

MITOGENOME ANNOUNCEMENT



## The mitochondrial genome of *Telenomus remus* (Hymenoptera: Platygasteridae)

Xiao-fei Li<sup>a,b</sup>, Ze-kai Li<sup>a,b</sup>, Jia-chen Zhu<sup>a,b</sup>, Bo-ying Zheng<sup>a,b</sup>, Pu Tang<sup>a,c,d,b</sup> and Xue-xin Chen<sup>a,c,d,b</sup>

<sup>a</sup>State Key Lab of Rice Biology, Zhejiang University, Hangzhou, China; <sup>b</sup>College of Agriculture and Biotechnology, Institute of Insect Sciences, Zhejiang University, Hangzhou, China; <sup>c</sup>Ministry of Agriculture Key Lab of Molecular Biology of Crop Pathogens and Insects, Zhejiang University, Hangzhou, China; <sup>d</sup>Zhejiang Provincial Key Laboratory of Biology of Crop Pathogens and Insects, Zhejiang University, Hangzhou, China

### ABSTRACT

*Telenomus remus* Nixon, 1937 is an important parasitoid of lepidopterans. We sequenced the mitochondrial genome of *T. remus*, 15,500 bp in size, and possessed all 37 typical mitochondrial genes. A few tRNAs show gene arrangements compared with the ancestral gene order, mainly involving in the four tRNA clusters (*E-C-Y-Q-I-A*, *D-K*, *N-F-S1-R*, and *M-V*). The nucleotide sequences of 13 protein-coding genes of this sequence and another seven species from Platygasteridae were used for phylogenetic analysis by MrBayes, with two species from Cynipoidea as an outgroup. The topology demonstrated that *T. remus* was most closely related to *Telenomus* sp.

### ARTICLE HISTORY

Received 26 December 2020  
Accepted 26 January 2021

### KEYWORDS

Mitochondrial genome;  
*Telenomus remus*;  
Platygasteridae; phylogeny

*Telenomus remus* Nixon, 1937, an important egg parasitoid of various Lepidoptera species, belonging to Platygasteridae of Hymenoptera. It originated from peninsular Malaysia, thought to be an excellent biological control agent and was introduced against *Spodoptera* spp. to various parts of the world (Nixon 1937; Bennett et al. 1985). In this study, we got the mt-genome of *T. remus* by the next-generation sequencing for the first time. It will facilitate a deeper understanding and exploitation of this species.

The sample of *T. remus* was collected from Ningbo (121°04'E, 30°15'N), Zhejiang Province, China in July 2019. The voucher specimen was stored in 100% ethanol and kept in the Parasitic Hymenoptera Collection of Institute of Insect Sciences, Zhejiang University (ZJUH\_2020401, Pu Tang, [ptang@zju.edu.cn](mailto:ptang@zju.edu.cn)). The whole genomic DNA was extracted from one female adult specimen using DNeasy tissue kit (Qiagen, Hilden, Germany). The library was constructed by VAHTS™ Universal DNA Library Prep Kit for Illumina® v3, and sequenced by Illumina HiSeq sequencer (150 bp paired-end). The reads were filtered by local BLAST with *E* value  $1 \times 10^{-5}$  referencing to all apocritan mitochondrial genomes dataset (downloaded from the GenBank database on 10 April 2020), and was assembled by Celera Assembler (Myers et al. 2000) and IDBA (Peng et al. 2012). The two assemblies were combined by GENEIOUS (Biomatters Ltd., San Diego, CA) to obtain a nonredundant set.

The mitochondrial genome of *T. remus* is 15,500 bp in length (GenBank accession MT906647), containing 13 protein-coding genes (PCGs), 22 transfer RNA (tRNA) genes, and two ribosomal RNA (rRNA) genes, but part of the control

region (D-loop) failed to be sequenced or assembled. The overall base composition of the mitogenome was estimated to be A 44.6%, T 40.4%, C 8.8%, and G 6.3%, with a high A + T content of 85.0% and a little higher than that of *Telenomus* sp. (MF776884, 84.6%). All 13 PCGs of *T. remus* have the conventional ATN start codons for invertebrate mitochondrial PCGs (seven ATA, four ATT, and two ATG). All of the PCGs terminate with the stop codon TAA or TAG. The lengths of 16S rRNA and 12S rRNA in *T. remus* were 1289 and 770 bp, with the AT contents of 88.6% and 88.2%, respectively.

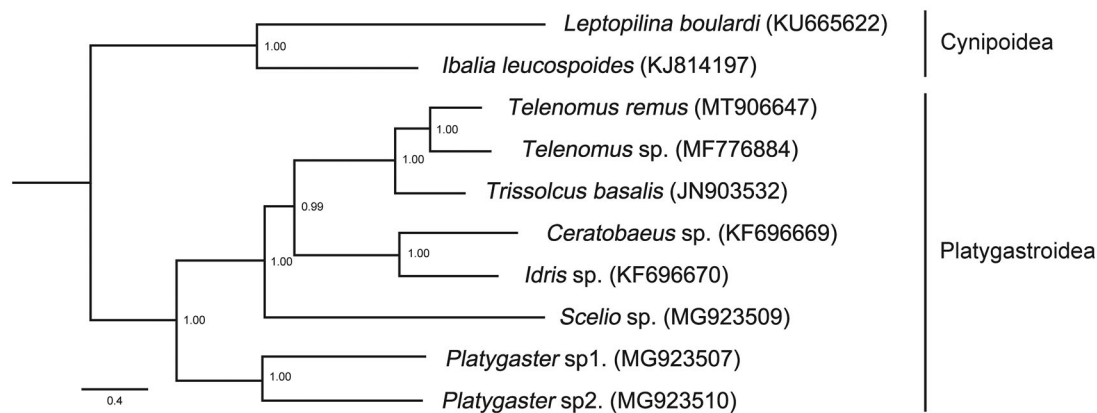
A few arrangements of tRNA genes compared with the ancestral gene order, where four main rearrangement events of tRNA clusters (*E-C-Y-Q-I-A*, *D-K*, *N-F-S1-R*, and *M-V*) were found in the sequenced region. All of these junctions are 'hot spots' where gene rearrangement occurs frequently in hymenopterans (Dowton and Austin 1999; Dowton et al. 2003; Wei et al. 2014). The arrangements are nearly identical to that of *Telenomus* sp., except for *E* moving from the cluster *N-E-F-S1-R* (*Telenomus* sp.) to *E-C-Y-Q-I-A* (*T. remus*).

To explore the phylogenetic position of *T. remus*, we used 13 PCGs of another seven mitochondrial genomes of Platygasteridae and two species of Cynipoidea as an outgroup to perform phylogenetic analysis. The nucleotide sequences were aligned by using MAFFT v7.271. PartitionFinder v1.1.1 (Lanfear et al. 2012) was used to identify best partition scheme and substitution models, and the phylogenetic tree was constructed by MrBayes v3.2.549 (Ronquist et al. 2012). Phylogenetic analysis showed that *T. remus* was most closely related to *Telenomus* sp. Besides, *Telenomus* is closely related

**CONTACT** Pu Tang ✉ [ptang@zju.edu.cn](mailto:ptang@zju.edu.cn) 📧 State Key Lab of Rice Biology, Zhejiang University, Hangzhou, China

© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



**Figure 1.** Phylogenetic tree of MrBayes using 13 PCGs in mitochondrial genomes of eight species in Platygastroidea with two species in Cynipoidea as outgroup. The numbers at the nodes are the Bayesian posterior probabilities.

to *Trissolcus*, which was consistent with other results from the analyses based on morphological characters and several sequences before (Figure 1) (Kononova 2008; Taekul et al. 2014; Shen et al. 2019).

## Disclosure statement

No potential conflict of interest was reported by the authors.

## Funding

This work was supported by the National Key R&D Program of China (2019YFD0300104, 2019YFD1002100), the Key R&D Program of Zhejiang Province (2020C02003, 2021C02045), the Public Welfare Technology Application Research Project of Zhejiang Province (LGN21C140008), the General Program of National Natural Science Foundation of China (32070467, 31702035), and the Fundamental Research Funds for the Central Universities.

## Data availability statement

The data that support the findings of this study are openly available in GenBank of NCBI at <https://www.ncbi.nlm.nih.gov>. Mitochondrial genome data accession number MT906647. All high-throughput sequencing data files are available from the GenBank Sequence Read Archive (SRA) accession number PRJNA691080.

## References

Bennett F, Hughes IW, Simmonds FG, Yaseen M. 1985. A review of biological control of pests in the commonwealth Caribbean and Bermuda up to 1982. Oxfordshire (UK): Commonwealth Agricultural Bureaux.

- Dowton M, Austin AD. 1999. Evolutionary dynamics of a mitochondrial rearrangement "hot spot" in the hymenoptera. *Mol Biol Evol.* 16(2): 298–309.
- Dowton M, Castro LR, Campbell SL, Bargon SD, Austin AD. 2003. Frequent mitochondrial gene rearrangements at the hymenopteran *nad3-nad5* junction. *J Mol Evol.* 56(5):517–526.
- Kononova SV. 2008. New species of Telenominae (Hymenoptera, Scelionidae) from the Palaearctic Fauna. *Entomol Rev.* 88(1):61–67.
- Lanfear R, Calcott B, Ho SY, Guindon S. 2012. Partitionfinder: combined selection of partitioning schemes and substitution models for phylogenetic analyses. *Mol Biol Evol.* 29(6):1695–1701.
- Myers EW, Sutton GG, Delcher AL, Dew IM, Fasulo DP, Flanigan MJ, Kravitz SA, Mobarry CM, Reinert KH, Remington KA, et al. 2000. A whole-genome assembly of *Drosophila*. *Science.* 287(5461):2196–2204.
- Nixon GEJ. 1937. Some Asiatic Telenominae (Hym., Proctotrupoidea). *Ann Mag Nat Hist.* 20(118):444–475.
- Peng Y, Leung HCM, Yiu SM, Chin FYL. 2012. IDBA-UD: a de novo assembler for single-cell and metagenomic sequencing data with highly uneven depth. *Bioinformatics.* 28(11):1420–1428.
- Ronquist F, Teslenko M, Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst Biol.* 61(3):539–542.
- Shen ZC, Chen L, Chen L, Li YX. 2019. Information from the mitochondrial genomes of two egg parasitoids, *Gonatocerus* sp. and *Telenomus* sp., reveals a controversial phylogenetic relationship between Mymaridae and Scelionidae. *Genomics.* 111(5):1059–1065.
- Taekul C, Valerio AA, Austin A, Klompen H, Johnson NF. 2014. Molecular phylogeny of telenomine egg parasitoids (Hymenoptera: Platygastriidae s.l.: Telenominae): evolution of host shifts and implications for classification. *Syst Entomol.* 39(1):24–35.
- Wei SJ, Li Q, van Achterberg K, Chen XX. 2014. Two mitochondrial genomes from the families Bethyilidae and Mutillidae: independent rearrangement of protein-coding genes and higher-level phylogeny of the Hymenoptera. *Mol Phylogenet Evol.* 77:1–10.