



The Effect of the 10-Valent Pneumococcal Nontypeable Haemophilus influenzae Protein D Conjugate Vaccine on H. influenzae in Healthy Carriers and Middle Ear Infections in Iceland

Hildigunnur Sveinsdóttir,^{a,b,c} Jana Birta Björnsdóttir,^{a,b,c} Helga Erlendsdóttir,^{a,b} Martha Á. Hjálmarsdóttir,^{a,b,c} Birgir Hrafnkelsson,^d Ásgeir Haraldsson,^{a,e} Karl G. Kristinsson,^{a,b,c} ©Gunnsteinn Haraldsson^{a,b,c}

^aUniversity of Iceland, Faculty of Medicine, Reykjavík, Iceland

^bDepartment of Clinical Microbiology, Landspitali University Hospital, Reykjavík, Iceland

^cBioMedical Centre, University of Iceland, Reykjavík, Iceland

^dDepartment of Mathematics, University of Iceland, Reykjavík, Iceland

eChildren's Hospital Iceland, Reykjavík, Iceland

ABSTRACT Vaccinations with the 10-valent pneumococcal conjugated vaccine (PHiD-CV) started in Iceland in 2011. Protein D (PD) from H. influenzae, which is coded for by the hpd gene, is used as a conjugate in the vaccine and may provide protection against PD-positive H. influenzae. We aimed to evaluate the effect of PHiD-CV vaccination on H. influenzae in children, both in carriage and in acute otitis media (AOM). H. influenzae was isolated from nasopharyngeal swabs collected from healthy children attending 15 day care centers in 2009 and from 2012 to 2017 and from middle ear (ME) samples from children with AOM collected from 2012 to 2017. All isolates were identified using PCR for the hpd and fuck genes. Of the 3,600 samples collected from healthy children, 2,465 were culture positive for H. influenzae (68.5% carriage rate); of these, 151 (6.1%) contained hpd-negative isolates. Of the 2,847 ME samples collected, 889 (31.2%) were culture positive for H. influenzae; of these, 71 (8.0%) were hpd negative. Despite the same practice throughout the study, the annual number of ME samples reduced from 660 in 2012 to 330 in 2017. The proportions of hpd-negative isolates in unvaccinated versus vaccinated children were 5.6% and 7.0%, respectively, in healthy carriers, and 5.4% and 7.8%, respectively, in ME samples. The proportion of hpd-negative isolates increased with time in ME samples but not in healthy carriers. The number of ME samples from children with AOM decreased. The PHiD-CV had no effect on the proportion of the hpd gene in H. influenzae from carriage, but there was an increase in hpd-negative H. influenzae in otitis media. The proportions of hpd-negative isolates remained similar in vaccinated and unvaccinated children.

KEYWORDS *Haemophilus influenzae*, Iceland, carriage, epidemiology, otitis media, protein D, vaccines

aemophilus influenzae is a common colonizer of the upper respiratory tracts of healthy individuals, especially children. However, *H. influenzae* can cause various infections in the respiratory tract and connected body sites, especially in young children (1).

A conjugated vaccine targeting *H. influenzae* type b (Hib) has almost eradicated invasive diseases caused by *H. influenzae* in countries where the vaccine is widely used. Before vaccination, serotype b caused up to 96% of all *H. influenzae* invasive diseases (2). Nowadays, the most common *H. influenzae* infections are respiratory tract infec-

Citation Sveinsdóttir H, Björnsdóttir JB, Erlendsdóttir H, Hjálmarsdóttir MÁ, Hrafnkelsson B, Haraldsson Á, Kristinsson KG, Haraldsson G. 2019. The effect of the 10-valent pneumococcal nontypeable *Haemophilus influenzae* protein D conjugate vaccine on *H. influenzae* in healthy carriers and middle ear infections in Iceland. J Clin Microbiol 57:e00116-19. https://doi.org/10.1128/JCM .00116-19.

Editor Daniel J. Diekema, University of Iowa College of Medicine

Copyright © 2019 Sveinsdóttir et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Gunnsteinn Haraldsson, gah@hi.is.

Received 25 January 2019 Returned for modification 24 February 2019

Accepted 27 April 2019 Accepted manuscript posted online 8 May 2019

Published 25 June 2019

tions, e.g., otitis media caused by nonencapsulated strains (NTHi). NTHi are the most common cause of acute otitis media (AOM) after pneumococci (3).

In Iceland, Hib vaccine has been in included in the national immunization program since 1989 (4), but the 10-valent pneumococcal conjugated vaccine (PHiD-CV; Synflorix) was introduced into the program in April 2011 in a 2-plus-1 schedule without catch-up. No other protein-conjugated pneumococcal vaccine had previously been included (5). Protein D (PD) from NTHi is used as the conjugate for 8 of the 10 pneumococcal serotypes in PHiD-CV (6). PD is a highly conserved outer membrane protein found in most, but not all, *H. influenzae* strains (7). The inclusion of PD in the vaccine may provide protection in young children against infections caused by PD-positive *H. influenzae* (8).

The aim of this study was to evaluate the effect of vaccination with the PHiD-CV on *H. influenzae* carriage in healthy children attending day care centers (DCC) and from the middle ears (MEs) of children with AOM.

MATERIALS AND METHODS

Sampling and isolation. Nasopharyngeal swabs were collected from healthy children 1 to <7 years of age attending 15 DCCs in the greater Reykjavík area in March 2009 and from 2012 to 2017, as previously described (5). When *H. influenzae* isolates showing different colony morphologies were seen from DCC samples, isolates of each respective form were included in the study.

All ME samples from children <7 years of age with suspected otitis media that were submitted to the Department of Clinical Microbiology, Landspitali University Hospital, in 2009 to 2017 as previously described were included (5). Samples were usually collected with a swab from the outer ear after a rupture in the tympanic membrane following AOM or if discharge was secreted through a tympanostomy tube. When two or more *H. influenzae* isolates were identified from ME samples from the same patient within 30 days, only the first isolate was included in the study.

All samples were selectively cultured for *H. influenzae* using chocolate blood agar with bacitracin discs and incubated in 5% CO₂ overnight.

Identification. The identification of all isolates was confirmed using PCR for the *hpd* gene, which codes for PD. Isolates negative with the hpd#1 primers were further tested with the hpd#3 primers to increase sensitivity, and isolates negative with both *hpd* primers were confirmed as *H. influenzae* using PCR for the *fucK* gene, which codes for fuculose kinase, using two sets of primers to increase sensitivity (9–11). The identification of isolates negative for both genes was confirmed as non-*H. influenzae* using a matrix-assisted laser desorption ionization–time of flight (MALDI-TOF) system.

In addition, all isolates were tested for the presence of capsular genes by using PCR for the *bexA* gene, and all positive isolates were further typed using type-specific primers (12, 13). All primers used in the study are listed in Table S1 in the supplemental material. Some of the primers and the PCRs were slightly modified for endpoint PCR.

Statistical analysis. The primary service area for the Landspitali University Hospital was considered to be within 100 km driving distance from the hospital. To correct the number of ME samples with regard to the population, the population demographic information for children <7 years of age in this region was obtained from Statistics Iceland (www.statice.is).

All children born before 31 December 2010 were grouped as unvaccinated, and children born 1 January 2011 or later were grouped as vaccinated.

GraphPad QuickCalcs (GraphPad Software, San Diego, CA) two-sided Fisher's exact test was used to compare groups. A likelihood ratio test was used to test the time trend in the number of ME samples, carriage rates, and proportions of *hpd*-negative isolates per year during the sampling period. All levels of significance were set at 0.05.

Ethics. Informed consent was obtained from parents of all children in the DCC studies. The study was approved by The National Bioethics Committee (VSNb2013010015/03.07), the appropriate authorities at the Landspitali University Hospital, and the DCC's directors.

RESULTS

Bacterial isolates. In total, 3,600 nasopharyngeal swab samples were collected from healthy children (aged 1 to 6 years; median age, 4.3), of which, 2,465 samples were culture positive for *H. influenzae* (68.5% overall carriage rate). Carriage rates fluctuated between years but did not change with relation to time, although it was highest in 2009 (81.6%) and lowest in 2017 (57.1%). The carriage rate was highest in children less than 2 years of age and gradually declined with age (see Table S2 in the supplemental material) but did not change with relation to vaccination status (see Table S3). Of the 2,465 nasopharyngeal swab samples positive for *H. influenzae*, 114 contained *H. influenzae* showing two types of colony morphology. In these cases, both isolates were tested for the presence of the *hpd* and *bexA* genes. Of the 2,465 nasopharyngeal swab

TABLE 1 Samples from the nasopharynx of healthy children

Category	2009	2012	2013	2014	2015	2016	2017	Total
No. of samples	520	465	471	566	533	539	506	3,600
Samples from vaccinated children (no. [%])	0 (0)	0 (0)	18 (3.8)	126 (22.3)	252 (47.3)	377 (69.8)	506 (100)	1,279 (35.5)
Samples positive for <i>H. influenzae</i> (no. [% carriage])	421 (81.6)	282 (60.6)	339 (72.0)	355 (62.7)	395 (74.1)	384 (71.1)	289 (57.1)	2,465 (68.5)
Samples containing <i>hpd</i> -negative isolates (no. [%])	27 (6.4)	14 (5.0)	16 (4.7)	21 (5.9)	24 (6.1)	27 (7.0)	22 (7.6)	151 (6.1)
No. of samples containing <i>bexA</i> -positive isolates	3	5	0	5	9	0	6	28
Type(s) (n)	b (2), e (1)	e (2), f (3)		f (5)	e (2), f (7)		e (1), f (5)	b (2), e (6), f (20)

samples positive for *H. influenzae*, 151 (6.1%) contained *H. influenzae* isolates that were negative in the *hpd* PCR but positive in *fucK* PCR. The numbers of isolates from each year are listed in Table 1.

Of the 114 nasopharyngeal swab samples containing two types of colony morphology, 15 samples (13.1%) contained both *hpd*-positive and -negative isolates, and 5 samples (4.4%) contained both capsulated isolates and NTHi.

The numbers of ME samples received for culture at the Department of Microbiology remained stable for the years 2009 to 2011, but these samples were not stored and were therefore not included in the study.

During 2012 to 2017, 2,847 ME samples from children (aged 0 to 6 years; median age, 1.7 years) were collected, of which, 889 (31.2%) were positive for *H. influenzae*. The highest number of ME samples was collected in 2013 (n = 683), and then the number of ME samples gradually declined during the period (P < 0.001) and was lowest in 2017 (n = 330). The proportion of ME samples culture positive for *H. influenzae* was highest in 2012 (36.1%) but lowest in 2016 (23.7%). The numbers of samples and the proportions of culture-positive samples were highest in children less than 2 years of age and gradually declined with age (Table S2). The numbers of ME samples from vaccinated children reduced with time as expected (Table S3). Of the 889 ME samples culture positive for *H. influenzae*, 64 (7.2%) were negative in the *hpd* PCR but positive in *fucK* PCR. The numbers of isolates from each year are listed in Table 2.

During the first half of the study period, 2012 to 2014, the number of ME samples was 1,835 from an average of 26,027 children in the uptake area (annual incidence rate of 23.5 cases/1,000 children). During the second half of the study period, 2015 to 2017, the number of ME samples was 1,012 from an average of 25,294 children (annual incidence rate of 13.3 cases/1,000 children) (P < 0.001). The proportion of culture-positive ME samples decreased from 34.6% during 2012 to 2014 (649/1,835) to 25.2% (262/1,011) during 2015 to 2017 (P < 0.001), and the annual incidence rate of culture-positive ME samples was reduced from 8.1/1,000 children to 3.4/1,000 children, respectively.

No difference was seen in the proportions of samples containing *hpd*-negative isolates with relation to time in healthy carriers, but an increase in the proportion of *hpd*-negative isolates was seen in ME samples (P < 0.005). However, no difference was seen in the proportions of *hpd*-negative isolates in unvaccinated children versus vaccinated children, in either sample group (Table 3).

The proportions of encapsulated strains in both sample groups were low, with type f being the most common serotype (Tables 1 and 2). Three isolates of *H. influenzae* type b were found, two in the nasopharynx of healthy boys, 4.8 and 6.2 years old, attending the same DCC in 2009, and one isolate was found in an ME sample from a 9-month-old boy in 2015.

DISCUSSION

In this study, we investigated almost 3,400 isolates of *H. influenzae* for the presence of the *hpd* gene, and we showed that most of the isolates had the gene. If vaccination with PD induces immune response against PD-positive *H. influenzae*, the proportion of PD-negative isolates would likely increase over time in vaccinated children, both in carriage and in disease. We saw an increase in the proportion of *hpd*-negative isolates over time in ME samples but not in samples from healthy carriers. However, the

ge
fa
Ö
years of age
$^{\succ}$
children
ild
÷
of
ME of
from ME of
from
s from
2 Samples from
2 Samples from
from

Category	2009	2010	2011	2012	2013	2014	2015	2016	2017	Total since 2012
No. of samples	725	664	717	660	683	492	336	346	330	2,847
Samples from vaccinated children (no. [%])				225 (34.1)	487 (71.3)	421 (85.6)	296 (88.0)	340 (98.3)	325 (98.5)	2,094 (73.6)
Population under 7 years of age in the uptake area (n)				25,918	26,030	26,134	25,899	25,274	24,711	
Samples positive for <i>H. influenzae</i> (no. [%])	239ª (33.0)	233 ^a (35.1)	238 ^a (33.2)	235 (35.6)	230 (33.7)	169 (34.3)	80 (23.8)	82 (23.7)	93 (28.2)	889 (31.2)
Samples containing <i>hpd</i> -negative isolates (no [%])				12 (5.1)	13 (5.7)	12 (7.1)	5 (6.3)	9 (11.0)	13 (14.0)	64 (7.2)
No. of samples containing bexA-positive isolates				-	0	-	2	0	0	4
Type(s) (n)				e		f	b, e			b (1), e (2), f (1)
	-									

alsolates were not stored and were therefore not included in the study.

	DCC samples			ME samples				
lsolate	No. (%) from unvaccinated children	No. (%) from vaccinated children	P value	No. (%) from unvaccinated children	No. (%) from vaccinated children	P value		
hpd negative		63 (7.0)	0.163	12 (5.4)	52 (7.8)	0.294		
<i>hpd</i> positive Total	1,482 (94.4) 1,570 (100)	832 (93.0) 895 (100)		210 (94.6) 222 (100)	615 (92.2) 667 (100)			

TABLE 3 Numbers of hpd-positive and -negative samples from unvaccinated and vaccinated children in each sample group

percentages of isolates lacking the gene were similar in unvaccinated and vaccinated children, both in carriage and in disease.

Vaccination with the PHiD-CV started in Iceland in 2011, and the vaccine uptake was high; >97% received at least two doses (14). The first fully vaccinated children entered the DCCs in autumn 2012 and were sampled for the first time in 2013. In 2017, all children in the DCCs were fully vaccinated. Since the introduction of the PHiD-CV, we have only seen minor fluctuation in the carriage rate of *H. influenzae* in Iceland, which is similar to other studies (15). The carriage rate of *H. influenzae* in Iceland remained high during the study and did not seem to change with relation to the vaccine, although the lowest rate was found in 2017. The carriage rate of pneumococci in the same sampling group was also lowest in 2017 (5). In Brazil, carriage rates of NTHi increased after vaccination with PHiD-CV and protection against NTHi was not observed (16). Carriage rates of *H. influenzae* seem to differ largely in various studies, and the carriage rate in Iceland is considerably higher than was found in most other studies (16–20), possibly because of the different climate and/or different time period when samples were collected; however, the lowest carriage rate was found to be similar to that in Holland (15).

Before 2012, the total number of ME samples and the proportion of *H. influenzae*positive samples remained similar. A significant reduction of ME samples was seen after the vaccination with PHiD-CV, and in addition, the proportion of *H. influenzae*-positive ME samples was also reduced significantly. Similar results were seen with the pneumococci: the proportion of samples positive for pneumococci from middle ear samples decreased significantly (5). It is possible that the PD conjugate vaccine provides protection against AOM infections caused by *H. influenzae* (21). However, it is also possible that the decrease in sampling and the isolation of *H. influenzae* from ME infections in children are a combination of a direct effect from the PD, which is used as a conjugate in the vaccine, and reduced pneumococcal ME infections after vaccination (5, 22).

The vast majority of the *H. influenzae* isolates, both from carriage and ME samples, were NTHi. The most common serotype was serotype f, as seen in other studies (15, 19). Although Hib has almost disappeared from invasive infections (2), we found two isolates of *H. influenzae* type b in healthy carriers and one in an ME sample. It is therefore clear that despite high uptake of Hib vaccine (23), Hib can still be found in low numbers in the community.

The main weakness of the study is having isolates from only 1 year prior to the start of the PHiD-CV vaccination. However, the inclusion of almost 3,400 isolates, almost 1,800 isolates thereof from unvaccinated children, and the testing all isolates for the presence of the *hpd* gene can be considered strengths in the study.

Considering the global burden of respiratory diseases and repeated exacerbations among patients, it is important to monitor possible changes of *H. influenzae* in healthy carriers and in disease. The role of NTHi in respiratory diseases justifies the need for a vaccine in order to reduce repeated use of antibiotics.

Conclusions. The PHiD-CV had no effect on the prevalence of the *hpd* gene in *H. influenzae* from healthy carriers, but there was an increase in *hpd*-negative *H. influenzae* in otitis media. The proportions of isolates lacking the *hpd* gene remained similar in unvaccinated and vaccinated children. However, the number of ME samples and the

proportion of *H. influenzae*-positive samples from children with AOM submitted to the Department of Clinical Microbiology decreased.

SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at https://doi.org/10.1128/JCM .00116-19.

SUPPLEMENTAL FILE 1, PDF file, 0.4 MB.

ACKNOWLEDGMENTS

We thank the staff at the Department of Clinical Microbiology for collecting the ME isolate and the students that collected the DCC samples. We also thank the teachers and staff at the DCCs, the children participating in the carriage studies, and their parents. We also thank other members of the VIce study group.

This study was funded by The Icelandic Centre for Research (grant number 152047-051) and the Landspitali University Hospital Research Fund. The collection of DCC samples was part of an investigator-initiated study, supported by GlaxoSmithKline Biologicals SA.

REFERENCES

- Bakaletz LO, Novotny LA. 2018. Nontypeable Haemophilus influenzae (NTHi). Trends Microbiol 26:727–728. https://doi.org/10.1016/j.tim.2018 .05.001.
- Ladhani S, Slack MP, Heath PT, von Gottberg A, Chandra M, Ramsay ME. 2010. Invasive *Haemophilus influenzae* disease, Europe, 1996–2006. Emerg Infect Dis 16:455–463. https://doi.org/10.3201/eid1603.090290.
- Murphy TF, Faden H, Bakaletz LO, Kyd JM, Forsgren A, Campos J, Virji M, Pelton SI. 2009. Nontypeable *Haemophilus influenzae* as a pathogen in children. Pediatr Infect Dis J 28:43–48. https://doi.org/10.1097/INF .0b013e318184dba2.
- The Directorate of Health. 2016. *Haemophilus influenzae* sjúkdómur af gerð b (Hib). https://www.landlaeknir.is/smit-og-sottvarnir/smitsjukdomar/ sjukdomur/item12480/Haemophilus-influenzae-sjukdomur-af-gerd-b -(Hib). Accessed 4 March 2019.
- Quirk SJ, Haraldsson G, Erlendsdóttir H, Hjálmarsdóttir MÁ, van Tonder AJ, Hrafnkelsson B, Sigurdsson S, Bentley SD, Haraldsson Á, Brueggemann AB, Kristinsson KG. 2018. Effect of vaccination on pneumococci isolated from the nasopharynx of healthy children and the middle ear of children with otitis media in Iceland. J Clin Microbiol 56:e01046-18. https://doi.org/10.1128/JCM.01046-18.
- Forsgren A, Riesbeck K, Janson H. 2008. Protein D of *Haemophilus* influenzae: a protective nontypeable *H. influenzae* antigen and a carrier for pneumococcal conjugate vaccines. Clin Infect Dis 46: 726–731. https://doi.org/10.1086/527396.
- Smith-Vaughan HC, Chang AB, Sarovich DS, Marsh RL, Grimwood K, Leach AJ, Morris PS, Price EP. 2014. Absence of an important vaccine and diagnostic target in carriage- and disease-related nontypeable *Haemophilus influenzae*. Clin Vaccine Immunol 21:250–252. https://doi.org/10 .1128/CVI.00632-13.
- Schuerman L, Borys D, Hoet B, Forsgren A, Prymula R. 2009. Prevention of otitis media: now a reality? Vaccine 27:5748–5754. https://doi.org/10 .1016/j.vaccine.2009.07.070.
- Binks MJ, Temple B, Kirkham LA, Wiertsema SP, Dunne EM, Richmond PC, Marsh RL, Leach AJ, Smith-Vaughan HC. 2012. Molecular surveillance of true nontypeable *Haemophilus influenzae*: an evaluation of PCR screening assays. PLoS One 7:e34083. https://doi.org/10.1371/ journal.pone.0034083.
- 10. Meyler KL, Meehan M, Bennett D, Cunney R, Cafferkey M. 2012. Development of a diagnostic real-time polymerase chain reaction assay for the detection of invasive *Haemophilus influenzae* in clinical samples. Diagn Microbiol Infect Dis 74:356–362. https://doi.org/10 .1016/j.diagmicrobio.2012.08.018.
- Wang X, Mair R, Hatcher C, Theodore MJ, Edmond K, Wu HM, Harcourt BH, Carvalho Mda G S, Pimenta F, Nymadawa P, Altantsetseg D, Kirsch M, Satola SW, Cohn A, Messonnier NE, Mayer LW. 2011. Detection of bacterial pathogens in Mongolia meningitis surveillance with a new real-time PCR assay to detect *Haemophilus influenzae*. Int J Med Microbiol 301:303–309. https://doi.org/10.1016/j.ijmm.2010.11.004.
- July 2019 Volume 57 Issue 7 e00116-19

- Falla TJ, Crook DW, Brophy LN, Maskell D, Kroll JS, Moxon ER. 1994. PCR for capsular typing of *Haemophilus influenzae*. J Clin Microbiol 32:2382–2386.
- Wroblewski D, Halse TA, Hayes J, Kohlerschmidt D, Musser KA. 2013. Utilization of a real-time PCR approach for *Haemophilus influenzae* serotype determination as an alternative to the slide agglutination test. Mol Cell Probes 27:86–89. https://doi.org/10.1016/j.mcp.2012.11.003.
- Eythorsson E, Hrafnkelsson B, Erlendsdóttir H, Gudmundsson SA, Kristinsson KG, Haraldsson Á. 2018. Decreased acute otitis media with treatment failure after introduction of the ten-valent pneumococcal *Haemophilus influenzae* protein D conjugate vaccine. Pediatr Infect Dis J 37: 361–366. https://doi.org/10.1097/INF.000000000001870.
- van den Bergh MR, Spijkerman J, Swinnen KM, Francois NA, Pascal TG, Borys D, Schuerman L, Ijzerman EP, Bruin JP, van der Ende A, Veenhoven RH, Sanders EA. 2013. Effects of the 10-valent pneumococcal nontypeable *Haemophilus influenzae* protein D-conjugate vaccine on nasopharyngeal bacterial colonization in young children: a randomized controlled trial. Clin Infect Dis 56:e30–e39. https://doi.org/10.1093/cid/ cis922.
- Brandileone M-CDC, Zanella RC, Almeida SCG, Brandao AP, Ribeiro AF, Carvalhanas T-RMP, Sato H, Andrade A-L, Verani JR, Pneumococcal Carriage Study Group. 2016. Effect of 10-valent pneumococcal conjugate vaccine on nasopharyngeal carriage of *Streptococcus pneumoniae* and *Haemophilus influenzae* among children in São Paulo, Brazil. Vaccine 34:5604–5611. https://doi.org/10.1016/j.vaccine.2016.09.027.
- 17. Giufre M, Daprai L, Cardines R, Bernaschi P, Rava L, Accogli M, Raponi M, Garlaschi ML, Ciofi Degli Atti ML, Cerquetti M. 2015. Carriage of *Haemophilus influenzae* in the oropharynx of young children and molecular epidemiology of the isolates after fifteen years of *H. influenzae* type b vaccination in Italy. Vaccine 33:6227–6234. https://doi.org/10.1016/j .vaccine.2015.09.082.
- Cleary D, Devine V, Morris D, Osman K, Gladstone R, Bentley S, Faust S, Clarke S. 2018. Pneumococcal vaccine impacts on the population genomics of non-typeable *Haemophilus influenzae*. Microb Genom 4:e000209. https://doi.org/10.1099/mgen.0.000209.
- Puig C, Marti S, Fleites A, Trabazo R, Calatayud L, Linares J, Ardanuy C. 2014. Oropharyngeal colonization by nontypeable *Haemophilus influenzae* among healthy children attending day care centers. Microb Drug Resist 20:450–455. https://doi.org/10.1089/mdr.2013.0186.
- Ortiz-Romero MDM, Espejo-García MP, Alfayate-Miguelez S, Ruiz-López FJ, Zapata-Hernandez D, Gonzalez-Pacanowska AJ, Collaborators of Study Group of Infectious Diseases in the Child in Cartagena. 2017. Epidemiology of nasopharyngeal carriage by *Haemophilus influenzae* in healthy children: a study in the Mediterranean coast region. Pediatr Infect Dis J 36:919–923. https://doi.org/10.1097/INF.000000000001625.
- Leach AJ, Wigger C, Hare K, Hampton V, Beissbarth J, Andrews R, Chatfield M, Smith-Vaughan H, Morris PS. 2015. Reduced middle ear infection with non-typeable *Haemophilus influenzae*, but not *Streptococ*-

cus pneumoniae, after transition to 10-valent pneumococcal non-typeable *H. influenzae* protein D conjugate vaccine. BMC Pediatr 15:162. https://doi.org/10.1186/s12887-015-0483-8.

22. Sigurdsson S, Eythorsson E, Hrafnkelsson B, Erlendsdóttir H, Kristinsson KG, Haraldsson Á. 2018. Reduction in all-cause acute otitis media in children <3 years of age in primary care following vaccination with

10-valent pneumococcal *Haemophilus influenzae* protein-D conjugate vaccine: a whole-population study. Clin Infect Dis 67:1213–1219. https://doi.org/10.1093/cid/ciy233.

23. The Directorate of Health. 2018. Þátttaka í almennum bólusetningum barna á Íslandi 2017: Uppgjör 2018. The Directorate of Health, Reykjavik, Iceland.