SSTR5-AS1 functions as a ceRNA to regulate CA2 by sponging miR-15b-5p for the development and prognosis of HBV-related hepatocellular carcinoma

JING XU[1], JING ZHANG[2], FENGLIAN SHAN[1,3], JIE WEN[3] and YUE WANG[1]

[1]Infectious Diseases Division; [2]Department of Gastroenterology; [3]Respiratory Medicine,
Affiliated Hospital of Jining Medical University, Jining, Shandong 272029, P.R. China

Abstract. Long non-coding RNAs (lncRNAs) have been implicated in the development and progression of cancer. However, the mechanisms of lncRNAs in hepatitis B virus (HBV) infection-induced hepatocellular carcinoma (HCC) remain unclear. The study aimed to reveal the roles of lncRNAs for HBV-HCC based on the hypothesis of competing endogenous RNA (ceRNA). The lncRNA (GSE27462), miRNA (GSE76903) and mRNA (GSE121248) expression profiles were collected from the Gene Expression Omnibus database. Differentially expressed lncRNAs (DELs), genes (DEGs) and miRNAs (DEMs) were identified using the LIMMA or EdgeR package, respectively. The ceRNA network was constructed based on interaction pairs between miRNAs and mRNAs/lncRNAs. The functions of DEGs in the ceRNA network were predicted using the DAVID database, which was overlapped with the known HCC pathways of Comparative Toxicogenomics Database (CTD) to construct the HCC-related ceRNA network. The prognosis values [overall survival, (OS); recurrence-free survival (RFS)] of genes were validated using the Cancer Genome Atlas (TCGA) data with Cox regression analysis. The present study screened 38 DELs, 127 DEMs and 721 DEGs. A ceRNA network was constructed among 17 DELs, 12 DEMs and 173 DEGs, including the FAM138B-hsa-miR-30c-CCNE2/RRM2 and SSTR5-AS1-hsa-miR-15b-5p-CA2 ceRNA axes. Function enrichment analysis revealed the genes in the ceRNA network that participated in the p53 signaling pathway [cyclin E2 (CCNE2), ribonucleotide reductase M2 subunit (RRM2)] and nitrogen metabolism [carbonic anhydrase 2 (CA2)], which were also included in the pathways of the CTD. Univariate Cox regression analysis revealed that six RNAs (2 DELs: FAM138B, SSTR5-AS1; 2 DEMs: hsa-miR-149, hsa-miR-7; 2 DEGs: CCNE2, RRM2) were significantly associated with OS; while seven RNAs (1 DEL: LINC00284; 3 DEMs: hsa-miR-7, hsa-miR-15b, hsa-miR-30c-2; and 3 DEGs: RRM2, CCNE2, CA2) were significantly associated with RFS. In conclusion, FAM138B-hsa-miR-30c-CCNE2/RRM2 and the SSTR5-AS1-hsa-miR-15b-5p-CA2 ceRNA axes may be important mechanisms for HBV-related HCC.

Introduction

Despite the advent of effective vaccines and antiviral therapy, hepatitis B virus (HBV) infection remains one of the most important risk factors for the development of hepatocellular carcinoma (HCC), accounting for at least 50% of cases of primary liver tumors (1,2). HCC is the most common malignant disease and is associated with a poor 5-year survival rate (<20%) in most patients (3). Therefore, there still is a great need for exploring the molecular mechanisms and developing more effective therapeutic strategies for HBV-related HCC.

It is reported that in addition to ~20,000 protein-coding genes, ~90% of the human genome is non-protein-coding RNAs, including microRNAs (miRNAs, 21 nt) and long noncoding RNAs (lncRNAs, 200 nt in length) (4). Thus, aberrant expression of miRNAs (5,6) and lncRNAs (7,8) may be potential mechanisms for the progression of HBV-related HCC. miRNAs have been extensively studied to play important roles in HBV-related HCC by negatively regulating protein-coding mRNAs by directly binding to their 3'-untranslated region (3'UTR). For example, Du *et al* utilized miRNA microarray to reveal that miR-382-5p was upregulated in hepatitis B virus core (HBc) protein-expressed HCC cells. Transfection of miR-382-5p inhibitors abrogated the enhanced effects of HBc on cell migration and invasion by regulating the expression of deleted in liver cancer 1 (DCL-1) (9). Li *et al* revealed that miR-125a-5p was evidently downregulated in HBV-related HCC. Overexpression of miR-125a-5p markedly inhibited cell proliferation and induced cell apoptosis by reducing both the mRNA and protein levels of its target gene erb-b2 receptor tyrosine kinase 3 (ErbB3) (10).

The understanding of the lncRNA functions in HBV-related HCC remains limited except for a few studies that have elucidated their influence on the expression of related mRNAs. For example, Liu *et al* used RNA deep sequencing to demonstrate that lnc-HUR1 was significantly upregulated in HBV transgenic HepG2-4D14 cells (11). lnc-HUR1 could enhance cell proliferation by interacting with p53 to block the transcription of the pro-apoptotic gene B cell lymphoma 2-associated X (Bax) (11). A study by Jin *et al* revealed that hepatitis B virus x protein (HBx) induced upregulation of ZEB2-AS1 by 3.55-fold. Inhibition of ZEB2-AS1 could reverse the expression of epithelial-mesenchymal transition (EMT) markers regulated by HBx and then prevent migration and invasion of HCC cells (12). Recently, accumulating evidence has indicated that lncRNAs may also suppress miRNA functions by acting as competing endogenous RNAs (ceRNAs) in HCC (13,14). Lv *et al* revealed that lnRNA Unigene56159 promoted the migration and invasion of HCC cells by acting as a ceRNA for miR-140-5p to de-repress the expression of Slug and induce EMT (15). Fan *et al* observed that lncRNA n335586 accelerated HCC cell migration and invasion by facilitating the expression of its host gene CKMT1A by competitively binding miR-924 (16). Therefore, identification of the lncRNA-associated ceRNA axes may provide further insights into the development of HBV-HCC, which to date remains rarely reported.

The aim of the present study was to specifically identify HBV-HCC related lncRNA ceRNA axes by using microarray or sequencing data collected from the public database. The study results may provide potential diagnostic, prognostic and therapeutic biomarkers for HBV-associated HCC.

Materials and methods

Data collection and preprocessing. The HBV-HCC datasets were downloaded from the Gene Expression Omnibus (GEO) database (http://www.ncbi.nlm.nih.gov/geo/) on November 2018, including: i) GSE27462 (17) which analyzed the lncRNA expression profiling in 5 HBV-HCC and 5 matched adjacent normal tissues using microarray (Platform: GPL11269, Arraystar Human lncRNA array version 1); ii) GSE76903 (18) which explored the miRNA expression profiling in 20 HBV-HCC and 20 matched adjacent normal tissues using high throughput sequencing (Platform: GPL16791, Illumina HiSeq 2500, Homo sapiens); and iii) GSE121248 which investigated the mRNA expression profiling in 70 HBV-HCC and 37 matched adjacent normal tissues using a microarray technique [Platform: GPL570, (HG-U133_Plus_2) Affymetrix Human Genome U133 Plus 2.0 Array].

The raw data were respectively preprocessed using the Linear Models for Microarray Data (LIMMA) package (version 3.34.0; https://bioconductor.org/packages/release/bioc/html/limma.html) in R (version 3.4.1; http://www.R-project.org/) (19) and oligo package (version 1.41.1; http://www.bioconductor.org/packages/release/bioc/html/oligo.html) (20) in R for the GSE27462 and GSE121248 microarray datasets. The downloaded data had been saved using the normalized_count function for GSE76903 and preprocessing was not necessary.

Differential expression analysis. The differentially expressed genes (DEGs) and lncRNAs (DELs) between HBV-HCC and adjacent normal tissues were identified using the LIMMA package (19). The differentially expressed miRNAs (DEMs) between HBV-HCC and adjacent normal tissues were screened using the EdgeR package of R software (version 3.22.3; http://www.bioconductor.org/packages/release/bioc/html/edgeR.html) (21). The |logFC(fold change)|>0.5 and false discovery rate (FDR) <0.05 were considered as the threshold value. The pheatmap package (version: 1.0.8; https://cran.r-project.org/web/packages/pheatmap) in R was used to perform hierarchical clustering based on Euclidean distance.

lncRNA-miRNA-mRNA ceRNA regulatory network construction. The DIANA-LncBase database (version 2.0; http://carolina.imis.athena-innovation.gr/diana_tools/web/index.php?r=lncbasev2%2Findex-experimental) (22) was used to screen the interactions between DELs and DEMs. Only the DEL-DEM interactions with opposite expression trend were retained. The target genes of DEMs were predicted using starBase (version 2.0; http://starbase.sysu.edu.cn/index.php) which provided the prediction information from five frequently used algorithms [TargetScan (http://www.targetscan.org/vert_72/), picTar (https://pictar.mdc-berlin.de), RNA22 (https://cm.jefferson.edu/rna22/), PITA (https://genie.weizmann.ac.il/pubs/mir07/mir07_prediction.html) and miRanda (http://www.microrna.org/microrna/home.do)]. The target genes of DEMs were selected if they were predicted by at least one algorithm, and were then overlapped with the DEGs to obtain the DEM-DEG interactions. In addition, only the negative interaction pairs between DEMs and DEGs were retained. The DEL-DEM and DEM-DEG interaction pairs were integrated to obtain the DEL-DEM-DEG ceRNA axes which were used to construct the ceRNA network and visualized using the Cytoscape software (version 3.6.1; www.cytoscape.org/) (23).

The Database for Annotation, Visualization and Integrated Discovery (DAVID) online tool (version 6.8; http://david.abcc.ncifcrf.gov) (24) was used for predicting the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways and Gene Ontology (GO) terms of DEGs in the ceRNA network. The pathways or terms with P-value <0.05 were considered as statistically significant. In addition, all known HCC related pathways updated to 2017 were also downloaded from the Comparative Toxicogenomics Database (CTD; http://ctd.mdibl.org/) using the key word 'hepatocellular carcinoma' (25), which was then overlapped with the pathways enriched by the DEGs in the ceRNA network to obtain an HCC-related ceRNA network.

Identification of prognosis-related DELs, DEMs and DEGs in the ceRNA network. The miRNA and mRNA expression profile data of the HBV-HCC samples were collected from the Cancer Genome Atlas (TCGA; https://gdc-portal.nci.nih.gov/) database. Univariate Cox regression analysis was performed to screen overall survival (OS) and recurrence-free survival (RFS)-related DELs, DEMs and DEGs using the survival package (version 2.42.6; https://cran.r-project.org/package=survival). Then, survival curves were drawn for the significant prognosis-related DELs, DEMs and DEGs

Table I. Differentially expressed lncRNAs, miRNAs and mRNAs between HCC and adjacent normal tissues.

mRNA			miRNA			lncRNA		
Symbol	FDR	logFC	Symbol	FDR	logFC	Symbol	FDR	logFC
HAMP	3.03E-15	-4.16	hsa-miR-9-5p	2.36E-15	4.35	KIAA0087	6.00E-04	0.86
KCNN2	4.13E-24	-3.99	hsa-miR-431-5p	1.71E-13	4.02	SNHG9	1.73E-03	-1.44
CXCL14	7.95E-28	-3.98	hsa-miR-301b	8.99E-13	3.87	LINC02203	2.01E-03	1.81
CNDP1	1.25E-19	-3.75	hsa-miR-216b-5p	1.59E-12	3.87	H19	3.55E-03	0.97
FCN2	6.81E-23	-3.64	hsa-miR-483-3p	1.51E-11	3.61	WWC2-AS2	5.30E-03	0.82
IGFBP3	1.61E-16	-2.08	hsa-miR-7974	1.45E-10	3.48	HNF1A-AS1	1.00E-02	0.87
RRM2	1.78E-15	2.25	hsa-miR-323a-3p	3.79E-10	3.37	TTTY6B	1.10E-02	0.70
STEAP3	4.73E-13	-1.48	hsa-miR-483-5p	4.78E-10	3.29	FAM138B	1.27E-02	1.16
ALDH1B1	9.83E-12	-1.16	hsa-miR-200c-3p	8.07E-10	3.25	WEE2-AS1	1.30E-02	0.54
ACSL4	1.83E-11	2.81	hsa-miR-183-5p	1.92E-09	3.15	HLA-F-AS1	1.38E-02	-0.90
CCNE2	7.91E-10	1.42	hsa-miR-410-3p	2.90E-09	3.11	LINC00284	1.80E-02	-0.61
GLS2	1.03E-09	-2.40	hsa-miR-182-5p	8.38E-07	2.53	SNHG15	1.83E-02	-0.54
CA2	6.67E-09	-1.24	hsa-miR-34c-5p	3.03E-06	2.46	DLGAP1-AS1	2.01E-02	-0.71
GADD45B	7.04E-09	-1.25	hsa-miR-7-5p	3.77E-05	2.14	KMT2E-AS1	2.12E-02	-0.68
ACAA2	2.51E-08	-1.01	hsa-miR-299-3p	4.17E-05	2.11	LINC00339	2.14E-02	-0.72
FAS	2.03E-07	-1.03	hsa-miR-15b-5p	1.37E-04	1.94	MAP3K14-AS1	2.19E-02	0.76
ACSL1	6.24E-07	-1.09	hsa-miR-30c-2-3p	1.97E-03	-1.61	LINC00909	2.72E-02	-0.60
THBS1	1.46E-06	-1.56	hsa-miR-149-5p	9.85E-03	1.35	SRRM2-AS1	3.78E-02	-0.99
ACADSB	1.30E-05	-1.03	hsa-miR-214-5p	1.89E-02	-1.22	LINC00928	4.00E-02	-0.75
CPS1	1.75E-03	-1.24	hsa-miR-128-3p	2.36E-02	1.17	RUSC1-AS1	4.06E-02	-1.03

lncRNAs, long non-coding RNAs; miRNAs, microRNAs; HCC, hepatocellular carcinoma; FC, fold change; FDR, false discovery rate; CCNE2, cyclin E2; RRM2, ribonucleotide reductase M2 subunit.
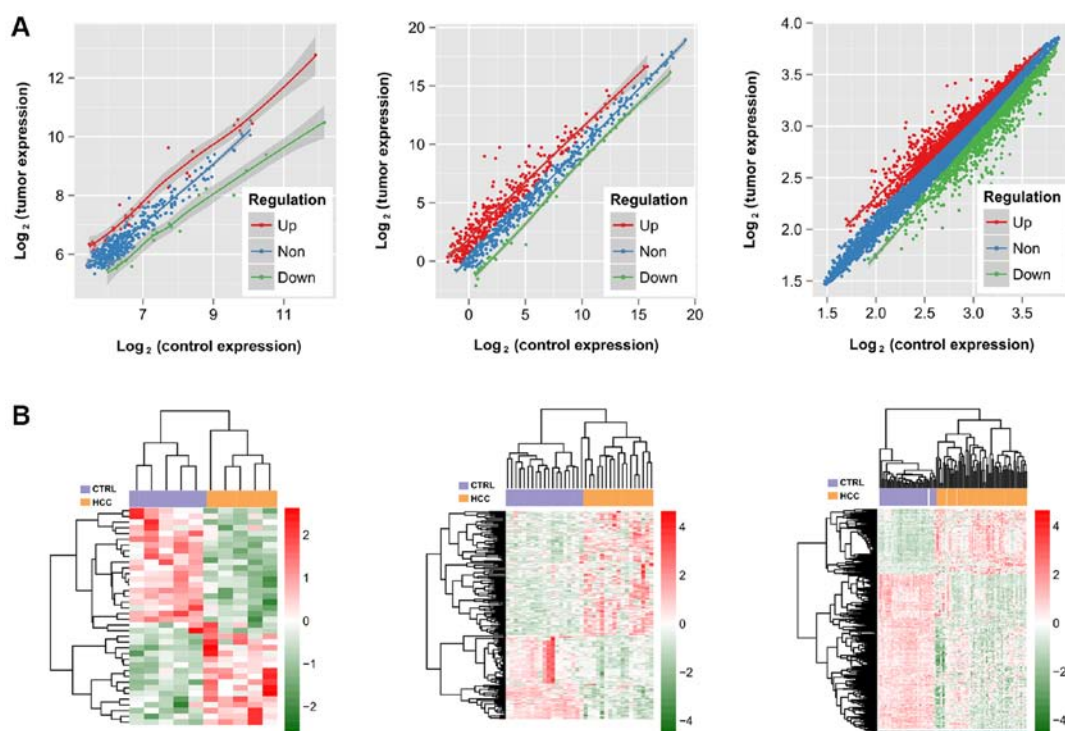

Figure 1. Expression pattern of lncRNAs, miRNAs and mRNAs in HBV-HCC (HCC group) and adjacent normal control tissues (CTRL group). (A) Scatter plots of lncRNAs (left), miRNAs (middle) and mRNAs (right) between two groups. The values of the x and y axes in the scatter plot are the averaged normalized signal values of the group (log₂ scaled); (B) Heat map showing differentially expressed lncRNAs (left), miRNAs (middle) and mRNAs (right) between two groups. Red and blue denote upregulated and downregulated expression, respectively. lncRNAs, long non-coding RNAs; miRNAs, microRNAs; HBV, hepatitis B virus; HCC, hepatocellular carcinoma.
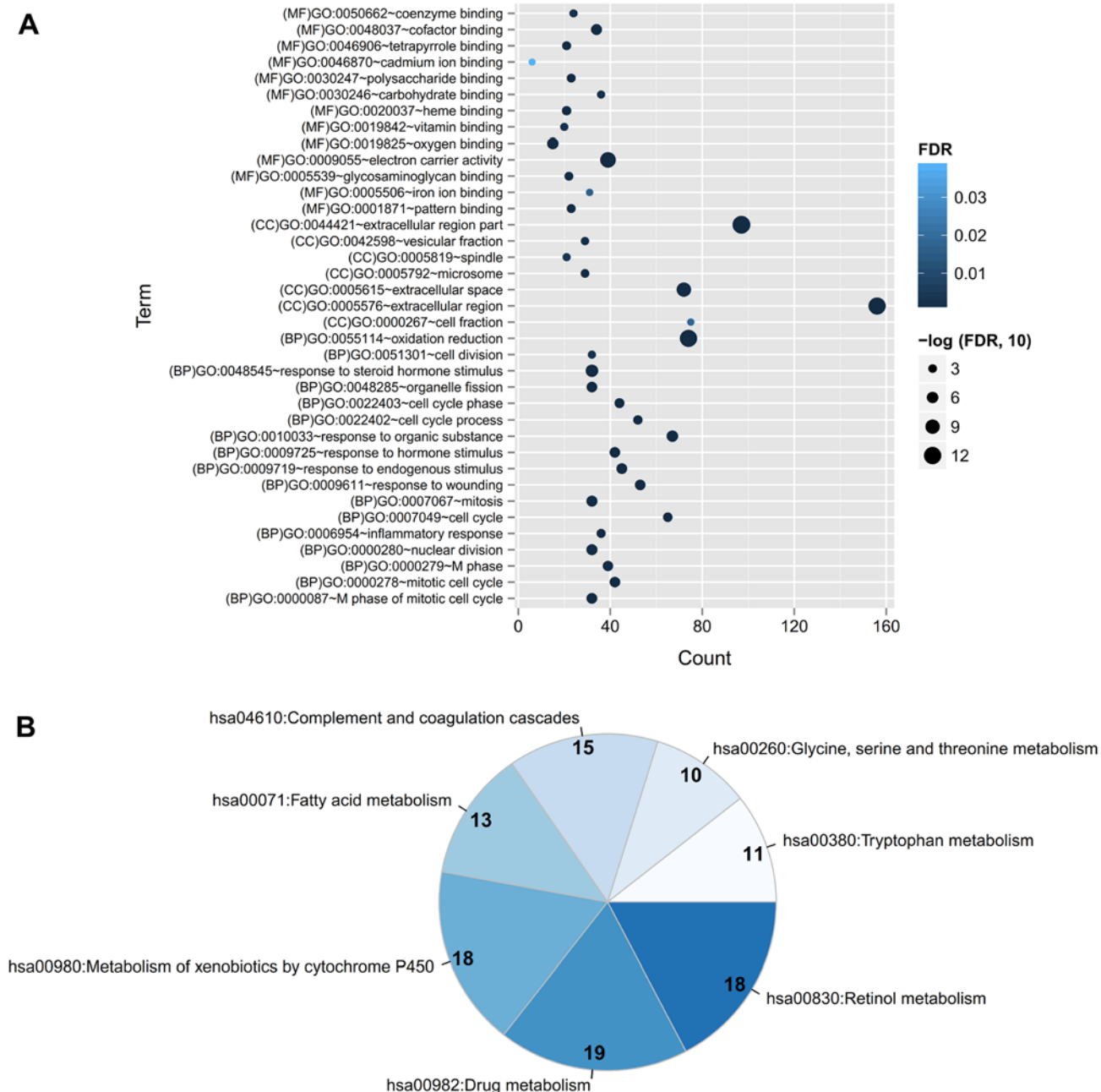
Figure 2. Function enrichment analyses for the differentially expressed genes. (A) GO enrichment. (B) KEGG pathway enrichment. GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes and Genomes.

by the Kaplan-Meier method with the log-rank test using the survival package. P<0.05 was considered to indicate a statistically significant difference.

Results

Differential expression analysis. A total of 38 DELs (16 downregulated and 22 upregulated), 127 DEMs (17 downregulated and 110 upregulated) and 721 DEGs (506 downregulated and 215 upregulated) were identified between HBV-HCC and adjacent normal tissues (Fig. 1A; Table I). The heat map analysis revealed that the DELs, DEMs and DEGs could be used to distinguish the HBV-HCC samples from normal samples completely (Fig. 1B).

Function enrichment for DEGs. GO terms and KEGG pathway enrichment analyses were performed for the DEGs to predict their functions by using DAVID software. The results revealed that 37 GO terms (including 17 biological processes, 7 cellular components and 13 molecular functions) were enriched for all the DEGs, mainly involving GO:0055114~oxidation reduction [ribonucleotide reductase M2 subunit (RRM2)], GO:0048545~response to steroid hormone stimulus [carbonic anhydrase 2 (CA2)] and GO:0007049~cell cycle [cyclin E2 (CCNE2)] (Fig. 2). Furthermore, 6 KEGG pathways were also enriched, including hsa00830:Retinol metabolism, hsa00982:Drug metabolism, hsa00980:Metabolism of xenobiotics by cytochrome P450, hsa00071:Fatty acid metabolism, hsa04610:Complement
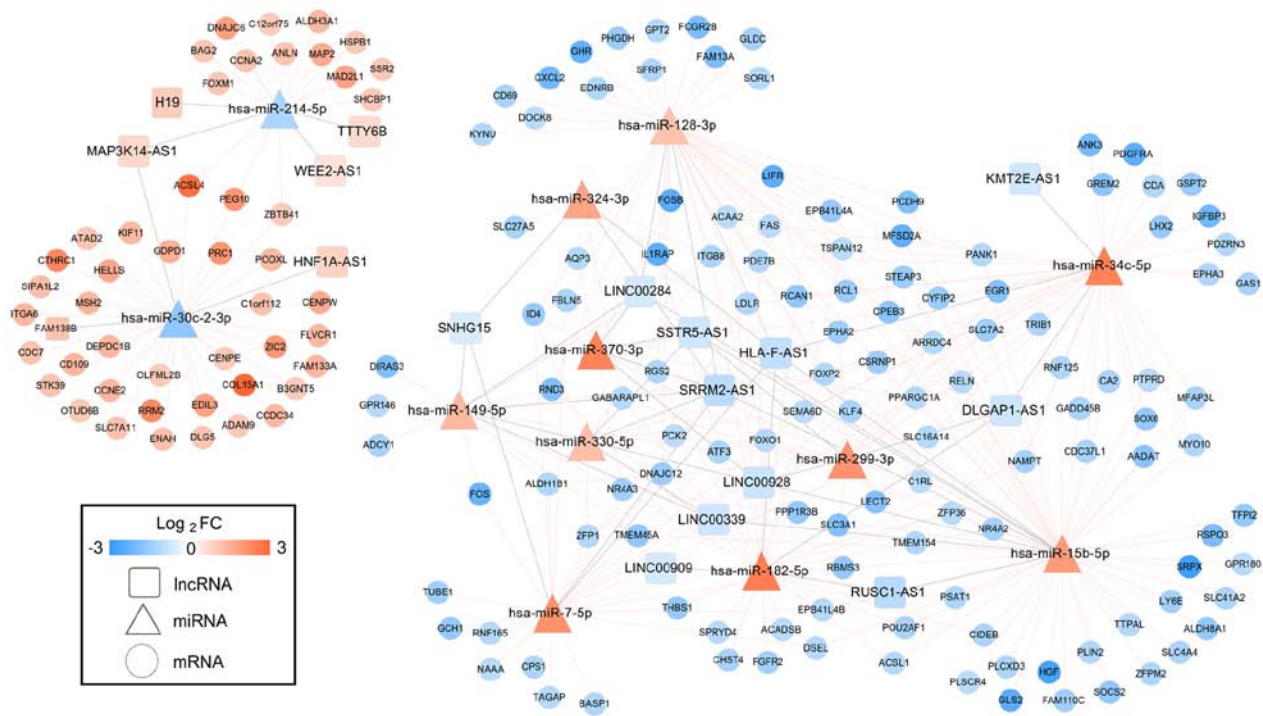
Figure 3. ceRNA interaction network of lncRNA-miRNA-mRNA. Square nodes represent lncRNAs; triangle nodes represent miRNAs; circular nodes represent mRNAs. Orange, upregulated; blue, downregulated. ceRNA, competing endogenous RNA; lncRNAs, long non-coding RNAs; miRNAs, microRNAs.
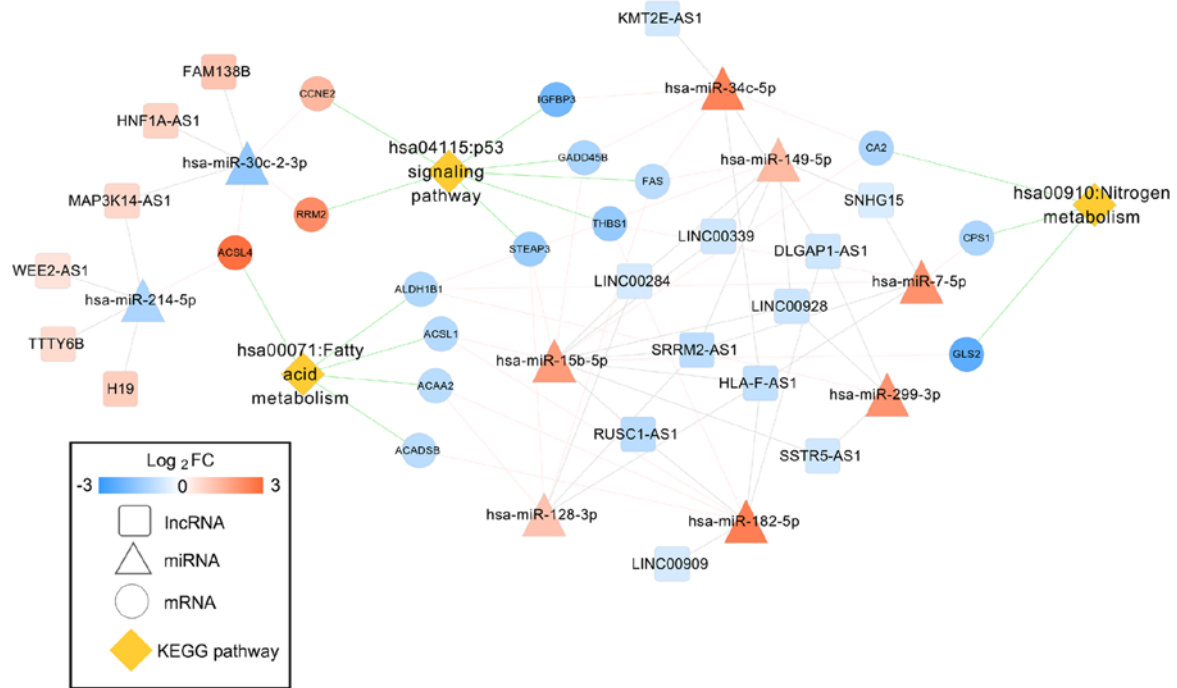

Figure 4. HCC-related ceRNA interaction network of lncRNA-miRNA-mRNA. Diamond nodes represent lncRNAs; triangle nodes represent miRNAs; circular nodes represent mRNAs; rhombus nodes represent HCC pathways. Orange, upregulated; blue, downregulated. HCC, hepatocellular carcinoma; ceRNA, competing endogenous RNA; lncRNAs, long non-coding RNAs; miRNAs, microRNAs.

and coagulation cascades and hsa00260:Glycine, serine and threonine metabolism (Fig. 2).

CeRNA network construction. By searching the DIANA-LncBasev2 database, 74 DEL-DEM interaction pairs with the opposite expression trend were predicted between 18 DELs (12 downregulated and 6 upregulated) and 20 DEMs (2 downregulated and 18 upregulated). Subsequently, the target genes of the aforementioned 20 DEMs were predicted with the starBase database. After removing the DEMs and DEGs with the consistent expression trend, 287 DEM-DEG interaction pairs between 12 DEMs (2 downregulated and

Table II. KEGG pathways enriched for the genes in the ceRNA network.

Term	P-value	Genes
hsa04115:p53 signaling pathway	2.83E-04	STEAP3, CCNE2, RRM2, FAS, GADD45B, THBS1, IGFBP3
hsa00071:Fatty acid metabolism	1.96 E-03	ACAA2, ACADSB, ACSL1, ALDH1B1, ACSL4
hsa00910:Nitrogen metabolism	3.82 E-02	GLS2, CA2, CPS1

KEGG, Kyoto Encyclopedia of Genes and Genomes; ceRNA, competing endogenous RNA; CCNE2, cyclin E2; RRM2, ribonucleotide reductase M2 subunit.

10 upregulated) and 173 DEGs (126 downregulated and 47 upregulated) were retained. By integrating the DEL-DEM and DEM-DEG interaction pairs, a DEL-DEM-DEG ceRNA network was constructed, which consisted of 202 nodes (17 DELs: 2 downregulated and 15 upregulated; 12 DEMs: 2 downregulated and 10 upregulated; 173 DEGs: 126 downregulated and 47 upregulated) (Fig. 3). In this ceRNA network, the FAM138B-hsa-miR-30c-CCNE2/RRM2 and SSTR5-AS1-has-miR-15b-5p-CA2 ceRNA axes were included.

Function enrichment analysis with DAVID revealed that the genes in the ceRNA network were involved in 3 significant KEGG pathways, including the hsa04115:p53 signaling pathway (CCNE2, RRM2), hsa00071:Fatty acid metabolism and hsa00910:Nitrogen metabolism (CA2) (Table II). These three pathways were also included in the 244 KEGG pathways that had been demonstrated to be associated with HCC in the CTD database. Thus, the DEL-DEM-DEG interaction pairs that were associated with these three pathways were extracted to form the HCC-related ceRNA network (Fig. 4).

Identification of prognosis-related DELs, DEMs and DEGs in the ceRNA network. Eighty HBV-related HCC samples having OS and RFS information were collected from TCGA database. Univariate Cox regression analysis was used to screen OS- and RFS-related DELs, DEMs and DEGs from the HCC-related ceRNA network in these samples. The results revealed that six RNAs (2 DELs: FAM138B and SSTR5-AS1; 2 DEMs: hsa-miR-149 and hsa-miR-7; 2 DEGs: CCNE2 and RRM2) were significantly associated with OS, and seven RNAs (1 DEL: LINC00284; 3 DEMs: hsa-miR-7, hsa-miR-15b, hsa-miR-30c-2; and 3 DEGs: RRM2, CCNE2, CA2) were significantly associated with RFS (Table III). Survival curves were drawn for these significant prognosis-related DELs, DEMs and DEGs by Kaplan-Meier according to their expression levels in the sequencing data. As anticipated, patients with a high expression of FAM138B, hsa-miR-149-5p, hsa-miR-7-5p, hsa-miR-15b-5p, CCNE2 and RRM2 had poor survival, while patients with low expression of SSTR5-AS1 and CA2 possessed poor survival (Figs. 5 and 6). However, patients with low expression of LINC00284 and hsa-miR-30c-2 had excellent survival, which was not consistent with what we anticipated (Fig. 6). Accordingly, our results indicated the SSTR5-AS1-hsa-miR-15b-5p-CA2 ceRNA axis may be especially important for the development and prognosis of HBV-related HCC by influencing nitrogen metabolism.

Table III. Univariate Cox regression analysis to screen OS and RFS-related genes.

OS		RFS	
ID	Cox-p	ID	Cox-p
FAM138B	0.0012	LINC00284	0.011
SSTR5-AS1	0.011	hsa-miR-7-1	0.0019
hsa-miR-149	5.00E-04	hsa-miR-15b	0.021
hsa-miR-7-2	0.037	hsa-miR-30c-1	0.0335
CCNE2	0.017	RRM2	0.025
RRM2	0.042	CA2	0.028
		CCNE2	0.046

OS, overall survival; RFS, recurrence-free survival; CCNE2, cyclin E2; RRM2, ribonucleotide reductase M2 subunit.

Discussion

In the present study, the expression of lncRNAs, miRNAs and mRNAs in HBV-related HCC was comprehensively analyzed and two lncRNA-associated ceRNA axes were identified: FAM138B-hsa-miR-30c-CCNE2/RRM2 and SSTR5-AS1-has-miR-15b-5p-CA2. They were involved in HBV-related HCC by influencing the p53 signaling pathway, the cell cycle and nitrogen metabolism. In addition, all the genes in these two axes were significantly associated with the OS or RFS of patients. These findings indicated that these genes may be important prognostic biomarkers and therapeutic targets for HBV-related HCC.

Although there are studies that have demonstrated the roles of lncRNAs in cancer development (including HCC) (16,26), there are almost no studies focusing on FAM138B except for one which confirmed its prognostic value in lung adenocarcinoma (27). However, in contrast to a study by Li *et al* (27), FAM138B was upregulated in HCC, not downregulated. This indicates that it is necessary to verify the specific roles of FAM138B in HCC. In present study, FAM138B was predicted to possibly be involved in HBV-HCC by regulating cell cycle-related genes (CCNE2 and RRM2) by sponging hsa-miR-30c, resulting in poor prognosis. Cyclin E2 (CCNE2) is the second member of the E-type cyclin family that forms a complex with cyclin-dependent kinase 2 to promote G1/S cell cycle phase transition (28). Thus, upregulation of CCNE2
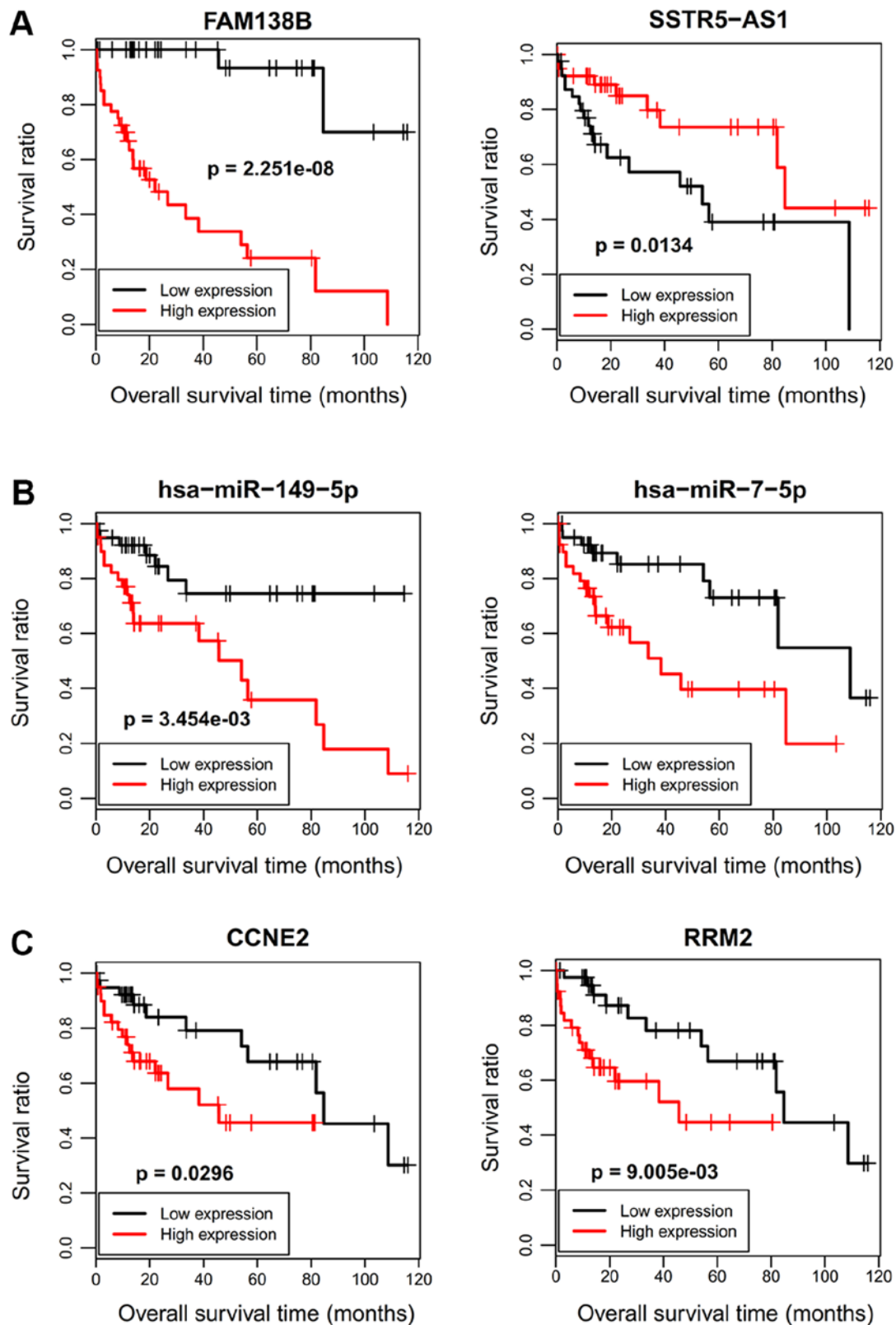
Figure 5. Kaplan-Meier analysis displaying the correlation of differentially expressed (A) lncRNAs, (B) miRNAs and (C) genes with overall survival outcomes for patients with HBV-related HCC. lncRNAs, long non-coding RNAs; miRNAs, microRNAs; HBV, hepatitis B virus; HCC, hepatocellular carcinoma.

may facilitate cell cycle progression and then excessive cell proliferation, ultimately leading to the development of cancer. These hypotheses have been demonstrated by previous studies. For example, Payton et al observed that the level of CCNE2 was significantly increased in primary breast

tumor samples relative to normal breast tissue controls (29). Kumari et al demonstrated that CCNE2 was frequently overexpressed in the early stages of gastric carcinoma and its expression was significantly correlated with histological type, tumor location, tumor differentiation, tumor invasion
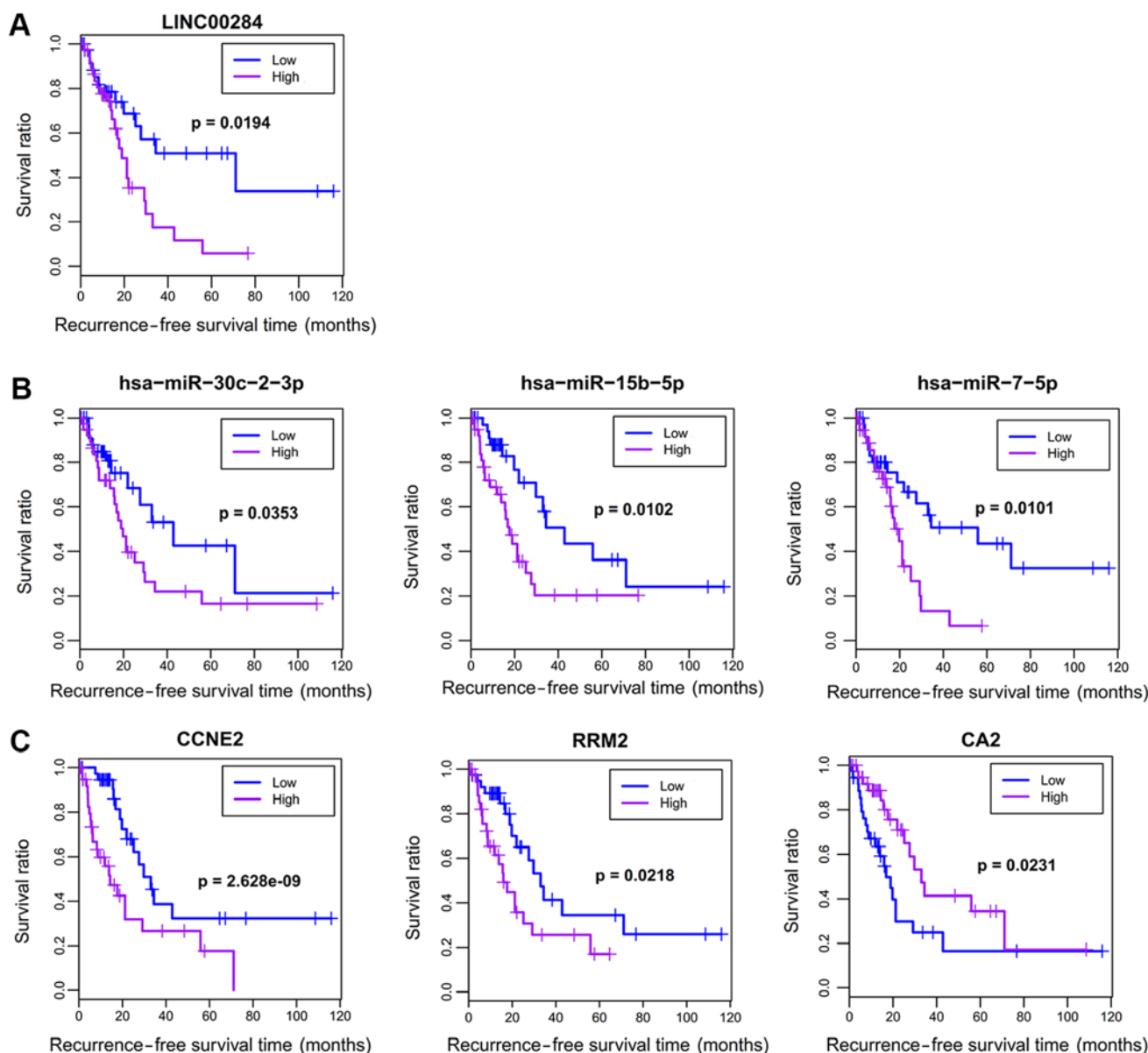
Figure 6. Kaplan-Meier analysis displaying the correlation of differentially expressed (A) lncRNAs, (B) miRNAs and (C) genes with recurrence-free survival outcomes for patients with HBV-related HCC. lncRNAs, long non-coding RNAs; miRNAs, microRNAs; HBV, hepatitis B virus; HCC, hepatocellular carcinoma.

and tumor metastasis (30). Xie *et al* demonstrated that high CCNE2 expression was associated with worse OS (hazard ratio: 1.36, 95% confidence intervals: 1.15-1.6, P<0.01) (31). The CCNE2-CDK2 complex was also reported to mediate SAMHD1 phosphorylation and abrogate the inhibitory roles of SAMHD1 on HBV replication in hepatoma cells (32). In line with these studies, our study also revealed that CCNE2 was upregulated in tumor samples of patients with HBV-related HCC and the high expression of CCNE2 predicted poor OS and RFS. Ribonucleotide reductase M2 subunit (RRM2) is a subunit of the ribonucleotide reductase that is responsible for the reduction of ribonucleotide 5'-diphosphates into 2'-deoxyribonucleotides for DNA synthesis and replication. In addition, RRM2 also plays an active role in tumor progression, which has been confirmed in several cancers. For example, Morikawa *et al* revealed that RRM2 was overexpressed in 72 cases (64.3%), but rarely expressed in normal gastric

mucosa. RRM2 overexpression was significantly associated with presence of muscularis propria invasion and presence of Epstein-Barr virus (33). Overexpression of RRM2 promoted their invasiveness (34), while suppression of RRM2 inhibited the growth of gastric cancer cells (33). A study by Wang *et al* revealed that human papillomavirus E7 induced the upregulation of RRM2 and promoted the development of cervical cancer (35). In accordance with these studies, in the present study RRM2 was predicted to be induced by HBV to promote the development of HCC and the high expression of RRM2 may serve as a poor prognostic factor. Previously, overexpression of miR-30c was reported to significantly inhibit cell proliferation and delay G1/S phase transition in hepatoma cells by blocking the HBV replication and the expression of HBV pgRNA, capsid-associated virus DNA and Hbx, indicating its tumor suppressive roles (36). As anticipated, miR-30c was also revealed to be downregulated in HBV-HCC samples.

However, its association with RFS was not in line with what was anticipated, which may be attributed to the small sample size. One study indicated that transfection with mimics of the miR-30 family member, miR-30b, significantly downregulated CCNE2 expression, while the miR-30b inhibitor upregulated CCNE2 in breast cancer cell lines (37). RRM2 was also predicted to be a direct target of miR-30d in lung adeno-carcinoma (38). However, there was no direct evidence to demonstrate the interaction between FAM138B and miR-30c as well as between miR-30c and CCNE2 and RRM2 in HCC.

A recent study by Ramnarine *et al* (39) revealed that SSTR5-AS1 was expressed at a low level in treatment-induced neuroendocrine prostate cancer, an aggressive variant of late-stage metastatic castrate-resistant prostate cancer compared with prostate adenocarcinoma; SSTR5-AS1 was able to iden-tify patients more likely to develop metastatic disease from those that did not; its underlying mechanism was to interact with KDM4B, a histone demethylase that could regulate and epigenetically activate the N-Myc oncogene and lead to cancer development (40). However, the function of SSTR5-AS1 in cancer remains not fully elucidated. In the present study, it was predicted that downregulated SSTR5-AS1 may be involved in HBV-HCC by regulating CA2 via sponging of miR-15b-5p, resulting in poor prognosis. CA2 encodes one of isozymes of carbonic anhydrase which catalyzes reversible hydration of carbon dioxide and plays a pivotal role in tissue pH homeo-stasis. Several studies have revealed that carbonic anhydrase members (i.e., CA1, CA9 and CA12) may be upregulated to promote cancer cell growth, survival and invasiveness (41-43). The use of carbonic anhydrase inhibitors significantly inhibits the malignant characteristics of cancers and improves chemotherapy sensitivity (44,45). However, the expression of CA2 appears to be controversial in different cancers, such as upregulated in urinary bladder cancers (46), but downregulated in esophageal adenocarcinoma (47). A recent study performed by Zhang *et al* also implied that downregulation of CA2 promoted HCC metastasis and invasion (48). Consistent with the study by Zhang *et al* (48) and Nortunen *et al* (47), the present results also revealed that CA2 was significantly expressed at a lower level in HBV-HCC samples compared with the normal control, suggesting that CA2 may be a tumor suppressor gene for HCC. Extensive studies have revealed that miR-15b-5p was upregulated in cancer samples to promote tumorigenicity and progression (49,50), including HCC (51). Consistently, it was also revealed in the present study that miR-15b-5p was highly expressed in HBV-HCC tissues. However, there was no direct evidence to demonstrate the interaction between SSTR5-AS1 and miR-15b-5p as well as between miR-15b-5p and CA2 in HCC.

Some limitations are present in the present study. First, the sample size of our study was not large, which may be a reason leading to the expression difference in some targets (LINC00284 and hsa-miR-30c-2) with what was anticipated. Thus, more clinical samples should be further collected to confirm their expression (quantitative-polymerase chain reaction analysis) and clinical values. Second, this study was only performed to preliminarily identify the potential ceRNA mechanisms for HBV-related HCC. Subsequent cells (RNA pull-down) and animal (knockout or overexpression) experiments should be performed to validate the interaction

relationships between DEMs and DELs/DEGs in HBV-related HCC.

In conclusion, the present study preliminarily indicated that FAM138B and SSTR5-AS1 may be novel prognostic biomarkers and therapeutic targets for HBV- associated HCC. They function as ceRNAs to sponge hsa-miR-30c and miR-15b-5p to regulate CCNE2/RRM2 and CA2.

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Availability of data and materials

All the microarray data were downloaded from the GEO database in NCBI (http://www.ncbi.nlm.nih.gov/geo/). The mRNA and miRNA Seq-data were obtained from The Cancer Genome Atlas (TCGA; https://tcga-data.nci.nih.gov/).

Authors' contributions

JX was involved in the conception and design of the study, drafted and revised the manuscript. JX, JW and YW collected and analyzed the data. JZ and FS contributed to the interpre-tation of the data and revised the manuscript critically for important intellectual content. All authors read and approved the manuscript and agree to be accountable for all aspects of the research in ensuring that the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Ethics approval and informed consent

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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