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Fatal COVID-19 infections: Is NK cell dysfunction a link with autoimmune HLH?

Dear Editor.

Shoenfeld [1] postulates that in COVID-19 infected patients a clinical constellation of cytokine storm, respiratory failure and eventually death is reminiscent of a "hyperferritinemic syndrome" a condition that resembles a hemophagocytic lymphohistiocytosis (HLH)-like syndrome. One of the most prominent immunological features of patients with primary or infection associated HLH (also referred to as macrophage activation syndrome) is the loss of natural killer (NK) cell effector functions in most types of HLH [2–4].

NK cells, unlike T lymphocytes, are governed by the balance of activating and inhibitory receptors, and as initially described by Kärre [5], the loss of inhibitory signals potentiate NK cell effector functions such as cytotoxic capacity (which require perforin (PERF) and granzyme B) and the release of interferon gamma (IFN_γ) [6]. A functional NK cell population is not only important in the direct response to the virus through the elimination of virally infected cells, but it is also critical in limiting the systemic inflammatory response by killing activated inflammatory dendritic cells, monocytes, and T cells [7,8] which are the primary drivers of this "hyperferritinemic syndrome" when other immunomodulatory mechanisms such as those mediated by tolerogenic dendritic cells are dysfunctional [9]. The importance of NK cell cytotoxic functions is highlighted in both infection-related HLH and HLH-related to inflammatory diseases. For instance, patients with influenza and systemic juvenile idiopathic arthritis that developed severe HLH more frequently were heterozygote carriers for mutations in genes implicated in NK cell cytotoxicity (e.g. PERF and LYST (lysosomal trafficking regulator) [10,11]. Similarly, NK cells may develop decreased cytotoxicity in the presence of circulating pathogenic autoantibodies targeting surface inhibitory receptors [12].

The inflammatory microenvironment may shift the balance to reduce NK cell effector functions in both COVID-19 and inflammatory forms of secondary HLH. For instance, during an acute viral infection affecting the lungs, the microenvironment becomes relatively hypoxic. Hypoxia may result in diminished cytolysis, although it does not impact antibody dependent cellular cytotoxicity (ADCC) by NK cells [13]. Also, elevated IL-6 and IL-10 levels, as observed in SARS-CoV2-infected patients [14], have the capacity to directly reduce NK cell cytotoxicity and the expression of PERF and granzyme B [15,16]. IL-6 may also reduce the expression of NKG2D, which is important in killing virally infected ones [17]. Alternatively, as it has been shown that SARS-CoV-2 binds to ACE2 [18], COVID-19 may infect NK cells to suppress their functions, as NK cells express angiotensin converting enzyme 2 (ACE2) [19]. Although not published in COVID-19, other RNA viruses that cause acute pulmonary infections such as influenza A, promote NK cell apoptosis and reduce their cytotoxicity following their infection [20,21]. With all of these mechanisms that tip the balance to favour suppression of NK cell functions (as summarized in Fig. 1), and thereby potentiate HLH, are there available therapies which target mechanisms that can help restore this balance in patients with SARS-Cov-2 infections?

As suggested by Shoenfeld [1], intravenous immunoglobulin (IVIG), a relatively safe non-immunosuppressive intervention, may help restore NK cell functions – although there have been no studies to date that have assessed the effects of IVIG on NK cell functions in the setting of an acute viral infection. IVIG (2 g/kg ideal body weight) may help improve outcomes in patients with COVID-19 [22]. In a pre-clinical model for graft-vs-host disease, IVIG promotes the expansion of functional NK cells [23]. In patients with Kawasaki's disease, who also develop a cytokine storm, treatment with IVIG increases NKG2D expression which may promote their cytolytic functions [17]. It may also reduce the re-

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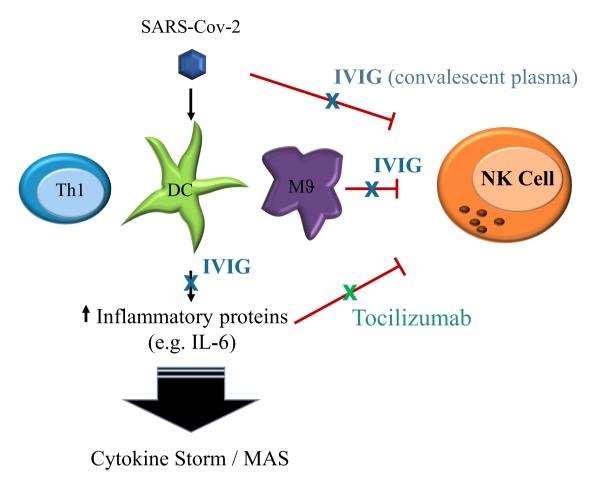


Fig. 1. SARS-CoV-2 and the subsequent immune cell inflammatory responses suppress NK cell cytotoxicity which promotes a severe cytokine release syndrome, and inadequate termination of immune responses. Intravenous immunoglobulin (IVIG) promotes NK cell cytotoxicity by promoting ADCC and subsequent dendritic cell apoptosis, reducing circulating viral particles (convalescent plasma), and by reducing inflammatory cytokine levels. Tocilizumab reduces IL-6-dependent suppression of NK cells.

lease of IL-6 by activated inflammatory cells [24]. Moreover, IVIG may directly promote NK cell-mediated termination of inflammatory responses by promoting NK-cell mediated ADCC of activated dendritic cells [25]. A different form of IVIG which may be most beneficial when given prophylactically, particularly to patients most at risk for complications is convalescent serum, which may reduce circulating viral particles and possibly promote NK cell ADCC towards virally infected cells (26–28). Other therapies that are currently used in patients with HLH due to COVID-19 include anti-IL6 receptor monoclonal antibodies (29) Anti-IL6R may increase NK proliferation and cytotoxicity [26]. Future studies defining the effects of IVIG and possibly anti-IL6R on NK cells in patients that are infected with COVID-19 may provide further insight if and how they may limit the cytokine storm that is observed in severe cases.

References

- Shoenfeld Y. Corona (COVID-19) Time musings our involvement in COVID-19 pathogenesis, diagnosis, treatment and vaccine planning. Autoimmunity Reviews 102538. elsevier; 2020. https://doi.org/10.1016/j.autrev.2020.102538. in press.
- [2] Vandenhaute J, Wouters CH, Matthys P. Natural killer cells in systemic autoinflammatory diseases: a focus on systemic juvenile idiopathic arthritis and macrophage activation syndrome. Front Immunol 2019;10:3089.
- [3] Crayne CB, Albeituni S, Nichols KE, Cron RQ. The immunology of macrophage activation syndrome. Front Immunol 2019;10:119.
- [4] Terrell CE, Jordan MB. Perforin deficiency impairs a critical immunoregulatory loop involving murine CD8(+) T cells and dendritic cells. Blood 2013;121(26):5184–91.
- [5] Karre K. NK cells, MHC class I molecules and the missing self. Scand J Immunol 2002;55(3):221–8.
- [6] Gianchecchi E, Delfino DV, Fierabracci A. NK cells in autoimmune diseases: linking innate and adaptive immune responses. Autoimmun Rev 2018;17(2):142–54.

- [7] Marcenaro E, Ferranti B, Moretta A. NK-DC interaction: on the usefulness of autoaggression. Autoimmun Rev 2005;4(8):520–5.
- [8] Vitale M, Cantoni C, Della Chiesa M, Ferlazzo G, Carlomagno S, Pende D, et al. An historical overview: the discovery of how NK cells can kill enemies, recruit defense troops, and more. Front Immunol 2019;10:1415.
- [9] Toubi E, Vadasz Z. Innate immune-responses and their role in driving autoimmunity. Autoimmun Rev 2019;18(3):306–11.
- [10] Schulert GS, Zhang M, Fall N, Husami A, Kissell D, Hanosh A, et al. Whole-exome sequencing reveals mutations in genes linked to hemophagocytic lymphohistiocytosis and macrophage activation syndrome in fatal cases of H1N1 influenza. J Infect Dis 2016;213(7):1180–8.
- [11] Vastert SJ, van Wijk R, D'Urbano LE, de Vooght KM, de Jager W, Ravelli A, et al. Mutations in the perforin gene can be linked to macrophage activation syndrome in patients with systemic onset juvenile idiopathic arthritis. Rheumatology (Oxford) 2010;49(3):441–9.
- [12] Segerberg F, Lundtoft C, Reid S, Hjorton K, Leonard D, Nordmark G, et al. Autoantibodies to killer cell immunoglobulin-like receptors in patients with systemic lupus erythematosus induce natural killer cell hyporesponsiveness. Front Immunol 2019;10:2164.
- [13] Balsamo M, Manzini C, Pietra G, Raggi F, Blengio F, Mingari MC, et al. Hypoxia downregulates the expression of activating receptors involved in NK-cell-mediated target cell killing without affecting ADCC. Eur J Immunol 2013;43(10):2756–64.
- [14] Pedersen SF, Ho Y. SARS-CoV-2: a storm is raging. J Clin Invest 2020;130(5):2202–5.
- [15] Cifaldi L, Prencipe G, Caiello I, Bracaglia C, Locatelli F, De Benedetti F, et al. Inhibition of natural killer cell cytotoxicity by interleukin-6: implications for the pathogenesis of macrophage activation syndrome. Arthritis Rheumatol 2015;67(11):3037–46.
- [16] Lassen MG, Lukens JR, Dolina JS, Brown MG, Hahn YS. Intrahepatic IL-10 maintains NKG2A + Ly49- liver NK cells in a functionally hyporesponsive state. J Immunol 2010;184(5):2693–701.
- [17] Ge X, Li CR, Yang J, Wang GB. Aberrantly decreased levels of NKG2D expression in children with Kawasaki disease. Scand J Immunol 2013;77(5):389–97.
- [18] Rivellese F, Prediletto E. ACE2 at the centre of COVID-19 from paucisymptomatic infections to severe pneumonia. Autoimmunity Reviews 2020. https://doi.org/10. 1016/j.autrev.2020.102536. in press.

- [19] Yanyan Zhu MJ, Gao Liang, Huang Xiaoyun. Single Cell Analysis of ACE2 Expression Reveals the Potential Targets for 2019-nCoV Preprints 2020, 2020020221 (doi: 1020944/preprints2020020221v1). 2020.
- [20] Mao H, Tu W, Liu Y, Qin G, Zheng J, Chan PL, et al. Inhibition of human natural killer cell activity by influenza virions and hemagglutinin. J Virol 2010;84(9):4148–57.
- [21] Wong CK, Lam CW, Wu AK, Ip WK, Lee NL, Chan IH, et al. Plasma inflammatory cytokines and chemokines in severe acute respiratory syndrome. Clin Exp Immunol 2004;136(1):95–103.
- [22] Wei Cao XL, Bai Tao, Fan Hongwei, Hong Ke, Song Hui, Han Yang, et al. High-dose intravenous immunoglobulin as a therapeutic option for deteriorating patients with coronavirus disease 2019. Open Forum Infect Dis 2020;7(3):1–6.
- [23] Gregoire-Gauthier J, Fontaine F, Benchimol L, Nicoletti S, Selleri S, Dieng MM, et al. Role of natural killer cells in intravenous immunoglobulin-induced graft-versus-host disease inhibition in NOD/LtSz-scidIL2rg(-/-) (NSG) mice. Biol Blood Marrow Transplant 2015;21(5):821–8.
- [24] Gupta M, Noel GJ, Schaefer M, Friedman D, Bussel J, Johann-Liang R. Cytokine modulation with immune gamma-globulin in peripheral blood of normal children

and its implications in Kawasaki disease treatment. J Clin Immunol 2001;21(3):193-9.

- [25] Tha-In T, Metselaar HJ, Tilanus HW, Groothuismink ZM, Kuipers EJ, de Man RA, et al. Intravenous immunoglobulins suppress T-cell priming by modulating the bidirectional interaction between dendritic cells and natural killer cells. Blood 2007;110(9):3253–62.
- [26] Daien CI, Gailhac S, Audo R, Mura T, Hahne M, Combe B, et al. High levels of natural killer cells are associated with response to tocilizumab in patients with severe rheumatoid arthritis. Rheumatology (Oxford) 2015;54(4):601–8.

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