

# 2D printed multicellular devices performing digital and analogue computation

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Much effort has been expended on building cellular computational devices for different applications. Despite the significant advances, there are still several addressable restraints to achieve the necessary technological transference. These improvements will ease the development of end-user applications working out of the lab. In this study, we propose a methodology for the construction of printable cellular devices, digital or analogue, for different purposes. These printable devices are designed to work in a 2D surface, in which the circuit information is encoded in the concentration of a biological signal, the so-called carrying signal. This signal diffuses through the 2D surface and thereby interacts with different device components. These components are distributed in a specific spatial arrangement and perform the computation by modulating the level of the carrying signal in response to external inputs, determining the final output. For experimental validation, 2D cellular circuits are printed on a paper surface by using a set of cellular inks. As a proof-of-principle, we have printed and analysed both digital and analogue circuits using the same set of cellular inks but with different spatial topologies. The proposed methodology can open the door to a feasible and reliable industrial production of cellular circuits for multiple applications.

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To date, most cellular computational devices have been designed according to a standard architecture that organizes their components into three different layers: the input layer, where external signals are detected; the processing layer, where computation is performed; and the output layer, where the final output is produced<sup>1–4</sup>. However, when this architecture is applied to biological substrates<sup>3,5</sup>, different constraints such as the so-called wiring problem<sup>6–8</sup>, the genetic complexity<sup>8–10</sup> or the metabolic burden<sup>11–14</sup> limit the development of complex devices based on this architecture and these limitations<sup>15–17</sup> can compromise the development of end-user applications<sup>18–20</sup>. Although there are some studies proposing optimal methodologies and software solutions to facilitate complex circuit designs inspired on electronic design automation (EDA)<sup>21,22</sup>, issues regarding scalability, reusability, engineering complexity and technological transference remain a challenge. In previous works, the idea of multicellularity has been applied to design complex computational circuit<sup>23</sup>. However, co-culturing different cell strains has some limitations when considering temporal stability, easy experimental setup and reproducibility for potential applications.

In this study, we explore a framework for computational devices based on biological substrates completely different from the standard architecture, as no distinctions between input and processing layers are considered. By breaking with the conventional architecture and introducing the space<sup>24</sup> as an additional computational element<sup>25,26</sup>, we demonstrate that simple, robust and scalable multicellular computational devices can be built. This methodology, inspired from printed electronics<sup>27–31</sup>, has enabled us to construct 2D printable cellular devices capable to undertake any type of computation, either digital or analogic. These circuits can be printed in flexible substrates such as paper, allowing the creation of versatile devices. For example, complex single-use living biosensors can be industrially produced at a low cost with potential applications in different market segments.

## Results

**Circuits design.** In this study, two main issues needed to be addressed for the development of 2D printed circuits: (i) the optimal circuit architecture and (ii) the methodology for fast and reliable printing.

Regarding the first issue, we propose a multicellular architecture. Considering that computation is, in essence, a matter of information processing<sup>32</sup>, our approach encodes the information in the concentration of a unique biological signal, the carrying signal (CS)<sup>33,34</sup>.

We constructed our devices in a 2D surface through which the CS, which is produced at a specific location in the device, can diffuse. While diffusing, the CS can interact with the different device components which are distributed on the surface in a specific spatial arrangement. The main role of these components is to modulate the flow of the CS in one of two different ways: these can either allow, or even amplify the flow of the CS across them, or can reduce or abolish the CS flow. When these modulatory elements act by only allowing or blocking the CS flow, the device performs digital computations. On the other hand, when regulatory elements increase or reduce the concentration of the CS, and these variations are meaningful in terms of output production, the device performs analogic computations.

The ability of codifying the computational complexity in the topology of modulatory elements, i.e. in their specific spatial arrangement, enables the construction of systems that have very low genetic complexity but that are capable of implementing complex computations<sup>6</sup>. It should be noted that in these devices, the spatial sequence in which the CS meets the different modulatory elements defines the computation. Thus, the same

modulatory elements can implement different computations by just altering their spatial position.

The use of a multicellular implementation for computational devices has been shown to be very useful for complex circuits<sup>23,24,26,35</sup> as each component is implemented in a different cell type. Moreover, this embodiment is appealing because it simplifies the genetic engineering required, thereby minimizing the emergence of unexpected interactions with the host cell and significantly reducing the metabolic burden associated with foreign genes<sup>9,36</sup>. Furthermore, the growth of different cell types in different spatial locations provides devices with high stability since negative competition effects<sup>37,38</sup>, which emerge when multiple cell types grow in the same media, are avoided.

**Development of a library of engineered cells.** In order to experimentally validate our approach, we built a library of engineered cells. The genetic architecture of each cell type is schematically shown in Supplementary Fig. 1 and each specific genetic part is summarized in Supplementary Table 1 (see Supplementary Information for details). First, four cell types, S<sub>1</sub>, S<sub>2</sub>, S<sub>3</sub> and S<sub>4</sub> were engineered to produce constitutively (S<sub>1</sub>), or upon external induction (S<sub>2</sub> upon arabinose, S<sub>3</sub> upon rhamnose and S<sub>4</sub> upon mercury), a small extracellular molecule that would act as the CS. This molecule is the bacterial 3OC6HSL acyl homoserine lactone (AHL) which is involved in the natural quorum sensing mechanism<sup>39,40</sup>. AHL molecules can diffuse along the paper surface and thereby encounter the different modulatory elements. In our design, unlike in standard methods, there are no explicit connections between the different components of the circuit. Instead, each component contributes independently to the final computation by partially modifying the AHL flow. In consequence, this approach overcomes the so-called wiring problem.

Regarding the modulatory elements, these elements were implemented in differently engineered cell types that express, or do not express AiiA, an enzyme that degrades AHL<sup>41,42</sup>, in response to external inputs spread over the whole device surface. More specifically, some modulators (M<sup>–</sup> cells) reduce or even abolish the flow of AHL by inducing AiiA expression upon the presence of an external inducer. In contrast, in other modulators (M<sup>+</sup> cells), AiiA is constitutively expressed and external inducers block this constitutive expression, thereby increasing the flow of AHL. Finally, to compensate the AHL concentration decrease associated with the diffusion process, an auto-amplifier element (CA) was also created. This auto-amplifier cell can produce AHL only when AHL is detected and in this way, restores the AHL signal.

The final overall circuit response was embodied in a specific cell type (reporter cell, CR) that produces the output only when the CS concentration exceeds a given threshold. As a proof-of-principle we expressed a green fluorescent reporter protein (GFP) as the final output. Supplementary Fig. 2 shows the relationship between AHL concentration and GFP expression. It should be noted that each cell type also constitutively expresses red fluorescent protein (RFP) in order to quantitatively assess the cell population growth on the surface. Supplementary Fig. 3 shows the temporal change in RFP expression associated with cell growth in the various cell types (see Supplementary Information for details regarding experimental measurements and quantifications). Experimental results indicate similar dynamic growths for the different cell types.

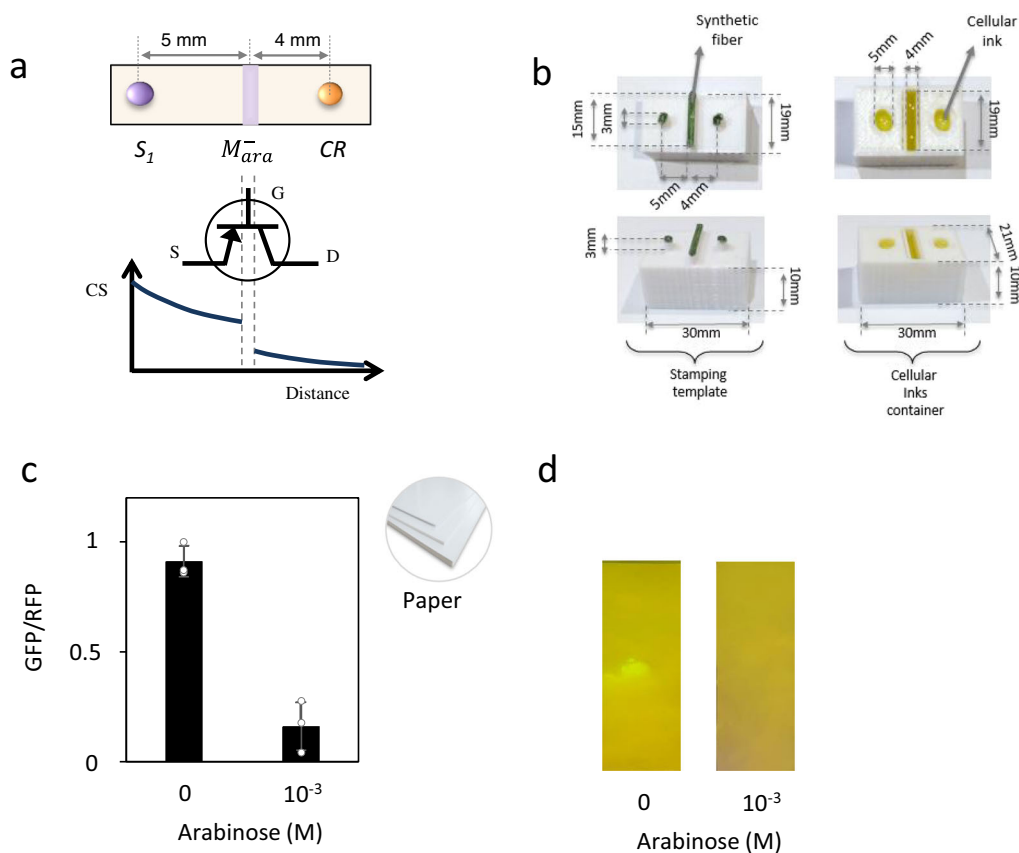
**Circuit substrates.** Despite there being several suitable materials that could act as the working substrate, paper was selected due to its low cost and its ability to allow bacterial growth and AHL diffusion. It is worth mentioning that previous studies have also

used paper for different cell-based applications or organic transistors due to these optimal properties<sup>43–45</sup>. To take full advantage of our approach, we also developed a set of cellular inks composed of a single cell type mixed with cellular nutrients and agar (used as a thickener). These inks were used to draw the different circuits in the paper (see Supplementary Information for details regarding the cellular inks and paper used).

**Printing devices.** The second issue addressed was the methodology for circuit printing. In this study, we explored the construction of different cellular devices by stamping them on paper surfaces. For it, we used a container filled with different cellular inks. Then, a stamping template was soaked with the different cellular inks and stamped on a paper surface according to a given topology, codifying the computation to be performed. Finally, the stamped paper was deposited on an LB-agar plate mixed with the different inputs, and it was subsequently incubated at 37 °C for 24 h. Stamping offers numerous advantages over previous methods for cell printing such as 3D printers<sup>46</sup>. For instance, stamping is faster than 3D printing, does not require complex printing devices and templates can be easily customized, which makes stamping suitable for industrial production potentially at very low cost and with minimal requirements. Supplementary Fig. 4 shows a schematic representation of the stamping pipeline.

To endorse the proposed methodology, we first analysed cell deposition and cell growth on a paper substrate, as well as CS transmission along the surface. Supplementary Fig. 5a shows a printed pattern composed of a constitutive AHL supply ( $S_1$ ) at one end of the strip and reporter cells (CR) uniformly distributed along the rest of the surface. Quantification of the GFP produced in the reporter cells indicates that  $S_1$  cells can grow in the surface and secrete AHL, which efficiently diffuses along the paper and can be detected by the CR cells at large distances, i.e. up to 20 mm away from the AHL cell source. The ability of AHL auto-amplification by CA cells, printed between  $S_1$  and CR cells, to restore the signal decay due to diffusion is also analysed in Supplementary Fig. 5b. Experimental results demonstrate that CA cells can extend the range of the diffusing CS.

**Biosensor prototype responding to mercury.** In order to prove potential applications of paper-based cellular devices, we created a very simple paper-based biosensor prototype responding to mercury. This device is based on the minimal architecture shown in Supplementary Fig. 5a, composed by a CS supply cell ( $S_4$ ) that secretes AHL in the presence of mercury combined with a reporter cell CR expressing GFP in response to AHL (see Supplementary Fig. 1 and Supplementary Table 2 for details about the genetic architecture of the involved cells). CR cells have been

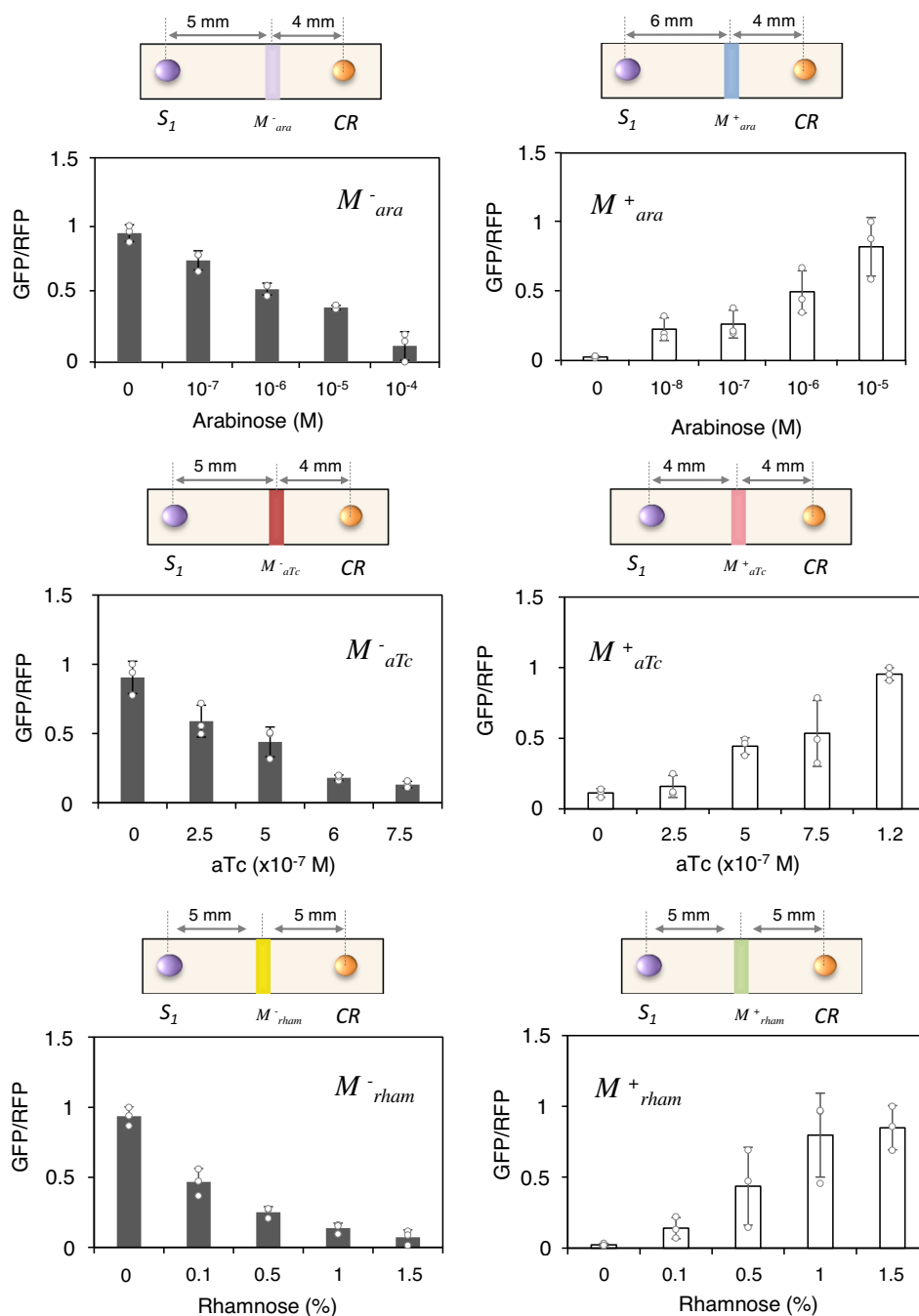


**Fig. 1 Printed circuit implementing a transistor-like device.** **a** Mapping of a general transistor architecture on a cellular printed pattern obtained using a stamping template. Similar to the transistor architecture, the cellular pattern is composed of three main components: source ( $S_1$  cells), gate ( $M^-$  cells) that responds to external inputs and a drain (CR cells) as the final output responding to the presence of the carrying signal (CS). **b** Stamping template used to create the circuit made of PLA with a layer of synthetic fibre (green). Cellular inks (yellow) are in their corresponding containers. Before stamping, the synthetic fibre is soaked with the different cell types. Finally, the stamping template is pressed against the paper surface, depositing all cells. **c** Circuit response. In the absence of external input, i.e. arabinose, the CS encoded in the production of AHL molecules by  $S_1$  cells diffuses along the surface, inducing GFP expression in reporter cells CR. In the presence of  $10^{-3}$  M arabinose (Ara), the modulatory element  $M^-_{ara}$  produces the AHL cleaving enzyme AiiA, which degrades the CS. Error bars are the standard deviation (SD) of three independent experiments. Data are presented as mean values  $\pm$  SD. Experiments are performed on paper strips. The average fold change is 5.6x. **d** Photography of the device. Source data are provided as a Source Data file.

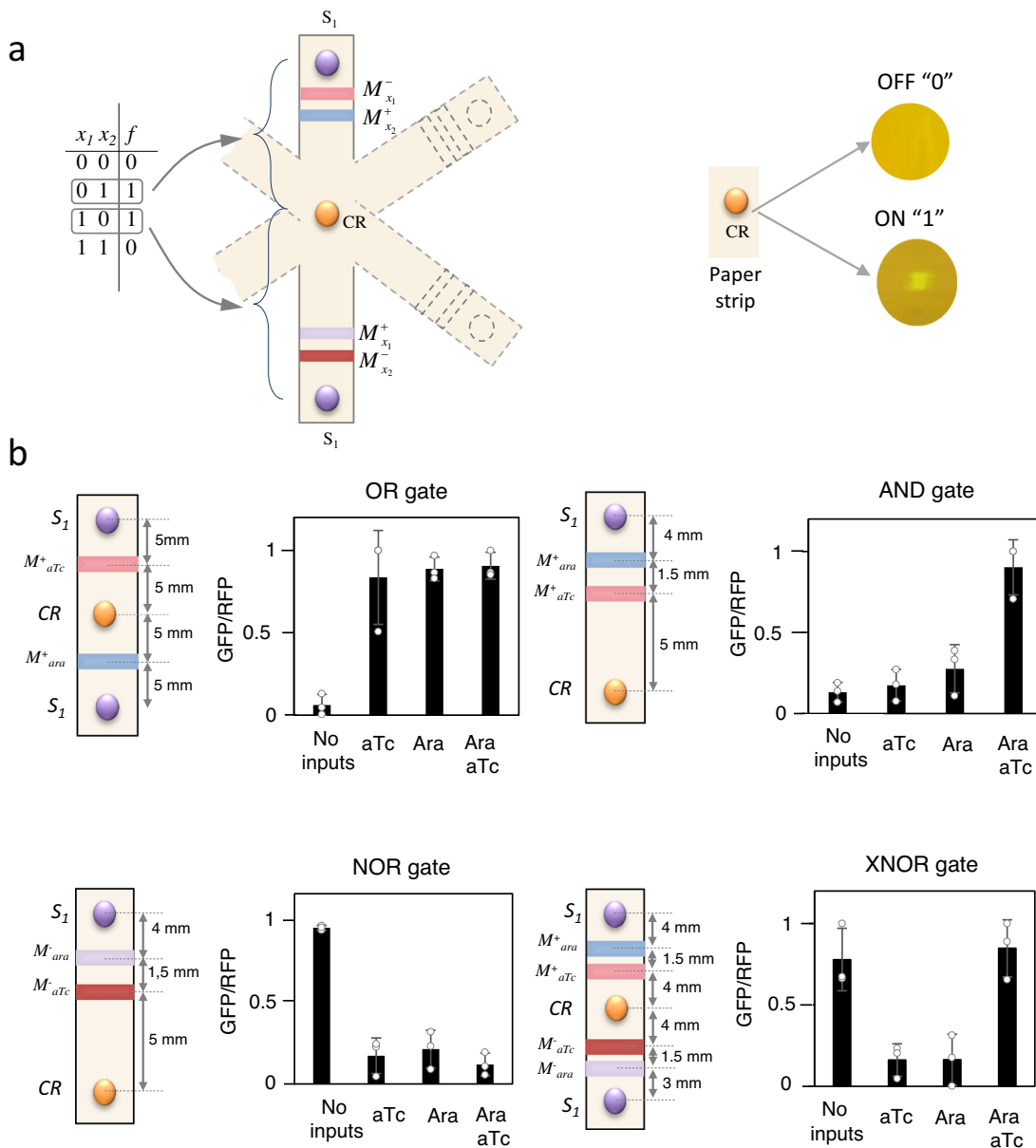
distributed in dots along the paper strip. Depending on the mercury concentration, different amounts of AHL are produced, i.e. higher AHL production implies larger AHL diffusion. In consequence, the number of dots expressing GFP (green dots) correlates with the mercury concentration and can visually be estimated. Experimental results are shown in Supplementary Fig. 6. Supplementary Fig. 6a shows the correlation between mercury concentration and number of green dots observed. Supplementary Fig. 6b shows photographs of the visual readout at different mercury concentrations. Results demonstrate a responsive range between 0.013 and 2.71 ppm. It is established that the

allowed amount of mercury in most commercialized products is between 0.1 and 1 ppm. Thus, this prototype could be optimal for a fast and easy product screening to ensure safety.

**Transistor-like devices.** To implement more complex devices, we analysed the effect of CS modulation on the final output by printing different modulatory elements between  $S_1$  and CR. The resulting circuits can be compared to electronic transistor architecture, in which the CS, i.e. AHL molecules, enter into the device (source), are modulated in response to an external signal (gate) and then exit the device (drain)<sup>47</sup>. Figure 1 shows an example of a



**Fig. 2 Relationship between input and output in transistor-like circuits.** Output modulation in response to different input concentrations. Modulatory elements  $M^-$  (black bars) and  $M^+$  (white bars) cells respond to the external inputs, arabinose ( $M^+_{ara}/M^-_{ara}$ ), aTc ( $M^+_{aTc}/M^-_{aTc}$ ) and rhamnose ( $M^+_{rham}/M^-_{rham}$ ).  $S_1$  cells produce AHL constitutively. GFP levels are normalized by RFP, which correlates with cell population. CR are the reporter cells. Error bars are the standard deviation (SD) of three independent experiments. Data are presented as mean values  $\pm$  SD. Source data are provided as a Source Data file.



**Fig. 3** General multi-branch architecture for implementation of digital circuits. **a** Schematic representation of the multi-branch implementation of a truth table. **b** Implementation of different logic gates. A schematic representation of the cells used in each paper strip and their corresponding distance points is given (Left). Gates with two sources of  $S_1$  (OR and XNOR gates) are circuits carrying two branches, while the other gates (NOR and AND gates) can be implemented with just one branch. Input concentrations are  $Ara = 10^{-3} M$  and  $aTc = 10^{-6} M$ .  $M^+_{aTc}$  and  $M^-_{aTc}$  are, respectively, positive and negative modulatory cells responding to aTc.  $M^+_{ara}$  and  $M^-_{ara}$  are, respectively, positive and negative modulatory cells responding to arabinose.  $S_1$  cells produce AHL constitutively and CR are the reporter cells. Error bars are the standard deviation (SD) of three independent experiments. The average fold change has been obtained from the mean of ON and OFF states from each circuit. OR gate 14.31x, AND gate 6.21x, NOR gate 6.58x, XNOR gate 5.6x. Source data are provided as a Source Data file.

transistor-like circuit where all components can be directly mapped onto our circuit topology. Circuits were assembled by stamping cells using the template shown in Fig. 1b. This template was created in polylactic acid plastic (PLA) with a 3D printer combined with a synthetic fibre tissue (80% polyester and 20% polyamide) in which cells are embedded. Figure 1c shows the behaviour of the device. In the absence of the external input, i.e. arabinose, the CS (AHL molecules) is constitutively produced by  $S_1$  cells and diffuses along the surface, activating GFP production by the CR reporter cells. In the presence of arabinose ( $10^{-3} M$ ) the modulatory element  $M^-_{ara}$ , inserted between the  $S_1$  and CR cells,

produces Aiaa, which degrades AHL, thereby preventing further AHL flow.

Although this study focuses on the use of paper as a substrate, other different materials can be used for printed circuits. As a proof-of-principle, we have printed the same circuit shown in Fig. 1a on nylon fabric. As seen in Supplementary Fig. 7, the circuit printed on nylon has also a good performance.

It is worth mentioning that printed circuits can be either used directly to perform the desired computation or can be stored, in the fridge at  $4^\circ C$  or frozen at  $-80^\circ C$ , for future use. To evaluate the temporal stability of the transistor-like circuit described in

Fig. 1, we analysed the performance evolution of this printed circuit along time, storing them at 4 °C for several days. Experimental results, shown in Supplementary Fig. 8, indicate that circuits stored at 4 °C exhibit a progressive reduction of the fold-change over time, i.e. up to 10 days. Despite this reduction, the device exhibits a proper behaviour. On the other hand, frozen circuits do not exhibit significant changes in their performance, maintaining an optimal functionality, leading to larger durability.

From this first transistor-like architecture, we analysed the CS modulation. Specifically, we tested six different modulatory elements that respond to one of two external signals, aTc, arabinose and rhamnose. Three of the modulatory elements ( $M^+_{aTc}$ ,  $M^+_{ara}$  and  $M^+_{rham}$  cells) allow the CS flow across the circuit when the level of the external signal increases, whereas the other three modulatory elements ( $M^-_{aTc}$ ,  $M^-_{ara}$  and  $M^-_{rham}$  cells) reduce the CS flow when the level of the external signal decreases. The experimental results shown in Fig. 2 demonstrate that these modulations are, in essence, analogue, allowing the tuning of CS levels in a continuous manner. This figure displays the output data relative to cell population, i.e. ratio of GFP/RFP. In consequence, by properly combining these modulatory elements, either analogue or digital circuits can be implemented.

Previous to the creation of more complex computational devices, the effect of introducing multiple modulatory elements between  $S_1$  and CR cells was analysed. In absence of modulatory elements, the optimal distance between  $S_1$  and CR cells is 10 mm. However, as Supplementary Fig. 9 shows, when an increasing number of modulatory elements are introduced between  $S_1$  and CR cells, a reduction of the CS levels is observed due to their basal activity. However, this reduction can be restored by introducing CA cells, which allow the introduction of more modulatory elements, overcoming this limitation.

**Development of printed digital circuits.** To develop printed digital circuits, we explored a general method for mapping any arbitrary truth table into a spatial pattern. Specifically, we proposed the multi-branch circuit that is schematically represented in Fig. 3a. In this topology, each input combination in a truth table associated with output 1 can be encoded in a different branch. At one end of the branch, there are AHL producer cells, whereas at the other end, there is the output, produced by CR cells. In the middle, different bands containing modulatory cells are printed. Logic inputs “1” in the truth table are encoded in  $M^+$  cell types, which allow CS diffusion only if the input is present. Logic inputs “0” are encoded in  $M^-$  cell types, which allow CS diffusion only when the input is absent. We analysed the scalability of this topological implementation. For circuits responding to  $N$  inputs, the maximum size of the cell library implementing modulatory elements is  $2 \cdot N$ , i.e. two types of modulatory element,  $M^+$  and  $M^-$ , for each input. Moreover, the maximum number of branches is  $2^{N-1}$ . However, our computational analysis, shown in Supplementary Fig. 10, indicates that most of the circuits require less branches. To perform this analysis, the truth tables of all circuits responding to 2 (Supplementary Fig. 10a), 3 (Supplementary Fig. 10b) and 4 inputs (Supplementary Fig. 10c) were simplified to its minimal configuration using the standard Karnaugh maps method<sup>48</sup> and directly mapped into a topological configuration, according to the method represented in Fig. 3a.

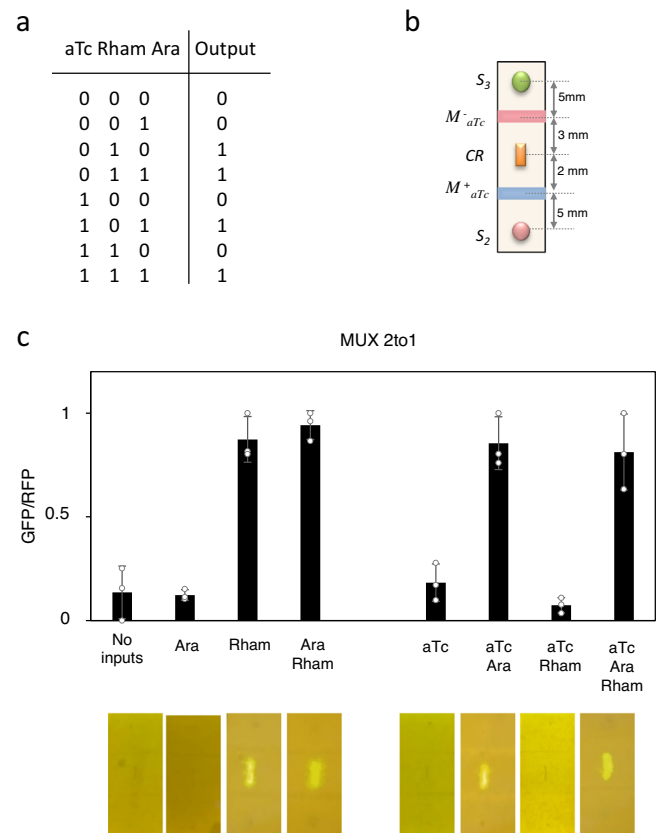
These results indicate that large scalability is achievable with this multi-branched architecture. As a proof-of-principle, we implemented several examples of logic gates using the multi-branch architecture. Relative GFP/RFP values obtained from these examples are shown in Fig. 3b.

Once logic gates responding to two inputs were constructed, we introduced an additional input, i.e. rhamnose, in order to create a

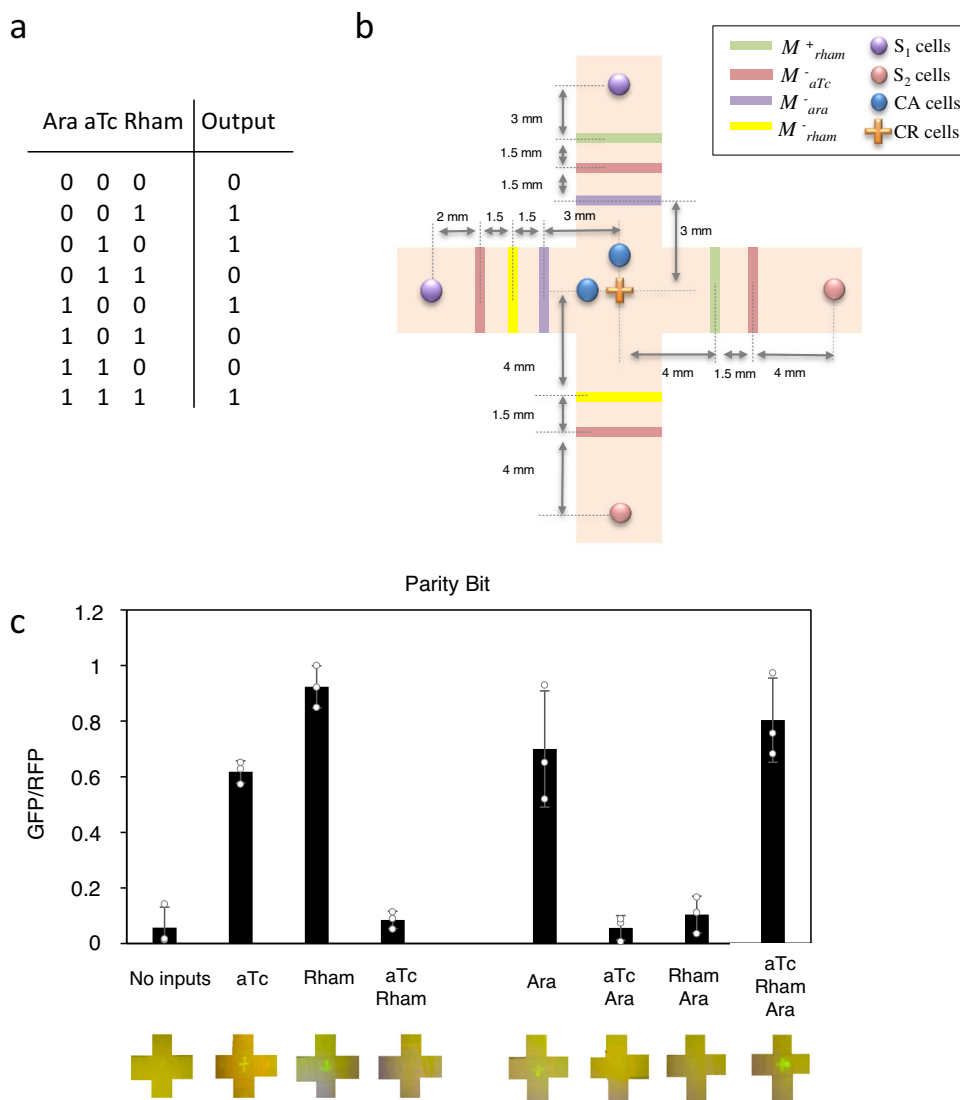
3-input device. As a proof-of-concept, we implanted a 2 to 1 multiplexer (MUX2to1). This device involves two branches producing AHL by  $S_3$  and  $S_2$  cells in response to rhamnose and arabinose, and different modulatory elements responding to aTc. Figure 4a shows the truth table describing the behaviour of this device and Fig. 4b displays the specific spatial topology. Figure 4c shows the circuit response upon different input combinations. As seen in this figure, the device behaves according to the truth table, implementing the expected response.

To explore more complex topologies, an even parity bit circuit was implemented. This circuit, responding to three different inputs, involves four different branches with up to three modulatory elements per branch. Due to this large number of modulators, an auto-amplifier cell (CA) was introduced close to the output cells CR in order to restore AHL levels. Figure 5a shows the truth table describing circuit's logics, which is translated into a printed pattern (Fig. 5b). Experimental results are shown in Fig. 5c, indicating a proper circuit response.

**Development of analogic circuits.** Not only digital, but also analogic computations were considered to prove the versatility of



**Fig. 4** Multiplexer 2 to 1. **a** Truth table of the 2 to 1 multiplexer. **b** Spatial pattern of cells encoding the multiplexer device with their corresponding distances.  $M^+_{aTc}$  and  $M^-_{aTc}$  are, respectively, positive and negative modulatory cells responding to aTc.  $S_2$  cells produce AHL upon arabinose induction and  $S_3$  cells produce AHL upon rhamnose induction. CR are the reporter cells. **c** Multiplexer response upon input of the indicated combinations. Input concentrations are Ara =  $10^{-3}$  M, aTc =  $10^{-6}$  M and Rham = 1.5%. Error bars are the standard deviation (SD) of three independent experiments. Data are presented as mean values  $\pm$  SD. The average fold change has been obtained from the mean of ON and OFF states from each input combination. Multiplexer 2 to 1 fold change: 6.76x. Source data are provided as a Source Data file.



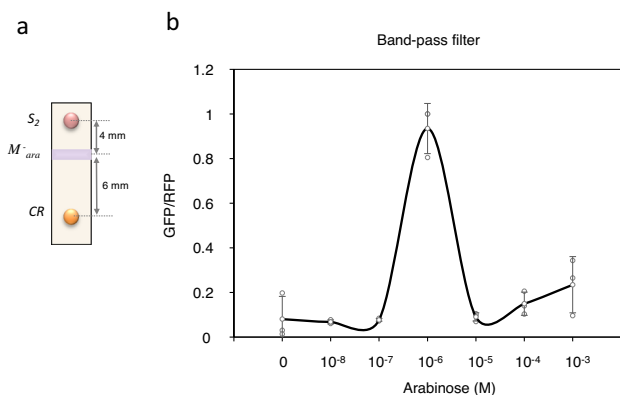
**Fig. 5 Parity bit.** **a** Truth table of the even parity bit circuit. **b** Spatial pattern of the cells encoding the truth table. This circuit involves four branches, with the final output in the middle. The output cells CR were printed as a cross to visualize from which branch comes dominantly the AHL signal.  $M^{+}_{aTc}$  and  $M^{-}_{aTc}$  are, respectively, positive and negative modulatory cells responding to aTc.  $M^{+}_{rham}$  and  $M^{-}_{rham}$  are, respectively, positive and negative modulatory cells responding to rhamnose.  $S_1$  cells produce AHL constitutively and  $S_2$  cells produce AHL upon arabinose induction. **c** Multiplexer response upon input of the indicated combinations. In this circuit, due to the amount of modulatory elements per branch, auto-amplifier cells (CA) were incorporated close to reporter cells (CR). These cells amplify the signal on the presence of AHL. Input concentrations are  $ara = 10^{-3}$  M,  $aTc = 10^{-6}$  M and  $rham = 1.5\%$ . Error bars are the standard deviation (SD) of three independent experiments. Data are presented as mean values  $\pm$  SD. The average fold change has been obtained from the mean of ON and OFF states from each input combination. Parity bit fold change: 10.47x Source data are provided as a Source Data file.

printed circuits. For it, an example of an analogic circuit, i.e. a band-pass filter, was implemented using the same elements described in Fig. 1, except that cell type  $S_1$  (constitutive production of AHL) was replaced by  $S_2$  (arabinose-inducible production of AHL). This allows the introduction of an additional modulatory element as AHL production depends on the external input, in this case arabinose. At low arabinose concentrations, AHL levels are low due to the low production in  $S_2$  cells. The combination of low AHL synthesis with the reduction of AHL due to its diffusion does not allow AHL-induced output production by CR cells. In contrast, at high arabinose concentrations, although the higher AHL secretion, it also induces the production of AiiA by the modulatory element ( $M^{-}_{ara}$  cells) located between  $S_2$  and CR cells, which degrades the diffusing signal and therefore, blocks the AHL flow and the output is not produced. However, at intermediate arabinose concentrations, the level of AHL secreted

from  $S_2$  cells is sufficient for CR activation but the arabinose concentration is not high enough for  $M^{-}_{ara}$  activation. Thus, AHL concentration induces output production in CR cells. Figure 6 shows the GFP/RFP levels at different arabinose concentrations, indicating a clear band-pass response.

**Discussion**

Multiple studies have demonstrated that multicellular embodiments are optimal to implement complex computational devices. However, co-culturing different cell types in the same medium induces the emergence of problems related to competition for nutrients, which can compromise large temporal stability. This problem has been overcome introducing spatial segregation<sup>24</sup>. However, in general, this methodology requires multiple wiring molecules coexisting, which can limit the scalability. Other approaches only require a single wiring molecule but the



**Fig. 6 Band-pass filter circuit.** **a** Band-pass filter implementation is outlined by a schematic representation of the cells used for the paper strips and their corresponding distance points.  $M^-_{ara}$  are negative modulatory cells responding to arabinose.  $S_2$  cells produce AHL upon arabinose induction and CR are the reporter cells. **b** Circuit response upon increasing concentrations of the external input, i.e. arabinose. Error bars are the standard deviation (SD) of three independent experiments. Data are presented as mean values  $\pm$  SD. Source data are provided as a Source Data file.

experimental setup is complex and the composition and stability of the multicellular consortia must be carefully controlled<sup>23</sup>. The methodology presented in this study combines the benefits of the previous approaches but overcoming some of their limitations, namely only a single molecule is required for computation and different cell types can grow separately, avoiding competition effects.

Although other approaches based on cell-free systems<sup>49</sup> are suitable for paper-based devices, the limited number of biochemical reactions available in these systems could limit the creation of complex devices involving multiple circuit components. The present results are just a small piece of the potentially far-reaching implications and possibilities of this approximation. For instance, it has been demonstrated that very simple but useful biosensors can be easily implemented in paper-based circuits (e.g. mercury detector shown in Supplementary Fig. 6) and used out of the lab, allowing a rapid estimation of mercury concentration directly from the visual reporter. Additionally, more complex devices able to detect and integrate different signals to produce the output only upon certain combination have also been built.

Although our system works for three inputs, future work should be devoted to (i) scaling up this system to enlarge the possible computations, (ii) automatize the stamping of the complex circuits for large-scale industrial production and (iii) reduce time response.

In summary, printable cellular circuits could represent an interesting alternative for paper-based devices with appealing potential advantages such as the possibility of producing low-cost cellular devices based on a simple experimental setup methodology. Further development is required to explore this potential and meet further market needs.

## Methods

**Strains, media and growth conditions.** Cloning and expression experiments were performed in *Escherichia Coli* Top10 (Invitrogen, USA) and *Escherichia Coli* ZN1 (ExpresSys, Germany). Cells were grown in Lysogeny Broth (LB) at 37 °C and selected with appropriate antibiotics (chloramphenicol 35  $\mu$ g/ml, kanamycin 35  $\mu$ g/ml Sigma, USA). Bacterial strains were preserved in LB glycerol 20% (v/v) at  $-80^\circ\text{C}$ . Cells inoculated from single colonies from streaked glycerol stocks were grown at 37 °C with shaking (450 revolutions per minute (rpm)) for about 5 h until reaching an OD between 0.15–0.2.

Experiments were performed in petri dishes (dabiolab) pouring 20 ml of LB broth with agar (Sigma Aldrich, USA) supplemented with the antibiotic and the appropriate inducers before depositing the paper strips to print the circuit.

The inducers used were  $L^- (+)$ -arabinose 98% (Sigma Aldrich, USA), N-3-oxo-tetradecanoyl-L-Homoserine lactone (Cayman Chemical, USA) and Anhydrotetracycline 98% (Cayman Chemical, USA).

**Cellular inks.** Cellular inks were elaborated by preparing a cell culture from single colonies obtained from streaked glycerol stocks and grown until reaching an optical density (OD) of 0.2. From these cultures, two different cellular inks were elaborated, one for ready-to-use circuits and another one to have circuits stored at  $-80^\circ\text{C}$ . For the first ones, these cell cultures were mixed with LB agar, prepared by adding 0.6 g of agar to 50 mL of LB and boiling it until having everything diluted. After cooling to  $\sim 50^\circ\text{C}$  properly, antibiotic selection and cell cultures were added to obtain each cellular ink. The use of agar in the mixture prevents cell culture dispersion when deposited on paper surface, providing some cellular confinement and to have a better control of the positioning of each cellular element. For circuits stored at  $-80^\circ\text{C}$ , the same principles were used but, in this case, a 20% v/v glycerol was used for the mixture, rather than the agar. Glycerol cryoprotects cells deposited on paper, so circuits can be thawed and used later.

More specifically, for the modulatory elements, a volume of 100  $\mu$ l of cell culture was mixed with 50  $\mu$ l of liquid LB-agar mixture. For the rest of elements, i.e. the  $S_1$ ,  $S_2$ , CA and CR cells, a volume of 50  $\mu$ l of cell culture was mixed with 50  $\mu$ l of LB-agar, also in liquid state.

**Paper properties.** The paper used for all experiments was 75 gsm White Paper (Pack of 2500) 59908. It has an optimal quality due to its eucalypto globulus fibres mixed with calcium carbonate. This combination allows to produce a 75  $\text{g}/\text{m}^2$  paper with necessary rigidity and opacity.

**Cell growth.** Because cell population grown in a paper surface cannot be directly estimated by optical density measures, all cell strains constitutively express RFP. We assume that cell population has a linear relationship with RFP levels as RFP expression at single-cell level is at the steady state. Supplementary Fig. 2 shows the temporal evolution of RFP levels associated to cell growth in a paper surface. Experimental results indicate that the different cell types have similar growth dynamics.

**GFP quantification.** Reporter cell CR expresses GFP in response to AHL diffused throughout paper surface. In order to characterize its induction range, several LB plates containing different concentrations of AHL, from  $10^{-4}$  M to  $10^{-10}$  M were prepared. Later, small pieces of paper were placed on the top of each plate and a dot of 1  $\mu$ l of the reporter cellular ink was added. The plate was grown at 37 °C overnight and after 24 h, a surface scanning was carried out, measuring GFP and RFP intensities. Measures were performed with *Synergy HXT- Hybrid Multi-Mode Reader* (BioTek Instruments, USA). Experimental data were collected with Gen5 software package and analysed with Microsoft Excel 2013. Supplementary Fig. 4 shows the CR response at different AHL concentrations. GFP levels were normalized by RFP that is assumed proportional to cell population. Experimental data was mathematically fitted to a Hill function

$$\text{GFP/RFP} = \alpha_{\text{AHL}} + \frac{k_0 \cdot [\text{AHL}]^n}{k_1 + [\text{AHL}]^n} \quad (1)$$

Fitting parameters are  $\alpha_{\text{AHL}} = 1$ ,  $k_0 = 33 \text{ M}^{-1}$ ,  $k_1 = 4 \times 10^7 \text{ M}$  and  $n = 1$ . Fluorescent measures for GFP: excitation wavelength:  $475 \pm 9 \text{ nm}$ , emission wavelength:  $509 \pm 9 \text{ nm}$ . Fluorescence measures for RFP: excitation wavelength:  $578 \pm 9 \text{ nm}$ , emission wavelength:  $616 \pm 9 \text{ nm}$ .

**Reporting summary.** Further information on research design is available in the Nature Research Reporting Summary linked to this article.

## Data availability

Plasmids and strains used in this study and other materials are available upon request to authors. Correspondence and requests for materials should be addressed to J.M. (javier.macia@upf.edu). DNA sequences for all constructs are provided as Supplementary Information. Genetic parts used in this work are available at Registry of Standard Biological Parts (<http://parts.igem.org/>). Source data are provided with this paper.

## Code availability

Scripts used to analyse the scalability of the circuits are available at [https://github.com/jmacia-upf/printed\\_circuits.git](https://github.com/jmacia-upf/printed_circuits.git).

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## Author contributions

J.M. developed the theoretical background for printed cellular devices. Circuits were designed by S.M. and J.M. S.M. and E.G. did the experimental designs. J.M. and S.M. wrote the paper.

## Competing interests

The authors declare no competing interests.

## Additional information

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