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Genetics and therapy for pediatric eye diseases

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1. Introduction

Amongst the plethora of ocular disorders, congenital anomalies and early-onset diseases have particular significance. From a clinical perspective, these pediatric conditions frequently destine children to a lifetime of severe vision impairment and/or blindness, with longlasting societal effects not least in terms of future employment. Equally, scientific importance of these largely heritable phenotypes is reflected by molecular entry points for characterizing developmental mechanisms relevant to the eye and frequently multiple other organ systems, as well as their substantial contribution to adult-onset diseases. Significance is further enhanced by the eye being an optically clear and uniquely accessible part of the central nervous system. These attributes facilitate phenotyping at the cellular level *in vivo* and have accelerated advancement of therapeutic approaches in multiple areas of medicine.

Appreciation of the mechanistic basis of pediatric disorders, presented here, is augmented by an understanding of key steps in eye development. In vertebrates, the eye's multiple components include the light-focusing cornea and lens of the anterior segment, and the posterior light-sensitive neural retina which captures, integrates and processes visual information before the optic nerve delivers it to the brain (Fig. 1A and 1B). First steps in human eye specification begin

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ABSTRACT

Ocular morphogenesis in vertebrates is a highly organized process, orchestrated largely by intrinsic genetic programs that exhibit stringent spatiotemporal control. Alternations in these genetic instructions can lead to hereditary or nonhereditary congenital disorders, a major cause of childhood visual impairment, and contribute to common late-onset blinding diseases. Currently, limited treatment options exist for clinical phenotypes involving eye development. This review summarizes recent advances in our understanding of early-onset ocular disorders and highlights genetic complexities in development and diseases, specifically focusing on coloboma, congenital glaucoma and Leber congenital amaurosis. We also discuss innovative paradigms for potential therapeutic modalities.

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during the third and fourth week of gestation. These stages are stringently regulated, frequently in a reiterative manner, by molecular networks that include eye-field transcription factors as well as Hedgehog, Retinoic acid (RA), Wingless and int-1 (Wnt), Transforming growth factor beta (TGF- β), and other signaling pathways [1,2]. After initial specification of the eye field and its subsequent Hedgehog-mediated separation into two domains, the earliest morphological evidence of ocular development is the evagination of the optic vesicles from the prosencephalon (Fig. 1C) [1,3]. Patterning along a proximal-distal axis yields the optic stalk that connects the optic vesicle to the forebrain and eventually forms the optic nerve. The distal portion of the optic vesicle later invaginates to form a bilayered optic cup, and the neural retina and retinal pigment epithelium ultimately differentiate from these laminae.

Among other essential phases of eye development is the contact of the optic vesicle with the overlying surface ectoderm, triggering a sequence of inductive events: first generating the lens placode, then the lens vesicle, and ultimately the crystalline lens that focuses incident light onto the retina. Extensive remodeling is also required for the optic cup to form an intact spherical globe, a process that involves closure of a large fissure on the ventral or inferior portion of the eye. Failure of this choroid fissure to close results in coloboma (plural colobomata) — a spectrum of pediatric defects and malformations that represent important causes of visual impairment and blindness. The tightly regulated interval during which the fissure is open provides transient intraocular access for migrating neural crest cells [4]. These stem cells delaminate from edges of the cranial neural tube, and after a lengthy migratory route, envelop the eye and contribute

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Fig. 1. Cross-sectional images of the human eye (A) and the light-sensitive retina (B). (C) Development of the human eye. Eye field specification starts at around three weeks gestation in the anterior neural plate, followed by the formation of optic vesicle, lens, cornea and retina at various stages. ONL, outer nuclear layer; OPL, outer plexiform layer; INL, inner nuclear layer; IPL, inner plexiform layer; GCL, ganglion cell layer.

to numerous extra- and intra-ocular structures. Consequently, perturbation of neural crest function results in optic fissure closure defects [5,6] as well as diverse congenital ocular diseases, including Axenfeld-Rieger syndrome and congenital glaucoma.

Another key facet of ocular development concerns the processes by which retinal progenitor cells (RPCs), derived from neuroepithelium of the optic cup, form six distinct neuronal cell types and one type of glia in an ordered and overlapping sequence (Fig. 1B and 1C). A series of transcription factors and signaling molecules endow RPCs with competence to generate specific retinal cell types [7]. Intriguingly, many of these regulatory factors sub-serve the same function in other tissues, such as the cerebral cortex, highlighting conserved determination of cell fate across diverse Central Nervous System (CNS) tissues [8]. Transcriptome profiles of developing human retina have provided new insights into temporal and regional cell fate specification by suggesting distinct trajectories of neuronal birth in the fovea versus the peripheral retina [9]. Mutations in retinal developmental genes are key causes of profound pediatric vision loss [10], resulting both in Leber congenital amaurosis (LCA) and juvenile forms of retinitis pigmentosa [11]. Identification of disease-causing genes and elucidation of respective pathogenic mechanisms offer opportunities for developing novel treatment modalities.

2. Congenital eye diseases

Congenital eye defects account for up to 60% of blindness among infants and an estimated 1.4 million visually impaired children under the age of 16 worldwide [10,12]. Among over 450 reported clinical manifestations of congenital eye disorders in the Online Mendelian Inheritance in Man database (OMIM; https://omim.org), the etiology of many remains elusive. In this review, we briefly discuss the genetic basis of three common forms of pediatric eye disease – coloboma, congenital glaucoma and LCA and describe current treatment strategies, or those in progress, to alleviate the phenotypes and/or restore vision.

2.1. Ocular coloboma

Coloboma is a congenital anomaly which is estimated to account for 11% of pediatric blindness and characterized by an inferior or ventrally located gap in one or more tissues, extending between the cornea and the optic nerve [5,6]. Cases may be unilateral or bilateral, usually with a genetic etiology, and comprise a clinical spectrum that includes congenitally reduced ocular size (microphthalmia), and in severe cases, absence of one or both eyes (anophthalmia). Patients with unilateral anophthalmia and contralateral colobomatous microphthalmia demonstrate that these disorders represent a phenotypic continuum [13]. Although the mechanisms by which coloboma-causing mutations induce unilateral disease remain undefined, their identification is expected to signify a key step in determining therapeutic targets.

Coloboma is readily explicable by perturbed morphogenesis – failure of choroid fissure fusion. The severity broadly correlates with involvement of essential retinal structures, such as the macula. Consequently, iris colobomata that primarily intensify light entry are associated with a relatively mild vision impairment (20/30 to 20/60 acuity), while those affecting the retina, and particularly the macula and optic nerve, result in profound reductions in vision (potentially 20/200 to 'counting fingers' levels) (Fig. 2A). The last two decades have seen substantial advances in deciphering the genetic bases of coloboma, which is estimated to have a heritability of at least 80% in developed countries. Interestingly, extensive genetic heterogeneity exists, with mutations in almost 40 genes molecularly explaining only a minority of cases (Table 1A). Consequently, elucidation of molecules and pathways involved in optic fissure closure continues to uncover disease-causing coloboma genes.

Ocular morphogenesis is guided by combinatorial interactions of transcription factors and gradients of signaling molecules; therefore, such components are frequently associated with coloboma (and of micro/anophthalmia) (Table 1A). For instance, Pax2 and Pax6 have essential antagonistic roles in the dorsal-ventral partitioning of the developing optic vesicle, respectively delineating the optic stalk and optic cup. Mutations of PAX2 induce optic nerve colobomata (and renal anomalies), while PAX6 mutations can lead to an extensive anomaly spectrum that includes coloboma and microphthalmia [14]. Similarly, perturbation of every phase of eve development, from eve field specification through migration of retinal progenitor cells and axis formation to the migration of the neural crest-derived periocular mesenchyme, may impair choroid fissure closure. Indeed, reflecting the fundamental nature of these processes, causative mutations have now been identified in members of most developmental pathways such as those corresponding to Hedgehog, RA, Bone morphogenetic protein (BMP), TGF- β , Fibroblast growth factor (FGF), Wnt and Hippo signaling [6,15]. The main exception appears to be Notch signaling, where ligand mutation induces coloboma in murine but leads to discrete anterior segment phenotypes in patients [16]. Genes involved in cell proliferation/migration/death signaling pathways are also involved in epithelial remodeling at the fissure, yet apoptosis has not been detected by a recent transcriptome analysis of optic fissure closure signature genes using human samples, thereby suggesting distinctions among species [15,17]. It is also important to highlight that the process of tissue fusion required for choroid fissure closure is not unique to the eye and occurs at multiple sites including the neural tube, palate and lip. Likely reflecting evolutionary parsimony, genetic pathways implicated in coloboma, neural tube defects and cleft palate are largely conserved and therefore treatment strategies developed in one tissue may have applicability to others.

2.2. Anterior segment dysgenesis

The anterior segment of the eye comprises the tissues (from cornea to lens) that lie in front of the vitreous. Their key roles include refracting and focusing incident light onto the retina and circulating aqueous humor, which is essential for maintaining clarity of the avascular cornea and lens. Maldevelopment of the anterior segment frequently results in early-onset glaucoma. One of the subtypes, congenital glaucoma, is characterized by chronic intraocular pressure (IOP) elevation. Affected infants exhibit ocular enlargement which manifests as increased corneal diameter and frequently splits in Descemet's membrane (Haab striae) [18] as well as increased lacrimation (Fig. 2B). Additional features include angle anomalies, IOP elevation to 30-50 mm of mercury, and optic disc cupping that may be partially reversible with prompt normalization of IOP [19]. Iris alterations are observed in some molecular subtypes, along with a range of systemic anomalies in syndromic cases. Clinical management of the patients is usually surgical and is reviewed in [20].

Although exhibiting some common etiologies with coloboma, congenital glaucoma can be caused by alteration of transcription factors or signaling pathways crucial for the development of the anterior segment. From a genetics standpoint, the congenital glaucoma phenotype is intriguing for its molecular diversity, which encompasses alterations in RA, TGF- β and angiopoietin signaling as well as transcriptional networks involving PAX6, FOXC1 and PITX2 [21-25] (Table 1B). These developmental pathways control the formation of anterior segment of the eye, and their perturbation impairs aqueous humor drainage and induces IOP elevation. A majority of cases can be attributed to mutations in CYP1B1 [26,27], which encodes a cytochrome P450 oxygenase for stepwise conversion of retinol (vitamin A) to retinal and then to RA [28]. Biallelic CYP1B1 mutations result in near complete loss of function of this RA-synthesizing enzyme and thus disrupt RA morphogen gradients needed for the genesis of anterior segment, perturbing the peri-ocular mesenchyme from which Coloboma



В

Congenital glaucoma



С

Leber congenital amaurosis



Fig. 2. Etiology and phenotypes of pediatric eye diseases. (A) Coloboma. Optic fissure closure defects affecting the formation of the optic nerve (upper) and iris (lower) as shown by graphic illustration (left), clinical images (middle) and fundus images (right). The blue circles and arrows highlight the approximate location of the optic fissure closure defects in the eye. In the upper panel, there is colobomatous enlargement of the optic nerve without iris involvement. In contrast, both the iris and the retina are impacted in the lower panel. (B) Congenital glaucoma. Increased intraocular pressure causes corneal enlargement with splits in Descemet's membrane. Also present are a detached posterior embryotoxon (white arrow) and irido-corneal adhesions (middle and right). (C) Leber congenital amaurosis (LCA) is characterized by dysfunction or loss light-sensitive photoreceptors. In this patient, the death of photoreceptors is caused by a loss-of-function mutation of *RPE65* leading to visual cycle defect in the retinal pigment epithelium. White arrows in the fundus images (middle and right) indicate the affected regions.

Α

 Table 1

 Genes implicated in pathogenesis of pediatric eye diseases

A Coloboma*

	1 0	<u>.</u>			
OMIM number	Gene symbol	Protein	Phenotype	Mode of inheritance	Syndromic (S) or isolated (I)
102502	1000	A still and starting of		40	<u> </u>
102560 107480	ACTG1 SALL1	Actin gamma 1 Spalt like transcription factor 1	Deafness, autosomal dominant 20/26 Townes-Brocks branchiootorenal-like syndrome; Townes-Brocks syndrome 1	AD AD	S S
107580	TFAP2A	Transcription factor AP-2 alpha	Branchiooculofacial syndrome	AD	S
107773	NR2F2	Nuclear receptor subfamily 2 group F member 2	46, XX sex reversal 5; Congenital heart defects, multiple types, 4	AD	S
109400	PTCH2	Patched 2	Basal cell nevus syndrome	AD	S
109400	PTCH1	Patched 1	Basal cell nevus syndrome	AD	S
109400	SUFU	SUFU negative regulator of hedgehog signaling	Basal cell nevus syndrome	AD	S
112262	BMP4	Bone morphogenetic protein 4	Microphthalmia, syndromic 6; Orofacial cleft 11	AD	S
113620	TFAP2A	Transcription factor AP-2 alpha	Branchiooculofacial syndrome	AD	S
120330	PAX2	Paired box 2	Papillorenal syndrome	AD	S
120433	YAP1	Yes1 associated transcriptional regulator	Coloboma, ocular, with or without hearing impairment, cleft lip/palate, and/or mental retardation	AD	S
132890	NR2F1	Nuclear receptor subfamily 2 group F member 1	Bosch-Boonstra-Schaaf optic atrophy syndrome	AD	S
142993	VSX2	Visual system homeobox 2	Microphthalmia with coloboma 3; Microphthal- mia, isolated 2	AR	Ι
145410	GBBB2/ SPECC1L	Sperm antigen with calponin homology and coiled-coil domains 1 like	Opitz GBBB syndrome, type II	AD	S
147950	FGFR1	Fibroblast growth factor receptor 1	Hypogonadotropic hypogonadism 2 with or without anosmia	AD	S
154400	SF3B4	Splicing factor 3b subunit 4	Acrofacial dysostosis 1, Nager type	AD	S
154500	TCOF1	Treacle ribosome biogenesis factor 1	Treacher Collins syndrome 1	AD	S
156845	MITF	Melanocyte inducing transcription factor	COMMAD syndrome; Tietz albinism-deafness syndrome; Waardenburg syndrome, type 2A; Waardenburg syndrome/ocular albinism, digenic	AD or AR	S
157170	SIX3	SIX homeobox 3	Holoprosencephaly 2; Schizencephaly	AD	S or I
163200	NRAS	NRAS proto-oncogene, GTPase	Schimmelpenning-Feuerstein-Mims syndrome, somatic mosaic	AD	S
163200	HRAS	HRAS proto-oncogene, GTPase	Schimmelpenning-Feuerstein-Mims syndrome, somatic mosaic	AD	S
163200	KRAS	KRAS proto-oncogene, GTPase	Schimmelpenning-Feuerstein-Mims syndrome, somatic mosaic	AD	S
164950	FGF3	Fibroblast growth factor 3	Deafness, congenital with inner ear agenesis, microtia, and microdontia	AR	S
167409	PAX2	Paired box 2	Glomerulosclerosis, focal segmental, 7	AD	S
177075	MAF	MAF bZIP transcription factor	Cataract 21, multiple types; Ayme-Gripp syndrome	AD	S or I
184429	SOX2	SRY-box transcription factor 2	Optic nerve hypoplasia and abnormalities of the central nervous system; Microphthalmia, syn- dromic 3	AD	S
191170	TP53	Tumor protein p53	Adrenocortical carcinoma, pediatric; Basal cell carcinoma 7; Choroid plexus papilloma; Colo- rectal cancer; Glioma susceptibility 1; Osteo- sarcoma; Bone marrow failure syndrome 5; Breast cancer, somatic; Hepatocellular carci- noma, somatic; Li-Fraumeni syndrome; Naso- pharyngeal carcinoma, somatic; Pancreatic cancer, somatic	AD, SMu	S
200990	KIF7	Kinesin family member 7	Joubert syndrome 12; Acrocallosal syndrome; Al- Gazali-Bakalinova syndrome; Hydrolethalus syndrome 2	AR	S
206700	ITPR1	Inositol 1,4,5-trisphosphate receptor type 1	Gillespie syndrome	AD or AR	S
206900	SOX2	SRY-box transcription factor 2	Optic nerve hypoplasia and abnormalities of the central nervous system; Microphthalmia, syn- dromic 3	AD	S
213300	INPP5E	Inositol polyphosphate-5-phosphatase E	Joubert syndrome 1	AR	S
216360	TMEM67	Transmembrane protein 67	COACH syndrome 1	AR	S
216820	SALL2	Spalt like transcription factor 2	Coloboma, ocular	AR	Ι
218340	C12orf57		Temtamy syndrome	AR	S
221900	ATOH7	Atonal bHLH transcription factor 7	Persistent hyperplastic primary vitreous	AR	Ι
235730	ZEB2	Zinc finger E-box binding homeobox 2	Mowat-Wilson syndrome	AD	S
243310	ACTB	Actin beta	CHARGE syndrome; Hypogonadotropic hypogo- nadism 5 with or without anosmia	AD	S
243605	CENPF	Centromere protein F	Stromme syndrome	AR	S
244450	UBE3B	Ubiquitin protein ligase E3B	Single median maxillary central incisor	AD	S
248190	CLDN19	Claudin 19	Hypomagnesemia 5, renal, with ocular involvement	AR	S

Table 1 (Continued)

OMIM	Gene	Protein	Phenotype	Mode of	Syndromic (S)
number	symbol			inheritance	or isolated (I)
248390	POLR1C	RNA polymerase I and III subunit C	Treacher Collins syndrome 3	AR	S
248450 263750	FKEM1 DHODH	FKAST related extracellular matrix 1 Dibydroorotate debydrogenase	wanitoba oculotrichoanal syndrome Miller syndrome	AK AR	5 5
203730	DIIODII	(quinone)	while synchonic	711	5
270420	SPINT2	Serine peptidase inhibitor, Kunitz type 2	Diarrhea 3, secretory sodium, congenital and chorioretinal coloboma	AR	S
274270	DPYD	Dihydropyrimidine dehydrogenase	Dihydropyrimidine dehydrogenase deficiency; 5- fluorouracil toxicity	AR	S
300166	BCOR	BCL6 corepressor	Microphthalmia, syndromic 2	XLD	1
300244	FLNA	Filamin A	Terminal osseous dysplasia	XLD	S
300307	GZF1	GDNF inducible zinc finger protein 1	Cleft palate with ankyloglossia	AR	S
300463	PQBPI	Polyglutamine binding protein 1	Kenpenning syndrome		S
500472	IGDF I		dation. ocular coloboma and micrognathia	ALK	3
300915	HMGB3	High mobility group box 3	Microphthalmia, syndromic 13	XL	S
309500	PQBP1	Polyglutamine binding protein 1	Renpenning syndrome	XLR	S
309800	NAA10	N-alpha-acetyltransferase 10, NatA cata- lytic subunit	Microphthalmia, syndromic 1	XL	I
600037	OTX2	Orthodenticle homeobox 2	Microphthalmia, syndromic 5; Retinal dystrophy, early-onset, with or without pituitary dysfunc- tion; Pituitary hormone deficiency, combined, 6	AD	I
600251	SPECC1L	Sperm antigen with calponin homology and coiled-coil domains 1 like	Facial clefting, oblique, 1	AD	S
600463	ALDH1A3	Aldehyde dehydrogenase 1 family mem- ber A3	Microphthalmia, isolated 8	AR	Ι
600725	SHH	Sonic hedgehog signaling molecule	Holoprosencephaly 3; Microphthalmia with colo- boma 5; Schizencephaly; Single median maxil- lary central incisor	AD	S or I
601147	GDF6	Growth differentiation factor 6	Klippel-Feil syndrome 1;	AD or AR	S
601147	GDF6	Growth differentiation factor 6	Microphthalmia with coloboma 6, digenic; Leber congenital amaurosis 17; Microphthalmia, iso- lated 4; Multiple synostoses syndrome 4	AD or AR	I
601186	STRA6	Signaling receptor and transporter of ret- inol STRA6	Microphthalmia, isolated, with coloboma 8, or syndromic 9	AR	S or I
601707	SMO	Smoothened, frizzled class receptor	Curry-Jones syndrome, somatic mosaic	Unclear	S
601723	FZD5	Frizzled class receptor 5	Microphthalmic coloboma	AD	I
601723	FZD5	Frizzled class receptor 5	Microphthalmic coloboma	AD	l
602000	FADD	Fas associated via death domain	Infections, recurrent, with encentral on the	AD	S
002437	mbb		hepatic dysfunction, and cardiovascular malformations	7 III	5
603714	SIX3	SIX homeobox 3	Holoprosencephaly 2; Schizencephaly	AD	S or I
604219	CRYAA	Crystallin alpha A	Cataract 9, multiple types	AD or AR	I
604294 605124	VAX I SDINTO	Ventral anterior nomeodox 1 Serine pentidase inhibitor. Kunita tune 2	MICTOPHTHAIMIA, SYNDTOMIC 11 Diarrhea 3, secretory sodium, congenital and	AR	l S
003124	51 1112	serine peptidase minisitor, kunitz type z	chorioretinal coloboma		5
605452	ABCB6	ATP binding cassette subfamily B mem- ber 6	Microphthalmia, isolated, with coloboma 7; dys- chromatosis universalis hereditaria 3; pseudo- hyperkalemia, familial, 2, due to red cell leak	AD	S or I
605947	PIGL	Phosphatidylinositol glycan anchor bio- synthesis class L	CHIME syndrome	AR	S
606522	GDF3	Growth differentiation factor 3	Microphthalmia with coloboma 6, or isolated 7; Klippel-Feil syndrome 3	AD	S or I
607086	AAT1	Aortic aneurysm, familial thoracic 1	Aortic aneurysm, familial thoracic 1	Unknown	S
607108	PAX6	Paired box gene 6	Coloboma of optic nerve; Coloboma, ocular; Morning glory disc anomaly; Aniridia; Anterior segment dysgenesis 5, multiple subtypes; Cata- ract with late-onset corneal dystrophy; Foveal hypoplasia 1; Keratitis; Optic nerve hypoplasia	AD	I
607906	ALG2	ALG2 alpha-1,3/1,6-mannosyltransferase	Congenital disorder of glycosylation, type li	AR	S
608166	SEMA3E	Semaphorin 3E	CHARGE syndrome	AD	S
608296	FIBP	FGF1 intracellular binding protein	Thauvin-Robinet-Faivre syndrome	AR	S
608488	SMOC1	SPARC related modular calcium binding	Microphthalmia with limb anomalies	AR	5
608553	NMNAT1	Nicotinamide nucleotide adenylyltrans- ferase 1	Leber congenital amaurosis 9	AR	I
608572	TXNL4A	Thioredoxin like 4A	Burn-McKeown syndrome	AR	S
008892	CHD7	protein 7	CHARGE syndrome; Hypogonadotropic hypogo- nadism 5 with or without anosmia	AD	3
608944	FREM1	FRAS1 related extracellular matrix 1	Bifid nose with or without anorectal and renal anomalies; Manitoba oculotrichoanal syn- drome; Trigonocephaly 2	AD or AR	S
609637	ZIC2	Zic family member 2	Holoprosencephaly 5	AD	I

Table 1 (Continued)

OMIM number	Gene symbol	Protein	Phenotype	Mode of inheritance	Syndromic (S) or isolated (I)
610256	FOXE3	Forkhead box E3	Anterior segment dysgenesis 2, multiple subtypes	AR	I
610688	TMEM67	Transmembrane protein 67	Joubert syndrome 6	AR	S
610745	STRA6	Signaling receptor and transporter of ret- inol STRA6	Microphthalmia, isolated, with coloboma 8, or syndromic 9	AR	S or I
610937	RPGRIP1L	RPGRIP1 like	COACH syndrome 3; Joubert syndrome 7; Meckel syndrome 5	Unknown	S
611254	KIF7	Kinesin family member 7	Joubert syndrome 12; Acrocallosal syndrome; Al- Gazali-Bakalinova syndrome; Hydrolethalus syndrome 2	AR	S
611499	GUSB	Glucuronidase beta	Mucopolysaccharidosis VII	AR	S
612109	HMX1	H6 family homeobox 1	Oculoauricular syndrome	AR	S
612379	SRD5A3	Steroid 5 alpha-reductase 3	Congenital disorder of glycosylation, type Iq	AR	S
612713	SRD5A3	Steroid 5 alpha-reductase 3	Kahrizi syndrome	AR	S
612779	DPYD	Dihydropyrimidine dehydrogenase	Dihydropyrimidine dehydrogenase deficiency; 5- fluorouracil toxicity	AR	S
613001	FGFR1	Fibroblast growth factor receptor 1	Encephalocraniocutaneous lipomatosis, somatic mosaic	Unknown	S
613456	ALX1	ALX homeobox 1	Frontonasal dysplasia 3	AR	S
613477	PDE6D	Phosphodiesterase 6D	Developmental and epileptic encephalopathy 5	AD	S
613477	PDF6D	Phosphodiesterase 6D	Developmental and epileptic encephalopathy 5	AD	S
613517	MCOP6/ PRSS56	Serine protease 56	Microphthalmia, isolated 6	AR	I
613674	SOX17	SRY-box transcription factor 17	Vesicoureteral reflux 3	AD	S
613717	POLR1D	RNA polymerase I and III subunit D	Treacher Collins syndrome 2	AD or AR	S
613842	GZF1	GDNF inducible zinc finger protein 1	Joint laxity short stature and myonia	AR	S
614583	ACTG1	Actin gamma 1	Baraitser-Winter syndrome 2	AD	S
615009	PACS1	Phosphofurin acidic cluster sorting pro-	Schuurs-Hoeijmakers syndrome	AD	S
615113	ALDH1A3	Aldehyde dehydrogenase 1 family mem- ber A3	Microphthalmia, isolated 8	AR	Ι
615140	C12orf57	berns	Temtamy syndrome	AR	S
615145	TENM3	Teneurin transmembrane protein 3	Microphthalmia, isolated, with coloboma 9, or syndromic 15	AR	S or I
615147	RBP4	Retinol binding protein 4	Retinal dystrophy, iris coloboma, and comedo- genic acne syndrome	AR	S or I
615665	PDF6D	Phosphodiesterase 6D	Joubert syndrome 22	AR	S
615665	PDF6D	Phosphodiesterase 6D	Joubert syndrome 22	AR	s
615877	MAB21L2	Mab-21 like 2	Microphthalmia/coloboma and skeletal dysplasia	AD, AR	S
615877	MAB21L2	Mab-21 like 2	Microphthalmia/coloboma and skeletal dysplasia syndrome	AD or AR	S or I
616428	RBP4	Retinol binding protein 4	Microphthalmia, isolated, with coloboma 10	AD	I
616490	KIAA0586	KIAA0586	Joubert syndrome 23	AR	S
616722	MIR204	microRNA 204	Retinal dystrophy and iris coloboma with or without cataract	AD	I
616789	MED13L	Mediator complex subunit 13L	Mental retardation and distinctive facial features with or without cardiac defects	AD	S
617107	FIBP	FGF1 intracellular binding protein	Thauvin-Robinet-Faivre syndrome	AR	S
617662	GZF1	GDNF inducible zinc finger protein 1	Joint laxity, short stature and myonia	AR	S
618586	WDR37	WD repeat domain 37	Neurooculocardiogenitourinary syndrome	AD	S
618652	WDR37	WD repeat domain 37	Neurooculocardiogenitourinary syndrome	AD	S
618939	POLR1R	RNA polymerase I subunit B	Treacher-Collins syndrome 4	AD	S
619113	RPGRID11	RPCRIP1 like	COACH syndrome 3: Joubert syndrome 7: Meckel	Unknown	S
515115	M GMI IL	A SAILT INC	syndrome 5	OIIKIIUWII	5

*There are 317 entries in OMIM for coloboma. This table only lists those with strong association to coloboma or diseases with coloboma as one of the major clinical characteristics. AD, autosomal dominant; AR, autosomal recessive; DR, digenic recessive; SMu, Somatic mutation; XL, X-linked; XLD, X-linked dominant; XLR, X-linked recessive.

the ciliary body, trabecular meshwork, iris, corneal stroma and endothelium are derived. RA also directly regulates *FOXC1* and *PITX2* [29], two transcription factors with fundamental roles in anterior segment development [21,23,24], likely explaining why ocular anomalies induced by *FOXC1* and *PITX2* mutations can phenocopy congenital glaucoma.

As a recessively inherited disorder, the incidence of congenital glaucoma is strongly influenced by geographic prevalence of consanguinity. Consanguineous cases are frequently bilateral, and more often associated with greater disease severity. In these patients, higher IOPs and the associated decrease in corneal clarity may preclude angle surgery (called goniotomy), necessitating more complex surgical approaches. Non-consanguineous cases are more often unilateral and amenable to goniotomy, suggesting that modifier loci may contribute to phenotypic severity. Notably, intermediate levels of RA resulting from a heterozygous *CYP1B1* mutation induce milder disease, with such mutations explaining 5% of the related juvenile open-angle glaucoma phenotypes diagnosed between 5 and 40 years of age [30]. Thus, studies of early-onset and comparatively extreme pediatric phenotypes exemplify the power to decipher genetic mechanisms in common later-onset diseases.

A similar paradigm is illustrated by angiopoietin-Tie (ANG-TIE) signaling pathway, an essential regulator of blood and lymphatic development, in the genesis of congenital glaucoma. Mouse mutants with conditional deletion of the Angiopoietin 1 and 2 (Angpt1/2) ligands that bind to the TIE receptors develop ocular enlargement and optic neuropathy, in addition to complete absence of Schlemm's canal, a modified lymphatic that drains aqueous humor from the eye

Table 1b
Congenital glaucoma#

OMIM number	Gene symbol	Protein	Phenotype	Mode of inheritance	Syndromic (S) or isolated (I)
105650	RPS19	Ribosomal protein S19	Diamond-Blackfan anemia 1	AD	S
137600	PITX2	Paired like homeodomain 2	Anterior segment dysgenesis 4	AD	Ι
175780	COL4A1	Collagen type IV alpha 1 chain	Brain small vessel disease with or without ocular anomalies including congenital glaucoma	AD	S
180849	CREBBP	CREB binding protein	Rubinstein-Taybi syndrome 1	AD	S
236670	POMT1	Protein O-mannosyltransferase 1	Muscular dystrophy-dystroglycanopathy (con- genital with brain and eye anomalies), type A, 1; Muscular dystrophy-dystroglycanopathy (congenital with mental retardation), type B, 1; Muscular dystrophy-dystroglycanopathy (limb-girdle), type C, 1	AR	S
249420	SH3PXD2B	SH3 and PX domains 2B	Frank-ter Haar syndrome with or without glaucoma	AR	S
251750	LTBP2	Latent transforming growth factor beta binding protein 2	Weill-Marchesani syndrome 3; Glaucoma 3, pri- mary congenital, D; Microspherophakia and/or megalocornea, with ectopia lentis and with or without secondary glaucoma	AR	S or I
253280	POMGNT1	Protein O-linked mannose N-acetylglu- cosaminyltransferase 1 (beta 1,2-)	Muscular dystrophy-dystroglycanopathy (con- genital with brain and eye anomalies), type A, 3	AR	S
600221	TEK	TEK receptor tyrosine kinase	Glaucoma 3, primary congenital, E; Venous mal- formations, multiple cutaneous and mucosal	AD	S or I
601090	FOXC1	Forkhead box C1	Anterior segment dysgenesis 3, multiple sub- types; Axenfeld-Rieger syndrome, type 3	AD	S or I
601631	FOXC1	Forkhead box C1	Anterior segment dysgenesis 3, multiple sub- types; Axenfeld-Rieger syndrome, type 3	AD	S or I
601652	MYOC	Myocilin	Glaucoma 1A, primary open angle	AD	Ι
601771	CYP1B1	Cytochrome P450 family 1 subfamily B member 1	Anterior segment dysgenesis 6, multiple sub- types; Glaucoma 3A, primary open angle, con- genital, juvenile, or adult onset	AR	Ι
602091	LTBP2	Latent transforming growth factor beta binding protein 2	Weill-Marchesani syndrome 3; Glaucoma 3, pri- mary congenital, D; Microspherophakia and/or megalocornea, with ectopia lentis and with or without secondary glaucoma	AR	S or I
603474	RPS19	Ribosomal protein S19	Diamond-Blackfan anemia 1	AD	S
604563	SBF2	SET binding factor 2	Charcot-Marie-Tooth disease, type 4B2	AR	S
607423	POMT1	Protein O-mannosyltransferase 1	Muscular dystrophy-dystroglycanopathy (con- genital with brain and eye anomalies), type A, 1; Muscular dystrophy-dystroglycanopathy (congenital with mental retardation), type B, 1; Muscular dystrophy-dystroglycanopathy (limb-girdle), type C, 1	AR	S
610192	GLIS3	GLIS family zinc finger 3	Diabetes mellitus, neonatal, with congenital hypothyroidism. Additional features include congenital glaucoma	AR	S
610199	GLIS3	GLIS family zinc finger 3	Diabetes mellitus, neonatal, with congenital hypothyroidism. Additional features include congenital glaucoma	AR	S
613150	POMT2	Protein O-mannosyltransferase 2	Muscular dystrophy-dystroglycanopathy (con- genital with brain and eye anomalies), type A, 2	AR	S
613293	SH3PXD2B	SH3 and PX domains 2B	Frank-ter Haar syndrome with or without glaucoma	AR	S
617315	CYP1B1	Cytochrome P450 family 1 subfamily B member 1	Anterior segment dysgenesis 6, multiple sub- types; Glaucoma 3A, primary open angle, con- genital invenile or adult onset	AR	Ι

#There are 55 entries in Online Mendelian Inheritance in Man (OMIM, https://www.ncbi.nlm.nih.gov/omim) for congenital glaucoma. This table only lists those with strong association to congenital glaucoma or diseases with congenital glaucoma as one of the major clinical features. AD, autosomal dominant; AR, autosomal recessive; XLR, X-linked recessive.

[31]. Heterozygous *TIE2* mutations cause ~5% of congenital glaucoma [32], with a few disrupting TIE2 receptor clustering that is required for pathway activation [33]. *TIE2* haploinsufficiency can recapitulate variable expressivity of *CYP1B1*-mediated congenital glaucoma in terms of age of onset and unilateral disease prevalence [32]. Subsequent investigations have identified *ANGPT1* mutations in congenital glaucoma and, equally importantly, suggest the requirement of ANG-TIE signaling for maintaining the Schlemm's canal [34,35]. Intriguingly, aging-associated decline in functional integrity of Schlemm's canal can be rescued by an agonistic Tie2 antibody, at least in mice [34]. In light of Schlemm's canal's central role, additional ANG-TIE components are predicted to be involved in glaucoma pathogenesis, offering novel targets for intervention.

2.3. Leber congenital amaurosis

In vertebrates, photons are captured by light-sensitive photoreceptors in the neural retina, integrated and processed by interneurons (the horizontal, bipolar and amacrine cells) and transmitted to the brain through the optic nerve composed of the retinal ganglion cell axons and glia (Fig. 1B). Vision loss stems from obstruction of the light path to the neural retina and inability of the retina to detect

Table 1c	
Leber Congenital Amaurosis (LCA)	^

OMIM number	Gene symbol	Protein	Phenotype	Mode of inheritance	Syndromic (S) or isolated (I)
204000 601777	GUCY2D	Guanylate cyclase 2D, retinal	Leber congenital amaurosis 1 Cone-rod dystrophy 6	AD or AR	Ι
204100 613794	RPE65	Retinoid isomerohydrolase RPE65	Leber congenital amaurosis 2 Retinitis pigmentosa 20	AR	Ι
618697			Retinitis pigmentosa 87 with choroidal involvement	AD	
604232	SPATA7	Spermatogenesis associated 7	Leber congenital amaurosis 3; Retinitis pigmen- tosa, juvenile	AR	Ι
604393	AIPL1	Aryl hydrocarbon receptor interacting protein like 1	Cone-rod dystrophy; Retinitis pigmentosa, juve- nile; Leber congenital amaurosis 4	AD or AR	Ι
604537	LCA5	Lebercilin LCA5	Leber congenital amaurosis 5	AR	Ι
613826	RPGRIP1	RPGR interacting protein 1	Leber congenital amaurosis 6	AR	Ι
613829	CRX	Cone-rod homeobox	Leber congenital amaurosis 7	AD	Ι
613835	CRB1	Crumbs cell polarity complex component	Leber congenital amaurosis 8	AR	Ι
600105		1	Retinitis pigmentosa-12		
608553	NMNAT1	Nicotinamide nucleotide adenylyltrans- ferase 1	Leber congenital amaurosis 9	AR	Ι
611755	CEP290	Centrosomal protein 290	Leber congenital amaurosis 10	AR	I or S
610188			Joubert syndrome 5		
610189			Senior-Loken syndrome 6		
613837	IMPDH1	Inosine monophosphate dehydrogenase	Leber congenital amaurosis 11	AR	Ι
180105		1	Retinitis pigmentosa 10	AD	
610612	RD3	RD3 regulator of GUCY2D	Leber congenital amaurosis 12	AR	Ι
612712	RDH12	Retinol dehydrogenase 12	Leber congenital amaurosis 13	AD or AR	Ι
613341	LRAT	Lecithin retinol acyltransferase	Retinitis pigmentosa, juvenile; Leber congenital amaurosis 14; Retinal dystrophy, early-onset severe	AR	I
613843 600132	TULP1	TUB like protein 1	Leber congenital amaurosis 15	AR	Ι
614186	KCNJ13	Potassium inwardly rectifying channel subfamily J member 13	Leber congenital amaurosis 16	AR	Ι
615360	GDF6	Growth differentiation factor 6	Leber congenital amaurosis 17	AR	Ι
608133	PRPH2	Peripherin 2	Leber congenital amaurosis 18; Retinitis pigmen- tosa 7 and digenic form	AD or AR	I or S
169150			Macular dystrophy, patterned, 1		
618513	USP45	Ubiquitin specific peptidase 45	Leber congenital amaurosis 19	AR	Ι
617879	TUBB4B	Tubulin beta 4B class IVb	Leber congenital amaurosis with early-onset deafness	AD	S
609237	IQCB1	IQ motif-containing protein B1	Leber congenital amaurosis	AR	I or S
609254			Senior-Loken syndrome 5	AR	

[^]There are 101 entries in OMIM (https://www.ncbi.nlm.nih.gov/omim) for LCA. This table only lists the genes with strong association to LCA or diseases with LCA as one of the major clinical attributes. AD, autosomal dominant; AR, autosomal recessive; DR, digenic recessive; XL, X-linked.

and/or transmit light-triggered signals to the brain. In retinal degenerative diseases, the irreversible blindness is largely attributable to dysfunction or death of photoreceptor cells [11] (Fig. 2C). Amongst such disorders, LCA is a group of severe and early-onset retinal diseases responsible for childhood blindness [36]. LCA patients exhibit profound visual impairment at infancy or in childhood, with almost undetectable full field electroretinogram, nystagmus (eye movements), poor pupillary light responses, and oculodigital reflex [37], [38]. LCA accounts for approximately 5% of all retinal dystrophies, affecting 2–3 per 100,000 live births with an estimated prevalence of 180,000 individuals worldwide [37].

Molecular diagnosis of LCA can be challenging because of phenotypic variability and genetic heterogeneity. Mutations in at least 28 genes have been reported to cause 70-75% of LCA, which is largely inherited as an autosomal recessive trait though dominant mutations have been reported in *IMPDH1*, *OTX2* and *CRX* genes [39] (Table 1C). LCA genes can be categorized in discrete functional classes, including transcriptional regulation and primary cilia (the photoreceptor outer segment is a modified cilium) biogenesis and transport. Pleiotropic effects of mutations in LCA genes add another level of genetic complexity; e.g., defects in cilia-associated genes (such as *CEP290*) can lead to syndromic diseases that usually include photoreceptor degeneration [40]. Genetic modifiers can also influence clinical features of ciliopathy gene defects; the phenotype of *CEP290* mutations is amenable modifications by its interaction partners [41,42]. Although the rd16 mouse model phenocopies CEP290-LCA with rapid degeneration of photoreceptors [43], a humanized knock-in mouse model carrving most common CEP290-LCA the mutation (c.2991 + 1655A > G) does not show obvious phenotypes [44], suggesting unique mechanisms of disease pathogenesis in humans. Notably, retinal organoids differentiated from induced pluripotent stem cells (iPSCs) of a CEP290-LCA patient demonstrate disease-associated cilia findings in vitro [45]. Persistence of photoreceptor cell bodies in some LCA patients with advanced disease, despite undetectable visual function, suggest possibilities of vision restoration by targeted therapies [38].

3. Therapeutic strategies

As a transparent, compartmentalized and immune-privileged organ, it is self-evident why the eye offers exceptional opportunities for evaluating different treatment paradigms. However, designing treatment for congenital eye disease is still challenging. This reflects clinical presentation months after the period of perturbed *in utero* development, the involvement of multiple ocular tissues and cell types, as well as tremendous genetic heterogeneity. Each of the three diseases discussed here poses unique complexities regarding the timing and methods of delivery to the therapeutic targets. Retinal dystrophies are currently the most tractable for correction, benefitting from many decades of fundamental research in retinal genetics and cell biology, and the retina comprising of a single tissue. Despite its fiendishly complex neuronal composition and synaptic organization, the retina has been a focus for therapeutic trials because of its unique accessibility within the CNS. In contrast, congenital glaucoma involves anterior and posterior segment tissues, requiring therapeutic modification of multiple cell types including dysfunctional and dving lateral geniculate nucleus-projecting retinal ganglion cells. The challenges are further augmented in colobomata, which at a minimum manifest many months after arising during early development. Prevention of colobomata requires the identification of disease-causing genetic profiles and their precise correction before the initiation of the optic fissure closure at gestation week 16; such corrections are not feasible currently since tissue regeneration would be required at an unprecedented scale, with existing stem cells coaxed into repopulating the affected area. Cell transplantation represents another theoretical approach; however, it is unclear whether signaling cues required for donor cell engraftment, division and differentiation persist to recapitulate such lengthy periods of normal development. As a result, the following section largely focuses on therapeutic approaches developed in retinal disorders (Table 2), which provide a broader exemplar for the CNS, and that in time may be extended to other pediatric eye disorders. Considerable progress has been made, in part, because of a longer window of intervention opportunity for retinal diseases.

3.1. Gene therapy

Gene therapy harnesses different vectors/vehicles for delivering desired gene products into affected tissues and/or cell types. A widely used approach for gene delivery in eye tissues is the use of viral vectors simply by injection at the preferred site and with low risk of immune response [46]. The low rate of integration into the host genome makes adeno-associated viral (AAV) vectors a promising platform for gene therapy [47]. One successful example of this

Table 2

Pros and cons of major therapeutic approaches

approach is the first FDA-approved drug for treatment of LCA caused by RPE65 loss-of-function mutations [38,48]; however, we should mention that the long-term data from clinical trials have been less encouraging [49]. A second potentially exciting approach is CRISPR/ Cas9-mediated genome editing [50], which can potentially correct disease-causing mutations in multiple scenarios (from retinal explants, humanized mice, non-human primates to patient iPSCderived retinal organoids) [51]. However, the technique is currently constrained by limited editing efficiency [52] and off-target mutations that include induced chromosomal anomalies [53]. Another promising methodology is use of antisense oligonucleotides (AON) [54], which induce quite persistent suppression of pathological RNA transcripts by exon skipping and other mechanisms. Several of these are in therapeutic use for pediatric neurological disorders such as Duchenne Muscular Dystrophy and Spinal Muscular Atrophy, while AON-based therapy for CEP290-LCA has yielded encouraging results [55], with vision improvement without serious adverse effect reported in one clinical trial [56].

A key limitation of gene therapy for congenital eye diseases is the temporal window for effective treatment. AAV vectors cannot reach the target cells of fetus, and a vast majority of early-onset disorders already exhibit severe developmental defects or cell loss at birth [5]. The small packaging limit of AAV (<5kb) also restricts its application for diseases caused by larger genes. In such cases, alternative approaches include gene augmentation by delivering parts of the gene [57,58], use of lentiviral vectors with larger packaging capacity [59], or splitting the transgene into two separate AAV vectors [60]; however, the efficiency and/or safety of these approaches in humans require further investigations. In any case, it would be time-consuming and currently prohibitively expensive to tailor gene therapy for each causative mutation, particularly since a therapy effective for one mutation may not be readily extrapolated to phenotypes caused by another [61]. Thus, innovative mutation-independent strategies are needed to maintain cell survival or restore visual function. One encouraging example is provided by CRISPR-mediated knockdown of a key transcriptional regulator Nrl, which has generated longer

	Successful examples	Strengths	Limitations
Gene therapy	 FDA approved the first gene therapy drug Luxturna for RPE65-LCA CRISPR/Cas9-mediated genome edit- ing precisely corrected disease-caus- ing mutations in preclinical models of CEP290-LCA Antisense oligonucleotide-based ther- apy partially restored CEP290-LCA patient vision in one clinical trial 	 High specificity to mutations and relevant tissues Clear evidence of clinical efficacy Low risk of immune response in the eyes 	 Packing limit of adeno-associated viruses (AAV) Complexity and cost of manufacturing and production Unclear effect of long-term expression of genome editors or augmented genes Off-target effect of genome editors Accessibility to correct early-onset congenital diseases
Large and small molecule drugs	 Readthrough drug PCT124 in Phase II clinical trial for treatment of <i>PAX6</i>-coloboma Therapeutic antibody against vascular endothelial growth factor (VEGF) in Phase IV clinical trial for treatment of age-related macular degeneration 	 Small molecule Ease in administration and dosage control High scale of synthesis Low cost Be able to cross blood barriers or placenta Be able to target multiple tissues simultaneously Large molecule (antibody) High specificity High stability 	 Low tissue specificity High demand on physiology-relevant models to evaluate pharmacological effects and pharmacokinetics
Cell replacement therapy	 Transplantation of stem cell-derived retinal pigment epithelium in preclini- cal and clinical trials for age-related macular degeneration 	Large treatment window	 Consistency and quality of cell-based therapies Safety and ethical issues of cell source Exchange of cytoplasmic materials between the host and graft cells

survival of (dysfunctional) rod photoreceptors while preserving the cones that are essential for the macula and thus fine vision [62]. Viral-mediated expression of rod-derived cone viability factor (RdCVF) also shows promising effect in the maintenance of cone and rod photoreceptors in multiple mouse models of retinal degeneration [63]. Optogenetic therapies deliver light-activated ion channels to surviving retinal cell types (bipolar cell and retinal ganglion cells), restoring a degree of photosensitivity in animal models [64,65]; and clinical trials are in progress. In a related manner, engraftment of optogenetically engineered photoreceptors has achieved partial recovery of visual function in the murine retina [66]. Such results are naturally very encouraging; however, it remains to be determined whether this promise translates into long-lasting restoration of retinal function in humans.

3.2. Large and small molecule drugs

Small molecule drugs offer an effective and widely used approach, reflecting ease in administration and dosage control, stability, scale of synthesis and low cost [67]. Importantly, the ability of many small molecule drugs to cross the blood-brain (or even placental) barrier may facilitate treatment of early-onset ocular/neurological diseases. Pharmaceutical modulation of signaling pathways contributing to cell death is a more specific way to preserve their survival and function [68]. Photoswitchable ion channel blockers can also offer potential for restoring vision [69]. Small molecule drugs have been applied for reading through a stop codon, correcting the structure of mutated proteins or circumventing functions of abnormal proteins [5,70]. Notably, the readthrough drug PCT124 was reported to be effective in a Pax6 mutation iris coloboma model, leading to initiation of a phase II clinical trial [5]. Such approaches hold great potential for treating developmental defects caused by misregulation of signaling pathways, such as the ANG-TIE signaling pathway for congenital glaucoma. Antioxidants (e.g., vitamin A, vitamin B3, docosahexaenoic acid, lutein), anti-apoptotic factors (e.g., tauroursodeoxycholic acid, rasagiline, norgestrel, and myriocin) and neurotrophic factors (e.g., ciliary neurotrophic factor (CNTF), Brain-derived neurotrophic factor (BDNF)) have been evaluated in the treatment of retinal degenerative diseases [40]. Therapeutic antibodies have been extensively used to neutralize bioactive factors, as illustrated by intravitreally administered monoclonals to vascular endothelial growth factor (VEGF) that are effective in treatments of neovascular age-related macular degeneration [71].

A major challenge for developing relevant drug targets is identification of appropriate molecules with excellent pharmacological benefit and pharmacokinetics and low off-target effects [67], especially in case of small molecules that can penetrate various tissues. However, ninety percent of drug candidates fail to progress from Phase I trials to clinical use [72], partly because a majority of the drugs are identified using adherent cell culture or small animal models, which, although offering valuable mechanistic insights, do not fully recapitulate human pathobiology. Recent advances in three-dimensional human retinal organoids that structurally and functionally, at least in part, mimic *in vivo* tissues can provide a promising platform for complementing the existing strategies for identifying drug candidates [73]. A recent breakthrough of deep-learning program for determining three-dimensional shapes of proteins without crystallography should accelerate the process of drug design and discovery [74].

3.3. Cell replacement therapy

When affected cells are lost or grossly abnormal at infancy, regenerative medicine may offer a plausible approach for restoring at least partial vision. A few attempts have been made to stimulate regeneration of lost cells from other cell types [75,76], whereas others have generated desired cell types from pluripotent stem cells and

transplanted the products into the eye [77]. In LCA and early-onset retinal degeneration, the need to replace photoreceptors for restoring vision requires donor cell survival, maturation (including development of the outer segment) and functional integration to form synapses with host retinal interneurons. Transplantation of photoreceptors was previously demonstrated to improve visual function in animal models, yet recent studies indicate transfer of cytoplasmic material between the donor and host cells, potentially offering unanticipated opportunities for therapeutic delivery [73,78]. In contrast, transplantation of stem cell-derived retinal pigment epithelium that can be produced at high efficiency and purity offers hope in preclinical and clinical trials for age-related macular degeneration [79,80]. In congenital glaucoma, the loss of retinal ganglion cells (RGCs) requires the elongation of axons, integration into the optic nerve and projection to the lateral geniculate nucleus. Despite efficient generation of functional RGCs from pluripotent stem cells, transplantation of these cells has yet to yield desirable results, with extensive investigations continuing in preclinical models [81].

A major concern in using iPSC-derived products is related to genomic stability [82]. Although no adverse effects are reported at this early stage, possible deleterious outcomes, such as tumor formation or degeneration of transplanted cells, may take a longer time frame to unravel. Embryonic stem cells have raised ethical concerns and may trigger immune response, whereas the use of iPSCs may not be feasible for congenital diseases. CRISPR-based approaches have also been utilized for correcting mutations in patient iPSC-derived retinal organoids [83]. Given the immense interest in stem cells, rapid advancements are expected in generation of photoreceptors from stem cells [84] and in gene-editing. Further investigations are necessary to overcome numerous challenges for correct functional integration of transplanted cells into the host retina with minimal undesirable consequences [85].

Outstanding questions

Several questions remain. Can we accelerate gene discovery for pediatric eye diseases? Is high variability of congenital eye phenotypes caused by gene/pathway interactions? How to efficiently and safely design prevention or treatment paradigms for congenital diseases *in utero*? Is it possible to selectively manipulate signaling pathways to expedite and improve the specificity of treatments? What would criteria/standards be for devising such therapies for distinct early onset diseases affecting the children?

Search strategy and selection criteria

The references were identified using Google Scholar and PubMed search engines with the search terms "coloboma" or "congenital glaucoma" or "Leber congenital amaurosis" or "congenital eye diseases" or "coloboma AND genetics" or "congenital glaucoma AND genetics" or "Leber congenital amaurosis AND genetics" or "gene therapy AND congenital eye diseases" or "small molecule AND congenital eye diseases" or "cell replacement therapy AND congenital eye diseases" or "human eye development" or "treatment AND congenital diseases". Disease-associated genes were identified by the Online Mendelian Inheritance in Man database (OMIM; https://www.ncbi.nlm.nih.gov/ omim) using keywords "coloboma" or "congenital glaucoma" or "Leber congenital amaurosis" and further selected using Google Scholar and Genecards engines with the search terms "[gene name] AND [disease]". All references were considered with preference for the most recently published works.

Conclusions

Congenital ocular diseases exhibit extensive genotypic and phenotypic heterogeneity. Despite identification of many diseasecausing genes, genetic defects remain to be discovered in a large number of congenital eye diseases. We believe a gene-independent approach will be desirable for developing therapies of such a divergent cohort. A network-based approach by modulating pathways associated with congenital eye diseases and a combination of gene and small-molecule based therapies would likely have promising impact on treating early-onset eye diseases. Nonsense suppression therapy, neurotrophic and antiapoptotic as well as other small molecule drugs might help in maintaining the survival of defective cells and achieve at least a partially desirable treatment outcome.

Declaration of Competing Interest

All authors declare that they have no competing interests.

Contributors

All authors conceived the original draft and edited the final manuscript. All authors read and approved the final version of the manuscript.

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References

- [1] Graw J. Eye development. Curr Top Dev Biol 2010;90:343-86.
- [2] Cardozo MJ, Almuedo-Castillo M, Bovolenta P. Patterning the vertebrate retina with morphogenetic signaling pathways. Neuroscientist 2020;26(2):185–96.
- [3] Miesfeld JB, Brown NL Eye organogenesis: a hierarchical view of ocular development. Curr Top Dev Biol 2019;132:351–93.
- [4] Heavner W, Pevny L. Eye development and retinogenesis. Cold Spring Harb Perspect Biol 2012;4(12).
- [5] Ohuchi H, Sato K, Habuta M, Fujita H, Bando T. Congenital eye anomalies: more mosaic than thought? Congenit Anom (Kyoto) 2019;59(3):56–73.
- [6] Patel A, Sowden JC. Genes and pathways in optic fissure closure. Semin Cell Dev Biol 2019;91:55–65.
- [7] Kohwi M, Doe CQ. Temporal fate specification and neural progenitor competence during development. Nat. Rev Neurosci 2013;14(12):823–38.
- [8] Manuel MN, Mi D, Mason JO, Price DJ. Regulation of cerebral cortical neurogenesis by the Pax6 transcription factor. Front Cell Neurosci 2015;9:70.
- [9] Hoshino A, Ratnapriya R, Brooks MJ, Chaitankar V, Wilken MS, Zhang C, et al. Molecular anatomy of the developing human Retina. Dev Cell 2017;43:763–79.
- [10] Graw J. The genetic and molecular basis of congenital eye defects. Nat Rev Genet 2003;4(11):876–88.
- [11] Wright AF, Chakarova CF, Abd El-Aziz MM, Bhattacharya SS. Photoreceptor degeneration: genetic and mechanistic dissection of a complex trait. Nat Rev Genet 2010;11(4):273–84.
- [12] Gogate P, Gilbert C, Zin A. Severe visual impairment and blindness in infants: causes and opportunities for control. Middle East Afr J Ophthalmol 2011;18 (2):109–14.
- [13] Warburg M. Genetics of microphthalmos. Int Ophthalmol 1981;4(1-2):45–65.
- [14] Azuma N, Yamaguchi Y, Handa H, Tadokoro K, Asaka A, Kawase E, et al. Mutations of the PAX6 gene detected in patients with a variety of optic-nerve malformations. Am J Hum Genet 2003;72(6):1565–70.
- [15] ALSomiry AS, Gregory-Evans CY, Gregory-Evans K. An update on the genetics of ocular coloboma. Human Genet 2019;138(8-9):865–80.
- [16] Rowan S, Conley KW, Le TT, Donner AL, Maas RL, Brown NL. Notch signaling regulates growth and differentiation in the mammalian lens. Dev Biol 2008;321 (1):111–22.
- [17] Patel A, Anderson G, Galea G, Balys M, Sowden JC. A molecular and cellular analysis of human embryonic optic fissure closure related to the eye malformation coloboma. Development; 2020.

- [18] Badawi AH, Al-Muhaylib AA, Al Owaifeer AM, Al-Essa RS, Al-Shahwan SA. Primary congenital glaucoma: an updated review. Saudi J Ophthalmol 2019;33(4):382–8.
- [19] He Z, Bui BV, Vingrys AJ. The rate of functional recovery from acute IOP elevation. Investig Ophthalmol Vis Sci 2006;47(11):4872–80.
- [20] Ko F, Papadopoulos M, Khaw PT. Primary congenital glaucoma. Prog Brain Res 2015;221:177–89.
- [21] Gage PJ, Camper SA. Pituitary homeobox 2, a novel member of the bicoid-related family of homeobox genes, is a potential regulator of anterior structure formation. Hum Mol Genet 1997;6(3):457–64.
- [22] Mears AJ, Jordan T, Mirzayans F, Dubois S, Kume T, Parlee M, et al. Mutations of the forkhead/winged-helix gene, FKHL7, in patients with Axenfeld-Rieger anomaly. Am J Hum Genet 1998;63(5):1316–28.
- [23] Smith RS, Zabaleta A, Kume T, Savinova OV, Kidson SH, Martin JE, et al. Haploinsufficiency of the transcription factors FOXC1 and FOXC2 results in aberrant ocular development. Hum Mol Genet 2000;9(7):1021–32.
- [24] Lehmann OJ, Ebenezer ND, Jordan T, Fox M, Ocaka L, Payne A, et al. Chromosomal duplication involving the forkhead transcription factor gene FOXC1 causes iris hypoplasia and glaucoma. Am J Hum Genet 2000;67(5):1129–35.
- [25] Jamieson RV, Perveen R, Kerr B, Carette M, Yardley J, Heon E, et al. Domain disruption and mutation of the bZIP transcription factor, MAF, associated with cataract, ocular anterior segment dysgenesis and coloboma. Hum Mol Genet 2002;11 (1):33–42.
- [26] Stoilov I, Akarsu AN, Sarfarazi M. Identification of three different truncating mutations in cytochrome P4501B1 (CYP1B1) as the principal cause of primary congenital glaucoma (Buphthalmos) in families linked to the GLC3A locus on chromosome 2p21. Hum Mol Genet 1997;6(4):641–7.
- [27] Bejjani BA, Lewis RA, Tomey KF, Anderson KL, Dueker DK, Jabak M, et al. Mutations in CYP1B1, the gene for cytochrome P4501B1, are the predominant cause of primary congenital glaucoma in Saudi Arabia. Am J Hum Genet 1998;62(2):325– 33.
- [28] Chambers D, Wilson L, Maden M, Lumsden A. RALDH-independent generation of retinoic acid during vertebrate embryogenesis by CYP1B1. Development 2007;134(7):1369–83.
- [29] Matt N, Dupe V, Garnier JM, Dennefeld C, Chambon P, Mark M, et al. Retinoic aciddependent eye morphogenesis is orchestrated by neural crest cells. Development 2005;132(21):4789–800.
- [30] Melki R, Colomb E, Lefort N, Brezin AP, Garchon HJ. CYP1B1 mutations in French patients with early-onset primary open-angle glaucoma. J Med Genet 2004;41 (9):647–51.
- [31] Thomson BR, Heinen S, Jeansson M, Ghosh AK, Fatima A, Sung HK, et al. A lymphatic defect causes ocular hypertension and glaucoma in mice. J Clin Invest 2014;124(10):4320–4.
- [32] Souma T, Tompson SW, Thomson BR, Siggs OM, Kizhatil K, Yamaguchi S, et al. Angiopoietin receptor TEK mutations underlie primary congenital glaucoma with variable expressivity. J Clin Invest 2016;126(7):2575–87.
- [33] Leppanen VM, Saharinen P, Alitalo K. Structural basis of Tie2 activation and Tie2/ Tie1 heterodimerization. Proceed Natl Acad Sci USA 2017;114(17):4376–81.
- [34] Kim J, Park DY, Bae H, Park DY, Kim D, Lee CK, et al. Impaired angiopoietin/Tie2 signaling compromises Schlemm's canal integrity and induces glaucoma. J Clin Invest 2017;127(10):3877–96.
- [35] Thomson BR, Souma T, Tompson SW, Onay T, Kizhatil K, Siggs OM, et al. Angiopoietin-1 is required for Schlemm's canal development in mice and humans. J Clin Invest 2017;127(12):4421–36.
- [36] Kumaran N, Moore AT, Weleber RG, Michaelides M. Leber congenital amaurosis/ early-onset severe retinal dystrophy: clinical features, molecular genetics and therapeutic interventions. Br J Ophthalmol 2017;101(9):1147–54.
- [37] Koenekoop RK. An overview of Leber congenital amaurosis: a model to understand human retinal development. Surv Ophthalmol 2004;49(4):379–98.
- [38] Cideciyan AV, Jacobson SG. Leber Congenital Amaurosis (LCA): Potential for improvement of vision. Invest Ophthalmol Vis Sci 2019;60(5):1680–95.
- [39] Kondkar AA, Abu-Amero KK. Leber congenital amaurosis: Current genetic basis, scope for genetic testing and personalized medicine. Exp Eye Res 2019;189:107834.
- [40] Chen HY, Kelley RA, Li T, Swaroop A. Primary cilia biogenesis and associated retinal ciliopathies. Semin Cell Dev Biol 2020.
- [41] Rachel RA, Li T, Swaroop A. Photoreceptor sensory cilia and ciliopathies: focus on CEP290, RPGR and their interacting proteins. Cilia 2012;1(1):22.
- [42] Ramsbottom S, Miles C, Sayer J. Murine Cep290 phenotypes are modified by genetic backgrounds and provide an impetus for investigating disease modifier alleles. F1000Research. 2015;4:590.
- [43] Chang B, Khanna H, Hawes N, Jimeno D, He S, Lillo C, et al. In-frame deletion in a novel centrosomal/ciliary protein CEP290/NPHP6 perturbs its interaction with RPGR and results in early-onset retinal degeneration in the rd16 mouse. Hum Mol Genet 2006;15(11):1847–57.
- [44] Garanto A, van Beersum SE, Peters TA, Roepman R, Cremers FP, Collin RW. Unexpected CEP290 mRNA splicing in a humanized knock-in mouse model for Leber congenital amaurosis. PLoS One 2013;8(11):e79369.
- [45] Shimada H, Lu Q, Insinna-Kettenhofen C, Nagashima K, English MA, Semler EM, et al. In vitro modeling using ciliopathy-patient-derived cells reveals distinct cilia dysfunctions caused by CEP290 Mutations. Cell Rep 2017;20(2):384–96.
- [46] Sahel JA, Dalkara D. Gene therapy for retinal dystrophy. Nat Med 2019;25(2):198– 9.
- [47] Wang D, Tai PWL, Gao G. Adeno-associated virus vector as a platform for gene therapy delivery. Nat Rev Drug Discov 2019;18(5):358–78.
- [48] Russell S, Bennett J, Wellman JA, Chung DC, Yu ZF, Tillman A, et al. Efficacy and safety of voretigene neparvovec (AAV2-hRPE65v2) in patients with RPE65-

mediated inherited retinal dystrophy: a randomised, controlled, open-label, phase 3 trial. Lancet 2017;390(10097):849-60.

- [49] Gardiner KL, Cideciyan AV, Swider M, Dufour VL, Sumaroka A, Komaromy AM, et al. Long-term structural outcomes of late-stage RPE65 gene therapy. Mol Ther 2020;28(1):266–78.
- [50] Jang HK, Song B, Hwang GH, Bae S. Current trends in gene recovery mediated by the CRISPR-Cas system. Exp Mol Med 2020;52(7):1016–27.
- [51] Maeder ML, Stefanidakis M, Wilson CJ, Baral R, Barrera LA, Bounoutas GS, et al. Development of a gene-editing approach to restore vision loss in Leber congenital amaurosis type 10. Nat Med 2019;25(2):229–33.
- [52] Gapinske M, Luu A, Winter J, Woods WS, Kostan KA, Shiva N, et al. CRISPR-SKIP: programmable gene splicing with single base editors. Genome Biol 2018;19 (1):107.
- [53] Zuccaro MV, Xu J, Mitchell C, Marin D, Zimmerman R, Rana B, et al. Allele-specific chromosome removal after Cas9 Cleavage in Human Embryos. Cell 2020.
- [54] Bennett CF. Therapeutic antisense oligonucleotides are coming of age. Annu Rev Med 2019;70:307–21.
- [55] Dulla K, Aguila M, Lane A, Jovanovic K, Parfitt DA, Schulkens I, et al. Splice-Modulating Oligonucleotide QR-110 Restores CEP290 mRNA and Function in Human c.2991+1655A>G LCA10 Models. Mol Ther Nucleic Acids 2018;12:730–40.
- [56] Cideciyan AV, Jacobson SG, Drack AV, Ho AC, Charng J, Garafalo AV, et al. Effect of an intravitreal antisense oligonucleotide on vision in Leber congenital amaurosis due to a photoreceptor cilium defect. Nat Med 2019;25(2):225–8.
- [57] Zhang W, Li L, Su Q, Gao G, Khanna H. Gene therapy using a miniCEP290 fragment delays photoreceptor degeneration in a mouse model of leber congenital amaurosis. Hum Gene Ther 2018;29(1):42–50.
- [58] Mookherjee S, Chen HY, Isgrig K, Yu W, Hiriyanna S, Levron R, et al. A CEP290 Cterminal domain complements the mutant CEP290 of Rd16 Mice In Trans and Rescues Retinal Degeneration. Cell Rep 2018;25(3):611–23 e6.
- [59] Burnight ER, Wiley LA, Drack AV, Braun TA, Anfinson KR, Kaalberg EE, et al. CEP290 gene transfer rescues Leber congenital amaurosis cellular phenotype. Gene Ther 2014;21(7):662–72.
- [60] Allocca M, Doria M, Petrillo M, Colella P, Garcia-Hoyos M, Gibbs D, et al. Serotypedependent packaging of large genes in adeno-associated viral vectors results in effective gene delivery in mice. J Clin Invest 2008;118(5):1955–64.
- [61] Ghazi NG, Abboud EB, Nowilaty SR, Alkuraya H, Alhommadi A, Cai H, et al. Treatment of retinitis pigmentosa due to MERTK mutations by ocular subretinal injection of adeno-associated virus gene vector: results of a phase I trial. Hum Genet 2016;135(3):327–43.
- [62] Yu W, Mookherjee S, Chaitankar V, Hiriyanna S, Kim JW, Brooks M, et al. Nrl knockdown by AAV-delivered CRISPR/Cas9 prevents retinal degeneration in mice. Nat Commun 2017;8:14716.
- [63] Byrne LC, Dalkara D, Luna G, Fisher SK, Clerin E, Sahel JA, et al. Viral-mediated RdCVF and RdCVFL expression protects cone and rod photoreceptors in retinal degeneration. J Clin Invest 2015;125(1):105–16.
- [64] McGregor JE, Godat T, Dhakal KR, Parkins K, Strazzeri JM, Bateman BA, et al. Optogenetic restoration of retinal ganglion cell activity in the living primate. Nat Commun 2020;11(1):1703.
- [65] Cronin T, Vandenberghe LH, Hantz P, Juttner J, Reimann A, Kacso AE, et al. Efficient transduction and optogenetic stimulation of retinal bipolar cells by a synthetic adeno-associated virus capsid and promoter. EMBO Mol Med 2014;6 (9):1175–90.
- [66] Garita-Hernandez M, Lampic M, Chaffiol A, Guibbal L, Routet F, Santos-Ferreira T, et al. Restoration of visual function by transplantation of optogenetically engineered photoreceptors. Nat Commun 2019;10(1):4524.

- [67] Tambuyzer E, Vandendriessche B, Austin CP, Brooks PJ, Larsson K, Miller Needleman KI, et al. Therapies for rare diseases: therapeutic modalities, progress and challenges ahead. Nat Rev Drug Discov 2020;19(2):93–111.
- [68] Mockel A, Obringer C, Hakvoort TB, Seeliger M, Lamers WH, Stoetzel C, et al. Pharmacological modulation of the retinal unfolded protein response in Bardet-Biedl syndrome reduces apoptosis and preserves light detection ability. J Biol Chem 2012;287(44):37483–94.
- [69] Hull K, Benster T, Manookin MB, Trauner D, Van Gelder RN, Laprell L. Photopharmacologic Vision Restoration Reduces Pathological Rhythmic Field Potentials in Blind Mouse Retina. Sci Rep 2019;9(1):13561.
- [70] Chen Y, Chen Y, Jastrzebska B, Golczak M, Gulati S, Tang H, et al. A novel small molecule chaperone of rod opsin and its potential therapy for retinal degeneration. Nat Commun 2018;9(1):1976.
- [71] Holekamp NM. Review of neovascular age-related macular degeneration treatment options. Am J Manag Care 2019;25(10 Suppl):S172–S81.
- [72] Horvath P, Aulner N, Bickle M, Davies AM, Nery ED, Ebner D, et al. Screening out irrelevant cell-based models of disease. Nat Rev Drug Discov 2016;15(11):751– 69.
- [73] Kruczek K, Swaroop A. Pluripotent stem cell-derived retinal organoids for disease modeling and development of therapies. Stem Cells 2020;38(10):1206–15.
- [74] AlQuraishi M. End-to-End Differentiable Learning of Protein Structure. Cell Syst 2019;8(4):292–301 e3.
- [75] Jorstad NL, Wilken MS, Grimes WN, Wohl SG, VandenBosch LS, Yoshimatsu T, et al. Stimulation of functional neuronal regeneration from Muller glia in adult mice. Nature 2017;548(7665):103–7.
- [76] Fernandez-Nogales M, Murcia-Belmonte V, Chen HY, Herrera E. The peripheral eye: a neurogenic area with potential to treat retinal pathologies? Prog Retin Eye Res 2019;68:110–23.
- [77] West EL, Ribeiro J, Ali RR. Development of stem cell therapies for retinal degeneration. Cold Spring Harb Perspect Biol 2020;12(8).
- [78] Gasparini SJ, Llonch S, Borsch O, Ader M. Transplantation of photoreceptors into the degenerative retina: current state and future perspectives. Prog Retin Eye Res 2019;69:1–37.
- [79] da Cruz L, Fynes K, Georgiadis O, Kerby J, Luo YH, Ahmado A, et al. Phase 1 clinical study of an embryonic stem cell-derived retinal pigment epithelium patch in agerelated macular degeneration. Nat Biotechnol 2018;36(4):328–37.
- [80] Mandai M, Kurimoto Y, Takahashi M. Autologous induced stem-cell-derived retinal cells for macular degeneration. N Engl J Med 2017;377(8):792–3.
- [81] Zhao C, Wang Q, Temple S. Stem cell therapies for retinal diseases: recapitulating development to replace degenerated cells. Development 2017;144 (8):1368-81.
- [82] Henry MP, Hawkins JR, Boyle J, Bridger JM. The genomic health of human pluripotent stem cells: genomic instability and the consequences on nuclear organization. Front Genet 2018;9:623.
- [83] Deng WL, Gao ML, Lei XL, Lv JN, Zhao H, He KW, et al. Gene correction reverses ciliopathy and photoreceptor Loss in iPSC-derived retinal organoids from retinitis pigmentosa patients. Stem Cell Reports 2018;10(4):1267–81.
- [84] Mahato B, Kaya KD, Fan Y, Sumien N, Shetty RA, Zhang W, et al. Pharmacologic fibroblast reprogramming into photoreceptors restores vision. Nature 2020;581 (7806):83–8.
- [85] Singh MS, Park SS, Albini TA, Canto-Soler MV, Klassen H, MacLaren RE, et al. Retinal stem cell transplantation: Balancing safety and potential. Prog Retin Eye Res 2020;75:100779.