, that is, severe vasodilation and edematous changes of skin, was detected in xylene-treated control. However, these histological indicators of acute inflammation were dramatically decreased in all dosing groups as compared with xylene-treated control. In addition, dose-dependency was also demonstrated in the DHU001-treated groups (Fig. 1). Significant (p < 0.01) increases in the infiltrated neutrophils, thickness of induced ear (both anterior skin and full ear thicknesses) were detected in xylene-treated control compared to that of intact control. However, these increases in the infiltrated neutrophils and thickness of the induced ear were significantly (p < 0.01 or p < 0.05) decreased in all dosing groups compared to that of xylene-treated control (Table 3).

DISCUSSION

The xylene-induced acute inflammation of the mouse ear has generally been used as one of the classic methods for detecting the efficacy of anti-inflammatory agents (Hosseinzadeh et al., 2003; Kou et al., 2005). In the present study, the effects of DHU001, a mixed herbal formula consisted of 7 types aqueous extracts, on xylene-induced acute inflammation were observed. As the result of pretreatment with DHU001, the changes associated with acute inflammation such as the marked increases of induced ear weight, vasodilation, edematous changes in the skin and increases in the thickness of ear tissues were significantly and dosedependently decreased. These results are considered direct evidence that DHU001 inhibits the induction of the acute inflammatory response. Previously reported immunomodulatory effects of Platycodi radix (Yoon et al., 2003), Glycyrrhizae radix (Sun and Pan, 2006) and Zingiberis rhizome (Tan and Vanitha, 2004), and antioxidative effects of Ficis fructus (Pèrez et al., 2003), Liriopis tuber (Han et al., 1981), Platycodi radix (Lee et al., 2004; Wang et al., 2004), Schisandrae fructus (Ko and Lam, 2002), Glycyrrhizae radix (Zhou and Xu, 1992),

Table 2. Changes in ear weights in intact or xylene-applicated mi	Table	2.	Changes	in ear	weiahts	in	intact	or x	vlene-ap	plicated	mic
---	-------	----	---------	--------	---------	----	--------	------	----------	----------	-----

Croup	Ear weight (g)			
Group –	Intact ear	Induced ear	Difference	
Controls				
Intact	0.115 ± 0.009	0.117 ± 0.006	0.002 ± 0.006	
Xylene	0.115 ± 0.009	0.151 ± 0.011*	0.036 ± 0.017*	
References				
Dexamethasone	0.115 ± 0.013	$0.128 \pm 0.019^{\#}$	0.013 ± 0.017 [#]	
Indomethacin	0.114 ± 0.007	0.133 ± 0.009* ^{, ##}	0.018 ± 0.010** ^{,##}	
DHU001				
500 mg/kg	0.115 ± 0.007	0.133 ± 0.017** ^{,#}	0.018 ± 0.017** ^{,#}	
250 mg/kg	0.114 ± 0.012	0.133 ± 0.017** ^{,#}	0.019 ± 0.012* ^{,#}	
125 mg/kg	0.115 ± 0.011	0.134 ± 0.012 ^{*,##}	0.019 ± 0.018* ^{,#}	

Mean ± S.D. of nine mice; Differences = induced ear weight - intact ear weight; *p < 0.01 and **p < 0.05 compared with intact control; *p < 0.01 and **p < 0.05 compared with xylene control.

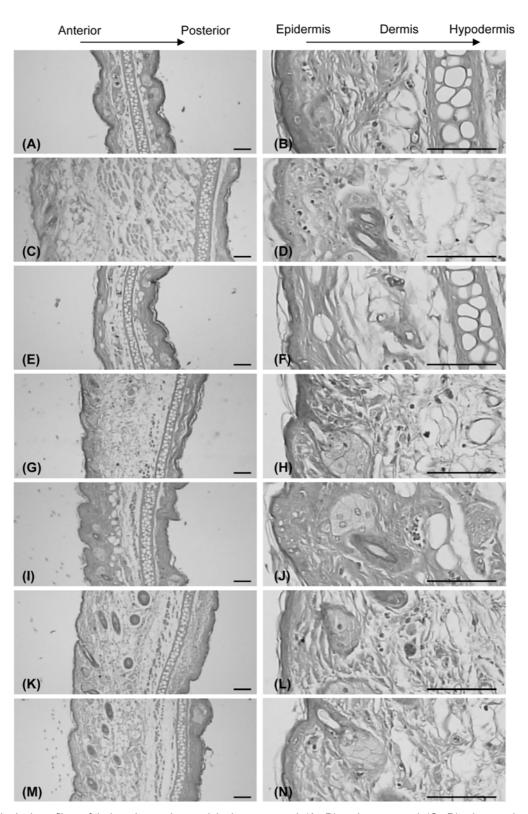


Fig. 1. Histological profiles of induced ear detected in intact control (A, B), xylene control (C, D), dexamethasone (E, F), indomethacin (G, H), 500 (I, J), 250 (K, L) and 125 (M, N) of DHU001-administered groups. Note the classic histological profile of acute inflammation as severe vasodilation and edematous changes of skin were detected in xylene control. However, these inflammation indicators were dramatically decreased in all dosing groups tested. All Hematoxylin & Eosin staining, Scale bars = 80 μ m.

Crown	Neutrophil numbers (cells/1 mm ² of field)	Thickness of ear (μm)			
Group		Full ear thickness	Anterior skin thickness		
Controls					
Intact	8.89 ± 4.17	766.765 ± 127.712	324.387 ± 94.258		
Xylene	75.56 ± 11.58*	1916.069 ± 147.049*	1495.320 ± 166.733*		
References					
Dexamethasone	7.78 ± 5.07 [#]	763.837 ± 103.992 [#]	328.823 ± 71.952 [#]		
Indomethacin	38.33 ± 12.75* ^{,#}	1059.186 ± 221.111* ^{,#}	739.264 ± 185.304* ^{,#}		
DHU001					
500 mg/kg	16.11 ± 8.21** ^{,#}	925.185 ± 115.760 [#]	561.230 ± 155.201*,#		
250 mg/kg	36.67 ± 12.50* ^{,#}	1317.571 ± 263.949* ^{,#}	980.569 ± 271.066*,#		
125 mg/kg	47.22 ± 10.93* ^{,#}	1543.921 ± 199.440* ^{,#}	1282.226 ± 221.894* ^{,##}		

Table 3. The histomorphometric analyses

Mean ± S.D. of nine mice; Anterior skin thickness = thicknesses from epidermis to dermis of induced ear skin; Full ear thickness = thicknesses from anterior to posterior ear of induced ear, *p < 0.01 and **p < 0.05 compared with intact control; *p < 0.01 and **p < 0.05 compared with intact control; *p < 0.01 and **p < 0.05 compared with xylene control.

Zingiberis rhizome (Masuda *et al.*, 2004) and Menthae herba (Baliga *et al.*, 2003) were considered one of the major mechanisms of the anti-inflammatory effect of DHU001. In the present study, the anti-inflammatory effect of DHU001 was detected, because immunomodulatory agents can reduce inflammation (Ramprasath *et al.*, 2006) and, furthermore, NO synthase inhibitors can reverse several classic inflammatory symptoms (Amin *et al.*, 1995).

After topical application of xylene, marked increases in ear weight were detected as a result of the acute inflammation response; these increases in ear weight have been used as valuable markers for anti-inflammatory effects (Al-Majed *et al.*, 2003; Ojewole 2005). In the present study, the increases in ear weight were dosedependently inhibited by DHU001 treatment. This inhibition was considered direct evidence that the DHU001 used in this study has favorable effects on reducing the acute inflammatory response.

Histopathologically, severe vasodilation and edematous changes of skin were detected as signs of acute inflammation after topical application of xylene (De La Puerta *et al.*, 1996; Kou *et al.*, 2003; Rotelli *et al.*, 2003; Kim *et al.*, 2007); as a result of these histopathological changes, the thickness of the ear tissues was also markedly increased. However, these histopathological changes and the thickness of ear tissues were dosedependently decreased after pretreatment with three different dosages of DHU001 in the present study. This inhibition was again considered direct evidence that the DHU001 used in this study has favorable effects on reducing the acute inflammatory response.

Based on the results, we conclude that the DHU001, a mixed herbal formula has somewhat favorable effect in the reduction of the acute inflammatory response induced by xylene application in mice.

REFERENCES

- Ahn, K.S., Noh, E.J., Zhao, H.L., Jung, S.H., Kang, S.S. and Kim, Y.S. (2005). Inhibition of inducible nitric oxide synthase and cyclooxygenase II by *Platycodon grandiflorum* saponins via suppression of nuclear factor-kappaB activation in RAW 264.7 cells. *Life Sci.*, **76**, 2315-2328.
- Aimbire, F., Penna, S.C., Rodrigues, M., Rodrigues, K.C., Lopes-Martins, R.A. and Sertié, J.A. (2007). Effect of hydroalcoholic extract of *Zingiber officinalis* rhizomes on LPS-induced rat airway hyperreactivity and lung inflammation. *Prostaglandins Leukot Essent Fatty Acids*, **77**, 129-138.
- Al-Majed, A.A., Khattab, M., Raza, M., Al-Shabanah, O.A. and Mostafa, A.M. (2003). Potentiation of diclofenacinduced anti-inflammatory response by aminoguanidine in carrageenan- induced acute inflammation in rats: the role of nitric oxide. *Inflamm. Res.*, **52**, 378-382.
- Amin, A.R., Vyas, P., Attur, M., Leszczynska-Piziak, J., Patel, I.R., Weissmann, G. and Abramson, S.B. (1995). The mode of action of aspirin-like drugs: effect on inducible nitric oxide synthase. *Proc. Natl. Acad. Sci. USA*, **92**, 7926-7930.
- Baliga, M.S., Jagetia, G.C., Rao, S.K. and Babu, K. (2003). Evaluation of nitric oxide scavenging activity of certain spices *in vitro*: a preliminary study. *Nahrung*, **47**, 261-264.
- Ghayur, M.N., Gilani, A.H. and Janssen, L.J. (2008). Ginger attenuates acetylcholine-induced contraction and Ca²⁺ signalling in murine airway smooth muscle cells. *Can. J. Physiol. Pharmacol.*, **86**, 264-271.
- Habashy, R.R., Abdel-Naim, A.B., Khalifa, A.E. and Al-Azizi, M.M. (2005). Anti-inflammatory effects of jojoba liquid wax in experimental models. *Pharmacol. Res.*, **51**, 95-105.
- Han, B.H., Yoo, S.Y., Park M.H. and Lee, H.J. (1981). Antioxidant activity screening on crude drugs. *J. Pharmacogn.*, **12**, 66.
- Hosseinzadeh, H. and Younesi, H.M. (2002). Antinociceptive and anti-inflammatory effects of *Crocus sativus* L. stigma and petal extracts in mice. *BMC Pharmacol.*, **2**, 7-12.
- Hosseinzadeh, H., Haddadkhodaparast, M.H. and Arash, A.R.

(2003). Antinociceptive, antiinflammatory and acute toxicity effects of *Salvia leriifolia* Benth seed extract in mice and rats. *Phytother. Res.*, **17**, 422-425.

- Jang, M.H., Kim, C.J., Kim, E.H., Kim, M.G., Leem, K.H. and Kim, J. (2006). Effects of *Platycodon grandiflorum* on lipopolysaccharide-stimulated production of prostaglandin E2, nitric oxide, and interleukin-8 in mouse microglial BV2 cells. *J. Med. Food*, **9**, 169-174.
- Kim, H.D., Cho, H.R., Moon, S.B., Shin, H.D., Yang, K.J., Park, B.R., Jang, H.J., Kim, L.S., Lee, H.S. and Ku, S.K. (2007). Effects of beta-glucan from *Aureobasidium pullulans* on acute inflammation in mice. *Arch. Pharm. Res.*, **30**, 323-328.
- Kim, J.Y., Hwang, Y.P., Kim, D.H., Han, E.H., Chung, Y.C., Roh, S.H. and Jeong, H.G. (2006). Inhibitory effect of the saponins derived from roots of *Platycodon grandiflorum* on carrageenan-induced inflammation. *Biosci. Biotechnol. Biochem.*, **70**, 858-864.
- Kim, S.S., Lee, S.C., Shin, H.D., Shin, M.K., Kim J.H. and Song, H.J. (2004). Studies on allergy asthma effect of Radix Platicodi. *Kor. J. Herbology*, **19**, 61-70.
- Ko, K.M. and Lam, B.Y. (2002). Schisandrin B protects against tert-butylhydroperoxide induced cerebral toxicity by enhancing glutathione antioxidant status in mouse brain. *Mol. Cell Biochem.*, **238**, 181-186.
- Kou, J., Ma, R., Zhu, D. and Yan, Y. (2003). Blood-activating and anti-inflammatory actions of *Polygala fallax*. *Zhong Yao Cai*, **26**, 268-271.
- Kou, J., Ni, Y., Li, N., Wang, J., Liu, L. and Jiang, Z.H. (2005). Analgesic and anti-inflammatory activities of total extract and individual fractions of Chinese medicinal ants *Polyrhachis lamellidens. Biol. Pharm. Bull.*, **28**, 176-180.
- Lee, K.J., Choi, C.Y., Chung, Y.C., Kim, Y.S., Ryu, S.Y., Roh, S.H. and Jeong, H.G. (2004). Protective effect of saponins derived from roots of *Platycodon grandiflorum* on tert-butyl hydroperoxide-induced oxidative hepatotoxicity. *Toxicol. Lett.*, **147**, 271-282.
- Masuda, Y., Kikuzaki, H., Hisamoto, M. and Nakatani, N. (2004): Antioxidant properties of gingerol related compounds from ginger. *Biofactors*, **21**, 293-296.
- Minghetti, P., Sosa, S., Cilurzo, F., Casiraghi, A., Alberti, E., Tubaro, A., Loggia, R.D. and Montanari, L. (2007). Evaluation of the topical anti-inflammatory activity of ginger dry extracts from solutions and plasters. *Planta Medica*, **73**, 1525-1530.
- Narimanian, M., Badalyan, M., Panosyan, V., Gabrielyan, E., Panossian, A., Wikman, G. and Wagner, H. (2005). Impact of Chisan (ADAPT-232) on the quality-of-life and its efficacy as an adjuvant in the treatment of acute nonspecific pneumonia. *Phytomedicine*, **12**, 723-729.

Ojewole, J.A. (2005). Antinociceptive, anti-inflammatory and

antidiabetic effects of *Bryophyllum pinnatum* (Crassulaceae) leaf aqueous extract. *J. Ethnopharmacol.*, **99**, 13-19.

- Ojewole, J.A. (2006). Analgesic, antiinflammatory and hypoglycaemic effects of ethanol extract of *Zingiber officinale* (Roscoe) rhizomes (Zingiberaceae) in mice and rats. *Phytother. Res.*, **20**, 764-772.
- Park, E., Kum, S., Wang, C., Park, S.Y., Kim, B.S. and Schuller-Levis, G. (2005). Anti-inflammatory activity of herbal medicines: inhibition of nitric oxide production and tumor necrosis factor-alpha secretion in an activated macrophage-like cell line. *Am. J. Chin. Med.*, **33**, 415-424.
- Park, J.H. and Geon, D.G. (2003). Pharmacognostical studies on the Chinese crude drug "Maig Moon Dong". Kor. J. Pharmacogn., 34, 6-9.
- Pèrez, C., Canal, J.R. and Torres, M.D. (2003). Experimental diabetes treated with *Ficus carica* extract: effect on oxidative stress parameters. *Acta Diabetol.*, **40**, 3-8.
- Ramprasath, V.R., Shanthi, P. and Sachdanandam, P. (2006). Immunomodulatory and anti-inflammatory effects of *Semecarpus anacardium* LINN. Nut milk extract in experimental inflammatory conditions. *Biol. Pharm. Bull.*, **29**, 693-700.
- Rhyu, M.R., Kim, E.Y., Yoon, B.K., Lee, Y.J. and Chen, S.N. (2006). Aqueous extract of *Schizandra chinensis* fruit causes endothelium-dependent and -independent relaxation of isolated rat thoracic aorta. *Phytomedicine*, **13**, 651-657.
- Shin, T.Y. (2003). Inhibition of immunologic and nonimmunologic stimulation-mediated anaphylactic reactions by the aqueous extract of *Mentha arvensis*. *Immunopharmacol. Immunotoxicol.*, 25, 273-283.
- Sun, H.X. and Pan, H.J. (2006). Immunological adjuvant effect of *Glycyrrhiza uralensis* saponins on the immune responses to ovalbumin in mice. *Vaccine*, **24**, 1914-1920.
- Tan, B.K. and Vanitha, J. (2004). Immunomodulatory and antimicrobial effects of some traditional chinese medicinal herbs: a review. *Curr. Med. Chem.*, **11**, 1423-1430.
- Wang, C., Schuller Levis, G.B., Lee, E.B., Levis, W.R., Lee, D.W., Kim, B.S., Park, S.Y. and Park, E. (2004). Platycodin D and D3 isolated from the root of *Platycodon grandiflorum* modulate the production of nitric oxide and secretion of TNF-alpha in activated RAW 264.7 cells. *Int. Immunopharmacol.*, **4**, 1039-1049.
- Yoon, Y.D., Han, S.B., Kang, J.S., Lee, C.W., Park, S.K., Lee, H.S., Kang, J.S. and Kim, H.M. (2003). Toll-like receptor 4-dependent activation of macrophages by polysaccharide isolated from the radix of *Platycodon grandiflorum*. *Int. Immunopharmacol.*, **3**, 1873-1882.
- Zhou, Y. and Xu, R. (1992). Antioxidative effect of Chinese drugs. *Zhongguo Zhong Yao Za Zhi*, **17**, 368-369.

194