Human papillomavirus infection and ocular surface disease (Review)

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Abstract. Human papillomavirus (HPV) infection has been implicated as a primary cause of lesions in the anogenital region, skin, oropharynx and respiratory tract. Additionally, the role of HPV in the pathogenesis of ocular surface disease has also been extensively studied. Conjunctival papilloma development has been strongly associated with the HPV infection of certain subtypes. On the other hand, the role of HPV in conjunctival pterygium, conjunctival intraepithelial neoplasia (CIN) and ocular surface squamous neoplasia (OSSN) remains controversial. Genetic predisposition and environmental factor is important in HPV hosts as regards the pathogenesis of ocular surface disease. Several studies have indicate a synergic role of HPV with ultraviolet radiation in pterygium establishment. A higher recurrence risk rate and more aggressive disease of ophthalmic pterygium is observed in cases of HPV infection. The purpose of this review was to provide a systematic review of the literature and to assist in a better understanding of the role of HPV in ocular surface disease.

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Abbreviations: HPV, human papillomavirus; CIN, conjunctival intraepithelial neoplasia; OSSN, ocular surface squamous neoplasia; SCCC, squamous cell carcinoma of the conjuctiva; UV, ultraviolet; MMC, mitomycin C; EV, Epidermodysplasia verruciformis

Key words: human papillomavirus, virus, conjunctiva, papilloma, neoplasia

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1. Introduction

Papillomaviruses (HPV) belong to the virus family of *Papovaviridae*. The papillomaviruses are highly host species-specific and tissue-restricted, and present epithelial tropism. More than 100 genotypes of human papillomaviruses have been fully sequenced, and more HPV types are continuously being found (1,2).

All recognized viruses require terminal differentiation for replication and virion production. HPVs cause a wide range of diseases from benign lesions to invasive tumors (3). They are grouped in 3 groups as follows: Cutaneous, mucocutaneous and associated with the autosomal recessive epidermodysplasia verruciformis (EV). According to their propensity for malignant progression in the cervix, mucocutaneous HPVs have been divided into low- and high-risk types (4).

Papillomaviruses are small, non-enveloped viruses with icosahedral symmetry and contain a double-strained circular DNA with approximately 8 open reading frames which are divided into 3 functional regions: The early (E) region encoding proteins E1- E7 responsible for viral replication, the late (L) region encoding structural proteins L1-L2, and the long control region (LCR) responsible for transcription and replication (5,6). Only the E6 and E7 proteins of high-risk HPV strains present transforming properties by interacting with the tumor suppressor genes p53 and with the retinoblastoma family of proteins pRb, involved in controlling cell proliferation (7,8). The viral proteins may also contribute to potentially higher number of abnormalities in the cell genetic material (9).

In recent years, HPV has been associated with benign and malignant lesions of not only the anogenital region, but also of the skin, oropharynx, respiratory tract and ocular surface with variation of different genotypes tropism in the various anatomical sites (10-12). The epithelium of the ocular surface is exposed to the environment and therefore it is susceptible to infections, particularly in cases when protective barriers of mucin, tears and superficial cellular layer are compromised. HPV has mainly been shown to be involved in the pathogenesis

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of ocular surface diseases, such as conjunctival papillomas, papillomas and carcinomas of the lacrimal sac, conjunctival intraepithelial neoplasia (CIN), ocular surface squamous neoplasia (OSSN) and conjunctival pterygium and even squamous cell carcinoma of the conjunctiva (SCCC) (12-14).

2. Data collection methods

For this review article, a thorough search on MEDLINE (through PubMed), EMBASE (through OVID) and SCOPUS was performed, from inception to February, 2019, in order to identify studies addressing the association between infection with HPV and ocular surface disease.

3. HPV detection in ophthalmic pterygium

Conjunctival pterygia are fibrovascular lesions of the bulbar conjunctiva, that can display an aggressive clinical behavior and, occasionally, threaten vision. They represent a proliferative disorder of the conjunctiva, characterized by the overgrowth of altered limbal cells centripetally towards the cornea in a wing-shaped manner. In advanced stages, they present Bowman's layer dissolution, epithelial and mesenchymal transition and stromal inflammation, neovascularization and matrix remodeling under the action of cytokines, growth factors matrix metalloproteinases and vascular endothelial growth factors. They have a predilection for nasal limbus and their growth can obscure the visual axis, and can cause irregular astigmatism and chronic inflammation (15). Its pathogenesis is considered a multifactorial process where ultraviolet (UV) radiation (16) and other environmental factors, genetic predisposition and oncogenic viruses may play a role (17). Increased UV-associated oxidative stress has been reported in pterygium, compared with normal conjunctiva, leading to the induction of proteins, such as survivin (18). The latter has been associated with DNA oxidation and the downregulation of p53 (18). Detorakis et al (19) previously identified potential viral co-morbidity in pterygium development and proposed a 'two-hit' theory for its establishment. The first hit is a damaging reaction mediated by UV radiation exposure that leads to genetic alterations or mutations, and the second hit is an oncogenic event mediated by viral infection in the compromised ocular barriers.

HPV has been extensively studied as a possible pathogenetic co-factor; however controversy exists between different studies (Table I). In their studies, McDonnell *et al* (20), Dushku *et al* (22), Chen *et al* (26), Schellini *et al* (28), Kuo *et al* (29), Otlu *et al* (32), Guthoff *et al* (34) and Hamed-Azzam *et al* (38) did not detect HPV in pterygia, while Sjo *et al* (13), Takamura *et al* (30) and Hsaio *et al* (36) noted a very low prevalence of the virus ranging from 3-4,8%.

On the contrary, a number of previously published studies have successfully detected HPV (21,23-25,27,31,33,35,37,39) The average HPV prevalence in human pterygium was found to be 18.6% (range 0-100%) (40). The lack of consensus between studies could reflect differences in methodology and sampling. Piras *et al* (25) proposed that geographic differences in the prevalence of the virus in the various countries may explain these findings, supporting its multifactorial pathogenesis.

The diagnosis of viral infection is based on the detection of HPV-DNA. The application of various detection techniques with varied sensitivity and specificity can significantly compromise the results (41). HPV-DNA can be directly isolated from a biopsy specimen with in situ hybridization (ISH), Southern blotting and dot blot hybridization. These techniques however, are laborious and lack sensitivity. By contrast, polymerase chain reaction (PCR) is highly associated with false-positive results due to its high sensitivity (42). Real-time PCR permits the rapid detection and quantification of the viral load (42). Reverse transcriptase-PCR is a qualitative assay that permits the identification of viral gene expression with the use of reverse transcriptase (42). A combination of the previous techniques can be applied in order to acquire qualitative and qualitative information of viral gene expression.

Pterygium treatment is based on surgical excision and the topical use of antimetabolites, such as mitomycin C (MMC) or 5-fluorouracil. Several surgical techniques have been described, such as bare sclera closure and sliding conjunctival flaps (43). The additional use of conjunctival autografts or amniotic membrane grafts, has significantly lowered the need for repetitive surgery, although recurrences may still occur (44).

4. HPV detection in conjunctival papilloma

Conjunctival papillomas represent one of the most common benign tumors of the squamous epithelium of the conjunctiva. They may display dysplasia, but rarely undergo malignant transformation (45). Despite their tendency to recur following surgical excision, spontaneous regression is possible. According to their growth pattern, conjunctival papillomas may present as exophytic, mixed or more rarely inverted. The exophytic growth pattern may be sessile or pedunculated (12). Although they can appear in both children and adults (46), they most frequently occur in adults aged 20-39 years, which also corresponds to the peak age of genital HPV infection in sexually active adults. Papillomas seem to appear with a male preponderance and their incidence declines with increasing age. They are usually detected medially and inferiorly on the conjunctiva (45).

Their pathogenetic role has not been fully clarified. HPV infection by auto-inoculation from contaminated fingers has been strongly associated with their development. Fetal passage through an HPV-infected birth canal may explain the presence of conjunctival papilloma in children. Several studies have investigated the role of HPV in the pathogenesis of conjunctival papilloma (Table II). In the majority of these cases, the low-risk HPV types 6 and 11, typically found in condylomata accuminata, are predominant among HPV-infected conjunctival papillomas with an incidence varying from 50-100%. Egbert and Kersten (61) also refer the detection of HPV 6 and 11 in a conjunctival papilloma of an infant whose mother suffered from vulvar HPV infection during pregnancy, indicating a possible vertical transmission during delivery. The detection of low-risk HPV types in conjunctival papillomas may explain the benign nature of these lesions. In addition, HPV infection has been also detected in epithelial lacrimal sac papillomas and carcinomas. The tear

Author (Refs.)	Date of publication	HPV prevalence	HPV type	Country of the study	Method of detection	Sample size
McDonell et al (20)	1992	0%	-	USA	PCR	6
Varinli et al (21)	1994	64%	-	Turkey	IHC	25
Dushku et al (22)	1999	0%	-	USA	PCR	13
Detorakis et al (23)	2001	24%	18	Greece	PCR	50
Gallagher et al (24)	2001	50%	6, 11, 16	UK	PCR	10
Piras et al (25)	2003	100% 21%	Types 52, 54, <i>cand</i> HPV90	Italy/ Equador	PCR, sequencing	41
Chen et al (26)	2003	0%	-	Taiwan	PCR	65
Ateenyi-Agaba et al (27)	2004	50%	11,37	Uganda	PCR, Southern blotting	10
Schellini et al (28)	2006	0%	-	Brazil	PCR	36
Kuo et al (29)	2006	0%	-	Taiwan	PCR	4
Sjö et al (13)	2007	4.4%	6	Denmark	PCR, ISH	90
Takamura <i>et al</i> (30)	2008	4.8%	-	Japan	PCR-HC II	42
Rodrigues et al (31)	2008	58.3%	1, 2, 16	Brazil	PCR	36
Otlu et al (32)	2009	0%	-	Turkey	Real-time PCR	40
Tsai et al (33)	2009	24%	16, 18	Taiwan	Nested PCR	129
Guthoff et al (34)	2009	0%	-	Germany	PCR, IHC	
Piecyk-Sidor et al (35)	2009	27.6%	5, 6, 11, 16, 18, 31, 52, 59	Poland	PCR	58
Hsiao et al (36)	2010	3%	18	Taiwan	PCR, ISH	65
Chong et al (37)	2014	64.4%	16, 18, 58, 59	Malaysia	Nested PCR	45
Hamed-Azzam <i>et al</i> (38)	2016	0%	-	Israel	IHC	100
Chalkia et al (39)	2018	42.86%	33, 39, 45, 56, 59, 66	Greece	Real-time PCR	21

Table L	HPV	in	ophthalmic	pterygium.
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HPV, human papillomavirus; IHC, immunohistochemistry; ISH, in situ hybridization; HC II, hybrid capture II.

flow on an HPV infected conjunctiva may be responsible for the development of such lesions.

Treatment options of conjunctival papillomas include mainly surgical excision, cryotherapy and carbon dioxide laser (62). Additionally, oral cimetidine, topical MMC and topical interferon- α have been implicated in the management of these benign lesions (63). However, despite the various therapies, the recurrence rate in conjunctival papilloma remains high (6-27%).

5. HPV detection in ocular surface squamous neoplasia

OSSN encompasses a wide range of conjunctival lesions that range histologically from dysplasia and carcinoma *in situ*, generally termed CIN, to invasive SCCC and is considered the most common ocular-surface malignancy (64). It involves more commonly the interpalpebral area, arising from the limbus and may extend to involve cornea (65). The incidence of OSSN varies widely with increased incidence in countries where HIV infection is an epidemic (66). In fact, immunosuppression due to HIV-infection has been strongly associasted with OSSN, as various studies have demonstrated a 10-fold increase for OSSN in these patients. Other risk factors associated with OSSN are an advanced age, the male sex, UV exposure, immunosuppression, atopic eczema and xeroderma pigmentosum (66,67). Conjunctival SCC represents the most severe form of OSSN, which if left untreated, can result in mortality (68). Metastasis to lymph nodes is common; thus, aggressive treatment with enucleation or exenteration should be considered (69).

A number of studies have been conducted in order to identify the presence of HPV in OSSN (Table III). However, the reported presence of the virus varies widely with a range from 0-100%. From the studies reported to date in Table III, an average prevalence rate of 33.8% (range, 0-100%) has been observed. It should also be noted that in the majority of these studies, the HIV status of the patients has not been disclosed, leaving the role of HIV as enhancer or confounder of HPV carcinogenicity in OSSN unclear. In a number of studies, mucosal, mainly high risk types 16 and 18 have been detected. Scott et al (75) isolated HPV16 and mRNA corresponding to the E6 region in CIN specimens. Notably, Ateenyi-Agaba et al (14) detected cutaneous HPV types in nearly half of OSSN cases in HIV-positive patients, but rarely in HIV-negative patients, and thus no association was found with mucosal types in both groups.

Author (Refs.)	Date of publication	HPV prevalence	HPV types	Method of detection	Sample size
Lass et al (47)	1983	50%	11	SB	2
Naghashfar et al (48)	1986	0%	-	SB/ISH	1
McDonnell et al (49)	1987	65%	6,11	ISH	23
Mäntyjärvi et al (50)	1989	0%	-	ISH	1
Fierlbeck et al (51)	1990	0%	-	ISH	1
Mincione et al (52)	1992	50%	6,11	ISH	4
Saegusa et al (53)	1995	100%	16	PCR	5
Michel et al (54)	1996	0%	-	ISH	1
Nakamura et al (55)	1997	50%	6	PCR	8
Assadoullina et al (56)	2000	0%	-	PCR	1
Sjo et al (57)	2001	92%	6, 11, 16	PCR	52
Minchiotti et al (58)	2006	100%	11	PCR	4
Sjö et al (59)	2007	81%	6, 11, 45	PCR	106
Takamura <i>et al</i> (30)	2008	100%	-	PCR/HC-II	8
Annadanam et al (60)	2017	100%	6,11	ISH	1

Table II. HPV in conjuctival papilloma.

Chauhan *et al* (84) also reported an improved disease-free survival of patients with HPV-infected OSSN. On the other hand, De Koning *et al* (81) noted a low prevalence of cutaneous HPV cases and no evidence of association of mucosal HPV types with OSSN. Tornesello *et al* (79) also noted an absence of high-risk types and low detection of EV-related types in their study and yet, another study (74) did not note significant correlation between abnormal p53 gene-product expression in OSSN and HPV infection. These discrepancies between the different studies render the role of HPV in OSSN uncertain.

Margin-free excision remains the treatment of choice for OSSN. The additional use of topical mitomycin, 5-fluorouracil, interferon and cryotherapy and radiation may be used to reduce the risk of recurrence and metastasis (87).

6. Conclusions and future perspectives

HPV is a known cause of intraepithelial damage which leads to squamous neoplasms on mucosal surfaces (88). Many types of cervical carcinoma and precancerous lesions have been attributed to HPV infection. In addition, HPV has been linked with dysplastic and malignant squamous lesions of the oropharynx (89). The associatoin between HPV and squamous neoplasms of the ocular surface and the conjunctiva is not completely understood. It appears that the HPV genotype as well as the presence of associated risk factors play a significant role in lesion pathogenesis (13,45).

Koilocytosis is the histological hallmark of HPV infection (90). The koilocyte is a superficial or intermediate mature squamous cell characterized by perinuclear vacuolation, densely staining peripheral cytoplasm, and a nucleus with an undulating nuclear membrane and a rope-like chromatin pattern (91). Viral antigen has been demonstrated in nuclei of koilocytes using broad spectrum papillomavirus antibodies (47).

HPV has been identified in several lesions of the ocular surface. A strong association between HPV types 6 and 11 and conjunctival papilloma has been established (49,52,57,59,60). The varied percentages of papillomas associated with HPV presence could be attributed to differences in genetic predisposition, lifestyle and environmental exposure (73). In cases of pterygium, the association between the virus presence is not clear. Based on current data, HPV appears to function as a pathogenetic co-factor in addition to genetic factors like p53 gene mutation (92), as well as environmental factors, such as UV radiation and HIV co-infection (18) and chemical exposure (73). Despite the controversies in the literature, HPV infections seem to be a crucial co-morbidity in susceptible hosts (33). Frequent recurrences of a pterygium following excision may be associated with the presence of HPV (24). The proposed pathogenesis involves p53 inactivation (33).

While there is a strong association between HIV and the risk of OSSN (66), the role of HPV is less conclusive. Previous studies have suggested that only the cutaneous HPV subtype, and not the mucosal, is correlated with the presence of OSSN (14,81). Furthermore, older individuals seem to be more prone to the development of the described lesions (40).

The discrepancies in the reported results may be attributed to the selection bias of different regions (25) and the different methods for HPV isolation. While there is no gold standard for measuring HPV, PCR is generally considered to be the most sensitive method (76). It should also be noted that the newer HPV subtypes are continuously sequenced; thus, the range of possible genotype identification is incomplete. Some unknown types of HPV could be involved in the pterygium pathogenesis (93). Chalkia *et al* (39) described PCR-mediated

Table III. HPV in OSSN.

Author (Refs.)	Date of publication	HPV prevalence	HPV types	Method of detection	Sample size	HIV status
Lauer et al (70)	1990	80%	16, 18	PCR	5	_
Tuppurainen et al (71)	1992	0%	-	PCR/ISH	4	-
McDonnell et al (20)	1992	88%	16	PCR	42	-
Tabrizi et al (72)	1997	39%	6, 11, 16, 18	PCR	88	-
Karcioglu et al (73)	1997	55.6%	16,18	Nested PCR, SB	45	-
Dushku et al (22)	1999	0	-	Nested PCR	8	-
Toth <i>et al</i> (74)	2000	22%	16, 18	PCR/IHC	23	-
Scott et al (75)	2002	100%	16, 18	PCR/ISH	10	-
Eng et al (76)	2002	0%	-	Nested PCR	20	-
Tulvatana <i>et al</i> (77)	2003	0%	-	PCR/dot hybridization	30	-
Ateenyi-Agaba et al (27)	2004	86%	EV-HPV types	PCR	21	_
Moubayed <i>et al</i> (78)	2004	93%	6, 11, 18	ISH immunomax		64,2%
Tornesello <i>et al</i> (79)	2006	19.8%	6, 18, EV-related HPVs, CJ198	PCR	86	65,1% (25% HPV ⁺)
Kuo <i>et al</i> (29)	2006	100%	6, 11, 16, 18, 33, 37, 58, 72	Nested PCR	9	-
Sen <i>et al</i> (80)	2007	0%	-	IHC	30	-
De Koning <i>et al</i> (81)	2008		38% Genital (both high and low risk). 22% cutaneous types	PCR	81	(48% HPV ⁺)
Manderwad et al (82)	2009	0%	-	PCR/ISH-CARD	57	-
Guthoff <i>et al</i> (34)	2009	0%	-	PCR/IHC	31	No HIV patients
Ateenyi-Agaba et al (14)	2010		Mucosal HPV: 6,4% SCC; 7,7% dysplasia cutaneous HPV: 44,7% SCC; 41% dysplasia	PCR	94 SCC 39 dysplasia	Uncertain role of HIV
Asadi-Amoli et al (83)	2011	88%	No type found	Nested PCR	50	-
Chauhan et al (84)	2012	11%	16	PCR	64	-
Woods et al (85)	2013	6.5%	16	Nested PCR	50	-
Afrogheh et al (86)	2016	30%	16	IHC, ISH, PCR	43	?

HPV, human papillomavirus; OSSN, ocular surface squamous neoplasia; ISH, in situ hybridization; CARD, catalyzed reporter deposition; IHC, immunohistochemistry; SCC, squamous cell carcinoma.

exfoliative cytology as a valuable detection method for HPV in ophthalmic pterygium, while others have used exfoliative cytology for OSSN and conjunctival papilloma (94). The use of an easy-applicable, reliable and cost effective method may offer a more detailed investigation of the role of HPV in ocular surface, that may permit the use of topical antiviral treatment in HPV related ocular surface diseases. In fact, two recent studies (95,96) refer the efficient topical use of Cidofovir in OSSN. Cidofovir is a nucleoside analog with activity against a broad spectrum of DNA viruses. It has been used efficiently in squamous papilloma of the oropharynx, condylomata acuminata, molluscum contagiosum, and Kaposi's sarcoma (97,98). Finally, it would be of interest to evaluate the potential effect of HPV vaccination on the prevalence of these diseases in the future.

To conclude, HPV infection seems to play an important role in several aspects of ocular surface disease. Further research is required to elucidate the specific pathogenetic mechanisms of HPV in various ocular surface disease entities and findings may be clinically important in view of the potential development of targeted therapies or preventive measures, such as HPV vaccines.

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Authors' contributions

AKC was involved in the design of the study, and in the acquisition of data, drafting and writing of the manuscript. GB was involved in the acquisition and analysis of the data from studies to be included in this review, and in the drafting and writing of the manuscript. ETD and DAS were involved in the conception of the study, and in the revision of the manuscript. All authors have read and approved the final version.

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Not applicable.

Patient consent for publication

Not applicable.

Competing interests

DAS is the Editor-in-Chief for the journal, but had no personal involvement in the reviewing process, or any influence in terms of adjudicating on the final decision, for this article. The other authors declare that they have no competing interests.

References

- Bottalico D, Chen Z, Dunne A, Ostoloza J, McKinney S, Sun C, Schlecht NF, Fatahzadeh M, Herrero R, Schiffman M, *et al*: The oral cavity contains abundant known and novel human papillomaviruses from the *Betapapillomavirus* and *Gammapapillomavirus* genera. J Infect Dis 204: 787-792, 2011.
- Köhler A, Gottschling M, Manning K, Lehmann MD, Schulz E, Krüger-Corcoran D, Stockfleth E and Nindl I: Genomic characterization of ten novel cutaneous human papillomaviruses from keratotic lesions of immunosuppressed patients. J Gen Virol 92: 1585-1594, 2011.
- Duensing S and Münger K: Mechanisms of genomic instability in human cancer: Insights from studies with human papillomavirus oncoproteins. Int J Cancer 109: 157-162, 2004.
- Cubie HA: Diseases associated with human papillomavirus infection. Virology 445: 21-34, 2013.
 Mistry N, Wibom C and Evander M: Cutaneous and mucosal
- Mistry N, Wibom C and Evander M: Cutaneous and mucosal human papillomaviruses differ in net surface charge, potential impact on tropism. Virol J 5: 118, 2008.
- 6. Graham SV: Human papillomavirus: Gene expression, regulation and prospects for novel diagnostic methods and antiviral therapies. Future Microbiol 5: 1493-1506, 2010.
- 7. Zheng ZM and Baker CC: Papillomavirus genome structure, expression, and post-transcriptional regulation. Front Biosci 11: 2286-2302, 2006.
- Münger K, Baldwin A, Edwards KM, Hayakawa H, Nguyen CL, Owens M, Grace M and Huh K: Mechanisms of human papillomavirus-induced oncogenesis. J Virol 78: 11451-11460, 2004.
- Nair S and Pillai MR: Human papillomavirus and disease mechanisms: Relevance to oral and cervical cancers. Oral Dis 11: 350-359, 2005.

- Ljubojevic S and Skerlev M: HPV-associated diseases. Clin Dermatol 32: 227-234, 2014.
- 11. Gélinas JF, Manoukian J and Côté A: Lung involvement in juvenile onset recurrent respiratory papillomatosis: A systematic review of the literature. Int J Pediatr Otorhinolaryngol 72: 433-452, 2008.
- Verma V, Shen D, Sieving PC and Chan CC: The role of infectious agents in the etiology of ocular adnexal neoplasia. Surv Ophthalmol 53: 312-331, 2008.
- Sjö NC, von Buchwald C, Prause JU, Norrild B, Vinding T and Heegaard S: Human papillomavirus and pterygium. Is the virus a risk factor? Br J Ophthalmol 91: 1016-1018, 2007.
- 14. Ateenyi-Agaba C, Franceschi S, Wabwire-Mangen F, Arslan A, Othieno E, Binta-Kahwa J, van Doorn LJ, Kleter B, Quint W and Weiderpass E: Human papillomavirus infection and squamous cell carcinoma of the conjunctiva. Br J Cancer 102: 262-267, 2010.
- Hilgers JH: Pterygium: Its incidence, heredity and etiology. Am J Ophthalmol 50: 635-644, 1960.
- Zhou WP, Zhu YF, Zhang B, Qiu WY and Yao YF: The role of ultraviolet radiation in the pathogenesis of pterygia (Review). Mol Med Rep 14: 3-15, 2016.
- Cárdenas-Cantú E, Zavala J, Valenzuela J and Valdez-García JE: Molecular basis of pterygium development. Semin Ophthalmol 31: 567-583, 2016.
- Maxia C, Perra MT, Demurtas P, Minerba L, Murtas D, Piras F, Corbu A, Gotuzzo DC, Cabrera RG, Ribatti D, *et al*: Expression of survivin protein in pterygium and relationship with oxidative DNA damage. J Cell Mol Med 12: 2372-2380, 2008.
- Detorakis ĚT, Drakonaki EE and Spandidos DA: Molecular genetic alterations and viral presence in ophthalmic pterygium. Int J Mol Med 6: 35-41, 2000.
 McDonnell JM, McDonnell PJ and Sun YY: Human papil-
- McDonnell JM, McDonnell PJ and Sun YY: Human papillomavirus DNA in tissues and ocular surface swabs of patients with conjunctival epithelial neoplasia. Invest Ophthalmol Vis Sci 33: 184-189, 1992.
- 21. Varinli S, Varinli I, Köksal Erkisi M and Doran F: Human papilomavirus in pterygium. Cent Afr J Med 40: 24-26, 1994.
- Dushku N, Hatcher SL, Albert DM and Reid TW: p53 expression and relation to human papillomavirus infection in pingueculae, pterygia, and limbal tumors. Arch Ophthalmol 117: 1593-1599, 1999.
- Detorakis ET, Sourvinos G and Spandidos DA: Detection of herpes simplex virus and human papilloma virus in ophthalmic pterygium. Cornea 20: 164-167, 2001.
- 24. Gallagher MJ, Giannoudis A, Herrington CS and Hiscott P: Human papillomavirus in pterygium. Br J Ophthalmol 85: 782-784, 2001.
- 25. Piras F, Moore PS, Ugalde J, Perra MT, Scarpa A and Sirigu P: Detection of human papillomavirus DNA in pterygia from different geographical regions. Br J Ophthalmol 87: 864-866, 2003.
- Chen KH, Hsu WM, Cheng CC and Li YS: Lack of human papillomavirus in pterygium of Chinese patients from Taiwan. Br J Ophthalmol 87: 1046-1048, 2003.
- 27. Ateenyi-Agaba C, Weiderpass E, Smet A, Dong W, Dai M, Kahwa B, Wabinga H, Katongole-Mbidde E, Franceschi S and Tommasino M: Epidermodysplasia verruciformis human papillomavirus types and carcinoma of the conjunctiva: A pilot study. Br J Cancer 90: 1777-1779, 2004.
- Br J Cancer 90: 1777-1779, 2004.
 28. Schellini SA, Hoyama E, Shiratori CA, Sakamoto RH and Candeias JM: Lack of papillomavirus (HPV) in pterygia of a Brazilian sample. Arg Bras Oftalmol 69: 519-521, 2006.
- Brazilian sample. Arq Bras Oftalmol 69: 519-521, 2006.
 29. Kuo KT, Chang HC, Hsiao CH and Lin MC: Increased Ki-67 proliferative index and absence of P16INK4 in CIN-HPV related pathogenic pathways different from cervical squamous intraepithelial lesion. Br J Ophthalmol 90: 894-899, 2006.
- Takamura Y, Kubo E, Tsuzuki S and Akagi Y: Detection of human papillomavirus in pterygium and conjunctival papilloma by hybrid capture II and PCR assays. Eye (Lond) 22: 1442-1445, 2008.
- Rodrigues FW, Arruda JT, Silva RE and Moura KK: TP53 gene expression, codon 72 polymorphism and human papillomavirus DNA associated with pterygium. Genet Mol Res 7: 1251-1258, 2008.
- 32. Otlu B, Emre S, Turkcuoglu P, Doganay S and Durmaz R: Investigation of human papillomavirus and Epstein-Barr virus DNAs in pterygium tissue. Eur J Ophthalmol 19: 175-179, 2009.
- DNAs in pterygium tissue. Eur J Ophthalmol 19: 175-179, 2009.
 33. Tsai YY, Chang CC, Chiang CC, Yeh KT, Chen PL, Chang CH, Chou MC, Lee H and Cheng YW: HPV infection and p53 inactivation in pterygium. Mol Vis 15: 1092-1097, 2009.

- 34. Guthoff R, Marx A and Stroebel P: No evidence for a pathogenic role of human papillomavirus infection in ocular surface squamous neoplasia in Germany. Curr Eye Res 34: 666-671, 2009.
- Piecyk-Sidor M, Polz-Dacewicz M, Zagórski Z and Zarnowski T: Occurrence of human papillomavirus in pterygia. Acta Ophthalmol 87: 890-895, 2009.
- Hsiao CH, Lee BH, Ngan KW, Chuang WY, Yeung L, Yeh LK, Tan HY, Hui-Kang D and Lin KK: Presence of human papillomavirus in pterygium in Taiwan. Cornea 29: 123-127, 2010.
 Chong PP, Tung CH, Rahman NA, Yajima M, Chin FW, Yeng CL,
- 37. Chong PP, Tung CH, Rahman NA, Yajima M, Chin FW, Yeng CL, Go ES, Chan CM, Yawata N and Yamamoto N: Prevalence and viral load of oncogenic human papillomavirus (HPV) in pterygia in multi-ethnic patients in the Malay Peninsula. Acta Ophthalmol 92: e569-e579, 2014.
- Hamed-Azzam S, Edison N, Briscoe D, Mukari A and Elmalah I: Identification of human papillomavirus in pterygium. Acta Ophthalmol 94: e195-e197, 2016.
- 39. Chalkia AK, Derdas S, Bontzos G, Sourvinos G and Detorakis ET: Non-invasive detection of HPV DNA in exfoliative samples from ophthalmic pterygium: A feasibility study. Graefes Arch Clin Exp Ophthalmol 256: 193-198, 2018.
- 40. Di Girolamo N: Association of human papilloma virus with pterygia and ocular-surface squamous neoplasia. Eye (Lond) 26: 202-211, 2012.
- 41. Abreu AL, Souza RP, Gimenes F and Consolaro ME: A review of methods for detect human *Papillomavirus* infection. Virol J 9: 262, 2012.
- Mackay IM: Real-time PCR in the microbiology laboratory. Clin Microbiol Infect 10: 190-212, 2004.
- 43. Hirst LW: The treatment of pterygium. Surv Ophthalmol 48: 145-180, 2003.
- Todani A and Melki SA: Pterygium: Current concepts in pathogenesis and treatment. Int Ophthalmol Clin 49: 21-30, 2009.
- 45. Šjö N, Heegaard S and Prause JU: Conjunctival papilloma. A histopathologically based retrospective study. Acta Ophthalmol Scand 78: 663-666, 2000.
- Elsas FJ and Green WR: Epibulbar tumors in childhood. Am J Ophthalmol 79: 1001-1007, 1975.
- Lass JH, Jenson AB, Papale JJ and Albert DM: Papillomavirus in human conjunctival papillomas. Am J Ophthalmol 95: 364-368, 1983.
- Naghashfar Z, McDonnell PJ, McDonnell JM, Green WR and Shah KV: Genital tract papillomavirus type 6 in recurrent conjunctival papilloma. Arch Ophthalmol 104: 1814-1815, 1986.
- 49. McDonnell PJ, McDonnell JM, Kessis T, Green WR and Shah KV: Detection of human papillomavirus type 6/11 DNA in conjunctival papillomas by in situ hybridization with radioactive probes. Hum Pathol 18: 1115-1119, 1987.
- 50. Mäntyjärvi M, Syrjänen S, Kaipiainen S, Mäntyjärvi R, Kahlos T and Syrjänen K: Detection of human papillomavirus type 11 DNA in a conjunctival squamous cell papilloma by in situ hybridization with biotinylated probes. Acta Ophthalmol (Copenh) 67: 425-429, 1989.
- 51. Fierlbeck G, Rassner G, Thiel HJ and Pfister H: Virus-induced papilloma of the conjunctiva. Detection of HPV 6a DNA. Z Hautkr 65: 497-499, 1990 (In German).
- 52. Mincione GP, Taddei GL, Wolovsky M, Calzolari A and Mincione F: Detection of human papillomavirus (HPV) DNA type 6/11 in a conjunctival papilloma by in situ hybridization with biotinylated probes. Pathologica 84: 483-488, 1992.
- 53. Saegusa M, Takano Y, Hashimura M, Okayasu I and Shiga J: HPV type 16 in conjunctival and junctional papilloma, dysplasia, and squamous cell carcinoma. J Clin Pathol 48: 1106-1110, 1995.
- 54. Michel JL, Guiguen Y, Léger F, Gain P, Valanconny C and Cambazard F: Human papillomavirus 6/11 in conjunctival papilloma. Ann Dermatol Venereol 123: 90-92, 1996 (In French).
- 55. Nakamura Y, Mashima Y, Kameyama K, Mukai M and Oguchi Ý: Detection of human papillomavirus infection in squamous tumours of the conjunctiva and lacrimal sac by immunohistochemistry, in situ hybridisation, and polymerase chain reaction. Br J Ophthalmol 81: 308-313, 1997.
- 56. Assadoullina A, Bialasiewicz AA, de Villiers EM and Richard G: Detection of HPV-20, HPV-23, and HPV-DL332 in a solitary eyelid syringoma. Am J Ophthalmol 129: 99-101, 2000.
- 57. Sjö NC, Heegaard S, Prause JU, von Buchwald C and Lindeberg H: Human papillomavirus in conjunctival papilloma. Br J Ophthalmol 85: 785-787, 2001.

- Minchiotti S, Masucci L, Serapiao Dos Santos M, Perrella E, Graffeo R, Lambiase A and Bonini S: Conjunctival papilloma and human papillomavirus: Identification of HPV types by PCR. Eur J Ophthalmol 16: 473-477, 2006.
 Sjö NC, von Buchwald C, Cassonnet P, Norrild B, Prause JU,
- 59. Sjö NC, von Buchwald C, Cassonnet P, Norrild B, Prause JU, Vinding T and Heegaard S: Human papillomavirus in normal conjunctival tissue and in conjunctival papilloma: Types and frequencies in a large series. Br J Ophthalmol 91: 1014-1015, 2007.
- 60. Annadanam A, Vizcaino MA, Eberhart CG, Khurshid GS and Gupta P: Long standing exophytic conjunctival papilloma infected with human papillomavirus. J Eye Ophthalmol 4: 1, 2017.
- Egbert JE and Kersten RC: Female genital tract papillomavirus in conjunctival papillomas of infancy. Am J Ophthalmol 123: 551-552, 1997.
- 62. Kalogeropoulos C, Koumpoulis I, Papadiotis E, Zioga A, Gkrepi K, Pappa C, Paschides C, Malamou-Mitsi V and Aspiotis M: Squamous cell papilloma of the conjunctiva due to human papillomavirus (HPV): Presentation of two cases and review of literature. Clin Ophthalmol 6: 1553-1561, 2012.
- 63. Shields CL and Shields JA: Tumors of the conjunctiva and cornea. Surv Ophthalmol 49: 3-24, 2004.
- Lee GA and Hirst LW: Ocular surface squamous neoplasia. Surv Ophthalmol 39: 429-450, 1995.
- Lee GA and Hirst LW: Retrospective study of ocular surface squamous neoplasia. Aust N Z J Ophthalmol 25: 269-276, 1997.
- 66. Gichuhi S, Sagoo MS, Weiss HA and Burton MJ: Epidemiology of ocular surface squamous neoplasia in Africa. Trop Med Int Health 18: 1424-1443, 2013.
- 67. Ahmed H, Hassan RY and Pindiga UH: Xeroderma pigmentosum in three consecutive siblings of a Nigerian family: Observations on oculocutaneous manifestations in black African children. Br J Ophthalmol 85: 110-111, 2001.
- Ogun GO, Ogun OA, Bekibele CO and Akang EE: Intraepithelial and invasive squamous neoplasms of the conjunctiva in Ibadan, Nigeria: A clinicopathological study of 46 cases. Int Ophthalmol 29: 401-409, 2009.
- 69. McKelvie PA, Daniell M, McNab A, Loughnan M and Santamaria JD: Squamous cell carcinoma of the conjunctiva: A series of 26 cases. Br J Ophthalmol 86: 168-173, 2002.
- Lauer SA, Malter JS and Meier JR: Human papillomavirus type 18 in conjunctival intraepithelial neoplasia. Am J Ophthalmol 110: 23-27, 1990.
- 71. Tuppurainen K, Raninen A, Kosunen O, Kankkunen JP, Kellokoski J, Syrjänen S, Mäntyjärvi M and Syrjänen K: Squamous cell carcinoma of the conjunctiva. Failure to demonstrate HPV DNA by in situ hybridization and polymerase chain reaction. Acta Ophthalmol (Copenh) 70: 248-254, 1992.
- 72. Tabrizi SN, McCurrach FE, Drewe RH, Borg AJ, Garland SM and Taylor HR: Human papillomavirus in corneal and conjunctival carcinoma. Aust N Z J Ophthalmol 25: 211-215, 1997.
- Karcioglu ZA and Issa TM: Human papilloma virus in neoplastic and non-neoplastic conditions of the external eye. Br J Ophthalmol 81: 595-598, 1997.
- 74. Toth J, Karcioglu ZA, Moshfeghi AA, Issa TM, Al-Ma'ani JR and Patel KV: The relationship between human papillomavirus and p53 gene in conjunctival squamous cell carcinoma. Cornea 19: 159-162, 2000.
- 75. Scott IU, Karp CL and Nuovo GJ: Human papillomavirus 16 and 18 expression in conjunctival intraepithelial neoplasia. Ophthalmology 109: 542-547, 2002.
- 76. Eng HL, Lin TM, Chen SY, Wu SM and Chen WJ: Failure to detect human papillomavirus DNA in malignant epithelial neoplasms of conjunctiva by polymerase chain reaction. Am J Clin Pathol 117: 429-436, 2002.
- 77. Tulvatana W, Bhattarakosol P, Sansopha L, Sipiyarak W, Kowitdamrong E, Paisuntornsug T and Karnsawai S: Risk factors for conjunctival squamous cell neoplasia: A matched case-control study. Br J Ophthalmol 87: 396-398, 2003.
- 78. Moubayed P, Mwakyoma H and Schneider DT: High frequency of human papillomavirus 6/11, 16, and 18 infections in precancerous lesions and squamous cell carcinoma of the conjunctiva in subtropical Tanzania. Am J Clin Pathol 122: 938-943, 2004.
- 79. Tornesello ML, Duraturo ML, Waddell KM, Biryahwaho B, Downing R, Balinandi S, Lucas SB, Buonaguro L and Buonaguro FM: Evaluating the role of human papillomaviruses in conjunctival neoplasia. Br J Cancer 94: 446-449, 2006.
- Sen S, Sharma A and Panda A: Immunohistochemical localization of human papilloma virus in conjunctival neoplasias: A retrospective study. Indian J Ophthalmol 55: 361-363, 2007.

- 81. de Koning MN, Waddell K, Magyezi J, Purdie K, Proby C, Harwood C, Lucas S, Downing R, Quint WG and Newton R: Genital and cutaneous human papillomavirus (HPV) types in relation to conjunctival squamous cell neoplasia: A case-control study in Uganda. Infect Agent Cancer 3: 12, 2008.
- Manderwad GP, Kannabiran C, Honavar SG and Vemuganti GK: Lack of association of high-risk human papillomavirus in ocular surface squamous neoplasia in India. Arch Pathol Lab Med 133: 1246-1250, 2009.
- 83. Asadi-Amoli F, Heidari AB, Jahanzad I and Jabbarvand M: Detection of human papillomavirus in squamous cell carcinoma of conjunctiva by nested PCR: A case control study in Iran. Acta Med Iran 49: 707-714, 2011.
- 84. Chauhan S, Sen S, Sharma A, Dar L, Kashyap S, Kumar P, Bajaj MS and Tandon R: Human papillomavirus: A predictor of better survival in ocular surface squamous neoplasia patients. Br J Ophthalmol 96: 1517-1521, 2012.
- 85. Woods M, Chow S, Heng B, Glenn W, Whitaker N, Waring D, Iwasenko J, Rawlinson W, Coroneo MT, Wakefield D, *et al*: Detecting human papillomavirus in ocular surface diseases. Invest Ophthalmol Vis Sci 54: 8069-8078, 2013.
- 86. Afrogheh AH, Jakobiec FA, Hammon R, Grossniklaus HE, Rocco J, Lindeman NI, Sadow PM and Faquin WC: Evaluation for high-risk HPV in squamous cell carcinomas and precursor lesions arising in the conjunctiva and lacrimal sac. Am J Surg Pathol 40: 519-528, 2016.
- Yin VT, Merritt HA, Sniegowski M and Esmaeli B: Eyelid and ocular surface carcinoma: Diagnosis and management. Clin Dermatol 33: 159-169, 2015.
- Hoffmann M, Ihloff AS, Görögh T, Weise JB, Fazel A, Krams M, Rittgen W, Schwarz E and Kahn T: p16(INK4a) overexpression predicts translational active human papillomavirus infection in tonsillar cancer. Int J Cancer 127: 1595-1602, 2010.
- 89. Quiroga-Garza G, Zhou H, Mody DR, Schwartz MR and Ge Y: Unexpected high prevalence of HPV 90 infection in an underserved population: Is it really a low-risk genotype? Arch Pathol Lab Med 137: 1569-1573, 2013.

- 90. Koss LG: Cytologic and histologic manifestations of human papillomavirus infection of the uterine cervix. Cancer Detect Prev 14: 461-464, 1990.
- Rosai J: Ackerman's Surgical Pathology. Rosai J (ed). Mosby, St. Louis, p1358, 1996.
- 92. Joanna R, Renata Z, Witold P, Małgorzata S, Bernaczyk P and Chyczewski L: The evaluation of human papillomavirus and p53 gene mutation in benign and malignant conjunctiva and eyelid lesions. Folia Histochem Cytobiol 48: 530-533, 2010.
- Bernard HU: The clinical importance of the nomenclature, evolution and taxonomy of human papillomaviruses. J Clin Virol 32 (Suppl 1): S1-S6, 2005.
- 94. Kayat KV, Correa Dantas PE, Felberg S, Galvão MA and Saieg MA: Exfoliative cytology in the diagnosis of ocular surface squamous neoplasms. Cornea 36: 127-130, 2017.
- 95. Ip MH and Coroneo MT: Treatment of previously refractory ocular surface squamous neoplasia with topical cidofovir. JAMA Ophthalmol 135: 500-502, 2017.
- 96. Ip MH, Robert George CR, Naing Z, Perlman EM, Rawlinson W and Coroneo MT: Topical cidofovir for treatment-refractory ocular surface squamous neoplasia. Ophthalmology 125: 617-619, 2018.
- 97. Bielamowicz S, Villagomez V, Stager SV and Wilson WR: Intralesional cidofovir therapy for laryngeal papilloma in an adult cohort. Laryngoscope 112: 696-699, 2002.
- 98. Little RF, Merced-Galindez F, Staskus K, Whitby D, Aoki Y, Humphrey R, Pluda JM, Marshall V, Walters M, Welles L, et al: A pilot study of cidofovir in patients with kaposi sarcoma. J Infect Dis 187: 149-153, 2003.
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