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Assessment of Superparamagnetic Iron Oxide Nanoparticle Poly(Ethylene Glycol) Coatings on Magnetic Resonance Relaxation for Early Disease Detection

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ABSTRACT *Objective*: Superparamagnetic Iron Oxide Nanoparticles (SPIONs) are widely researched as contrast agents in clinical magnetic resonance imaging (MRI). SPIONs are frequently coated with anti-biofouling substances such as poly(ethylene glycol) (PEG) to prevent protein deposition and improve circulation time *in vivo*. The aim of this study is to optimize SPION MR properties with respect to physicochemical properties of the core SPION and the polymeric coating to better understand the interaction of these parameters and the efficacy of the designed agent. *Methods*: We used different methods of chemical attachment of a polymer, polymer chain length, and polymer coating density and examined their effects on the MR relaxivities of SPIONs. *Results*: These studies indicate that the chemical composition and, in particular, the hydrophobicity/hydrophilicity of the chemical group linking PEG chains to a SPION core may play a larger role in the resulting MR relaxivities than other variable properties such as SPION core size and PEG chain length. *Conclusions*: The method of SPION fabrication and chemical composition of the coating play a significant role in the MR relaxivities of the resulting particles. These results should be considered in the fabrication of particles for clinical purposes, particularly when optimization of the MR relaxivities is desired.

INDEX TERMS Magnetic resonance imaging (MRI), magnetic relaxation, poly(ethylene glycol), SPIONs.

IMPACT STATEMENT To our knowledge, this is the first controlled study examining the impact of PEGylation method on MR relaxivities, which is key for optimizing targeted MR agents for early disease detection.

I. INTRODUCTION

Superparamagnetic iron oxide nanoparticles (SPIONs) are used as contrast agents for clinical magnetic resonance imaging (MRI) [1]–[3]. SPION variability in terms of iron core size, surface functionalization, and targeting moieties makes them potentially well-suited for detection of a wide array of diseases [4]–[9]. SPIONs hold specific advantages over widely-used gadolinium-based MR contrast agents: each nanoparticle contains thousands of iron atoms and approaches saturation magnetization in magnetic fields typically used

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for MR imaging, i.e., each nanocrystal can generate signal contrast several orders of magnitude higher than gadolinium chelates [10]. Additionally, SPIONs have demonstrably lower toxicity *in vivo* than gadolinium-based contrast agents [11]–[15]. These two features make SPIONs particularly wellsuited for early detection or diagnosis of diseases ranging from cancer to atherosclerosis.

Upon injection into the bloodstream, SPIONs immediately encounter a high-protein environment where adsorption of various proteins onto the SPION surface can ultimately lead to rapid clearance, reducing their effectiveness as a contrast agent [16]–[18]. A frequently used strategy to prevent protein deposition is to coat SPIONs with an antibiofouling layer such as poly(ethylene glycol) (PEG) [14], [19]. PEG is thought to display good resistance against nonspecific protein adsorption in large part due to its extensive hydrogen bonding with water [20]. SPIONs are often coated with PEG in addition to targeting moieties such as antibodies, peptides, or aptamers, [7], [21] and have been thoroughly reported in the literature for *in vivo* imaging of early disease markers.

One challenge of using molecular-targeted contrast agents at early disease stages is that molecular markers are usually present in small quantities [22], [23]. Detecting small quantities of these markers is challenging in MRI even with the benefit of SPION-enhanced contrast; thus maximal MR signal is needed to improve disease detection sensitivity. Clinically, increasing MR relaxivities can yield several benefits, including a greater signal-to-noise ratio and therefore increased resolving power that may be particularly beneficial in applications such as early disease detection. Moreover, increased MR relaxivities may allow lower administered doses to yield equivalent signal-to-noise ratios, meaning that agents with increased MR relaxivities may carry a cost and/or safety benefit to patients.

Few previous studies have comprehensively examined optimization of SPION MR properties with respect to physicochemical properties.

SPIONs create contrast in MR imaging by affecting two properties of nearby water protons: [24] T_1 (spin-lattice relaxation time) and T_2 (spin-spin relaxation time). The corresponding relaxivities r_1 and r_2 are in part determined by translational diffusion of water molecules in the inhomogeneous magnetic field caused by the SPIONs [10]. Although SPIONs impact r_1 and r_2 , they are primarily used as T_2 contrast agents because of their larger r_2 relaxivities [25], [26]. Two design parameters of SPIONs and their chemical coatings known to impact relaxivities are (1) iron oxide core (size and material), and (2) polymer coating surrounding the SPION core, which includes coating density (i.e., polymer chains per nm²), thickness (i.e., polymer chain length), and chemical composition.

Some prior work has explored the first aspect of SPION contrast agents. It is known that for a fixed total amount of iron in the core, r_2 relaxivity of SPIONs increases with magnetization and total core size (i.e., iron oxide plus other elements that may be added to the core) [4], [27]. Altering

SPION magnetization by doping iron oxide with other magnetic elements (e.g., nickel, cobalt, and manganese) leads to enhancement of measured relaxivities [5], [27], [28].

However, effects of changes in polymer coating in terms of density, thickness, and chemical composition on relaxivities of the resulting SPION agents has not been well-characterized to date. Of the limited prior studies, none have controlled for or measured coating density, which is thought to have a direct effect on the resultant relaxivities [15], [p. 2], [29]. The importance of understanding how altering the SPION core coating may impact MR signal is in part due to the fact that the strength of the magnetic field surrounding the SPION core diminishes rapidly with distance; thus, the strongest part of the magnetic field surrounding the SPION often falls within the polymer coating [29], [30]. Moreover, SPION functionalization using different coating schemes, particularly in terms of method of chemical attachment of the coating to the core and polymer chain length, may affect their MR properties even if the coating density remains constant. The aim of the present study is to determine effects of different methods of chemical attachment of a polymer, polymer chain length, and polymer coating density on SPION MR relaxivities, thereby contributing to a better understanding of the interaction of these parameters and the efficacy of the designed agent. These results will aid in optimization of SPION MR properties with the aim of improving SPION contrast agents intended for clinical imaging, especially in early disease detection.

II. RESULTS

Both 4nm and 10nm SPION cores were successfully functionalized using 3 different fabrication schemes (Fig. 1).

Dynamic light scattering (DLS), zeta potential measurements, and thermogravimetric analysis (TGA) were used to characterize and confirm success of SPION fabrication for each type of SPION produced (Table 1).

In phosphine oxide (PO)-PEG SPIONs, nanoparticle effective diameter appears to increase as PEG chain length increases, yet these differences were not statistically significant across multiple batches (p > 0.05). For 1,2-Distearoyl-snglycero-3-phosphoethanolamine (DSPE)-PEG SPIONs, the 5k PEG samples had a smaller effective diameter than either the 1k or the 2k PEG, but these differences were not significant (p > 0.05). This trend was also observed for 4nm NH₂-PEG SPIONs, with 3.4k NH₂-PEG SPIONs being significantly larger (p < 0.05) than 1k, 2k, and 5k NH₂-PEG SPIONs. For 10nm NH₂-PEG SPIONs, this did not appear to be the case: 5k NH₂-PEG SPIONs were significantly larger than 1k, 2k, and 3.4k NH₂-PEG SPIONs. For both PO-PEG and NH₂-PEG SPIONs, the effective diameters of the 4nm samples were significantly smaller (p < 0.05) than the equivalent 10nm samples. For DSPE-PEG SPIONs, this was not the case. Finally, it can be observed for all samples that for a given core size and PEG chain length, the effective diameter observed between different PEGylation methods was such that NH₂-PEG SPIONs < PO-PEG SPIONs < DSPE-PEG

TABLE 1. Summary of Characterization Results for all SPION Variants Produced. Results are Represented as Mean ± Standard Deviation, n = 3. Effective Diameter was Measured Using DLS, Zeta Potential Was Measured on the Same Machine, and Surface Density was Calculated From TGA Results. All PEG Chains Were Linear in Conformation

Diameter (nm)	PEGylation Method	PEG Size (kDa)	Effective Diameter (nm)	Zeta Potential (ζ) (mV)	Surface Density (PEG chains/nm ²)
4	PO-PEG (Method 1)	1000	18.66 ± 4.14	-14.75 ± 1.58	0.56 ± 0.06
		2000	22.67 ± 3.21	-5.63 ± 6.13	0.82 ± 0.09
		5000	22.86 ± 2.54	-1.62 ± 4.21	0.93 ± 0.23
	NH ₂ -PEG (Method 2)	1000	14.79 ± 3.74	-19.46 ± 1.71	0.68 ± 0.13
		2000	18.04 ± 4.24	-15.89 ± 1.11	0.93 ± 0.04
		3400	38.46 ± 6.16	-10.85 ± 1.29	0.87 ± 0.06
		5000	20.03 ± 4.42	-8.93 ± 1.21	0.86 ± 0.15
	DSPE-PEG (Method 3)	1000	64.24 ± 16.19	-23.55 ± 13.85	0.70 ± 0.05
		2000	82.40 ± 26.49	-18.36 ± 23.57	0.82 ± 0.05
		5000	52.98 ± 11.54	22.17 ± 19.41	1.13 ± 0.11
10	PO-PEG (Method 1)	1000	40.69 ± 5.42	-6.30 ± 1.29	0.41 ± 0.10
		2000	42.71 ± 7.45	-13.35 ± 3.16	0.67 ± 0.13
		5000	45.93 ± 5.47	-8.94 ± 0.89	0.66 ± 0.18
	NH ₂ -PEG (Method 2)	1000	30.22 ± 5.46	-22.79 ± 2.75	0.35 ± 0.31
		2000	27.33 ± 5.17	-13.79 ± 1.84	0.62 ± 0.08
		3400	31.12 ± 5.57	-20.67 ± 1.39	0.67 ± 0.03
		5000	41.21 ± 6.35	-13.54 ± 1.65	0.60 ± 0.14
	DSPE-PEG (Method 3)	1000	58.07 ± 15.21	-31.14 ± 31.40	0.69 ± 0.00
		2000	75.45 ± 28.67	-16.10 ± 33.98	0.69 ± 0.01
		5000	55.49 ± 11.70	-3.61 ± 16.48	0.77 ± 0.27



FIGURE 1. Schematic of particle fabrication. Three methods were used: (1) direct PEGylation via ligand exchange with phosphine oxide (PO)-PEG, (2) ligand exchange to coat particles with citric acid, followed by covalent addition of NH₂-PEG via EDC/NHS chemistry, and (3) PEGylation via hydrophobic interactions between oleic acid on the SPION surface and 1,2-Distearoyl-sn-glycero-3-phosphoethanolamine (DSPE) residues on DSPE-PEG. For each method of fabrication, at least 3 different PEG chain lengths were added to different particle samples to observe effects of polymer chain length on resulting MR relaxivities. Tested PEG chain lengths were: (1) for PO-PEG SPIONs – 1000, 2000, and 5000 Da, (2) for NH2-PEG SPIONs – 1000, 2000, 3400, and 5000 Da, and (3) for DSPE-PEG SPIONs – 1000, 2000, 5000 Da. In total, 20 unique SPIONs were fabricated, characterized, and tested.

SPIONs. These differences vary in statistical significance, but this trend was evident across all samples.

Zeta potential (ζ) results also reflected interesting trends. Generally speaking, ζ decreased with increasing PEG chain length (for 4nm PO-PEG SPIONs, 4nm NH₂-PEG SPIONs, and 10nm DSPE-PEG SPIONs). For 4nm DSPE-PEG SPI-ONs, ζ showed high error and no observable trend. For 10nm PO-PEG SPIONs, ζ showed low error, but no definitive trend. And for 10nm NH₂-PEG SPIONs, ζ appeared to be bimodal, with 1k and 3.4k samples having more similar ζ values, which were different than the similar ζ values shared by 2k and 5k samples. Given the same PEG chain length and PEGylation method, ζ for 4nm samples were generally closer to neutral than ζ for the equivalent 10nm sample. A definitive trend in ζ as a function of PEGylation method was not observed.

TGA results generally indicated averages similar to the targeted surface density of 0.7 PEG chains/nm². Importantly for comparison, there were no statistically significant differences (p > 0.05) in surface density between particles within the same PEGylation method and core size (e.g., all 10nm PO-PEG SPIONs) – with the exception of 4nm DSPE-PEG 5k samples, which exhibited a significantly higher surface density than either 1k or 2k DSPE-PEG samples of the same core size (p < 0.05). This suggests that we can attribute differences in MR relaxivities (described below) for particles with the same PEGylation method and core size to variations in PEG chain length, with the exception of results from the 4nm DSPE-PEG 5k samples. Similarly, there were no statistically significant differences in surface density between particles with the same PEG chain length and core size (e.g.,



FIGURE 2. (A) Impact of SPION core size and PEG chain length on r₁ for PO-PEG SPIONs, NH₂-PEG SPIONs, and DSPE-PEG SPIONs. Error bars are standard error in the calculated slope value. (B) Impact of SPION core size and PEG chain length on r₂ CPMG (see Supplemental Materials and Methods) for PO-PEG SPIONs, NH₂-PEG SPIONs, and DSPE-PEG SPIONs. Error bars are standard error in the calculated slope value.

all 4nm SPIONs with any 1k PEG). This suggests that we can attribute variations in MR relaxivities for particles with the same core size and PEG chain length to differences in PEGylation method and their resulting chemical differences.

The impact of SPION core size, PEGylation method, and PEG chain length on r_1 relaxivity was notable (Fig. 2A). DSPE-PEG SPIONS had markedly lower r_1 relaxivity than either NH₂- or PO-PEG SPIONs. For 4nm SPIONs with comparable PEG chain lengths, NH₂-PEG SPIONs had r_1 relaxivities at least 2.15x higher than PO-PEG SPIONs, and at least 8.23x higher than DSPE-PEG SPIONs for equivalent PEG chain lengths. For 10nm SPIONs, NH₂-PEG SPIONs had r_1 relaxivities of at least 2.09x higher than PO-PEG SPIONs, and at least 33.87x higher than DSPE-PEG SPIONs.

Interestingly, for NH₂- and PO-PEG SPIONs, the maximum effect imparted on r_1 relaxivity by changing PEG chain length was between 34-45% and was not markedly different for the two different core sizes. However, for DSPE-PEG SPIONs, r_1 relaxivity differed by a factor of 360 (10 nm core) to 480% (4 nm core) by changing PEG chain length.

For 10nm SPIONs, increasing the PEG chain length generally resulted in an increase in r_1 . For 4nm SPIONs, a trend was not apparent. With the notable exception of the 5k PO-PEG SPIONs (for which the 10nm SPION had a higher r_1 relaxivity), for all PEGylation methods and PEG chain lengths, 4nm SPIONs had higher r_1 relaxivities than their 10nm counterparts. This is the opposite effect than expected (and previously reported) for r_2 relaxivity. Differences in otherwise equivalent samples with different core sizes ranged from 0.91x (PO-PEG5k, where the r_1 of 10nm samples was greater than that of 4nm samples) to 2.84x (DSPE-PEG1k).

The impact of SPION core size, PEGylation method, and PEG chain length on r_2 (CPMG method) was significant (Fig. 2B), and only partly in line with what was observed with

the r_1 trends. The PEGylation method and resulting chemical composition of the layer closest to the SPION core had the most definitive impact on r_2 relaxivities. In line with r_1 results, across PEG chain lengths, NH₂-PEG SPIONs had the highest r_2 relaxivities; 1.41–4.67x higher than DSPE-PEG SPIONs and 1.06–7.02x higher than PO-PEG SPIONs.

For NH₂-PEG SPIONs with both 4nm and 10nm cores, increasing PEG chain length led to an increase in r₂ relaxivity. This was also the case only for DPSE-PEG SPIONs with 10 nm cores. For DSPE-PEG SPIONs with 4nm cores the largest PEG length also led to the largest r2, but there was a drop in r₂ at the intermediate PEG length. For PO-PEG SPIONs with 10 nm cores, there was not a definitive trend. However, for PO-PEG SPIONs with 4nm cores, increasing the PEG chain length led to a consistent decrease in r2 relaxivity. Additionally, for NH2-PEG SPIONs, r2 relaxivities were higher for the 10nm core sizes at each PEG chain length, with differences in equivalent samples with different core sizes ranging from 1.02x (PEG5k) to 1.30x (PEG2k). However, the opposite was true for PO-PEG SPIONs, where differences in equivalent samples with different core sizes ranged from 1.57x (PEG2k) to 3.56x (PEG1k). Differences in equivalent samples with different core sizes for DSPE-PEG SPIONs did not show a consistent trend.

Spin echo-derived r_2 relaxivities were higher than or comparable to (i.e., calculated values share an overlapping standard error range) CPMG relaxivities as expected (Fig. 3). Where they were greater, r_2 SE-derived relaxivities were 1.023x to 1.20x the r_2 CPMG-derived relaxivities, with the highest ratio observed for 5k PO-PEG, 10nm SPIONs.

With the exception of 4nm 1k PO-PEG SPIONs, NH_2 -PEG SPIONs had the highest r_2* relaxivities across PEG chain lengths (Fig 4). For NH_2 -PEG SPIONs with both 4nm and 10nm cores, increasing PEG chain length led to an increase



FIGURE 3. Comparison between relaxivities derived from spin echo (SE) measurements (blue) and CPMG measurements (orange). The general similarity of these results indicates that diffusion of particles in solution did not play a substantial role in altering measured relaxation rates. Error bars are standard error in the calculated slope value. For a particular data set (the 1k PO-PEG SPIONs with a 4nm core), r₂ rates are not reported for the SE method, as that method is even less robust to the presence of inhomogeneities, referred to in the Supplementary Methods and Materials.



FIGURE 4. Impact of SPION core size and PEG chain length on r_2* for PO-PEG SPIONs, NH₂-PEG SPIONs, and DSPE-PEG SPIONs. Error bars are standard error in the calculated slope value.

in r_2* relaxivity. This was also the case only for DPSE-PEG SPIONs with 10nm cores. For DSPE-PEG SPIONs with 4nm cores, and for PO-PEG SPIONs with 10nm cores, there was not a definitive trend. However, for PO-PEG SPIONs with 4nm cores, increasing the PEG chain length led to a consistent decrease in r_2* relaxivity. Additionally, for NH₂-PEG SPI-ONs, r_2* relaxivities were higher for 10nm core sizes at each PEG chain length, while the opposite was true for PO-PEG SPIONs. Differences in equivalent samples with different core sizes for DSPE-PEG SPIONs did not show a consistent trend. As expected, r_{2*} vs. r_{2} relaxivities for each given sample were either comparable to (with inclusion of standard error) or significantly greater than CPMG relaxivities (Fig. 5). Where they were greater, r_{2*} were 1.08x to 2.67x the r_{2} CPMGderived relaxivities, with the highest ratio observed for 1k PO-PEG, 4nm SPIONs.

III. DISCUSSION

It would generally be expected that, with equivalent core sizes and PEGylation methods, longer PEG chain lengths would lead to larger effective diameters, as observed with PO-PEG SPIONs (Table 1). However, in cases where this was not observed, it is possible that aggregation effects were at play. This seems to be a likely explanation particularly for DSPE-PEG SPIONs, as the effective diameter of 5k DSPE-PEG SPIONs was generally substantially lower than 2k DSPE-PEG SPIONs of the same core size. The larger standard deviation observed in DSPE-PEG SPION effective diameter measures could also be reflective of aggregation. For NH₂-PEG SPIONs, where the general size progression is 1k < 2k < 5k < 3.4k, the observed difference between 3.4k and 5k samples may be due to a difference in the preferred PEG chain conformation, or potentially to a small amount of aggregation occurring, as the standard deviations are not as high as those observed for **DSPE-PEG SPIONs.**

In comparing samples with similar PEGylation methods and PEG chain lengths, it would be expected that the effective diameter of 4nm samples would be smaller than 10nm samples. This was confirmed for both PO-PEG and NH₂-PEG SPIONs. However, this was not true for DSPE-PEG SPIONS. It is likely that, again, aggregation plays a role in these results.

Zeta potential measurements reflect the potential at the slipping/shear plane of particles in solution [31]. While PEG itself does not carry a high surface charge, SPIONs alone in solution tend to carry a high negative charge (e.g., CA-SPIONs have a $\zeta \sim -35$ mV). It is thus expected that coating with the more neutral PEG will raise the ζ of the particles closer to neutral (0mV) [31], [32]. This was observed in all cases.

The general increase towards neutral of ζ with increasing PEG chain length may be expected; it is possible that longer PEG chains help to better "mask" the highly negatively charged SPION core. The higher ζ in 4 nm samples as compared to 10 nm samples of the same PEGylation method and PEG chain length may be due to the same effect. Finally, it is intuitively understood that there was no observed trend in ζ as a function of PEGylation method, since solution-facing ends of each PEG chain were chemically identical regardless of PEGylation method.

The observed results from TGA tests indicated that the majority of samples have non-significant differences in terms of surface coverage (with the exception of 4nm DSPE-PEG 5k samples, which have a statistically significantly greater surface coverage). This indicates that, with the exception of 4nm DSPE-PEG 5k SPIONs, observed differences in MR relaxivities between samples will solely be due to differences in core size, PEG chain length, or PEGylation method. This





FIGURE 5. Relationship between r_{2*} and CPMG-derived r_2 values across all samples. Black lines indicate fit from linear regression; grey shading indicates 95% confidence interval. There is a strong positive correlation between r_{2*} and r_2 for each PEG attachment method (all R > 0.85). For NH₂-PEG SPIONs: r_{2*} vs r_2 (R = 0.986). Slopes of linear regression: 0.69 (DSPE-PEG), 1.13 (NH₂-PEG), and 2.54 (PO-PEG).

is an important confirmation step that has been omitted in previous literature examining the impact of these parameters on SPION MR properties [15], [29]. We also note that all surface densities are well above the critical surface density required for overlapping chains [33].

Perhaps the most salient trend in MR measurements is that r₁ and r₂ relaxivities are greater in NH₂-PEG SPIONs than either PO-PEG or DSPE-PEG SPIONs of the same core size and PEG chain length. It is hypothesized that this is due to the hydrophilic nature of the group closest to the core for these SPIONs (citric acid), in comparison to the less hydrophilic nature of the phosphine oxide and the hydrophobic nature of the DSPE chains. The more hydrophilic group near the core may allow water to move more easily closer to the core and spend more time in proximity to the magnetic field generated by the core. In comparing PO-PEG and DSPE-PEG SPIONs across particles with the same core size and PEG chain length, results appear to differ between r1 and r2 relaxivities. PO-PEG SPIONs show far higher r₁ relaxivities across the board. However, the difference between r2 relaxivities for PO-PEG and DSPE-PEG SPIONs do not show a consistent trend.

For NH₂-PEG and DSPE-PEG SPIONs, within a given SPION core, increasing PEG chain length from 1k to 5k generally led to an increase for r_1 and CPMG-derived r_2 relaxivities. This is consistent with results from Tong, et al. who considered PEG chain lengths over 1kDa [29]. This is likely because increased PEG chain length creates a larger corona, slowing movement of water molecules around the particle, thus increasing the amount of time water protons spend in proximity to the core. Interestingly, this is not true for PO-PEG SPIONs, which do not show a consistent trend with PEG chain length.

Results of this study do not necessarily corroborate some previous findings, [4], [27] i.e., SPIONs with larger core sizes yield higher r_2 relaxivities in samples with equivalent PEG chain length and PEGylation method. Although this is true for NH₂–PEG SPIONs, there is a less distinctive trend for both PO-PEG and DSPE-PEG SPIONs. The trend observed

in 5kDa PO-PEG SPIONs could potentially be linked to the increased surface coverage of 4nm samples as measured by TGA; however, this is not the case in 1kDa samples. On the other hand, SPIONs with larger core sizes generally yielded lower (or comparable) r_1 relaxivities in samples with equivalent PEG chain length and PEGylation method. This trend may be related to recent results demonstrating that SPIONs with sufficiently small core diameters can be used as T_1 contrast agents [34], [35]. Additional work is needed to further elucidate the impact of SPION core size on MR relaxivities with different PEG attachment methods.

IV. CONCLUSION

Our results suggest the chemical composition and, in particular, hydrophobicity/hydrophilicity of the group linking PEG chains to a SPION core may play a larger role in the resulting MR relaxivities than other variable properties such as SPION core size and PEG chain length. In addition, impact of SPION core size and PEG chain length on the resulting sample relaxivities is not a simple relationship, but appears to be conflated with the impact of PEGylation method and nature of the chemical group closer to the SPION core.

Such comprehensive and controlled studies of the impact of different fabrication parameters on MR relaxivities are necessary to provide a framework to optimize SPION MR properties and reveal the complexity of interactions involved in SPION functionalization. This work carries implied clinical significance in terms of the development of future MR contrast agents with properties that may be beneficial for the diagnosis and safety of patients.

V. MATERIALS AND METHODS

A number of SPIONs were synthesized to be physiochemically distinct in terms of their core size, PEG chain length, and the method of PEG attachment to the SPION core. SPION properties including size, zeta potential, and coating density of the attached PEG were evaluated. The MR relaxivities of these SPIONs were subsequently measured and compared.

SUPPLEMENTARY MATERIALS

For specific Materials and Methods information, please refer to the Supplementary Materials, which contain a thorough description of all methods and materials used for the experiments described in this manuscript.

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REFERENCES

- [1] S. Yoffe, T. Leshuk, P. Everett, and F. Gu, "Superparamagnetic iron oxide nanoparticles (SPIONs): Synthesis and surface modification techniques for use with MRI and other biomedical applications," *Current. Pharm. Des.*, vol. 19, no. 3, pp. 493–509, 2013.
- [2] Y.-X. J. Wang, "Superparamagnetic iron oxide based MRI contrast agents: Current status of clinical application," *Quantum Imag. Med. Surg.*, vol. 1, no. 1, pp. 35–40, Dec. 2011.
- [3] Y.-X. J. Wang, S. M. Hussain, and G. P. Krestin, "Superparamagnetic iron oxide contrast agents: physicochemical characteristics and applications in MR imaging," *Eur. Radiol.*, vol. 11, no. 11, pp. 2319–2331, Nov. 2001.
- [4] Y.-W. Jun *et al.*, "Nanoscale size effect of magnetic nanocrystals and their utilization for cancer diagnosis via magnetic resonance imaging," *J. Amer. Chem. Soc.*, vol. 127, no. 16, pp. 5732–5733, Apr. 2005.
- [5] J.-H. Lee *et al.*, "Artificially engineered magnetic nanoparticles for ultra-sensitive molecular imaging," *Nature Med.*, vol. 13, no. 1, pp. 95– 99, Jan. 2007.
- [6] N. Nitin, L. E. W. LaConte, O. Zurkiya, X. Hu, and G. Bao, "Functionalization and peptide-based delivery of magnetic nanoparticles as an intracellular MRI contrast agent," *J. Biol. Inorg. Chem. JBIC Publ. Soc. Biol. Inorg. Chem.*, vol. 9, no. 6, pp. 706–712, Sep. 2004.
- [7] M. Sardan, E. D. Eren, A. Ozdemir, A. B. Tekinay, and M. O. Guler, "Bioactive peptide functionalized superparamagnetic iron oxide nanoparticles (SPIONs) for targeted imaging with MRI," in *Proc. 5th Int. Workshop Magn. Particle Imag.*, 2015, pp. 1–1.
- [8] K. Hervé et al., "The development of stable aqueous suspensions of PE-Gylated SPIONs for biomedical applications," *Nanotechnology*, vol. 19, no. 46, 2008, Art. no. 465608.
- [9] M. Mahmoudi, S. Sant, B. Wang, S. Laurent, and T. Sen, "Superparamagnetic iron oxide nanoparticles (SPIONs): Development, surface modification and applications in chemotherapy," *Adv. Drug Del. Rev.*, vol. 63, no. 1–2, pp. 24–46, Jan. 2011.
- [10] Y. Gossuin, P. Gillis, A. Hocq, Q. L. Vuong, and A. Roch, "Magnetic resonance relaxation properties of superparamagnetic particles," *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.*, vol. 1, no. 3, pp. 299–310, Jun. 2009.
- [11] R. Chen *et al.*, "Parallel comparative studies on mouse toxicity of oxide nanoparticle- and gadolinium-based T1 MRI contrast agents," ACS Nano, vol. 9, no. 12, pp. 12425–12435, Dec. 2015.
- [12] M. Takahashi, H. Tsutsui, C. Murayama, T. Miyazawa, and B. Fritz-Zieroth, "Neurotoxicity of gadolinium contrast agents for magnetic resonance imaging in rats with osmotically disrupted blood-brain barrier," *Magn. Reson. Imag.*, vol. 14, no. 6, pp. 619–623, 1996.
- [13] H. S. Thomsen, "Gadolinium-based contrast media may be nephrotoxic even at approved doses," *Eur. Radiol.*, vol. 14, no. 9, pp. 1654–1656, Jun. 2004.
- [14] M. Yu, S. Huang, K. J. Yu, and A. M. Clyne, "Dextran and polymer polyethylene glycol (PEG) coating reduce both 5 and 30 nm iron oxide nanoparticle cytotoxicity in 2D and 3D cell culture," *Int. J. Mol. Sci.*, vol. 13, no. 5, pp. 5554–5570, May 2012.

- [15] Y. C. Park *et al.*, "Effect of PEG molecular weight on stability, T2 contrast, cytotoxicity, and cellular uptake of superparamagnetic iron oxide nanoparticles (SPIONs)," *Colloids Surf. B Biointerfaces*, vol. 119, pp. 106–114, Jul. 2014.
- [16] I. Lynch and K. A. Dawson, "Protein-nanoparticle interactions," *Nano Today*, vol. 3, no. 1, pp. 40–47, Feb. 2008.
- [17] M. Mahmoudi, I. Lynch, M. R. Ejtehadi, M. P. Monopoli, F. B. Bombelli, and S. Laurent, "Protein–Nanoparticle interactions: Opportunities and challenges," *Chem. Rev.*, vol. 111, no. 9, pp. 5610–5637, Sep. 2011.
- [18] T. Cedervall *et al.*, "Understanding the nanoparticle-protein corona using methods to quantify exchange rates and affinities of proteins for nanoparticles," *Proc. Nat. Acad. Sci. U. S. A.*, vol. 104, no. 7, pp. 2050– 2055, Feb. 2007.
- [19] R. Gref *et al.*, "Stealth' corona-core nanoparticles surface modified by polyethylene glycol (PEG): Influences of the corona (PEG chain length and surface density) and of the core composition on phagocytic uptake and plasma protein adsorption," *Colloids Surf. B Biointerfaces*, vol. 18, no. 3, pp. 301–313, Oct. 2000.
- [20] R. L. C. Wang, H. J. Kreuzer, and M. Grunze, "Molecular conformation and solvation of oligo (ethylene glycol)-terminated self-assembled monolayers and their resistance to protein adsorption," *J. Phys. Chem. B*, vol. 101, no. 47, pp. 9767–9773, Nov. 1997.
- [21] K. Mu *et al.*, "Monoclonal antibody-conjugated superparamagnetic iron oxide nanoparticles for imaging of epidermal growth factor receptor-targeted cells and gliomas," *Mol. Imag.*, vol. 14, 2015, doi: 10.2310/7290.2015.00002.
- [22] W. Guo, J. D. Morrisett, M. E. DeBakey, G. M. Lawrie, and J. A. Hamilton, "Quantification in situ of crystalline cholesterol and calcium phosphate hydroxyapatite in human atherosclerotic plaques by solid-state magic angle spinning NMR," *Arterioscler. Thromb. Vasc. Biol.*, vol. 20, no. 6, pp. 1630–1636, Jun. 2000.
- [23] R. Koos, A. H. Mahnken, A. M. Sinha, J. E. Wildberger, R. Hoffmann, and H. P. Kühl, "Aortic Valve Calcification as a Marker for Aortic Stenosis Severity: Assessment on 16-MDCT," *Amer. J. Roentgenol.*, vol. 183, no. 6, pp. 1813–1818, Dec. 2004.
- [24] Z. R. Stephen, F. M. Kievit, and M. Zhang, "Magnetite Nanoparticles for Medical MR Imaging," *Mater. Today Kidlington Engl.*, vol. 14, no. 7–8, pp. 330–338, 2011.
- [25] P. Gillis, F. Moiny, and R. A. Brooks, "On T(2)-shortening by strongly magnetized spheres: A partial refocusing model," *Magn. Reson. Med.*, vol. 47, no. 2, pp. 257–263, Feb. 2002.
- [26] P. Gillis and S. H. Koenig, "Transverse relaxation of solvent protons induced by magnetized spheres: Application to ferritin, erythrocytes, and magnetite," *Magn. Reson. Med.*, vol. 5, no. 4, pp. 323–345, Oct. 1987.
- [27] Y.-W. Jun, J.-W. Seo, and J. Cheon, "Nanoscaling laws of magnetic nanoparticles and their applicabilities in biomedical sciences," Acc. Chem. Res., vol. 41, no. 2, pp. 179–189, Feb. 2008.
- [28] U. I. Tromsdorf *et al.*, "Size and surface effects on the MRI relaxivity of manganese ferrite nanoparticle contrast agents," *Nano Lett.*, vol. 7, no. 8, pp. 2422–2427, Aug. 2007.
- [29] S. Tong, S. Hou, Z. Zheng, J. Zhou, and G. Bao, "Coating optimization of superparamagnetic iron oxide nanoparticles for high T2 relaxivity," *Nano Lett*, vol. 10, no. 11, pp. 4607–4613, 2010.
- [30] F. Reynolds, T. O'loughlin, R. Weissleder, and L. Josephson, "Method of determining nanoparticle core weight," *Anal. Chem.*, vol. 77, no. 3, pp. 814–817, Feb. 2005.
- [31] S. Bhattacharjee, "DLS and zeta potential–What they are and what they are not?," J. Controlled Release, vol. 235, pp. 337–351, Aug. 2016.
- [32] H. Kouchakzadeh, S. A. Shojaosadati, A. Maghsoudi, and E. V. Farahani, "Optimization of pegylation conditions for BSA nanoparticles using response surface methodology," *AAPS PharmSciTech*, vol. 11, no. 3, pp. 1206–1211, Sep. 2010.
- [33] Y. Park *et al.*, "Stability of superparamagnetic iron oxide nanoparticles at different pH values: Experimental and theoretical analysis," *Langmuir*, vol. 28, no. 15, pp. 6246–6255, Apr. 2012.
- [34] B. H. Kim *et al.*, "Large-scale synthesis of uniform and extremely small-sized iron oxide nanoparticles for high-resolution T1 magnetic resonance imaging contrast agents," *J. Amer. Chem. Soc.*, vol. 133, no. 32, pp. 12624–12631, Aug. 2011.
- [35] H. Wei *et al.*, "Exceedingly small iron oxide nanoparticles as positive MRI contrast agents," in *Proc. Nat. Acad. Sci.*, Feb. 2017, Art. no. 201620145.