

Review



2021; 11(17): 8234-8253. doi: 10.7150/thno.59293

Obesity and severe coronavirus disease 2019: molecular mechanisms, paths forward, and therapeutic opportunities

Tiantian Yan^{1™}, Rong Xiao¹, Nannan Wang¹, Ruoyu Shang², Guoan Lin^{1™}

1. Military Burn Center, the 990th Hospital of People's Liberation Army Joint Logistics Support Force, Zhumadian, Henan, China

2. State Key Laboratory of Trauma, Burns, and Combined Injury, Institute of Burn Research, the First Affiliated Hospital of Army Medical University (the Third Military Medical University), Chongqing Key Laboratory for Disease Proteomics, Chongqing, China

🖂 Corresponding authors: Tiantian Yan, tiantianyan27031@163.com. Guoan Lin, guoanlin169@163.com

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/). See http://ivyspring.com/terms for full terms and conditions.

Received: 2021.02.10; Accepted: 2021.06.20; Published: 2021.07.13

Abstract

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) appears to have higher pathogenicity among patients with obesity. Obesity, termed as body mass index greater than 30 kg/m², has now been demonstrated to be important comorbidity for disease severity during coronavirus disease 2019 (COVID-19) pandemic and associated with adverse events. Unraveling mechanisms behind this phenomenon can assist scientists, clinicians, and policymakers in responding appropriately to the COVID-19 pandemic. In this review, we systemically delineated the potential mechanistic links between obesity and worsening COVID-19 from altered physiology, underlying diseases, metabolism, immunity, cytokine storm, and thrombosis. Problematic ventilation caused by obesity and preexisting medical disorders exacerbate organ dysfunction for patients with obesity. Chronic metabolic disorders, including dyslipidemia, hyperglycemia, vitamin D deficiency, and polymorphisms of metabolism-related genes in obesity, probably aid SARS-CoV-2 intrusion and impair antiviral responses. Obesity-induced inadequate antiviral immunity (interferon, natural killer cells, invariant natural killer T cell, dendritic cell, T cells, B cell) at the early stage of SARS-CoV-2 infection leads to delayed viral elimination, increased viral load, and expedited viral mutation. Cytokine storm, with the defective antiviral immunity, probably contributes to tissue damage and pathological progression, resulting in severe symptoms and poor prognosis. The prothrombotic state, driven in large part by endothelial dysfunction, platelet hyperactivation, hypercoagulability, and impaired fibrinolysis in obesity, also increases the risk of severe COVID-19. These mechanisms in the susceptibility to severe condition also open the possibility for host-directed therapies in population with obesity. By bridging work done in these fields, researchers can gain a holistic view of the paths forward and therapeutic opportunities to break the vicious cycle of obesity and its devastating complications in the next emerging pandemic.

Key words: Obesity; Coronavirus disease 2019; Metabolism; Immunity; Inflammation; Thrombosis

Introduction

The coronavirus disease 2019 (COVID-19) pandemic continues deteriorating, especially in the winter months before the effects of vaccination become perceptible. As of June 18, 2021, World Health Organization (WHO) had announced more than 177.11 million confirmed cases, including 3.84 million deaths, posing substantial threats to human life and worldwide health systems [1]. The clinical

manifestations of COVID-19 range from asymptomatic infection to critical illness, and outcomes vary widely from recovery to intensive care unit (ICU) admission and death. Special attention should be paid to critically ill patients due to the high mortality rate. An in-depth understanding of risk factors and molecular mechanisms underlying COVID-19 severity and therapeutic choices can be therefore invaluable in reducing mortality and saving lives.

Obesity, termed as body mass index (BMI) greater than 30 kg/m^2 , is identified as one of the most important independent risk factors for severe COVID-19. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) appeared to be more virulent among patients with obesity. These patients showed a greater tendency to develop adverse events, including respiratory failure, admission to ICU, and invasive mechanical ventilation. In the United States, individuals aged under 60 years old with BMI between 30 and 35 kg/m² and greater than 35 kg/m² were 1.8 and 3.6 times more likely to be admitted to ICU, respectively, compared with patients with BMI less than 30 kg/m² [2]. A similar phenomenon was observed in a large prospective analysis of 5279 COVID-19 patients [3]. A French study found that BMI greater than 35 kg/m² was associated with a 7.36-fold increase in requiring invasive mechanical ventilation than those with BMI lower than 25 kg/ m^2 , even after adjusting for other underlying diseases such as diabetes and hypertension [4]. The adverse consequence of obesity was further substantiated by with an its association increased risk of COVID-related death. In China, 88.24% of the deceased had a BMI greater than 25 kg/m^2 , while only 18.95% of the survivors were overweight [5]. According to the health analytics platform in England, one of the largest cohort studies to date of clinical factors associated with COVID-19-related death, HR was 1.92 (1.72-2.13) for BMI above 40 kg/m² [6]. This phenomenon is reminiscent of the H1N1 pandemic in 2009, in which obesity was also known as important comorbidity for increased disease severity and mortality [7].

Existing evidence from H1N1 and COVID-19 pandemic should serve as a note of caution for modern society. Obesity is such an alarming health problem nowadays that it has been declared as a chronic, progressive, and metabolic disease with low-grade inflammation rather than just a risk factor for other diseases [8]. From 1975 to 2014, the proportion of men with obesity worldwide increased from 3.2% to 10.8%, while this data grew from 6.4% to 14.9% in women [9]. With a higher expression of the cellular receptor angiotensinconverting enzyme receptor 2 (ACE2) in adipose tissue (AT), people with obesity are more susceptible to SARS-CoV-2 infection [10, 11]. Meanwhile, AT can serve as a reservoir after infection and spread SARS-CoV-2 to other organs, just as in influenza A virus, human adenovirus Ad-36, and cytomegalovirus infection [12]. Additionally, the health condition of people with obesity is further complicated by the

common presence of physiological, metabolic, and immune abnormalities. Coexisting comorbidities such as hypertension, chronic kidney disease, and cardiovascular disease all contribute to severe COVID-19. Taken together, the high prevalence of obesity, susceptibility to SARS-CoV-2 infection, tendency to develop complications, and higher mortality have necessitated the attention to the obese to save lives during the COVID-19 pandemic.

Although obesity has been linked to disease progression in observational or descriptive articles, and the reasons have been briefly discussed in several papers, the mechanisms underlying the COVID-19 pandemic disproportionately affecting patients with obesity remain to be systematically addressed [13, 14]. In this review, we expound on the potential mechanisms aggravating disease severity of patients with obesity admitted to hospital for COVID-19 from physiological changes, metabolism, immunity, cytokine storm, and thrombosis, hoping a holistic view and thorough understanding could aid tailored treatments and maximize patient benefit (Figure 1).

Basic physiological changes induced by obesity increase the susceptibility to severe COVID-19.

Obesity-related respiratory dysfunction

Respiratory dysfunction of individuals with obesity is characterized by low respiratory muscle strength compliance, and decreased lung accompanied by reduced vital capacity, increased residual capacity, and alveolar hypopnea. Excessive AT around the chest and abdomen leads to increased intra-abdominal pressure on the diaphragm, further reducing vital capacity and lung hypoventilation [15]. The direct mechanical effect of fat deposition contributes to elevated resistance and premature airway closure, resulting in gas retention, positive end-expiratory pressure, and even respiratory diseases such as sleep apnea syndrome [15]. The compensatory respiratory capacity of the obese subject is therefore inadequate and predispose COVID-19 cases with obesity to exacerbated respiratory dysfunction and worsening outcome [16].

Obesity-related visceral fat deposition

Studies from Italy and Germany, respectively, found that an increase of one unit or 10cm^2 in visceral fat area measured by CT was associated with an OR of 2.47% (95% CI = 1.02-6.02) or 1.37 (95% CI = 1.07-1.89) for the need of intensive care. The close connection between visceral adipose mass and critical illness was also found in research in China [14]. Ectopic fat deposition, especially in abdominal visceral adipose



Figure 1. The potential mechanistic links between obesity and severe COVID-19. After infection with SARS-CoV-2, innate and acquired immune responses in lean people are effectively activated to clear pathogen and infected cells with minimal inflammation and lung damage. The integrity of their vascular endothelial cells is well-maintained, with vessels possessing normal functions of contraction and dilation, anti-inflammation, anti-oxidation, and anticoagulation. However, obese patients have a higher viral load when exposed to a virus amount equivalent to lean people. Deficiency of anti-irral immunopathology such as pulmonary edema and hyaline membrane. The prothrombotic state of obesity driven in large part by endothelial damage, platelet hyperactivation, hypercoagulability, and impaired fibrinolysis inevitably link obesity with severe COVID-19 by promoting widespread thrombosis. COVID-19, coronavirus disease 2019; CHO, cholesterol; G, glycosylated; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

tissue, is common for individuals with obesity. The location of fat distribution determines metabolic risk. In contrast to the metabolic protective effect of gluteofemoral/leg fat, visceral fat obviously worsens insulin resistance, lipid metabolism disorders, and hypertension [17, 18] and also expresses more inflammatory cytokines such as tumor necrosis factor α (TNF- α) and IL-6 [19]. This secretory profile different from subcutaneous adipose tissue probably contributes to their variations in endocrine function and COVID-19 progression. Even for individuals with normal BMI, increased visceral adipose mass is associated with a greater risk of metabolic disease and mortality. This highlights the limitation of BMI and suggests that the increase in visceral adipose mass is more pernicious in health terms and probably better predict COVID-19 severity than BMI.

Common presence of other comorbidities

About one in five individuals worldwide have at least one underlying medical condition that puts them at increased risk of severe COVID-19 [20]. The comorbidities normally presented alone or in combination in the obese include chronic respiratory disease, cardiovascular disease, chronic kidney disease, type 2 diabetes (T2DM), non-alcoholic fatty liver diseases, and metabolic associated fatty liver disease [20, 21]. These diseases were identified as independent risk factors for poor COVID-19 prognosis, each contributing to disease progression through different mechanisms [20, 22]. For example, physiological abnormalities in patients with NAFLD such as impaired glucose and lipid metabolism, platelet hyperactivation and hypercoagulable state [23], the underlying liver fibrosis [24], up-regulated ACE2 [25], and a pronounced inflammatory response [26] probably all contribute to severe COVID-19. In the obese with comorbidities, the mechanisms of comorbidities aggravating COVID-19 and the changes caused by obesity itself synergistically lead to disease progression and severe condition.

Chronic metabolic disorder of obesity leads to the worsening condition of COVID-19 patients.

Dyslipidemia

Dyslipidemia, a common abnormal state of lipid metabolism in obesity, is characterized by decreased high-density lipoprotein (HDL) cholesterol, increased total cholesterol, triglyceride, and lowdensity lipoprotein (LDL) cholesterol. Lipid homeostasis is essential for normal lung physiology. For example, HDL acts as the main source of antioxidant vitamin E in type II alveolar epithelial cells [27] and promotes the growth of pulmonary fibroblasts [28] and the production of pulmonary surfactant [29]. As a universal and possibly contributory event in chronic lung diseases, dyslipidemia may also be involved in the pathogenesis of severe COVID-19 caused by obesity [30, 31].

High cholesterol in the circulation alters the biophysical properties and function of pulmonary surfactant, leads to cholesterol accumulation in macrophages and other immune cells, and then negatively affects the innate and acquired immune response in the lungs [30, 31]. These changes are believed to be critical contributors to the pathogenesis of asthma, pneumonia, acute lung injury, and other lung diseases, as well as increased susceptibility to pulmonary pathogens [30, 31]. For diet-induced obesity (DIO), excess fat will be stored in and around normally lean tissues such as the skeletal muscle, liver, and heart. The additional AT leads to an increase of up to 40% of cholesterol loaded into the lung, the most prominent organ affected by COVID-19 [32]. Moreover, hypercholesterolemia causes circulating cholesterol to be loaded into cells through apolipoprotein Ε, while chronic inflammation induced by obesity inhibits the synergistically unloading process, increasing cholesterol in cells and forming lipid rafts [33]. Cell-loaded cholesterol is essential for the cellular entry of various coronaviruses, such as swine deltacoronavirus and mouse hepatitis virus, and even enhances their pathogenicity in vivo [34, 35]. The same rule holds true, in varying degrees, for SARS-CoV [36] and its highly similar analog SARS-CoV-2 [32]. In the context of COVID-19, high cholesterol facilitates cellular infection of SARS-CoV-2 by augmented lipid raft formation, increasd viral entry sites on the cell surface, and subsequent binding of ACE2 and virus [32]. For people with obesity, the high cholesterol in their tissue and cells can increase viral load [32]. This finding is validated by studies in which the rapid decreases of total cholesterol and LDL in the blood, defined as hypolipidemia, have been found to be negatively associated with COVID-19 severity and death [37, 38]. The reason for hypolipidemia being a predictor of disease progression is that it possibly represents severe loading of cholesterol from the circulation into tissues, hence facilitating SARS-CoV-2 infection and COVID-19 severity.

As a carrier for various bioactive lipid species and lipoxygenase, HDL exhibits significant biological activities, including vasodilation, cellular cholesterol efflux capacity, anti-thrombotic, anti-apoptotic, anti-oxidative, anti-inflammatory, and anti-infectious activities. Besides, HDL also regulates toll-like receptors (TLRs), major histocompatibility complex II, B, and T cell receptors by affecting cholesterol utilization in lipid rafts, serving as a platform for both innate and acquired immunity [39]. In the context of dyslipidemic and inflammatory states induced by obesity, HDL showed reduced levels and functional defects characterized by altered composition, abnormal metabolism, and impaired biological activities mentioned above [39]. Should people with obesity get infected with SARS-CoV-2, they could worsening conditions brought experience bv subnormal and dysfunctional HDL. This hypothesis was given credence by the clinical significance of HDL, whose concentration decreased sharply after infection and changed accordingly with the prognosis in critical COVID-19 patients. The level of serum HDL was positively and negatively correlated with lymphocyte count and C-reactive protein, respectively. Besides, the fluctuations of HDL level were found to be in step with the results acquired from computed tomography examinations and nucleic acid tests, indicating that HDL level was negatively related to COVID-19 severity [40, 41].

The up-regulated expression of lipid metabolism genes in infected pulmonary epithelial cells suggests that SARS-CoV-2, possibly like the H1N1 influenza virus, can target cellular lipid signaling, synthesis, and metabolism to reshape host cells into the optimal environment for viral infection, replication, assembly, secretion, etc. Specific metabolic signatures in serum, liver, lung, and bronchoalveolar lavage fluid (BALF) of H1N1-infected obese mice were discovered, compared with infected lean mice of comparable age [42]. The higher levels of phospholipids, cholesterol, and fatty acids in the lungs harvested from obese mice were proved to be closely associated with pulmonary inflammatory overreaction and immune dysfunction, thereby driving more severe pathological changes [42]. According to the lessons learned from H1N1, we reasonably infer that dyslipidemia inherent in obesity will be amplified by COVID-19 and lay the foundation for organ damage and increased mortality.

Hyperglycemia

Obesity-related ectopic fat deposition, especially in the liver, a key insulin-sensitive organ, triggers changes in insulin signaling pathways and insulin resistance. In the inflammatory context of obesity, low-grade chronic inflammation is also present in pancreatic islet, which witnesses the accumulation of mounting immune cells, such as replicating resident macrophages and circulating monocytes. These cells reprogram the immune microenvironment of the pancreatic islet, giving rise to impaired β cell function, islet fibrosis, insulin resistance, and possible hyperglycemia [43, 44]. Chronic hyperglycemia can contribute to extensive histopathological changes in lungs, including altered alveolus morphology, thickened alveolar epithelial cells, narrow alveolar space, changed mucus secretion, and increased pulmonary capillary basal lamina and microangiopathy. The consequent decrease in lung function is related to fasting blood glucose index and the duration and severity of glycemic abnormalities Compared with normal people, [45]. obese individuals are more prone to deteriorated blood glucose after infected with SARS-CoV-2. Insufficient islet function combined with the direct destructive effect of SARS-CoV-2 on β cells that express ACE2 collectively precipitate acute hyperglycemia [46]. Previous studies identified T2DM history and hyperglycemia as risk factors for SARS death and fasting blood glucose as an independent predictor of death [47]. Consistently, clinical data demonstrated that abnormally elevated blood glucose (median ≥ 6.1 mmol/L) independently mediated high risk of critical progression to illness/death from mild/moderate COVID-19 cases as well as increased mortality in critically ill patients [48]. It is therefore reasonable that patients with obesity who are more prone to develop hyperglycemia have worse conditions.

Hyperglycemia contributes to severe COVID-19 probably by affecting both virus invasion and body responses. Glucose is not normally present in the thin layer of fluid that covers the epithelium of the respiratory tract, namely airway surface fluid [49]. This homeostasis is achieved by the glucose reabsorption of respiratory epithelial cells through sodium-glucose cotransporters. However, hyperglycemia can disrupt this balance and give rise to aberrant glycosylation of proteins, DNA, and lipids in epithelial cells, including ACE2, through directly increasing the glucose concentration in the airway surface fluid [49]. The increased glycosylated ACE2 is believed to be responsible for increased binding with SARS-CoV-2 and thereby the severity of COVID-19 infection [50]. In addition, the spike (S) protein of SARS-CoV-2 is cleaved into S1 and S2 subunit by protease furin. S1 binds to ACE2, while the highly glycosylated S2 subunit is responsible for the fusion of the SARS-CoV-2-ACE2 complex with the cell membrane and subsequent entry [50]. Conceivably, the combination of highly glycosylated S protein and ACE2 brought by hyperglycemia favors the cellular

invasion of SARS-CoV2 and following infection of multiple organs. Meanwhile, hyperglycemia also impairs major components of the immune system that copes with infection [51]. For instance, high glucose inhibits the production of type I interferon (IFN), leading to decreased antiviral activity at the cellular level [52]. Human and animal hyperglycemia-related studies both demonstrated the reduced formation of extracellular traps [53], impaired neutrophil activity (e.g., chemotaxis, phagocytosis) in a protein kinase C-dependent way, and thus failure of pathogen elimination [54]. Glycosylation of immune proteins under a hyperglycemic state also poses threats to immune function; for example, glycosylated complement C3 with abnormal tertiary structure inhibits the fixation with pathogens and decreases phagocytosis [55]. Therefore, hyperglycemia probably aids SARS-CoV-2 intrusion and impairs body antiviral responses, thereby causing severe symptoms and worse outcomes in obese patients.

Vitamin D deficiency

Unlike other disputed vitamins in obesity, vitamin D deficiency in people with obesity is highly prevalent and well-documented, as evidenced by the negative impact of high body fat percentage and BMI on serum 25-hydroxyvitamin D (the major circulating vitamin D metabolite) level [56]. Multiple comorbidities of obesity, such as impaired pancreatic islet function and cardiac metabolic diseases, are all associated with vitamin D deficiency [57, 58]. Physiological effects of vitamin D in the lungs include induction of autophagy, generation of antimicrobial peptides that lower viral replication rate, and synthesis of reactive oxygen intermediates and reactive nitrogen intermediates. Besides, vitamin D regulates the renin-angiotensin system (RAS) and maintains pro- and anti-inflammatory cytokines balance, thereby helping control respiratory tract infection [59, 60]. Accordingly, vitamin D deficiency can worsen COVID-19 by impeding the activation of defense pathways above, triggering cytokine storms, and aiding immune dysfunction [59, 61, 62]. The effects of inadequate vitamin D in COVID-19 are substantiated by a preliminary study showing a striking correlation between disease severity and the epidemiology of vitamin D deficiency regarding population, climate, and regions [61]. That is, it is logical to speculate that obese COVID-19 victims without sufficient vitamin D are predisposed to an increased risk of severe conditions.

Polymorphisms of metabolism-related genes

The formation of obesity is influenced by, environmental and genetic factors such as diet and

physical activities. Individual differences in weight gain and the occurrence of different phenotypes of obesity under a similar obesogenic environment indicate the significance of genetic differences. Currently, over twenty genes have been found to be involved in metabolism and fat deposition, among which melanocortin 4 receptor (MC4R), fat mass and obesity-associated gene (FTO), and peroxisome proliferator-activated receptor y (PPARy) have been extensively studied [63]. As the most common monogenic cause of obesity, deficient MC4R and the central melanocortin pathway together regulate energy intake, expenditure, and homeostasis [64]. FTO single nucleotide polymorphism is established as a key player in obesity and genetic aging by regulating dietary behavior, telomere length, and cell nutrition perception through amino acids [65]. In addition to regulating glucose and lipid metabolism, PPARy is also part of the cytokine loop as an anti-inflammatory player after viral infection, promising to be a therapeutic target for COVID-19 [66]. Previous researches reported that several individual gene polymorphisms such as interleukin 1 (IL-1), IL-1a, and interferon regulatory factor 9 (IRF9) are associated with disease severity and/or clinical outcome of influenza virus infections [67] and SARS [68]. Considering the close connection between metabolic gene polymorphisms and obesity, the possibility that these targets (e.g., MC4R, FTO, PPARy) in obese patients are more directly related to severe COVID-19 than BMI cannot be excluded. It is worthwhile to keep track of whether and how metabolic gene polymorphisms are related to severe COVID-19, helping pinpoint populations vulnerable to severe infection.

Dysfunctional antiviral immunity induced by obesity fails to eliminate SARS-CoV-2 and might explain severe COVID-19.

Once infected with SARS-CoV-2, the human body will initiate an antiviral response that depends on the coordination of well-functioning innate and acquired immunity to eliminate pathogens. However, the ectopic fat deposition in immune tissues such as the thymus, spleen, bone marrow, and lymph nodes lead to destructed tissue integrity and an altered cellular environment [69]. Consistent with the dependence of defensive function conferred by the immune system on the proper development and maturity of immune cells, the lymphoid tissue microenvironment undermined by obesity gives rise to alterations in the development, phenotype, diversity, and activity of immune cells. These alterations complicate and further perpetuate impaired innate and acquired immunity as well as their interactions, individually or synergistically increasing the risk of worse prognosis in obese patients infected with SARS-CoV-2 (Figure 2) [69-71].

IFN

IFN, which is classified into three types, is a kind of secretory protein induced by innate immune perceiving pathogen-related molecular sensors patterns through pattern recognition receptors such as retinoic acid-inducible gene I receptors and TLRs. In IFN-I, the most widely expressed and clearly defined is IFN- α and IFN- β , while IFN-II family consists of a single gene product IFN-y, and IFN-III is a variety of subtypes of IFN- λ [72]. The ubiquitously distributed receptors of IFN-I and IFN-II determined that they can act on most of the cells with extensive overlap between their signal pathways. Taking IFN-a as an example, it exerts pleiotropic effects in the combat against infection, including but not limited to enhanced responses of dendritic cells (DCs), natural killer (NK) cells, T cells, and B cells through differential activation of signal transducer and activator of transcription (STAT) molecules. The induction of interferon-stimulated genes (ISGs) by IFN-I largely contributes to restricting the viral replication cycle, while the upregulation of the major histocompatibility complex augments the lysis of infected cells [72]. IFN-III receptors are preferentially expressed on the epithelial cells of the respiratory system, gastrointestinal tract, and reproductive system. When combined with the receptor, IFN-III could locally control the viral infection at the entry sites such as the respiratory tract [73]. Altogether, IFN constitutes the first line of defense against the virus by establishing antiviral cellular status and activating subsequent immune responses.

In the long struggle for survival, viruses, including SARS-CoV-2, have adopted several strategies against antiviral immunity. Being different from the Middle East respiratory syndrome human parainfluenza coronavirus, virus respiratory syncytial virus, and influenza A virus, SARS-CoV-2 showed an extremely low level of IFN-I and IFN-III along with mild ISG responses in human, animals and cells in vitro [74]. The potential mechanism was that viral membrane protein prevented components such as retinoic acid-inducible gene I and mitochondrial antiviral signaling from forming multiprotein complex, resulting in impeded nuclear translocation and activation of IRF3 [75]. Corresponding to the laboratory evidence, clinical studies found that COVID-19 patients with severe IFN-a deficiency had longer ICU stay, higher

viral load, and worse prognosis [76, 77]. In addition to viral factors, specific host factors are also involved in determining the outcome of IFN signal transduction. Patients with obesity fail to launch a robust IFN-I response, resulting in the deterioration of H1N1 infection [78, 79]. It is conjectured that this destructive IFN response may increase the susceptibility of individuals with obesity to severe COVID-19. Things like reduced IFN production, faulted IFN signaling, pathogenic ISG effectors, and possibly accelerated viral mutation in obesity collectively lead to impaired antiviral ability and extensive cytopathogenic effect induced by uncontrolled SARS-CoV-2 at the early stage of COVID-19.

IFN production may be significantly reduced at first. Compared with the control group, obese mice infected with H1N1 showed a dramatically decreased level of IFN- α and IFN- β in the BALF and culture supernatant of alveolar macrophages. The viral load and mortality were also higher [79]. In line with animal experiments, H1N1 patients with obesity showed impaired production of IFN-I, probably related to leptin [80]. This reduction could be caused by the imperfect development of immune cells and their shift towards inflammatory phenotypes [70, 71]. Coincidentally, the decrease in IFN- γ production in obese people infected with influenza A virus is related to dysfunctional $\gamma\delta T$ cells, whose T cell receptor consists of γ and δ chain. The subnormal number of $\gamma\delta T$ cells, together with the surviving ones being unresponsive to TLR ligands and inactive IFN responses synergistically contribute to IFN- γ deficiency [81].

Moreover, the IFN signal transduction cascade may be inhibited. Typically, the binding of existing IFNs and the corresponding receptors leads to Janus Kinase-STAT being phosphorylated and translocating into the nucleus in a complex with IRF9. This trimeric binds to the complex then interferon-stimulated response elements and causes transcriptional induction of antiviral ISGs. However, chronically increased leptin produced by AT in obesity interferes with the IFN signal transduction through increasing suppressor of cytokine signaling 3, which negatively regulates the Janus Kinase-STAT pathway, causing the limited IFN in vivo to fail in inducing antiviral ISGs transcription [80, 82].



Figure 2. Severely deficient anti-viral immunity and its subsequent catastrophic effect in patients with obesity encountered SARS-CoV-2. Patients with obesity probably have significant defects in multiple aspects of innate and acquired immunity. IFN, the first pivotal defense line against the virus, exhibits reduced production, faulted IFN signal transduction, and pathogenic interferon stimulating genes, leading to uncontrolled viral replication and transmission. DCs with numerical and functional defects in obesity demonstrate impaired ability to stimulate naïve T cell expansion. The resulting insufficient number and unbalanced subsets of T cells combining obesity-related thymic degeneration, T cell senescence, impaired initiation, delayed timing of T cell response relative to viral replication, and other adverse factors in the context of obesity fail to elicit a potent immune response. The altered number and function of B cells and decreased antibody titers cannot neutralize and inactivate SARS-CoV-2. Coupled with the abnormalities of other cells such as NK cells, iNK cells, macrophages, and γδT cells, deficient anti-viral immunity caused by obesity probably drives the formation of systemic cytokine storm and progression of immunopathology, possibly followed by dysregulation of tissue repair. DC, dendritic cell; IFN, interferon; iNKT cells, invariant natural killer T cells; NK cells, NK cells, SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

The disordered IFN-I is pathogenic. In addition to expression level, the timing of IFN production is also important. As mentioned above, obesity would cause severe symptoms and adverse prognosis of H1N1 patients because of disordered IFN production and ISG responses [78-80]. Previous studies showed that IFN responses were more likely to delay rather than disappear. Instead of being protective against viral infection as supposed, disordered IFN-I response emerged as a critical driver for developing severe lung injury in SARS mice, facilitating massive immune cell infiltration, excessive inflammatory responses, vascular infiltration, and alveolar edema formation [83]. The aberrant intensity and duration of ISGs even determined the viral load and disease severity of COVID-19 patients [84]. Therefore, based on other coronaviruses' parallel immunological and pathophysiological features, it can be speculated that pathogenic IFN-I response in obesity as shown in H1N1 can also drive the immunopathological changes, COVID-19 progression, and mortality. However, the dynamics and interaction of viral replication and the IFN signaling pathway are still an open question that deserves further attention. Identifying the turning point of IFN function from protective to pathogenic is crucial for determining the therapeutic window.

Finally, ineffective IFN responses facilitate viral mutation. As a typical RNA virus, SARS-CoV-2 shares the characteristic of genetic material being highly mutable. The influence of mutations on viral replication kinetics and infectivity in vitro and clinical significance in vivo has been demonstrated in mounting reports [85]. For example, strain B.1.1.7 is a new variant more transmissible and likely to escape from human immunity. Being recently brought to light and became the dominant strain in the London area, strain B.1.1.7 prompted renewed alarm and escalation of control measures [86]. As shown in obese mice with H1N1, decreased type I IFN responses were associated with the rapidly emerged viral variants exhibiting increased replication, enhanced pathogenic, and more diverse quasispecies [87, 88]. The results showed that obesity-related impaired IFN responses afford the emergence of a more virulent virus population capable of inducing greater disease severity. This phenomenon highlights the need for vigilance in the clinical management of COVID-19 patients with obesity, as the emergence of potentially pathogenic variants is not specific to a particular obesity model or virus strain but a regular pattern [87, 88]. The analogy between the H1N1 virus and SARS-CoV-2 makes people wonder whether SARS-CoV-2 mutation, infectivity, and virulence will be influenced by obesity. Thus, further independent

studies are needed to confirm or disprove this idea.

NK cells

NK cells directly kill infected targets by secreting particles containing apoptosis-inducing lytic granzymes and perforin, thus acting as major players in protective immunity against viral infections. It is shown that the increased risk of cancer and infections attributable to numerical and functional defects of NK cells in human and experimental animals with obesity [89-91]. The cytotoxic activity of NK cells in obese mice infected with influenza virus was 50% lower than that in the control group, and the NK cell-mediated killing was also naturally reduced [89]. This phenomenon is related to the abnormal metabolism of NK cells caused by obesity. Once mobilized, NK cells would show special metabolic flexibility, converting from oxidative phosphorylation to glycolysis to meet the increased energy needs [91, 92]. The increased PPARs in obesity drive the inhibition of glycolysis, downregulated production of IFN-y and cytotoxic granules in NK cells, leading to blunted antiviral activity [90]. Lipotoxic obese environment such as the chronic elevation of LDL, triglyceride, and circulating free fatty acids (FFAs), induces a large accumulation of lipid in NK cells, thus interferes with cellular bioenergetics and results in complete "paralysis" of cellular metabolism and transition [90]. However, the effect of obesity on NK cells goes far beyond metabolic reprogramming. High levels of IL-6 secreted by AT could downregulate NK cell cytotoxicity by inhibiting the expression of granzyme B and perforin without altered granule exocytosis in both obese human and murine settings [93]. In contrast to the central role of leptin in modulating NK cell number and activity in lean individuals, post-receptor signaling components (protein kinase B pT308, Janus kinase-2p) of leptin differentially abrogated despite were the up-regulated leptin and leptin receptors in subjects with obesity [94]. This functional desensitization is also responsible for the insufficient function of NK cells in obesity. In addition, the frequency of NKG2A-positive NK cells increased as BMI elevated in patients with new-onset of T2DM [95]. Binding to non-classical human leukocyte antigen E, NKG2A suppressed cytotoxicity of NK cells and contributed to the poor control of human immunodeficiency virus 1 infection. Analogously, BMI-related increased NKG2A in obesity may also facilitate the immune escape of SARS-CoV-2 [96].

In conclusion, NK cells in people with obesity cannot limit SARS-CoV-2 transmission as expected, nor restrain systemic inflammation by killing the activated inflammatory dendritic cells, monocytes, or T cells [97]. Although this hypothesis needs further clarification, it alludes to potential mechanisms for poor outcomes of COVID-19 patients with obesity.

Invariant natural killer T (iNKT) cell

iNKT cells constitute a distinct T lymphocyte subset co-expressing NK-lineage receptor and an invariant T-cell receptor. After being activated, the iNK T cell, as an immunomodulator, mobilizes other immune cells to participate in the anti-viral chorus by secreting cytokines such as IL-4, IL-10, and IFN-y. The expression of effector molecules on iNKT cells like perforin and Fas ligand also endows cytotoxic effects [98, 99]. Under physiological conditions, iNKT cells were found highly abundant in mice and human AT. However, the number of iNKT cells decreased as AT expanded during the development of obesity and correlated inversely with the infiltration of pro-inflammatory macrophages. This reduction can be restored by weight loss of humans and mice [100]. Moreover, the vitro studies showed that the cytokine secretion and resultant function of surviving iNKT cells were inhibited by leptin and leptin receptor pathways which also reduced iNKT cell number [101]. It is therefore conceivable that once confronted with SARS-CoV-2, iNKT cells with numerical and functional defects in patients with obesity will be unable to link antigen-presenting cells with local or acquired immune cells (NK cells, T cells, and B cells) and are not conducive to virus clearance [98].

DCs

DCs are critical immune sentinels in response to viral infection. As the most potent and exclusive professional antigen-presenting cells, DCs can trigger naïve T cells and function critically in initiating and maintaining cellular immune responses. The crosstalk between DCs and adipocytes maintains or even dominates the immune homeostasis of AT, indicating the critical role of DCs in AT [102, 103]. D O'Shea et al. found that circulating DCs in the obese showed significantly reduced count and functional defects featured by lower expression of CD83 following TLRs stimulation when compared with lean controls. As an essential molecule implicated in the elicitation of T cell responses, the insufficient level of CD83 leads to declined antiviral capacity and an increased likelihood of severe viral infection in patients with obesity [104]. Relatively high levels of cytokines such as granulocyte-macrophage colony-stimulating factor and IL-6 induced by obesity can result in the pre-activation of DCs with, however, increased risk of activation-induced dysfunction and apoptosis. Despite the similar phenotype in the control group, DCs in DIO mice showed blunt ability to stimulate

naïve T cell expansion because of upregulated serum cytokines and chemokines, including IL-1 α , IL-17, and TNF- α [105]. Moreover, obesity also impairs the migration of DCs to lymph nodes [106]. Therefore, DCs in the obese are not competent to initiate required immune responses to cope with the pathogen, especially lately emerged SARS-CoV-2 that carries novel antigens first encountered by the human body.

Further support for the impaired function of DCs in patients with obesity is suggested by their impaired antigen presentation and inadequate capability of guiding antiviral orchestration of T cells when encountered influenza virus. This defect was closely up-regulated associated with IL-6-driven proinflammatory state in the lungs of DIO mice [107, 108]. Similarly, one of the important immune features of severe COVID-19 was the dramatic reduction of the proportion of myeloid and plasmacytoid DC in BALF. In particular, the degree of plasmacytoid DC depletion correlated strongly with disease severity [109, 110]. In the meantime, COVID-19 also leads to low expression of maturation markers in DCs, high expression of programmed cell death ligand 1, inability to upregulate DC80 and CD86 in response to ssRNA and other stimuli, decreased secretion of IFN- α and IFN- β , and impaired ability to stimulate T cell proliferation [111, 112]. Considering the weakening of antiviral responses and viral clearance by altered DCs in people with obesity, the close causality between the development of severe COVID-19 and abnormal DCs is therefore reasonable.

T cells

As the indispensable components of the immune system, T cells harmonize and maintain many aspects of anti-viral responses. Recently, Cañete et al. have highlighted the importance of T cells in response to SARS-CoV-2 by showing that virus-specific memory T cells might achieve immunologic memory despite the blunt long-lived antibody responses in COVID-19 [113]. Hence, impaired antiviral responses and deteriorated COVID-19 may occur in the setting of detrimentally altered development, number, and function of T cells consequent to obesity.

Immune senescence, commonly seen in the elderly, is accelerated by obesity as it promotes thymic degeneration and T cell senescence [71, 114]. This aging immunity was even found in children with obesity [115]. T lymphocyte DNA hypermethylation in humans and animals with obesity was epigenetic evidence supporting obesity-related T cell senescence [116, 117]. Article published by Williamson et al., one of the largest cohort studies on clinical factors associated with COVID-19-related death so far,

showed that aging is strongly associated with the risk of COVID-19-related death [6]. The risk for patients aged \geq 80 years old increased more than 20-fold than for those aged 50-59 years old [6]. By analogy with the significant contribution of age-related immuno-senescence to the high mortality rate of elderly patients, it is reasonably speculated that similar deficient immunity in the COVID-19 patients with obesity may also confer susceptibility to disease exacerbation and fatal outcomes [118].

As mentioned above, malfunctioning DCs from DIO mice result in impaired initiation of specific T cell responses [105]. In high-fat diet-fed animals and humans, CD4+ T cells preferentially differentiated toward into a type of effector memory-like T cell which was independent of the crosstalk between DCs and T cells [119]. This non-specific differentiation depletes the number of naïve and memory T cells, leading to decreased proportion of specific T cells that respond effectively to foreign antigens. Multi-omics studies of patients suffering from COVID-19 demonstrated that CD4+ naïve T cells could also differentiate into a group of clonally expanded CD4+ cytotoxic phenotype [120]. nonspecific Insufficient initiation and nonspecific depletion combine to render T cells unable to mobilize rapidly and effectively when confronted with novel antigens like SARS-CoV-2, leading to insufficient supply of effector and helper T cells owning specific antiviral functions. In addition, previous data showed that DIO could lead to a 44% decrease in CD8+ T cells [121], while controversial studies claimed that the number and frequency of influenza virus-specific CD8+ T cells in the lung, rather than the total population, were affected by obesity [108]. One of the prominent immunological features among severe COVID-19 patients was lymphopenia with decreased CD4+, CD8+ T, and Treg cells. Obesity and lymphopenia, especially preferential decline in CD8+ T cells, can serve as predictors of poor prognosis in COVID-19 patients [122]. The decrease in T cells arising from coronavirus infection and obesity together conduce to a higher possibility of critical condition in patients with obesity. However, the reciprocal interplay between obesity and disease progression may not be unilateral. On the one hand, the immune defenses of patients with obesity are inherently weakened because of immune senescence and inadequate T cells. On the other hand, the declined naïve T cell response is crucial in an exuberant inflammatory response and cytokine storm [123]. It is worth keeping track of whether the number or frequency of CD4+ and/or CD8+ T cells in peripheral blood and pulmonary inflammatory tissue decrease in obese patients with severe COVID-19. More efforts are warranted to

ascertain the mechanistic role of these alterations in disease pathogenesis and progression.

Functional defect of T cells across multiple species with obesity is also a critical point to be reckoned. Compared with nonobese controls, both human and animal models with obesity consistently revealed attenuated proliferative capacity and increased exhaustion of T cells, as indicated by down-regulated ki67 and up-regulated PD-1, respectively. This is further corroborated by the reduced IFN-y and TNF-a production in stimulated polyclonal T cells from subjects with obesity [124]. These dysfunctions could increase cell apoptosis and prevent T cells from proliferating and activating effectively when exposed to acute viral infections. The chances of obese COVID-19 patients with obesity becoming seriously ill are bound to be significantly increased in the absence of effective antiviral T cell response. These concerns were substantiated by the runaway influenza virus and worsening illness experienced by obese individuals with influenza [125]. Viruses such as SARS-CoV-2 can negatively affect the T cell response. Two unusual subtypes of CD4+ T cells called clonally expanded cytotoxic phenotype, and proliferative exhausted phenotype were associated with disease severity in multi-omics studies, while the polyfunctionality and cytotoxicity of CD8+ T cells were attenuated in severe COVID-19 patients [120]. It thus logically follows that malfunctional T cells caused by a combination of host and viral factors predispose COVID-19 patients with obesity to the failure of virus control and more severe conditions. The reasons for T cell dysfunction in patients with obesity are various. First of all, high leptin levels led to the upregulation of phosphorylated STAT3, which interacted with the PD-1 gene promoter and induced its expression on T cells, leading to cell exhaustion [124]. Then T cells switch the balance of the metabolic program from catabolism in the quiescent state to anabolism in the activated state to meet the energy required by the effective immune response to kill pathogens. Compared with ordinary people, extra increases in glucose, FFAs, cholesterol, phospholipids, and other metabolites in people with obesity mechanically alter the metabolism of T cells, leading to activation disorder and declined activity [42, 119, 126]. Finally, T cell dysfunction in people with obesity is also partially driven by IFNs deficiency at the early stage of being of infection. Instead protective and stimulative in T cell response, subsequent delayed IFNs inhibit T cell proliferation, prevent its outflow from lymphatic organs, and cause T cell exhaustion and death [127].

The T cell immunity in response to SARS-CoV-2

can act as a double-edged sword being antiviral effective or driving immunopathological development [128, 129]. The choice of being "friend" or "foe" depends on its timing of initiation, duration, and level, possibly. T cell response was found to peak around one to two weeks after SARS-CoV-2 infection while the patient was turning to the early clinical stage from the onset of symptom [130]. For patients with obesity, further delay and incompetence of antiviral T cell response in the early stage of COVID-19 would happen because of the impaired initiation of naïve T cells after receiving antigen presentation, decreased proliferation, and impaired function of T cells. These T cells fail to control highly replicating and possibly mutating viruses, and tend to fuel cytokine storms instead. In addition, obesity can also lead to altered cell subsets, with increased pro-inflammatory Th1 and Th17 cells and decreased anti-inflammatory Th2 and Treg cells, contributing to excessive inflammatory reaction [131]. The occurrence of progression from mild to severe cases of COVID-19 mainly in the second week from the onset of symptoms coincided with the obesity-delayed T cell response. Researches revealed the heightened chemokines (CXCL8, CXCL9, CXCL10) induced by circulating IFN and activated cytolytic Th1 cells phenotype in severe COVID-19 patients, yet their virus-specific responses were similar to that in mild patients [132].

Epithelial $\gamma\delta$ T cells are worthy of attention because of their unique connection with epithelial tissue. Except for the protective antiviral function in influenza pneumonia, $\gamma\delta$ T cells can provide growth factors, recruit macrophages, and neutrophils during wound repair. $\gamma\delta$ T cells also can induce DC maturation, interact with Tregs, and participate in the restoration of epithelial barrier [133]. However, in COVID-19 patients with obesity, $\gamma\delta$ T cells are negatively subjected to the combination of viral and host factors possibly. A decreased number of $\gamma\delta$ T cells were found in COVID-19 cases [134]. In the inflammatory context induced by obesity, $\gamma\delta$ T cells showed reduced number, impaired homeostasis and antiviral function in humans due to their high sensitivity to inflammation, leading to poor repair of lung lesions and worse pathological changes [81]. In unraveling the mysteries in COVID-19 victims with obesity, more data are needed to ascertain the mechanistic role of altered vo T cell responses from the overlay of obesity and SARS-CoV-2 in immunopathology and disease progression.

B cell

Obese individuals infected with the H1N1 influenza virus showed lower levels of specific

antibodies and weaker neutralization ability [135, 136]. This can be ascribed to various reasons, such as the inflammatory environment in vivo, DNA hypermethylation of B cells, and abnormal leptin that fails to regulate the development, maturation, and activity of B cells [116, 117]. The peripheral B cell pool in subjects with obesity is characterized by the increased proportion of pro-inflammatory late/ exhausted memory B subsets and decreased anti-inflammatory transitional B cells [137]. In addition to the reduced number and imbalanced subgroups, functional defects of B cells in individuals with obesity possibly participate in the formation of cytokine storm by secreting more pro-inflammatory cytokines (e.g., IL-6). It is rational that obesity leads to severe COVID-19 by altering the number and function of B cells and their interaction with other lymphocytes (such as T follicular helper cells) through the inflammatory environment and adipokine imbalance.

SARS-CoV-2 infection elicits a potent humoral immunity critical for the clearance of cytopathic viruses which is proved by rapid and near-universal detection of virus-specific antibodies in the several days following infection [138]. There is little information regarding host response and viral dynamics following SARS-CoV-2 infection in populations with obesity. The higher risk of more severe clinical prognosis with higher titers of antibodies suggests that robust antibody response alone is insufficient to avoid severe illness [139]. Considering that antibody-dependent cellular cytotoxicity probably aggravates cytokine storm and disease conditions in mice with obesity, it is necessary to find out whether a similar situation is present in obese patients with COVID-19 [140].

Obesity creates a conducive ground for the cytokine storm in COVID-19 cases.

SARS-CoV-2 can elicit an exuberant cytokine storm both in vitro and in critically ill patients, a pathological state in which cytokines are rapidly released to form a hyperinflammation cascade. The occurrence of immunopathology, multiorgan failure, and even death in COVID-19 are at least partially to be blamed on cytokine storm [141, 142]. This finding parallels the situation described in SARS [143] and H1N1 [144], in which similar dysregulation drives the tissue damage and pathological progression. Data about the cytokines in COVID-19 patients with obesity are currently scarce. However, previous studies on H1N1 influenza have provided noteworthy findings. Compared with lean mice, the expressions of inflammatory cytokines such as IL-6, TNF-a, and IL-1 β and chemokines such as monocyte chemotactic protein 1 in the lungs of mice with obesity were

delayed and decreased in the early stage after infection [79, 89, 145]. However, excess cytokine secretion then occurs and resulted in cytokine storms in mice with obesity [42, 89, 145]. Similarly, obesity also predisposes patients infected with SARS-CoV-2 to systemic cytokine storm, possibly, owing to different but synergetic mechanisms.

1) Low-grade inflammation in obesity. Obesity is a chronic low-grade inflammatory disease featured by elevated levels of inflammatory cytokines. Initiators of this inflammation include increased circulating FFAs, mechanical stress caused by AT expansion, hypoxia, damage-associated molecular proteins adipocyte released by death, gut-derived lipopolysaccharide, etc. [21, 146]. NF-KB is considered as a signaling intersection in the consequent signal transduction of all potential initiators above, leading to the recruitment of immune cells and secretion of pro-inflammatory cytokines. These cumulative factors lead to hypercytokinemia and chronic systemic inflammation, conducive to hyper-inflammatory response in COVID-19 patients with obesity [142].

2) Impaired virus clearance and high viral load. physiological Chronic changes, disordered metabolism, and dysregulated immune responses in individuals with obesity probably converge to impaired clearance of coronavirus in the early stage of COVID-19. Compared with patients infected with normal BMI, patients with obesity have higher viral load and reach peak earlier, as evidenced in DIO mice infected with H1N1 [87, 145]. This impaired elimination and high viral load probably fuel extensive virus-induced direct cytopathic effects in the early stage and the resultant production of proinflammatory cytokines such as IL-6, TNF-a, and IL-1 β in infected epithelial cells [136].

3) High possibility of the virus spreading to the lower respiratory tract. Compared with control mice with the same viral titer, more influenza virus antigens were present in the bronchioles and alveolar regions of obese mice, suggesting increased virus transmission to the lower respiratory tract [145]. Studies in animal models, especially in mice, have demonstrated that different infection sites of human coronavirus can result in distinct outcomes. Compared with infection mainly distributed in respiratory epithelial cells, simultaneous infection of respiratory epithelial cells and alveolar epithelial cells were more likely to cause direct cytopathic effects and severe lesions [147]. SARS-CoV-2 in obesity probably also spreads more viruses to the lower respiratory tract and causes more serious lesions.

4) Hyperglycemia. Impaired pancreatic function and/or hyperglycemia existing in obesity probably are further worsened by SARS-CoV-2 infection which attacks ACE2-positive islet cells [46]. The resulting hyperglycemia can cause the glycation of proteins and lipids and generate advanced glycation end products that facilitate the production of reactive oxygen species (ROS) through NF-kB-dependent activation. Together with ROS produced by oxidative phosphorylation of excessive blood glucose, these metabolites activate NLRP3 inflammasomes and subsequent IL-1 system [44], thereby fueling cytokine storm, one of the most prominent findings related to hyperglycemia [51].

5) Disordered adipokines. In addition to acting as an energy-storage depot, AT can also secrete adipokines with an immunomodulatory effect as an endocrine organ. The excessive production of proinflammatory leptin and resistin juxtaposed to insufficient anti-inflammatory adiponectin in obesity partially justifies the imperfect immune responses to SARS-CoV-2 [145]. High plasma leptin in subjects with obesity, along with CXCL-10 and TNF-a, is a predictor of COVID-19 severity and disease progression. Mechanistic studies have revealed that leptin promotes inflammatory M1 polarization of monocytes through STAT3 and NF-KB signaling pathways and upregulates the secretion of cytokines such as IL-6, resulting in excessive inflammatory responses [148]. In LPS-induced acute lung injury, resistin increased the sensitivity of neutrophils to LPS exposure and promoted the formation of neutrophil extracellular trap, production of pro-inflammatory cytokines, and the aggravation of pulmonary edema [149]. The process of cytokine storm probably was further amplified by the intricate cross-talk between decreased adiponectin and macrophages. Adiponectin induced the production of IL-10 in macrophages, inhibited NF-KB, and directly regulated the expression of IL-1 receptor-associated kinase M which controlled the response and tolerance of macrophage to proinflammatory stimuli [150, 151]. Therefore, the immune system of people with low plasma adiponectin levels tends to overreact to pathogens. The lack of negative feedback anti-inflammatory mechanisms is probably conducive developing the cytokine storm to and the susceptibility to fatal outcomes following SARS-CoV-2 infection.

6) Imbalanced RAS. The overall expression level of ACE2 in people with obesity is higher than in their nonobese counterparts. ACE2 falls off after binding with the spike protein of SARS-CoV-2 and therefore fails to produce Angiotensin (1-7), which is supposed to act on the Mas receptor and being protective in vasodilation, anti-inflammation, anti-oxidation, etc. [152]. An estimated higher viral load and a possible greater drop of ACE2 in the obese after infection shift the balance of RAS to the pro-inflammatory axis of ACE/Angiotensin II/Angiotensin type 1 receptor, conducive to worse pulmonary vascular permeability, alveolar epithelial cell apoptosis, pulmonary inflammation, etc. [10].

7) Dysregulated IFN-I response. Instead of eliminating coronavirus, dysregulated, delayed, and persistent IFN-I in obesity can selectively induce the expression of transcription factor IRF1 in a STAT1-dependent manner. This is followed by the activation and transcription of pro-inflammatory cytokine genes, including IL-1 β , IL-6, and TNF- α , potentially contributing to the development of cytokine storm [153, 154].

8) Inflammatory macrophages. Macrophages account for 40-60% of immune cells in AT in mice with obesity, while only 10-15% in lean mice, standing out as the most abundant immune cells of concern in obesity [155]. In individuals with obesity, lipid droplet accumulation, FFAs, local hypoxia, and lipopolysaccharide all polarize macrophages to the pro-inflammatory phenotype M1(or more specifically, metabolism-activated macrophage phenotype) [156], secreting enormous inflammatory cytokines including IL-6, IL-1β, and TNF-a, and acting as key mediators of inflammation in AT [155]. Coexisting conditions in COVID-19 patients with obesity like dysregulated IFN-I response, delayed T cell response, disordered adipokines, and extensive virus-induced cytopathic effects orchestrate excessive infiltration of macrophages in the lung parenchyma. Coincidentally, severe/critical COVID-19 cases are featured by the enormous highly inflammatory macrophages that in turn bring about excessive cvtokines production of inflammatory [110]. Subsequent recruitment of T lymphocytes and their consequent crosstalk with macrophages through release of cytokines such as IFN exacerbate the inflammatory responses in a vicious circle and culminate in cytokine storm and disease progression. Speranza et al. supported this by finding that alveolar macrophages drove the inflammatory responses during SARS-CoV-2 infection [157]. In addition, the glycosylated Fc terminal of IgG shows a stronger affinity for the FcyRIIIa of macrophages, causing macrophage overactivation. Cytokines such as IL-1 β , IL-6, and TNF- α amplify the potential inflammatory storm, undermine the pulmonary endothelial integrity, and thus trigger microvascular thrombosis, facilitating the development of severe COVID-19 [158, Obesity-related hyperglycemia 159]. probably increases IgG glycosylation and sets the stage for uncontrolled cytokine storm [83, 89]. In summary, the question of whether and why the cytokine storm will form in subjects with obesity remains an enigma.

Therefore, more studies are needed to clarify the role of the pro-inflammatory events proposed above in SARS-CoV-2 infection.

The prothrombotic state in patients with obesity confer susceptibility to severe COVID-19 and poor prognosis.

Universal studies have consistently found that thrombotic microangiopathy, arterial thrombosis, and venous thrombosis are important causes of COVID-19 progression and increase mortality [160]. Post-mortem examination of patients with COVID-19 reveals extensive microthrombosis in the lungs, lower extremities, hands, brain, heart, liver, kidney, etc. Primary microthrombus in pulmonary vessels can explain the sudden progression to acute respiratory distress syndrome with marked pulmonary edema and hypoxemia, possibly, while systemic thrombosis contributes to loss of life through multiple organ dysfunction such as acute liver and kidney failure, neurological insults, cardiomyopathy, and mesenteric ischemia [161-163]. The hypercoagulable state in patients with COVID-19 is mainly manifested as hypofibrinolysis and increased thrombin generation [164, 165]. COVID-19 is also featured by the increased expressions of circulating markers for endothelial injuries, such as von Willebrand factor, angiopoietin 2, plasminogen activator 1, follistatin, and soluble thrombomodulin, among which the first three markers are particularly high in patients in ICU while the last four markers are positively correlated with mortality [160, 166]. Hypoxia, vascular endothelial injury, oxidative stress and other stressors in COVID-19 can cause platelet hyperactivation and apoptosis by affecting the metabolism and function of its mitochondria, facilitating platelet-rich thrombotic microangiopathy and critical disease. Additionally, the binding of activated platelets to neutrophils recruited in pulmonary vessels and the subsequent rolling on the endothelium plays a vital role in initiating immune thrombosis [167]. It logically follows that pre-existing risk factors in obesity, including endotheliopathy, platelet hyperactivation, hypercoagulability, and impaired fibrinolysis, probably lead to exacerbated thrombosis and severe COVID-19.

1) Endotheliopathy. Due to long-term exposure to inflammatory stimulation, systemic oxidative stress, hyperlipidemia, and other stimulating factors in obesity, endothelial dysfunction act as a principal determinant of microvascular dysfunction by tilting the vascular equilibrium towards constriction. Meanwhile, blood vessels show decreased antioxidant defense approaches and the loss of antithrombotic properties, owing to increased ROS production, adhesion molecules, and endothelin-1 [168]. 2) Platelet hyperactivation. The increased platelet size and volume in subjects with obesity represents higher susceptibility to activation, while elevated expression of CD40L and P-selectin in vivo confirms continued activation of platelets and endothelial cells [169]. Weight gain also weakens the sensitivity of platelets to insulin, prostacyclin, and nitric oxide and endows platelets a higher prothrombotic response to aggregators [170, 171]. The fatal combination of endotheliopathy and platelet hyperactivation in individuals with COVID-19 is highlighted by the association between increased positive ICU admission/death and elevated levels of biomarkers of this alliance [160, 166]. 3) Hypercoagulability. Elevated levels of tissue factor, von Willebrand factor, prothrombin, fibrinogen, and factors VII and VIII place patients with obesity in a mild to moderate hypercoagulable state [172]. 4) Impaired fibrinolysis. Obesity impairs the fibrinolytic system by significantly upregulating the expression of plasminogen activator inhibitor 1 which inhibits plasminogen activator and limits the dissolution of fibrin clots [173]. Together, concomitant events in obesity favor a prothrombotic environment and contribute to thrombosis following a positive COVID-19 diagnosis, possibly.

Potential therapeutic opportunities for COVID-19 individuals with obesity

Unfortunately, a proverbial "silver bullet" for severe COVID-19 has not been found yet due to its multifaceted causes. A multi-pronged approach is therefore needed to manage the current crisis. In addition to the medicines disrupting viral replication and dissemination, the enormous benefits of modifiable lifestyle like diet and physical training in obesity should not be marginalized. For example, moderate weight loss and physical activity can reshape AT distribution, increase muscle mass, HDL levels and insulin sensitivity, and even augment vitamin D synthesis when outdoor activities are involved.

Host-directed therapies that restrain abnormalities and restore required responses in patients with obesity are also available as alternatives. Good adherence to prior treatment and control of comorbidities is indispensable for people with underlying diseases. It was once suspected that angiotensin receptor blockers/angiotensin-converting enzyme inhibitors probably increased individual susceptibility to COVID-19 and worse outcomes through the upregulation of ACE2. However, discontinuation of these two types of drugs in hypertensive patients is not recommended according to the large-scale retrospective cohort studies that disproved this doubt [174, 175]. The contribution of dyslipidemia to COVID-19 susceptibility and severity in patients with obesity opens the possibility that lipid-lowering therapeutics probably ameliorate the consequences brought by high cholesterol in circulation and/or tissues. For example, statins, a class of shelved lipid-lowering medicine drug, was verified to be associated with lower mortality. The 28-day all-cause mortality in the matched non-statin and statin groups was 9.4% and 5.2%, respectively, supporting statins as adjuvant therapy for COVID-19, especially in patients with obesity [176]. As for hyperglycemia, improvement in morbidity by well-controlled blood glucose (fluctuating within 3.9-10.0 mmol/L) in patient outcome has been demonstrated by Zhu et al. who has proved that this glycemic variability is associated with lower mortality compared to those whose upper limit of blood glucose exceeded 10.0 mmol/L during hospitalization [177]. Therefore, early detection and glycemic control in the clinical management of COVID-19 is crucial. It should be noted that medications like glucocorticoids, severe illness, and infection often necessitate timely and reasonable adjustment of hypoglycemic drugs and/or insulin dosage based on their impairment of insulin sensitivity. In addition, observational studies showed the safety and merits of vitamin D supplementation in acute respiratory infections. The obese suffering from severe vitamin D deficiency possibly benefits the most from supplementation [60]. However, randomized clinical trials vielded controversial results [178]. Specific recommendations for vitamin D supplementation in COVID-19 patients still need to be clarified, but patients with obesity should be beyond the scope of the debate since correcting their vitamin D deficiency is what they need. Therefore, exposure to sunlight and, if possible, artificial ultraviolet Β, and vitamin D supplementation seem to be appealing adjunct therapies for obese patients.

Given the distinct roles of IFN-I in obese patients at different stages of COVID-19, an IFN-based therapeutic schedule should be refined in specific situations. During the acute phase of SARS-CoV-2 infection, prophylactic treatment with IFN-I, which is immuno-stimulatory and anti-viral effective, can confer maximum protection without causing significant systemic inflammation and appreciable pathology. Once delayed and persistent production of IFN-I drives excessive inflammatory responses and progressive tissue damage, the optimal option is to block its pathways downstream. However, the non-inflammatory, long-lasting, and focused expression pattern of IFN-III allows it to be applied through the whole course of treatment, providing lasting anti-viral protection at the entrance and preferential targets of SARS-CoV-2 (the upper respiratory tract and lungs) [87]. Notably, IFN-stimulated expression of ACE2 in the target cells of SARS-CoV-2, thus enhancing viral infectivity, has introduced more complexity to the clinical application of IFN in COVID-19 management [179]. Further unbiased and strictly controlled studies with granularity are warranted to develop rationale-based interventions and maximize therapeutic effect.

Obtaining the dynamics of cytokines in COVID-19 patients with obesity is pivotal since different degradation rates of various cytokines narrow down the therapeutic window for their respective antagonists. According to the development of cytokine storm responsible for severe COVID-19, strategies directed at attenuating specific processes are beneficial, possibly. This speculation has been supported by the clinical results obtained from the treatment of severe COVID-19 with tocilizumab, a monoclonal antibody targeting IL-6 pathways. Other potential therapeutic choices, including targeted IL-1, IL-2, IL-1 β , and TNF- α , also have a promising, and yet to be fully explored, role in clinical treatment. Luckily, the anti-inflammatory profiles of drugs with proven safety can be exploited to address the urgent need. For example, statins show anti-inflammatory effects and association with lower mortality [176]; meanwhile, hypoglycemic metformin limits the secretion of inflammatory cytokines by inhibiting ROS production and NF-KB signaling [180]. In a retrospective cohort metformin demonstrated analysis, a notable association with lower mortality in women with obesity or type 2 diabetes. This sex-specific anti-inflammatory effect resulted from the greater reduction of IL-6 and TNF-a, was possibly crucial to COVID-19 severity [181]. Although it has been argued that this favorable anti-inflammatory influence should be weighed up against the theoretical over-immunosuppression possibility of and consequently delayed virus clearance, such strategies can be particularly relevant for obese patients with pre-existing hyperglycemia and hyperlipidemia. After a long bitter dispute, WHO issued guidelines and suggested the systemic application of corticosteroids in critically unwell patients according to its association with reduced mechanical ventilation and short-term mortality [182]. Cai et al. further proposed the criteria for corticosteroid treatment should be defined as neutrophil-to-lymphocyte ratio>6.11 at admission and absence of T2DM [183].

As for the hypercoagulable state of obese patients with COVID-19, low-molecular-weight heparin is recommended for anticoagulation because of its beneficial effects on the cumulative incidence of thromboembolic events and death risk [184]. Levi et al. suggested that low-molecular-weight heparin should be prophylactically used in all hospitalized COVID-19 patients in the absence of medical contraindications. However, the evidence for this suggestion is insufficient, and further prospective randomized controlled trials are needed [162]. Given importance endotheliopathy the of and thrombocytopathy in the pathophysiological changes in COVID-19, patients with obesity probably benefit from therapies targeting endothelial cell and platelet activation, such as nitric oxide and prostacyclin.

The protracted war against the COVID-19 pandemic begins to dawn with vaccine development and widespread vaccination (2378.48 million vaccine doses have been administered as of 16 June 2021), which are particularly important for people with obesity considering their high risk of severe COVID-19 [185, 186]. However, lessons learned from influenza vaccination suggest that vaccines may not be a panacea for people with obesity. The obese probably also show reduced efficacy of SARS-CoV-2 vaccine similar to influenza vaccine due to immune senescence [71, 114]. Sheridan et al. also found that participants with obesity cannot maintain long-term antibody responses [148]. Compared with healthy weight individuals, participants with obesity show a greater decline in influenza antibody titers, CD8+ T cell activation, and expression of functional proteins (IFN-y and granzyme B) 12 months after influenza vaccination [148]. The suboptimal response of people with obesity to the vaccine suggests that a tailored vaccination with increased doses and/or times is necessary, and the possibility of infection with SARS-CoV-2 after vaccination cannot be ruled out given weakened protection of vaccines in people with obesity.

In the context of the obesity epidemic, unraveling the mechanism behind severe COVID-19 in individuals with obesity and a thorough understanding of their interactions are requisites for saving severe cases and reducing the death toll. Attributing any apparent public health success or failure to a single factor is certainly tempting, yet such a simplistic approach to the complexities of pathophysiology is unlikely to provide enough insights against COVID-19. By bridging work done in pathophysiology, metabolism, immunity, basic inflammation, and coagulation, researchers can gain a holistic view of how obesity affects the condition and outcome of COVID-19. Ultimately, lessons learned from COVID-19 in obesity provide, despite its inherent tragedy, interesting and new insights for fighting against possible pandemics in the future.

Abbreviations

ACE2: angiotensin-converting enzyme receptor 2; AT: adipose tissue; BALF: bronchoalveolar lavage fluid; BMI: body mass index; COVID-19: coronavirus disease 2019; DCs: dendritic cells; DIO: diet-induced obesity; FFAs: free fatty acids; FTO: fat mass and gene; HDL: obesity-associated high density lipoprotein; ICU: intensive care unit; IFN: interferon; IL: interleukin; iNKT: invariant natural killer T; IRF9: interferon regulatory factor 9; ISGs: interferon stimulating genes; LDL: low-density lipoprotein; MC4R: melanocortin 4 receptor; NK: natural killer; PPAR: peroxisome proliferator-activated receptor; RAS: renin-angiotensin system; ROS: reactive oxygen species; SARS-CoV-2: severe acute respiratory syndrome coronavirus 2; STAT: signal transducer and activator of transcription; T2DM: type 2 diabetes; TLRs: toll-like receptors; TNF-a: tumor necrosis factor a; WHO: World Health Organization.

Acknowledgments

We would like to specially thank Abe for her great help and company as a loyal and kind friend in those days. We wouldn't have made it this far without her support.

Funding

This work was supported by the People's Liberation Army Medical Science and Technology Project for Youth Cultivation (grant number 20QNPY074).

Author Contributions

Tiantian Yan and Guoan Lin put forward the idea for this article. Literature search and data analysis were performed by Tiantian Yan, Rong Xiao, Nannan Wang, and Ruoyu Shang. The manuscript was written by Tiantian Yan and further revised by Guoan Lin. All authors read and approved the final manuscript.

Competing Interests

The authors have declared that no competing interest exists.

References

- 1. WHO. Coronavirus disease (COVID-19) situation dashboard 2021 [updated 20 June 2021; cited 2021 20 June]. https://covid19.who.int/.
- Lighter J, Phillips M, Hochman S, Sterling S, Johnson D, Francois F, et al. Obesity in patients younger than 60 Years is a risk factor for COVID-19 hospital admission. Clin Infect Dis. 2020;71:896-7.
- Petrilli CM, Jones SA, Yang J, Rajagopalan H, O'Donnell L, Chernyak Y, et al. Factors associated with hospital admission and critical illness among 5279 people with coronavirus disease 2019 in New York City: prospective cohort study. BMJ. 2020;369:m1966.
- Simonnet A, Chetboun M, Poissy J, Raverdy V, Noulette J, Duhamel A, et al. High prevalence of obesity in severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) requiring invasive mechanical ventilation. Obesity. 2020;28:1195-9.

- Peng YD, Meng K, Guan HQ, Leng L, Zhu RR, Wang BY, et al. Clinical characteristics and outcomes of 112 cardiovascular disease patients infected by 2019-nCoV. Zhonghua xin xue guan bing za zhi. 2020;48:450-5.
- Williamson EJ, Walker AJ, Bhaskaran K, Bacon S, Bates C, Morton CE, et al. Factors associated with COVID-19-related death using OpenSAFELY. Nature. 2020;584:430-6.
- Sun Y, Wang Q, Yang G, Lin C, Zhang Y, Yang P. Weight and prognosis for influenza A(H1N1)pdm09 infection during the pandemic period between 2009 and 2011: a systematic review of observational studies with meta-analysis. Infect Dis (Lond). 2016;48:813-22.
- Bray GA, Kim KK, Wilding JPH, World Obesity F. Obesity: a chronic relapsing progressive disease process. A position statement of the World Obesity Federation. Obes Rev. 2017;18:715-23.
- Collaboration NCDRF. Trends in adult body-mass index in 200 countries from 1975 to 2014: a pooled analysis of 1698 population-based measurement studies with 19.2 million participants. Lancet. 2016;387:1377-96.
- Yan T, Xiao R, Lin G. Angiotensin-converting enzyme 2 in severe acute respiratory syndrome coronavirus and SARS-CoV-2: A double-edged sword? FASEB J. 2020;34:6017-26.
- Jia X, Yin C, Lu S, Chen Y, Liu Q, Bai J, et al. Two things about COVID-19 might need attention. Preprints. 2020:2020020315. doi:10.20944/ preprints202002.0315.v1. https://www.preprints.org/manuscript/202002 .0315/v1. Preprint.
- Bourgeois C, Gorwood J, Barrail-Tran A, Lagathu C, Capeau J, Desjardins D, et al. Specific biological features of adipose tissue, and their impact on HIV persistence. Front Microbiol. 2019;10:2837.
- 13. Pavlov VA. The evolving obesity challenge: targeting the vagus nerve and the inflammatory reflex in the response. Pharmacol Ther. 2021;222:107794.
- Stefan N, Birkenfeld AL, Schulze MB. Global pandemics interconnected obesity, impaired metabolic health and COVID-19. Nat Rev Endocrinol. 2021;17:135-49.
- Pepin JL, Timsit JF, Tamisier R, Borel JC, Levy P, Jaber S. Prevention and care of respiratory failure in obese patients. Lancet Respir Med. 2016;4:407-18.
- Huang JF, Wang XB, Zheng KI, Liu WY, Chen JJ, George J, et al. Letter to the Editor: Obesity hypoventilation syndrome and severe COVID-19. Metabolism. 2020;108:154249.
- Britton KA, Massaro JM, Murabito JM, Kreger BE, Hoffmann U, Fox CS. Body fat distribution, incident cardiovascular disease, cancer, and all-cause mortality. J Am Coll Cardiol. 2013;62:921-5.
- Stefan N. Causes, consequences, and treatment of metabolically unhealthy fat distribution. Lancet Diabetes Endocrinol. 2020;8:616-27.
- Kershaw EE, Flier JS. Adipose tissue as an endocrine organ. J Clin Endocrinol Metab. 2004;89:2548-56.
- Clark A, Jit M, Warren-Gash C, Guthrie B, Wang HHX, Mercer SW, et al. Global, regional, and national estimates of the population at increased risk of severe COVID-19 due to underlying health conditions in 2020: a modelling study. Lancet Glob Health. 2020;8:e1003-e17.
- Reilly SM, Saltiel AR. Adapting to obesity with adipose tissue inflammation. Nat Rev Endocrinol. 2017;13:633-43.
- Richardson S, Hirsch JS, Narasimhan M, Crawford JM, McGinn T, Davidson KW, et al. Presenting characteristics, comorbidities, and outcomes among 5700 patients hospitalized with COVID-19 in the New York city area. JAMA. 2020;323:2052-9.
- 23. Ji D, Zhang M, Qin E, Zhang L, Xu J, Wang Y, et al. Letter to the Editor: Obesity, diabetes, non-alcoholic fatty liver disease and metabolic dysfunction associated fatty liver disease are proinflammatory hypercoagulable states associated with severe disease and thrombosis in Covid-19. Metabolism. 2021;115:154437.
- Portincasa P, Krawczyk M, Smyk W, Lammert F, Di Ciaula A. COVID-19 and non-alcoholic fatty liver disease: Two intersecting pandemics. Eur J Clin Invest. 2020;50:e13338.
- Meijnikman AS, Bruin S, Groen AK, Nieuwdorp M, Herrema H. Increased expression of key SARS-CoV-2 entry points in multiple tissues in individuals with NAFLD. J Hepatol. 2021;74:748-9.
- Forlano R, Mullish BH, Mukherjee SK, Nathwani R, Harlow C, Crook P, et al. In-hospital mortality is associated with inflammatory response in NAFLD patients admitted for COVID-19. PLoS One. 2020;15:e0240400.
- Choy LN, Rosen BS, Spiegelman BM. Adipsin and an endogenous pathway of complement from adipose cells. J Biol Chem. 1992;267:12736-41.
- Bjorkerud S, Bjorkerud B. Lipoproteins are major and primary mitogens and growth promoters for human arterial smooth muscle cells and lung fibroblasts in vitro. Arterioscler Thromb. 1994;14:288-98.

- Pian MS, Dobbs LG. Lipoprotein-stimulated surfactant secretion in alveolar type II cells: mediation by heterotrimeric G proteins. Am J Physiol. 1997;273:L634-9.
- Gowdy KM, Fessler MB. Emerging roles for cholesterol and lipoproteins in lung disease. Pulm Pharmacol Ther. 2013;26:430-7.
- 31. Fessler MB. A new frontier in immunometabolism. Cholesterol in lung health and disease. Ann Am Thorac Soc. 2017;14:S399-S405.
- Wang H, Yuan Z, Pavel MA, Hansen SB. The role of high cholesterol in age-related COVID19 lethality. bioRxiv. 2020. doi: https://doi.org/10.1101/2020.05.09.086249. Preprint.
- Tall AR, Yvan-Charvet L. Cholesterol, inflammation and innate immunity. Nat Rev Immunol. 2015;15:104-16.
- 34. Jeon JH, Lee C. Cholesterol is important for the entry process of porcine deltacoronavirus. Arch Virol. 2018;163:3119-24.
- Thorp EB, Gallagher TM. Requirements for CEACAMs and cholesterol during murine coronavirus cell entry. J Virol. 2004;78:2682-92.
- 36. Glende J, Schwegmann-Wessels C, Al-Falah M, Pfefferle S, Qu X, Deng H, et al. Importance of cholesterol-rich membrane microdomains in the interaction of the S protein of SARS-coronavirus with the cellular receptor angiotensin-converting enzyme 2. Virology. 2008;381:215-21.
- Wei X, Zeng W, Su J, Wan H, Yu X, Cao X, et al. Hypolipidemia is associated with the severity of COVID-19. J Clin Lipidol. 2020;14:297-304.
- Fan J, Wang H, Ye G, Cao X, Xu X, Tan W, et al. Letter to the Editor: Low-density lipoprotein is a potential predictor of poor prognosis in patients with coronavirus disease 2019. Metabolism. 2020;107:154243.
- Kontush A, Chapman MJ. Functionally defective high-density lipoprotein: a new therapeutic target at the crossroads of dyslipidemia, inflammation, and atherosclerosis. Pharmacol Rev. 2006;58:342-74.
- Hu X, Chen D, Wu L, He G, Ye W. Declined serum high density lipoprotein cholesterol is associated with the severity of COVID-19 infection. Clin Chim Acta. 2020;510:105-10.
- 41. Wang G, Zhaog Q, Zhao X, Dong H, Wu C, Wu F, et al. Low high-density lipoprotein level is correlated with the severity of COVID-19 patients: an observational study. Lipids Health Dis. 2020;19:204.
- Milner JJ, Rebeles J, Dhungana S, Stewart DA, Sumner SC, Meyers MH, et al. Obesity increases mortality and modulates the lung metabolome during pandemic H1N1 influenza virus infection in mice. J Immunol. 2015;194:4846-59.
- Ying W, Lee YS, Dong Y, Seidman JS, Yang M, Isaac R, et al. Expansion of islet-resident macrophages leads to inflammation affecting beta cell proliferation and function in obesity. Cell Metab. 2019;29:457-74 e5.
- Donath MY, Dalmas É, Sauter NS, Böni-Schnetzler M. Inflammation in obesity and diabetes: islet dysfunction and therapeutic opportunity. Cell Metab. 2013;17:860-72.
- Lecube A, Simo R, Pallayova M, Punjabi NM, Lopez-Cano C, Turino C, et al. Pulmonary function and sleep breathing: two new targets for type 2 diabetes care. Endocr Rev. 2017;38:550-73.
- Liu F, Long X, Zhang B, Zhang W, Chen X, Zhang Z. ACE2 expression in pancreas may cause pancreatic damage after SARS-CoV-2 infection. Clin Gastroenterol Hepatol. 2020;18:2128-30 e2.
- 47. Yang JK, Feng Y, Yuan MY, Yuan SY, Fu HJ, Wu BY, et al. Plasma glucose levels and diabetes are independent predictors for mortality and morbidity in patients with SARS. Diabet Med. 2006;23:623-8.
- Wu J, Huang J, Zhu G, Wang Q, Lv Q, Huang Y, et al. Elevation of blood glucose level predicts worse outcomes in hospitalized patients with COVID-19: a retrospective cohort study. BMJ Open Diabetes Res Care. 2020;8:e001476.
- Philips BJ, Meguer JX, Redman J, Baker EH. Factors determining the appearance of glucose in upper and lower respiratory tract secretions. Intensive Care Med. 2003;29:2204-10.
- Brufsky A. Hyperglycemia, hydroxychloroquine, and the COVID-19 pandemic. J Med Virol. 2020;92:770-5.
- Turina M, Fry DE, Polk HC, Jr. Acute hyperglycemia and the innate immune system: clinical, cellular, and molecular aspects. Crit Care Med. 2005;33:1624-33.
- 52. Hu R, Xia CQ, Butfiloski E, Clare-Salzler M. Effect of high glucose on cytokine production by human peripheral blood immune cells and type I interferon signaling in monocytes: Implications for the role of hyperglycemia in the diabetes inflammatory process and host defense against infection. Clin Immunol. 2018;195:139-48.
- Joshi MB, Lad A, Bharath Prasad AS, Balakrishnan A, Ramachandra L, Satyamoorthy K. High glucose modulates IL-6 mediated immune homeostasis through impeding neutrophil extracellular trap formation. FEBS Lett. 2013;587:2241-6.
- Wierusz-Wysocka B, Wysocki H, Wykretowicz A, Klimas R. The influence of increasing glucose concentrations on selected functions of polymorphonuclear neutrophils. Acta Diabetol Lat. 1988;25:283-8.
- 55. Hostetter MK. Handicaps to host defense. Effects of hyperglycemia on C3 and Candida albicans. Diabetes. 1990;39:271-5.

- Pereira-Santos M, Costa PR, Assis AM, Santos CA, Santos DB. Obesity and vitamin D deficiency: a systematic review and meta-analysis. Obes Rev. 2015;16:341-9.
- Stokic E, Kupusinac A, Tomic-Naglic D, Zavisic BK, Mitrovic M, Smiljenic D, et al. Obesity and vitamin D deficiency: trends to promote a more proatherogenic cardiometabolic risk profile. Angiology. 2015;66:237-43.
- Corica D, Zusi C, Olivieri F, Marigliano M, Piona C, Fornari E, et al. Vitamin D affects insulin sensitivity and beta-cell function in obese non-diabetic youths. Eur J Endocrinol. 2019;181:439-50.
- Grant WB, Lahore H, McDonnell SL, Baggerly CA, French CB, Aliano JL, et al. Evidence that vitamin D supplementation could reduce risk of influenza and COVID-19 infections and deaths. Nutrients. 2020;12:988.
- Martineau AR, Jolliffe DA, Hooper RL, Greenberg L, Aloia JF, Bergman P, et al. Vitamin D supplementation to prevent acute respiratory tract infections: systematic review and meta-analysis of individual participant data. BMJ. 2017;356:i6583.
- Kara M, Ekiz T, Ricci V, Kara O, Chang KV, Ozcakar L. 'Scientific Strabismus' or two related pandemics: coronavirus disease and vitamin D deficiency. Br J Nutr. 2020;124:736-41.
- Quesada-Gomez JM, Entrenas-Castillo M, Bouillon R. Vitamin D receptor stimulation to reduce acute respiratory distress syndrome (ARDS) in patients with coronavirus SARS-CoV-2 infections: Revised Ms SBMB 2020_166. J Steroid Biochem Mol Biol. 2020;202:105719.
- Razquin C, Marti A, Martinez JA. Evidences on three relevant obesogenes: MC4R, FTO and PPARgamma. Approaches for personalized nutrition. Mol Nutr Food Res. 2011;55:136-49.
- 64. Koochakpoor G, Hosseini-Esfahani F, Daneshpour MS, Hosseini SA, Mirmiran P. Effect of interactions of polymorphisms in the Melanocortin-4 receptor gene with dietary factors on the risk of obesity and Type 2 diabetes: a systematic review. Diabet Med. 2016;33:1026-34.
- Przeliorz-Pyszczek A, Regulska-Ilow B. The role of macronutrient intake in reducing the risk of obesity and overweight among carriers of different polymorphisms of FTO gene. A review. Rocz Panstw Zakl Hig. 2017;68:5-13.
- Ciavarella C, Motta I, Valente S, Pasquinelli G. Pharmacological (or synthetic) and nutritional agonists of PPAR-gamma as candidates for cytokine storm modulation in COVID-19 disease. Molecules. 2020;25:2076.
- Short KR, Kedzierska K, van de Sandt CE. Back to the future: Lessons learned from the 1918 influenza pandemic. Front Cell Infect Microbiol. 2018;8:343.
- Chen WJ, Yang JY, Lin JH, Fann CS, Osyetrov V, King CC, et al. Nasopharyngeal shedding of severe acute respiratory syndrome-associated coronavirus is associated with genetic polymorphisms. Clin Infect Dis. 2006;42:1561-9.
- Kanneganti TD, Dixit VD. Immunological complications of obesity. Nat Immunol. 2012;13:707-12.
- Andersen CJ, Murphy KE, Fernandez ML. Impact of obesity and metabolic syndrome on immunity. Adv Nutr. 2016;7:66-75.
- Yang H, Youm YH, Vandanmagsar B, Rood J, Kumar KG, Butler AA, et al. Obesity accelerates thymic aging. Blood. 2009;114:3803-12.
- Barrat FJ, Crow MK, Ivashkiv LB. Interferon target-gene expression and epigenomic signatures in health and disease. Nat Immunol. 2019;20:1574-83.
- 73. Kotenko SV, Rivera A, Parker D, Durbin JE. Type III IFNs: Beyond antiviral protection. Semin Immunol. 2019;43:101303.
- Blanco-Melo D, Nilsson-Payant BE, Liu WC, Uhl S, Hoagland D, Moller R, et al. Imbalanced host response to SARS-CoV-2 drives development of COVID-19. Cell. 2020;181:1036-45.
- 75. Zheng Y, Zhuang MW, Han L, Zhang J, Nan ML, Zhan P, et al. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) membrane (M) protein inhibits type I and III interferon production by targeting RIG-I/MDA-5 signaling. Signal Transduct Target Ther. 2020;5:299.
- Combes AJ, Courau T, Kuhn NF, Hu KH, Ray A, Chen WS, et al. Global absence and targeting of protective immune states in severe COVID-19. Res Sq. 2020;rs.3.rs-97042. doi: 10.21203/rs.3.rs-97042/v1. Preprint.
- Trouillet-Assant S, Viel S, Gaymard A, Pons S, Richard JC, Perret M, et al. Type I IFN immunoprofiling in COVID-19 patients. J Allergy Clin Immunol. 2020;146:206-8.
- Namkoong H, Ishii M, Fujii H, Asami T, Yagi K, Suzuki S, et al. Obesity worsens the outcome of influenza virus infection associated with impaired type I interferon induction in mice. Biochem Biophys Res Commun. 2019;513:405-11.
- Teran-Cabanillas E, Montalvo-Corral M, Silva-Campa E, Caire-Juvera G, Moya-Camarena SY, Hernandez J. Production of interferon alpha and beta, pro-inflammatory cytokines and the expression of suppressor of cytokine signaling (SOCS) in obese subjects infected with influenza A/H1N1. Clin Nutr. 2014;33:922-6.

- Teran-Cabanillas E, Hernandez J. Role of leptin and SOCS3 in inhibiting the type I interferon response during obesity. Inflammation. 2017;40:58-67.
- Costanzo AE, Taylor KR, Dutt S, Han PP, Fujioka K, Jameson JM. Obesity impairs gammadelta T cell homeostasis and antiviral function in humans. PLoS One. 2015;10(3):e0120918.
- Almond MH, Edwards MR, Barclay WS, Johnston SL. Obesity and susceptibility to severe outcomes following respiratory viral infection. Thorax. 2013;68:684-6.
- Channappanavar R, Fehr AR, Vijay R, Mack M, Zhao J, Meyerholz DK, et al. Dysregulated type I interferon and inflammatory monocyte-macrophage responses cause lethal pneumonia in SARS-CoV-infected mice. Cell Host Microbe. 2016;19:181-93.
- Wei L, Ming S, Zou B, Wu Y, Hong Z, Li Z, et al. Viral invasion and type I interferon response characterize the immunophenotypes during COVID-19 infection. SSRN Electronic Journal. 2020. https://ssrn.com/abstract=3564998. Preprint.
- Yao H, Lu X, Chen Q, Xu K, Chen Y, Cheng M, et al. Patient-derived SARS-CoV-2 mutations impact viral replication dynamics and infectivity in vitro and with clinical implications in vivo. Cell Discov. 2020;6:76.
- Kupferschmidt K. Mutant coronavirus in the United Kingdom sets off alarms, but its importance remains unclear 2021 [updated 20 December 2020; cited 2021 20 December]. https://www.sciencemag.org/ news/2020/12/mutant-coronavirus-united-kingdom-sets-alarms-its-im portance-remains-unclear.
- Klinkhammer J, Schnepf D, Ye L, Schwaderlapp M, Gad HH, Hartmann R, et al. IFN-lambda prevents influenza virus spread from the upper airways to the lungs and limits virus transmission. eLife. 2018;7:e33354.
- Honce R, Karlsson EA, Wohlgemuth N, Estrada LD, Meliopoulos VA, Yao J, et al. Obesity-related microenvironment promotes emergence of virulent influenza virus strains. mBio. 2020;11:e03341-19.
- Smith AG, Sheridan PA, Harp JB, Beck MA. Diet-induced obese mice have increased mortality and altered immune responses when infected with influenza virus. J Nutr. 2007;137:1236-43.
- Michelet X, Dyck L, Hogan A, Loftus RM, Duquette D, Wei K, et al. Metabolic reprogramming of natural killer cells in obesity limits antitumor responses. Nat Immunol. 2018;19:1330-40.
- Jung YS, Park JH, Park DI, Sohn CI, Lee JM, Kim TI. Physical inactivity and unhealthy metabolic status are associated with decreased Natural Killer cell activity. Yonsei Med J. 2018;59:554-62.
- Donnelly RP, Loftus RM, Keating SE, Liou KT, Biron CA, Gardiner CM, et al. mTORC1-dependent metabolic reprogramming is a prerequisite for NK cell effector function. J Immunol. 2014;193:4477-84.
- Cifaldi L, Prencipe G, Caiello I, Bracaglia C, Locatelli F, De Benedetti F, et al. Inhibition of natural killer cell cytotoxicity by interleukin-6: implications for the pathogenesis of macrophage activation syndrome. Arthritis Rheumatol. 2015;67:3037-46.
- 94. Nave H, Mueller G, Siegmund B, Jacobs R, Stroh T, Schueler U, et al. Resistance of Janus kinase-2 dependent leptin signaling in natural killer (NK) cells: a novel mechanism of NK cell dysfunction in diet-induced obesity. Endocrinology. 2008;149:3370-8.
- Guo H, Xu B, Gao L, Sun X, Qu X, Li X, et al. High frequency of activated natural killer and natural killer T-cells in patients with new onset of type 2 diabetes mellitus. Exp Biol Med (Maywood). 2012;237:556-62.
- Zheng M, Gao Y, Wang G, Song G, Liu S, Sun D, et al. Functional exhaustion of antiviral lymphocytes in COVID-19 patients. Cell Mol Immunol. 2020;17:533-5.
- Wang F, Nie J, Wang H, Zhao Q, Xiong Y, Deng L, et al. Characteristics of peripheral lymphocyte subset alteration in COVID-19 pneumonia. J Infect Dis. 2020;221:1762-9.
- van Eijkeren RJ, Krabbe O, Boes M, Schipper HS, Kalkhoven E. Endogenous lipid antigens for invariant natural killer T cells hold the reins in adipose tissue homeostasis. Immunology. 2018;153:179-89.
- Carreño LJ, Kharkwal SS, Porcelli SA. Optimizing NKT cell ligands as vaccine adjuvants. Immunotherapy. 2014;6:309-20.
- 100. Lynch L, Nowak M, Varghese B, Clark J, Hogan AE, Toxavidis V, et al. Adipose tissue invariant NKT cells protect against diet-induced obesity and metabolic disorder through regulatory cytokine production. Immunity. 2012;37:574-87.
- 101. Venken K, Seeuws S, Zabeau L, Jacques P, Decruy T, Coudenys J, et al. A bidirectional crosstalk between iNKT cells and adipocytes mediated by leptin modulates susceptibility for T cell mediated hepatitis. J Hepatol. 2014;60:175-82.
- 102. Cho KW, Zamarron BF, Muir LA, Singer K, Porsche CE, DelProposto JB, et al. Adipose tissue dendritic cells are independent contributors to obesity-induced inflammation and insulin resistance. J Immunol. 2016;197:3650-61.
- 103. Macdougall CE, Wood EG, Loschko J, Scagliotti V, Cassidy FC, Robinson ME, et al. Visceral adipose tissue immune homeostasis is regulated by

the crosstalk between adipocytes and dendritic cell subsets. Cell Metab. 2018;27:588-601.

- 104. O'Shea D, Corrigan M, Dunne MR, Jackson R, Woods C, Gaoatswe G, et al. Changes in human dendritic cell number and function in severe obesity may contribute to increased susceptibility to viral infection. Int J Obes (Lond). 2013;37:1510-3.
- 105. James BR, Tomanek-Chalkley A, Askeland EJ, Kucaba T, Griffith TS, Norian LA. Diet-induced obesity alters dendritic cell function in the presence and absence of tumor growth. J Immunol. 2012;189:1311-21.
- 106. Weitman ES, Aschen SZ, Farias-Eisner G, Albano N, Cuzzone DA, Ghanta S, et al. Obesity impairs lymphatic fluid transport and dendritic cell migration to lymph nodes. PLoS One. 2013;8:e70703.
- Macia L, Delacre M, Abboud G, Ouk T-S, Delanoye A, Verwaerde C, et al. Impairment of dendritic cell functionality and steady-state number in obese mice. J Immunol. 2006;177:5997-6006.
- 108. Smith AG, Sheridan PA, Tseng RJ, Sheridan JF, Beck MA. Selective impairment in dendritic cell function and altered antigen-specific CD8+ T-cell responses in diet-induced obese mice infected with influenza virus. Immunology. 2009;126:268-79.
- 109. Laing AG, Lorenc A, Del Barrio IDM, Das A, Fish M, Monin L, et al. A dynamic COVID-19 immune signature includes associations with poor prognosis. Nat Med. 2020;26:1623-35.
- Liao M, Liu Y, Yuan J, Wen Y, Xu G, Zhao J, et al. Single-cell landscape of bronchoalveolar immune cells in patients with COVID-19. Nat Med. 2020;26:842-4.
- 111. Zhou R, To KK-W, Wong Y-C, Liu L, Zhou B, Li X, et al. Acute SARS-CoV-2 infection impairs dendritic cell and T cell responses. Immunity. 2020;53:864-77.
- 112. Parackova Z, Zentsova I, Bloomfield M, Vrabcova P, Smetanova J, Klocperk A, et al. Disharmonic inflammatory signatures in COVID-19: augmented neutrophils' but impaired monocytes' and dendritic cells' responsiveness. Cells. 2020;9:2206.
- 113. Cañete PF, Vinuesa CG. COVID-19 makes B cells forget, but T cells remember. Cell. 2020;183:13-5.
- Linton PJ, Dorshkind K. Age-related changes in lymphocyte development and function. Nat Immunol. 2004;5:133-9.
- 115. Spielmann G, Johnston CA, O'Connor DP, Foreyt JP, Simpson RJ. Excess body mass is associated with T cell differentiation indicative of immune ageing in children. Clin Exp Immunol. 2014;176:246-54.
- 116. Jacobsen MJ, Mentzel CMJ, Olesen AS, Huby T, Jørgensen CB, Barrès R, et al. Altered methylation profile of lymphocytes is concordant with perturbation of lipids metabolism and inflammatory response in obesity. J Diabetes Res. 2016;2016:8539057.
- 117. Simar D, Versteyhe S, Donkin I, Liu J, Hesson L, Nylander V, et al. DNA methylation is altered in B and NK lymphocytes in obese and type 2 diabetic human. Metabolism. 2014;63:1188-97.
- Salimi S, Hamlyn JM. COVID-19 and crosstalk with the hallmarks of aging. J Gerontol A Biol Sci Med Sci. 2020;75:e34-e41.
- 119. Mauro C, Smith J, Cucchi D, Coe D, Fu H, Bonacina F, et al. Obesity-induced metabolic stress leads to biased effector memory CD4+ T cell differentiation via PI3K p110δ-Akt-mediated signals. Cell Metab. 2017;25:593-609.
- 120. Su Y, Chen D, Yuan D, Lausted C, Choi J, Dai CL, et al. Multi-omics resolves a sharp disease-state shift between mild and moderate COVID-19. Cell. 2020;183:1479-95.
- 121. Li H, Wetchapinant C, Zhang L, Wu K. High-fat diet from weaning until early adulthood impairs T cell development in the thymus. Lipids. 2020;55:35-44.
- 122. Urra JM, Cabrera CM, Porras L, Ródenas I. Selective CD8 cell reduction by SARS-CoV-2 is associated with a worse prognosis and systemic inflammation in COVID-19 patients. Clin Immunol. 2020;217:108486.
- 123. Schwartz MD, Emerson SG, Punt J, Goff WD. Decreased naïve T-cell production leading to cytokine storm as cause of increased COVID-19 severity with comorbidities. Aging Dis. 2020;11:742.
- 124. Wang Z, Aguilar EG, Luna JI, Dunai C, Khuat LT, Le CT, et al. Paradoxical effects of obesity on T cell function during tumor progression and PD-1 checkpoint blockade. Nat Med. 2019;25:141-51.
- 125. Rojas-Osornio SA, Cruz-Hernández TR, Drago-Serrano ME, Campos-Rodríguez R. Immunity to influenza: impact of obesity. Obes Res Clin Pract. 2019;13:419-29.
- 126. Renner K, Geiselhöringer AL, Fante M, Bruss C, Färber S, Schönhammer G, et al. Metabolic plasticity of human T cells: preserved cytokine production under glucose deprivation or mitochondrial restriction, but 2-deoxy-glucose affects effector functions. Eur J Immunol. 2015;45:2504-16.
- 127. Acharya D, Liu G, Gack MU. Dysregulation of type I interferon responses in COVID-19. Nat Rev Immunol. 2020;20:397-8.
- 128. Griffin DE. Are T cells helpful for COVID-19: the relationship between response and risk. J Clin Invest. 2020;130:6222-4.

- 129. Jubb RW. Therapeutic progress-review: XXV Osteoarthritis-are we making progress? J Clin Pharm Ther. 1987;12:81-90.
- 130. Snyder TM, Gittelman RM, Klinger M, May DH, Osborne EJ, Taniguchi R, et al. Magnitude and dynamics of the T-cell response to SARS-CoV-2 infection at both individual and population levels. medRxiv 2020.07.31.20165647. doi: https://doi.org/10.1101/2020.07.31.20165647. Available from https://www.medrxiv.org/content/10.1101/2020.07.31 .20165647v3. Preprint.
- 131. Frydrych LM, Bian G, O'Lone DE, Ward PA, Delano MJ. Obesity and type 2 diabetes mellitus drive immune dysfunction, infection development, and sepsis mortality. J Leukoc Biol. 2018;104:525-34.
- 132. Tincati C, Cannizzo ES, Giacomelli M, Badolato R, Monforte AdA, Marchetti G. Heightened circulating interferon-inducible chemokines, and activated pro-cytolytic Th1-cell phenotype features Covid-19 aggravation in the second week of illness. Front Immunol. 2020;11:580987.
- Cheung KP, Taylor KR, Jameson JM. Immunomodulation at epithelial sites by obesity and metabolic disease. Immunol Res. 2012;52:182-99.
- 134. Saksena S, Chattopadhyay P. Illuminating the immunopathology of SARS-CoV-2. Cytometry B Clin Cytom. 2021;100:33-41.
- 135. Milner JJ, Sheridan PA, Karlsson EA, Schultz-Cherry S, Shi Q, Beck MA. Diet-induced obese mice exhibit altered heterologous immunity during a secondary 2009 pandemic H1N1 infection. J Immunol. 2013;191:2474-85.
- 136. Kim Y-H, Kim J-K, Kim D-J, Nam J-H, Shim S-M, Choi Y-K, et al. Diet-induced obesity dramatically reduces the efficacy of a 2009 pandemic H1N1 vaccine in a mouse model. J Infect Dis. 2012;205:244-51.
- Frasca D, Diaz A, Romero M, Blomberg BB. Ageing and obesity similarly impair antibody responses. Clin Exp Immunol. 2017;187:64-70.
- 138. Huang AT, Garcia-Carreras B, Hitchings MDT, Yang B, Katzelnick LC, Rattigan SM, et al. A systematic review of antibody mediated immunity to coronaviruses: antibody kinetics, correlates of protection, and association of antibody responses with severity of disease. Nat Commun. 2020;11:4704.
- 139. Zhao J, Yuan Q, Wang H, Liu W, Liao X, Su Y, et al. Antibody responses to SARS-CoV-2 in patients with novel coronavirus disease 2019. Clin Infect Dis. 2020;71:2027-34.
- Chandra RK. Cell-mediated immunity in genetically obese (C57BL/6J ob/ob) mice. Am J Clin Nutr. 1980;33:13-6.
- 141. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet. 2020;395:497-506.
- 142. Giamarellos-Bourboulis EJ, Netea MG, Rovina N, Akinosoglou K, Antoniadou A, Antonakos N, et al. Complex immune dysregulation in COVID-19 patients with severe respiratory failure. Cell Host Microbe. 2020;27:992-1000.
- 143. Thiel V, Weber F. Interferon and cytokine responses to SARS-coronavirus infection. Cytokine Growth Factor Rev. 2008;19:121-32.
- 144. Bermejo-Martin JF, de Lejarazu RO, Pumarola T, Rello J, Almansa R, Ramírez P, et al. Th1 and Th17 hypercytokinemia as early host response signature in severe pandemic influenza. Crit Care. 2009;13:R201.
- 145. Zhang AJX, To KKW, Li C, Lau CCY, Poon VKM, Chan CCS, et al. Leptin mediates the pathogenesis of severe 2009 pandemic influenza A (H1N1) infection associated with cytokine dysregulation in mice with diet-induced obesity. J Infect Dis. 2013;207:1270-80.
- 146. Böni-Schnetzler M, Boller S, Debray S, Bouzakri K, Meier DT, Prazak R, et al. Free fatty acids induce a proinflammatory response in islets via the abundantly expressed interleukin-1 receptor I. Endocrinology. 2009;150:5218-29.
- 147. Channappanavar R, Perlman S, editors. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. Semin Immunopathol. 2017;39:529-39.
- 148. Wang J, Xu Y, Zhang X, Wang S, Peng Z, Guo J, et al. Leptin correlates with monocytes activation and severe condition in COVID-19 patients. J Leukoc Biol. 2021. Online ahead of print.
- 149. Jiang S, Park DW, Tadie J-M, Gregoire M, Deshane J, Pittet JF, et al. Human resistin promotes neutrophil proinflammatory activation and neutrophil extracellular trap formation and increases severity of acute lung injury. J Immunol. 2014;192:4795-803.
- 150. Zacharioudaki V, Androulidaki A, Arranz A, Vrentzos G, Margioris AN, Tsatsanis C. Adiponectin promotes endotoxin tolerance in macrophages by inducing IRAK-M expression. J Immunol. 2009;182:6444-51.
- 151. Tsatsanis C, Margioris AN, Kontoyiannis DP. Association between H1N1 infection severity and obesity – adiponectin as a potential etiologic factor. J Infect Dis. 2010;202:459-60.
- 152. Datta PK, Liu F, Fischer T, Rappaport J, Qin X. SARS-CoV-2 pandemic and research gaps: Understanding SARS-CoV-2 interaction with the ACE2 receptor and implications for therapy. Theranostics. 2020;10:7448-64.

- 153. Forero A, Ozarkar S, Li H, Lee CH, Hemann EA, Nadjsombati MS, et al. Differential activation of the transcription factor IRF1 underlies the distinct immune responses elicited by type I and type III interferons. Immunity. 2019;51:451-64.
- 154. Kim JS, Lee JY, Yang JW, Lee KH, Effenberger M, Szpirt W, et al. Immunopathogenesis and treatment of cytokine storm in COVID-19. Theranostics. 2021;11:316-29.
- 155. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW. Obesity is associated with macrophage accumulation in adipose tissue. J Clin Invest. 2003;112:1796-808.
- 156. Kratz M, Coats BR, Hisert KB, Hagman D, Mutskov V, Peris E, et al. Metabolic dysfunction drives a mechanistically distinct proinflammatory phenotype in adipose tissue macrophages. Cell Metab. 2014;20:614-25.
- 157. Speranza E, Williamson BN, Feldmann F, Sturdevant GL, Pérez-Pérez L, Meade-White K, et al. Single-cell RNA sequencing reveals SARS-CoV-2 infection dynamics in lungs of African green monkeys. Sci Transl Med. 2021;13:eabe8146.
- Larsen MD, de Graaf EL, Sonneveld ME, Plomp HR, Nouta J, Hoepel W, et al. Afucosylated IgG characterizes enveloped viral responses and correlates with COVID-19 severity. Science. 2021; 371:eabc8378.
- 159. Hoepel W, Chen H-J, Allahverdiyeva S, Manz X, Aman J, Bonta P, et al. Anti-SARS-CoV-2 IgG from severely ill COVID-19 patients promotes macrophage hyper-inflammatory responses. Sci Transl Med. 2021;13:eabf8654.
- 160. Gu SX, Tyagi T, Jain K, Gu VW, Lee SH, Hwa JM, et al. Thrombocytopathy and endotheliopathy: crucial contributors to COVID-19 thromboinflammation. Nat Rev Cardiol. 2021;18:194-209.
- 161. Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. J Thromb Haemost. 2020;18:844-7.
- Levi M, Thachil J, Iba T, Levy JH. Coagulation abnormalities and thrombosis in patients with COVID-19. Lancet Haematol. 2020;7:e438-e40.
- 163. Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, et al. Endothelial cell infection and endotheliitis in COVID-19. Lancet. 2020;395:1417-8.
- 164. Panigada M, Bottino N, Tagliabue P, Grasselli G, Novembrino C, Chantarangkul V, et al. Hypercoagulability of COVID-19 patients in intensive care unit: A report of thromboelastography findings and other parameters of hemostasis. J Thromb Haemost. 2020;18:1738-42.
- 165. Nougier C, Benoit R, Simon M, Desmurs-Clavel H, Marcotte G, Argaud L, et al. Hypofibrinolytic state and high thrombin generation may play a major role in SARS-COV2 associated thrombosis. J Thromb Haemost. 2020;18:2215-9.
- 166. Goshua G, Pine AB, Meizlish ML, Chang CH, Zhang H, Bahel P, et al. Endotheliopathy in COVID-19-associated coagulopathy: evidence from a single-centre, cross-sectional study. Lancet Haematol. 2020;7:e575-e82.
- 167. Zarbock A, Polanowska-Grabowska RK, Ley K. Platelet-neutrophil-interactions: linking hemostasis and inflammation. Blood Rev. 2007;21:99-111.
- Vilahur G, Ben-Aicha S, Badimon L. New insights into the role of adipose tissue in thrombosis. Cardiovasc Res. 2017;113:1046-54.
- 169. De Pergola G, Pannacciulli N, Coviello M, Scarangella A, Di Roma P, Caringella M, et al. sP-selectin plasma levels in obesity: association with insulin resistance and related metabolic and prothrombotic factors. Nutr Metab Cardiovasc Dis. 2008;18:227-32.
- 170. Anfossi G, Russo I, Massucco P, Mattiello L, Doronzo G, De Salve A, et al. Impaired synthesis and action of antiaggregating cyclic nucleotides in platelets from obese subjects: possible role in platelet hyperactivation in obesity. Eur J Clin Invest. 2004;34:482-9.
- 171. Bath PM, Butterworth RJ. Platelet size: measurement, physiology and vascular disease. Blood Coagul Fibrinolysis. 1996;7:157-61.
- 172. Mertens I, Gaal LFV. Obesity, haemostasis and the fibrinolytic system. Obes Rev. 2002;3:85-101.
- 173. Skurk T, Lee Y-M, Hauner H. Angiotensin II and its metabolites stimulate PAI-1 protein release from human adipocytes in primary culture. Hypertension. 2001;37:1336-40.
- 174. Fosbøl EL, Butt JH, Østergaard L, Andersson C, Selmer C, Kragholm K, et al. Association of angiotensin-converting enzyme inhibitor or angiotensin receptor blocker use with COVID-19 diagnosis and mortality. JAMA. 2020;324:168-77.
- 175. Mackey K, King VJ, Gurley S, Kiefer M, Liederbauer E, Vela K, et al. Risks and impact of angiotensin-converting enzyme inhibitors or angiotensin-receptor blockers on SARS-CoV-2 infection in adults: a living systematic review. Ann Intern Med. 2020;173:195-203.
- 176. Zhang X-J, Qin J-J, Cheng X, Shen L, Zhao Y-C, Yuan Y, et al. In-hospital use of statins is associated with a reduced risk of mortality among individuals with COVID-19. Cell Metab. 2020;32:176-87.

- 177. Zhu L, She Z-G, Cheng X, Qin J-J, Zhang X-J, Cai J, et al. Association of blood glucose control and outcomes in patients with COVID-19 and pre-existing type 2 diabetes. Cell Metab. 2020;31:1068-77.
- 178. Pham H, Waterhouse M, Baxter C, Romero BD, McLeod DSA, Armstrong BK, et al. The effect of vitamin D supplementation on acute respiratory tract infection in older Australian adults: an analysis of data from the D-Health Trial. Lancet Diabetes Endocrinol. 2021;9:69-81.
- 179. Ziegler CGK, Allon SJ, Nyquist SK, Mbano IM, Miao VN, Tzouanas CN, et al. SARS-CoV-2 receptor ACE2 is an interferon-stimulated gene in human airway epithelial cells and is detected in specific cell subsets across tissues. Cell. 2020;181:1016-35.
- 180. Isoda K, Young JL, Zirlik A, MacFarlane LA, Tsuboi N, Gerdes N, et al. Metformin inhibits proinflammatory responses and nuclear factor-kB in human vascular wall cells. Arterioscler Thromb Vasc Biol. 2006;26:611-7.
- 181. Bramante CT, Ingraham NE, Murray TA, Marmor S, Hovertsen S, Gronski J, et al. Metformin and risk of mortality in patients hospitalised with COVID-19: a retrospective cohort analysis. Lancet Healthy Longev. 2021;2:e34-e41.
- 182. van Paassen J, Vos JS, Hoekstra EM, Neumann KMI, Boot PC, Arbous SM. Corticosteroid use in COVID-19 patients: a systematic review and meta-analysis on clinical outcomes. Crit Care. 2020;24:696.
- 183. Cai J, Li H, Zhang C, Chen Z, Liu H, Lei F, et al. The neutrophil-to-lymphocyte ratio determines clinical efficacy of corticosteroid therapy in patients with COVID-19. Cell Metab. 2021;33:258-269.
- 184. Jonmarker S, Hollenberg J, Dahlberg M, Stackelberg O, Litorell J, Everhov AH, et al. Dosing of thromboprophylaxis and mortality in critically ill COVID-19 patients. Crit Care. 2020;24:653.
- 185. Karpinski TM, Ozarowski M, Seremak-Mrozikiewicz A, Wolski H, Wlodkowic D. The 2020 race towards SARS-CoV-2 specific vaccines. Theranostics. 2021;11:1690-702.
- Liu X, Liu C, Liu G, Luo W, Xia N. COVID-19: Progress in diagnostics, therapy and vaccination. Theranostics. 2020;10:7821-35.