Review Article PPARy in Neuroblastoma

Alessandro Peri, Ilaria Cellai, Susanna Benvenuti, Paola Luciani, Silvana Baglioni, and Mario Serio

Endocrine Unit, Department of Clinical Physiopathology, Center for Research, Transfer and High Education on Chronic, Inflammatory, Degenerative and Neoplastic Disorders (DENOThe), University of Florence, 50139 Florence, Italy

Correspondence should be addressed to Alessandro Peri, a.peri@dfc.unifi.it

Received 28 February 2008; Accepted 14 April 2008

Recommended by Dipak Panigrahy

Neuroblastoma (NB) is the most common extracranial tumor in children and accounts for around 15% of all paediatric oncology deaths. The treatment of NB includes surgery, chemotherapy, and radiotherapy. Unfortunately, most children with NB present with advanced disease, and more than 60% of patients with high-risk features will have a poor prognosis despite intensive therapy. Agonists of the nuclear receptor peroxisome proliferator-activated receptor γ (PPAR γ) have been shown to have pleiotropic effects, including antineoplastic effects. The studies that addressed the role and the possible mechanism(s) of action of PPAR γ in NB cells are reviewed.

Copyright © 2008 Alessandro Peri et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

1. INTRODUCTION

Neuroblastoma (NB), the most common extracranial solid tumor in children, accounts for more than 7% of malignancies in patients younger than 15 years and around 15% of all paediatric oncology deaths [1]. The disease has a heterogeneous clinical presentation and course [2]. First of all, NB is a disease of the sympaticoadrenal lineage of the neural crest, and therefore tumors can develop anywhere in the sympathetic nervous system. The majority of NB is developed within the abdomen and at least 50% of these tumors arise in the adrenal medulla [2]. Other frequent localizations include the neck, chest, and pelvis [3]. The clinical presentation of the disease may be also highly variable and depends on the site of the primary tumor as well as on the presence or absence of metastatic disease (mostly haematogeneous dissemination to cortical bone, bone marrow, liver, and noncontiguous lymph nodes) or paraneoplastic syndromes. The diagnosis of NB is based on histopathological assessment of tumor tissue or on the detection of cancer cells in a bone marrow aspirate/biopsy, together with the presence of increased levels of urinary catecholamines [2]. Imaging studies for the localization of the disease include computed tomography, magnetic resonance, ⁹⁹mTc-diphosphonate, or metaiodobenzylguanidine (using ¹²³I) scintigraphy for the detection of bone metastases.

The treatment of NB includes surgery, chemotherapy (i.e., cisplatin, etoposide, doxorubicin, cyclophosphamide, vincristine) [4], and radiotheraphy. Unfortunately, although substantial improvement in outcome of certain subsets of patients has been observed during the past few decades [2], most children with NB present with advanced disease and more than 60% of patients with high-risk features will have a poor prognosis despite intensive therapy [5, 6]. Thus, research efforts to understand the biological basis of NB and to identify new and more effective therapies are essential to improve the outcome for these children. In the last years an expanding number of new agents have been developed for use in high-risk patients affected by recurrent disease. Cytotoxic agents, such as the topoisomerase 1 inhibitors topotecan and irinotecan, have an acceptable toxicity profile and are effectively used in early relapsing NB [7-10]. The delivery of radioactive molecules that are selectively concentrated in NB cells, such as metaiodobenzylguanidine, somatostatin analogues, anti-G_{D2} (a disialoganglioside) antibodies, has been used in clinical trials [11-22]. G_{D2}targeted therapies using monoclonal antibodies are under investigation in phase III trials [19, 23, 24], and other immunotherapeutic strategies (i.e., vaccination or cellular immunotherapy using engineered cytolityc T lymphocytes) are currently investigated [25, 26]. Similarly, angiogenesis [27-33] and tyrosine kinase [34-38] inhibitors appear as an

attractive therapeutic option and clinical trials are ongoing. Retinoids have been shown to interfere with cell growth and to induce apoptosis in NB cells [39, 40] and preliminary clinical trials with retinoids in NB resulted in improved event-free survival in high-risk patients, with limited toxic effects [41, 42]. Thiazolidinediones (TZDs) are a class of molecules that activate the nuclear receptor peroxisome proliferator-activated receptor γ (PPAR γ) [43] and promote association with the 9-cis retinoic X receptor (RXR) to form functional heterodimers that recognize its cognate DNA response element within target genes [44, 45]. TZDs have been shown to have antineoplastic effects, as extensively discussed in this issue of the journal, in agreement with the demonstration that PPARy/RXR signalling exerts an important role in inhibiting cell proliferation and/or in inducing apoptosis [46]. It has been also shown that PPARy and RXR ligands may have a synergistic effect in inducing cell differentiation [47, 48] and in inhibiting cell growth in different tumors, such as colon, lung, and breast cancer [49-51]. There is evidence that also PPAR α and PPAR β ligands may play a role in counteracting tumoral cell growth and in promoting cell differentiation, including neuroblastoma cells [52, 53]. However, most of the reports covering this issue, that have been published in the literature so far, deal with PPARy agonists. Therefore, the role of PPARy ligands as a possible therapeutic option in NB is reviewed and discussed here.

2. PPARy AND PPARy AGONISTS IN NEUROBLASTOMA

The first demonstration that PPARy is expressed in NB cells was provided by Han et al. in 2001 [54]. Using RT-PCR the authors showed that LA-N-5 NB cells express also PPAR β , but not PPAR α . Similarly, in sections from human primary NB immunostaining for PPARy was detected in the nucleus and occasionally in the cytoplasm of cells, particularly in those showing ganglionic differentiation. Sato et al. [55] addressed the possibility that the amount of expression of PPARy in NB might be correlated to patients' outcome. To this purpose, the level of mRNA was measured by semiquantitative RT-PCR in NB samples from 17 patients under the age of one year. In this subset of patients, spontaneous differentiation and regression are often observed [56], and some investigators suggested to observe these patients without surgery until there is an increase of vanilmandelic acid (VMA) or tumoral growth occurs [57, 58]. PPARy mRNA was present in 12 samples. No difference between the expression of PPARy and histology, age, staging, DNA ploidy was observed, yet a correlation with the change in urinary VMA was found. In fact, in samples resected from patients, who showed a reduction of VMA in the period of time preceding surgery (2–7 months), higher PPARy expression was detected compared to those patients in which VMA increased. The authors hypothesized that PPARy might play a role in the decrease of VMA and hence in the regression of early-onset NB. Thereafter, several studies addressed the potential role of endogenous or synthetic PPARy ligands in counteracting NB cell growth.

5-Deoxy- $\Delta^{12,14}$ -prostaglandin J_2 (15-deoxy-PGJ₂) is a naturally occurring downstream metabolite of PGD₂, that is produced by degradation of PGD₂ [59]. In contrast to classic prostaglandins, which act after binding to cell surface G-protein coupled receptors (GPCRs), 15-deoxy-PGJ₂ is a high-affinity endogenous ligand of PPARy. A pro-apoptotic effect of 15-deoxy-PGJ₂ in SH-SY5Y NB cells, that was reverted by the caspase inhibitor Z-VAD, was reported by Rohn et al. [60]. A subsequent study confirmed that 15deoxy-PGJ₂ was able to inhibit cell growth and to induce apoptosis via the activation of ERK2 in two additional NB cell lines (i.e., SK-N-SH and SK-N-MC). An increase of the expression of the pro-apoptotic proteins caspase-3, caspase-9, and Bax, together with the decrease of the anti-apoptotic protein Bcl-2, was also observed [61]. The PPARy antagonist GW9662 reverted the effects of 15-deoxy-PGJ₂, including the activation of ERK2. The authors concluded that 15-deoxy-PGJ₂ induced apoptosis in a PPARy-dependent manner through the activation of ERK pathway. Another study showed that the mechanism by which 15-deoxy-PGJ₂ arrests cell growth may vary depending on the content of lipids in the culture medium [62]. In particular, the delipidation of fetal calf serum, which removes known serum lipid mitogens including lysophosphatidic acid [63] and sphingosine 1phosphate [64], potentiated the degree of 15-deoxy-PGJ₂induced growth inhibition via PPARy-dependent apoptosis in the NB cell line IMR-32. Conversely, growth inhibition in the presence of complete medium occurred through programmed cell death typeII (autophagy).

PPARy-independent effects of 15-deoxy-PGJ₂ have been also described. Jung et al. reported that this PPARy ligand was able to increase NGF-induced differentiation of PC-12 NB cells, as assessed by neurite extension and expression of neurofilament [65]. Pretreatment with the PPARy antagonist bisphenol A diglycidyl either did not alter the differentiating activity of 15-deoxy-PGJ₂. The fact that PC-12 cells do not express PPARy further supported the hypothesis that the biological effects elicited by 15-deoxy-PGJ₂ were not mediated by this receptor. Conversely, 15-deoxy-PGJ₂ enhanced NGFinduced p38 MAP kinase expression and phosphorylation as well as the activation of transcription factor AP-1, that on turn were counteracted by a specific inhibitor of p38 MAP kinase (SB203580). Altogether, these data suggested that the promoting effect of 15-deoxy-PGJ₂ on cell differentiation may be mediated by the activation of p38 MAP kinase in conjunction with the AP-1 signalling pathway.

Other studies addressed the role of *synthetic PPARy ligands* in counteracting cell growth in NB. In the already mentioned work by Han et al., in which the presence of PPARy in NB cells was described for the first time, the authors also demonstrated that the synthetic PPARy agonist GW1929 induced the differentiation of LA-N-5 cells and inhibited cell proliferation [54]. A subsequent study of the same group showed that the prodifferentiating effect of GW1929 is mediated by PPARy, because it was inhibited by the cotreatment with specific antagonists [66]. The antiproliferative effects of the TZDs ciglitazone, pioglitazone, troglitazone, and rosiglitazone in different NB cell lines (i.e., LAN-1, LAN-5, LS, IMR-32, SK-N-SH, SH-SY5Y) were determined by Valentiner et al. [67]. In these cell lines, which express PPARy, the four ligands were able to markedly inhibit cell growth at the highest doses that were used (10 and $100\,\mu\text{M}$). Ciglitazone determined the strongest inhibitory effect (more than 90% inhibition). The potency of the different PPARy ligands was not related to the amount of expression of PPARy in NB cell lines. Thus, the authors hypothesized that the effects of the molecules that were used seem to be independent of the amount of PPARy protein in one particular cell line. Conversely, they concluded that the response to PPARy ligands may rather depend on various cellular conditions, which are associated with the function of the receptor, such as its activation, translocation to the nucleus and binding to PPAR response elements (PPRE). The role played by PPARy transactivation was confirmed by the finding that growth inhibition determined by 15-deoxy-PGJ₂ and ciglitazone in NB cells was counteracted by the repression of PPARy transactivation via retinoblastoma protein overexpression [68]. Further studies investigated whether the inhibitory effect of TZDs on cell growth was mediated, at least partially, by a stimulatory effect on apoptosis. Kato et al. found that in NB-1 cells troglitazone induced PPARydependent apoptosis [69]. Similar data were reported later on by Schultze et al. [70], who showed that in SHEP NB cells the pro-apoptotic effect of the death ligand TRAIL is reinforced by troglitazone. However, troglitazone-induced sensitization to TRAIL appeared to be PPARy-independent, because it was achieved at concentrations that failed to activate PPARy. Conversely, the authors highlighted the fact that troglitazone may induce apoptotic death by various PPARy-independent mechanisms. In particular, troglitazone led to a marked downregulation of the antiapoptotic protein Survivin, as well as to an upregulation of the agonistic TRAIL receptor TRAIL-R2.

Overall, these data strongly indicate that PPAR*y* ligands are able to effectively counteract cell growth and to induce apoptosis in NB cells. Undoubtedly, the role of PPAR*y* in eliciting these responses would be further clarified by studies designed for instance to manipulate gene expression (i.e., by small interfering RNA or dominant negative strategies). To our knowledge, there are only two reports from one Korean group showing, in contrast to the current opinion, that a PPAR*y* agonist (i.e., rosiglitazone) protects NB (SH-SY5Y) cells against the neurotoxins acetaldehyde and 1-methyl-4phenylpyridinium ion, through inhibition of apoptosis [71, 72].

3. DIFFERENTIAL PPAR γ TRANSACTIVATION IN NEUROBLASTOMA CELL LINES WITH A DIFFERENT PHENOTYPE: RELATIONSHIP WITH THE RESPONSE TO ROSIGLITAZONE

NB is a phenotypically heterogeneous tumor, displaying cells of neuronal, melanocytic, or glial/schwannian lineage. This cellular heterogeneity is also present in vitro, where cells of neuroblastic (N) or stromal (S) type may be identified. It has been hypothesized that the sensitivity to PPARy ligands may be, at least partially, dependent on the different cell phenotype. To this purpose, Servidei et al. examined the response of 8 different NB cell lines with N (SH-SY5Y, LA-N-5, SMS-KCNR, SK-N-DZ), mixed (SK-N-FI, LA-N-1), or S (SH-EP1, SK-N-AS) phenotype to PPARy agonists [73]. All the cell lines investigated expressed a functionally active PPARy. 15-deoxy-PGJ₂ and rosiglitazone inhibited cell growth in all cell lines, and the sensitivity appeared to be more related to the cell phenotype than to PPARy expression. In particular, the N type cells appeared the most sensitive to treatment. In this experimental setting, the cotreatment with PPARy ligands and the RXR ligand 9-cis retinoic acid did not determine any synergistic effect on growth inhibition. The more evident response of N type cells to PPARy ligands was in part related to their higher capability to undergo apoptosis, although only 15-deoxy-PGJ₂ appeared to effectively induce the apoptotic cascade in these cells. It has to be said that in this study some experimental observations (i.e., apoptosis and cell viability) were not performed in all the investigated NB cell lines.

In order to further clarify the mechanisms underlying the response of NB cells to PPARy agonists, we compared the response of two cell lines (SH-SY5Y, N type, and SK-N-AS, S type) to rosiglitazone. In contrast to the above-mentioned findings, we observed that micromolar concentrations of rosiglitazone inhibited cell proliferation and reduced cell viability more effectively in SK-N-AS than in SH-SY5Y [74]. The PPARy antagonist BADGE reverted the effect of rosiglitazone, thus suggesting a direct role of PPARy in mediating the effects of this agonist on cell proliferation and viability. In addition, we found that SK-N-AS cells were more sensitive to rosiglitazone in terms of reduction of cell adhesion and invasiveness. The latter effect was in agreement with rosiglitazone-dependent reduced expression of matrix metalloproteinase-9 (MMP-9). In addition, rosiglitazone determined a trend toward increased expression levels of tissue inhibitor of matrix metalloproteinase-1 (TIMP-1). MMPs, which promote the invasion of extracellular matrix by tumoral cells, have been related to the progression of different tumors, including NB [75, 76]. In our study, we also addressed the possible role of rosiglitazone in inducing apoptosis. We demonstrated that micromolar concentrations of this molecule were able to induce caspase-3 activation in SK-N-AS, but not in SH-SY5Y (up to $50 \,\mu$ M). Therefore, all our data indicated that rosiglitazone played an effective antitumoral role in the S type SK-N-AS, yet not in the N type SH-SY5Y NB cells. Although our study was limited to two cell lines, this apparent prevalent effect on a particular cell phenotype may have clinical resonance. In fact, it is known that in NB, following cytotoxic therapy, the residual tumor often shows a reduction of the neuroblastic elements and the persistence of stromal components [77]. Hence, a molecule that appears to have S type NB cells as a preferential target might be of interest in the setting of residual disease.

A further aim of our study was to determine the reason underlying the peculiar sensitivity to rosiglitazone displayed by SK-N-AS cells. Both SK-N-AS and SH-SY5Y expressed a similar amount of PPARy. However, in transient transfection experiments, in which a PPRE-thimidine kinase luciferase reporter plasmid was inserted, we observed that in SK-N-AS 20 μ M rosiglitazone induced a near three-fold increase of the



FIGURE 1: PPARy transcriptional activity in control untreated NB cells (C), in cells treated with rosiglitazone (RGZ) (20 μ M), and in cells transfected with PPARy and treated with RGZ. L/C: peroxisome proliferator response element-n7₃-tk-luciferase reporter activity, normalized for CAT activity. * = *P* < 0.05 *versus* C. ** = *P* < 0.05 *versus* C, and *versus* RGZ-treated cells, in the absence of PPARy transfection (from [74], modified).

reporter activity compared to untreated cells. Conversely, no effect was elicited in SH-SY5Y. Only when these cells were co-transfected with a human PPARy expression plasmid, the response to rosiglitazone was present. These data indicated that the original lack of response showed by SH-SY5Y was due to a very low or absent transactivation potential of the endogenous PPAR γ (Figure 1). The different efficacy of PPARy as a transcriptional activator in the two cell lines might be hypothetically due to the presence of a PPAR γ gene mutation. However, no mutation was found in the entire coding region of the gene. Conversely, we found that the amount of phosphorylated PPARy was markedly lower in SK-N-AS than in SH-SY5Y cells (Figure 2). There is evidence that phosphorylation reduces the activity of the receptor [78]. Therefore, our conclusion was that the higher efficacy of rosiglitazone in SK-N-AS cells was due to a reduced phosphorylation status, hence to increased activity, of PPARy. To our knowledge, this was the first demonstration that the response of NB cells to TZDs may be dependent on PPARy transactivation.

4. PPAR γ AGONISTS IN NEUROBLASTOMA XENOGRAFT MODELS

To our knowledge, no study on the in vivo effect of TZDs in neuroblastoma has been published so far. However, our very recent preliminary in vivo observations on CD-1 athymic nude mice, in which SK-N-AS cells were subcutaneously inoculated, appear to confirm our previous in vitro observations [74]. Rosiglitazone (150 mg/kg/day, in agreement with the average dose used in other in vivo studies addressing different tumors) was administered by gavage for 4 weeks.



FIGURE 2: Detection of total (anti-PPARy antibody) and phosphorylated (anti-P-Ser antibody) PPARy, by Western blot analysis after PPARy immunoprecipitation (from [74], modified).

Tumoral growth was markedly reduced compared to control mice, treated with the vehicle alone. At the end of treatment, the weight of the tumor in rosiglitazone-treated animals was about 60% less than in control animals [Cellai et al; unpublished data]. An extensive molecular characterization of tumor specimens is currently ongoing, in order to elucidate the mechanisms underlying the growth inhibitory effect of rosiglitazone observed in vivo in our xenograft model.

5. CONCLUSIONS

In the last few years in vitro studies have shown that PPAR γ agonists may inhibit NB cell growth by stimulating cell differentiation and/or by inducing apoptosis. The different molecules that have been tested have generally produced similar results. However, the mode of action may change depending on the agonist and/or on the different cell line used. In addition, both PPAR γ -dependent as well as PPAR γ -independent effects have been described. Our recent data suggest that PPAR γ transactivation, determined at least in part by the phosphorylation status of the receptor, may play an important role in determining the response of NB cells to PPAR γ agonists. However, the exact mechanisms of action and the possibility to predict the success or failure of the treatment of NB with these molecules are, at this time, matter of further in vitro as well as in vivo research.

ACKNOWLEDGMENTS

The authors wish to thank the people who collaborated to in vitro and in vivo studies [67, Cellai et al., unpublished data], and in particular Professor Andrea Galli (Gastroenterology Unit, Department of Clinical Physiopathology, University of Florence), Doctor Lisa Simi (Clinical Biochemistry Unit, Department of Clinical Physiopathology, University of Florence), Doctor Monica Muratori (Andrology Unit, Department of Clinical Physiopathology, University of Florence), Doctor Graziella Pratesi (Istituto Nazionale per lo Studio e la Cura dei Tumori, Milan), and Doctor Carol J Thiele (Pediatric Oncology Branch, Center for Cancer Research, National Cancer Institute, National Institutes of Health, Bethesda, MD, USA.) These studies were partially supported by granto from Enta Cassadi Risparimo, Firenze and from Regione Toscana Ctresor Project, Principal Investigator Professor Mario Seriol.

REFERENCES

- [1] National Cancer Institute, Surveillance, Epidemiology and End Results Database, November 2005, http://seer.cancer.gov.
- [2] J. M. Maris, M. D. Hogarty, R. Bagatell, and S. L. Cohn, "Neuroblastoma," *The Lancet*, vol. 369, no. 9579, pp. 2106– 2120, 2007.
- [3] G. M. Brodeur and J. M. Maris, "Neuroblastoma," in *Principles and Practice of Pediatric Oncology*, P. A. Pizzo and D. G. Poplack, Eds., pp. 933–970, JB Lippincott Company, Philadel-phia, Pa, USA, 5th edition, 2006.
- [4] N. K. Cheung, B. H. Kushner, M. LaQuaglia, et al., "N7: a novel multi-modality therapy of high risk neuroblastoma (NB) in children diagnosed over 1 year of age," *Medical and Pediatric Oncology*, vol. 36, no. 1, pp. 227–230, 2001.
- [5] B. De Bernardi, B. Nicolas, L. Boni, et al., "Disseminated neuroblastoma in children older than one year at diagnosis: comparable results with three consecutive high-dose protocols adopted by the Italian Co-Operative Group for Neuroblastoma," *Journal of Clinical Oncology*, vol. 21, no. 8, pp. 1592– 1601, 2003.
- [6] K. K. Matthay, J. G. Villablanca, R. C. Seeger, et al., "Treatment of high-risk neuroblastoma with intensive chemotherapy, radiotherapy, autologous bone marrow transplantation, and 13-cis-retinoic acid," *The New England Journal of Medicine*, vol. 341, no. 16, pp. 1165–1173, 1999.
- [7] L. M. Wagner, K. R. Crews, L. C. Iacono, et al., "Phase I trial of temozolomide and protracted irinotecan in pediatric patients with refractory solid tumors," *Clinical Cancer Research*, vol. 10, no. 3, pp. 840–848, 2004.
- [8] G. Vassal, F. Doz, D. Frappaz, et al., "A phase I study of irinotecan as a 3-week schedule in children with refractory or recurrent solid tumors," *Journal of Clinical Oncology*, vol. 21, no. 20, pp. 3844–3852, 2003.
- [9] R. L. Saylors III, K. C. Stine, J. Sullivan, et al., "Cyclophosphamide plus topotecan in children with recurrent or refractory solid tumors: a Pediatric Oncology Group phase II study," *Journal of Clinical Oncology*, vol. 19, no. 15, pp. 3463–3469, 2001.
- [10] A. Längler, A. Christaras, K. Abshagen, K. Krauth, B. Hero, and F. Berthold, "Topotecan in the treatment of refractory neuroblastoma and other malignant tumors in childhood—a phase-II-study," *Klinische Pädiatrie*, vol. 214, no. 4, pp. 153– 156, 2002.
- [11] S. Tepmongkol and S. Heyman, "¹³¹I MIBG therapy in neuroblastoma: mechanisms, rationale, and current status," *Medical and Pediatric Oncology*, vol. 32, no. 6, pp. 427–432, 1999.
- [12] K. K. Matthay, K. DeSantes, B. Hasegawa, et al., "Phase I dose escalation of 131I-metaiodobenzylguanidine with autologous bone marrow support in refractory neuroblastoma," *Journal of Clinical Oncology*, vol. 16, no. 1, pp. 229–236, 1998.
- [13] A. Garaventa, O. Bellagamba, M. S. Lo Piccolo, et al., "¹³¹Imetaiodobenzylguanidine (¹³¹I-MIBG) therapy for residual neuroblastoma: a mono-institutional experience with 43 patients," *British Journal of Cancer*, vol. 81, no. 8, pp. 1378– 1384, 1999.
- [14] G. A. Wiseman and L. K. Kvols, "Therapy of neuroendocrine tumors with radiolabeled MIBG and somatostatin analogues," *Seminars in Nuclear Medicine*, vol. 25, no. 3, pp. 272–278, 1995.
- [15] M. S. O'Dorisio, M. Hauger, and A. J. Cecalupo, "Somatostatin receptors in neuroblastoma: diagnostic and therapeutic impli-

cations," *Seminars in Oncology*, vol. 21, supplement 13, no. 5, pp. 33–37, 1994.

- [16] P. Borgström, M. Hassan, E. Wassberg, et al., "The somatostatin analogue octreotide inhibits neuroblastoma growth in vivo," *Pediatric Research*, vol. 46, no. 3, pp. 328–332, 1999.
- [17] A. L. Yu, M. M. Uttenreuther-Fischer, C. S. Huang, et al., "Phase I trial of a human-mouse chimeric antidisialoganglioside monoclonal antibody ch14.18 in patients with refractory neuroblastoma and osteosarcoma," *Journal of Clinical Oncology*, vol. 16, no. 6, pp. 2169–2180, 1998.
- [18] N. K. Cheung, B. H. Kushner, S. D. J. Yeh, and S. M. Larson, "3F8 monoclonal antibody treatment of patients with stage 4 neuroblastoma: a phase II study," *International Journal of Oncology*, vol. 12, no. 6, pp. 1299–1306, 1998.
- [19] B. H. Kushner, K. Kramer, and N.-K. V. Cheung, "Phase II trial of the anti-G_{D2} monoclonal antibody 3F8 and granulocytemacrophage colony-stimulating factor for neuroblastoma," *Journal of Clinical Oncology*, vol. 19, no. 22, pp. 4189–4194, 2001.
- [20] T. I. Kang, P. Brophy, M. Hickeson, et al., "Targeted radiotherapy with submyeloablative doses of 131I-MIBG is effective for disease palliation in highly refractory neuroblastoma," *Journal* of *Pediatric Hematology/Oncology*, vol. 25, no. 10, pp. 769–773, 2003.
- [21] J. P. Howard, J. M. Maris, L. S. Kersun, et al., "Tumor response and toxicity with multiple infusions of high dose ¹³¹I-MIBG for refractory neuroblastoma," *Pediatric Blood and Cancer*, vol. 44, no. 3, pp. 232–239, 2005.
- [22] S. G. DuBois, J. Messina, J. M. Maris, et al., "Hematologic toxicity of high-dose iodine-131-metaiodobenzylguanidine therapy for advanced neuroblastoma," *Journal of Clinical Oncology*, vol. 22, no. 12, pp. 2452–2460, 2004.
- [23] Z. C. Neal, J. C. Yang, A. L. Rakhmilevich, et al., "Enhanced activity of hu14.18-IL2 immunocytokine against murine NXS2 neuroblastoma when combined with interleukin 2 therapy," *Clinical Cancer Research*, vol. 10, no. 14, pp. 4839– 4847, 2004.
- [24] D. M. King, M. R. Albertini, H. Schalch, et al., "Phase I clinical trial of the immunocytokine EMD 273063 in melanoma patients," *Journal of Clinical Oncology*, vol. 22, no. 22, pp. 4463–4473, 2004.
- [25] A. Yu, A. Batova, D. Strother, P. Angelini, and R. P. Castleberry, "Promising results of a pilot trial of a G_{D2} directed antiidiotypic antibody as vaccine for high risk neuroblastoma," in *Proceedings of the 11th Conference on Advances in Neuroblastoma Research (ANR '04)*, vol. 75, Genoa, Italy, June 2004.
- [26] S. Gonzalez, A. Naranjo, L. M. Serrano, W.-C. Chang, C. L. Wright, and M. C. Jensen, "Genetic engineering of cytolytic T lymphocytes for adoptive T-cell therapy of neuroblastoma," *The Journal of Gene Medicine*, vol. 6, no. 6, pp. 704–711, 2004.
- [27] H. M. Katzenstein, A. W. Rademaker, C. Senger, et al., "Effectiveness of the angiogenesis inhibitor TNP-470 in reducing the growth of human neuroblastoma in nude mice inversely correlates with tumor burden," *Clinical Cancer Research*, vol. 5, no. 12, pp. 4273–4278, 1999.
- [28] S. Shusterman, S. A. Grupp, and J. M. Maris, "Inhibition of tumor growth in a human neuroblastoma xenograft model with TNP-470," *Medical and Pediatric Oncology*, vol. 35, no. 6, pp. 673–676, 2000.
- [29] E. Wassberg, S. Påhlman, J.-E. Westlin, and R. Christofferson, "The angiogenesis inhibitor TNP-470 reduces the growth rate of human neuroblastoma in nude rats," *Pediatric Research*, vol. 41, no. 3, pp. 327–333, 1997.

- [30] E. Nagabuchi, W. E. VanderKolk, Y. Une, and M. M. Ziegler, "TNP-470 antiangiogenic therapy for advanced murine neuroblastoma," *Journal of Pediatric Surgery*, vol. 32, no. 2, pp. 287–293, 1997.
- [31] H. N. Lode, T. Moehler, R. Xiang, et al., "Synergy between an antiangiogenic integrin α_v antagonist and an antibodycytokine fusion protein eradicates spontaneous tumor metastases," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 96, no. 4, pp. 1591–1596, 1999.
- [32] G. Klement, S. Baruchel, J. Rak, et al., "Continous lowdose therapy with vinblastine and VEGF receptor-2 antibody induces sustained tumor regression without over toxicity," *The Journal of Clinical Investigation*, vol. 105, no. 8, pp. 15–24, 2000.
- [33] A. Erdreich-Epstein, H. Shimada, S. Groshen, et al., "Integrins $\alpha_{\nu}\beta_{3}$ and $\alpha_{\nu}\beta_{5}$ are expressed by endothelium of high-risk neuroblastoma and their inhibition is associated with increased endogenous ceramide," *Cancer Research*, vol. 60, no. 3, pp. 712–721, 2000.
- [34] A. E. Evans, K. D. Kisselbach, D. J. Yamashiro, et al., "Antitumor activity of CEP-751 (KT-6587) on human neuroblastoma and medulloblastoma xenografts," *Clinical Cancer Research*, vol. 5, no. 11, pp. 3594–3602, 1999.
- [35] A. E. Evans, K. D. Kisselbach, X. Liu, et al., "Effect of CEP-751 (KT-6587) on neuroblastoma xenografts expressing TrkB," *Medical and Pediatric Oncology*, vol. 36, no. 1, pp. 181–184, 2001.
- [36] R. Ho, J. E. Minturn, T. Hishiki, et al., "Proliferation of human neuroblastomas mediated by the epidermal growth factor receptor," *Cancer Research*, vol. 65, no. 21, pp. 9868– 9875, 2005.
- [37] K. Beppu, J. Jaboine, M. S. Merchant, C. L. Mackall, and C. J. Thiele, "Effect of imatinib mesylate on neuroblastoma tumorigenesis and vascular endothelial growth factor expression," *Journal of the National Cancer Institute*, vol. 96, no. 1, pp. 46– 55, 2004.
- [38] R. Vitali, V. Cesi, M. R. Nicotra, et al., "c-Kit is preferentially expressed in *MYCN*-amplified neuroblastoma and its effect on cell proliferation is inhibited in vitro by STI-571," *International Journal of Cancer*, vol. 106, no. 2, pp. 147–152, 2003.
- [39] G. Melino, M. Draoui, L. Bellincampi, et al., "Retinoic acid receptors α and γ mediate the induction of "tissue" transglutaminase activity and apoptosis in human neuroblastoma cells," *Experimental Cell Research*, vol. 235, no. 1, pp. 55–61, 1997.
- [40] A. Voigt and F. Zintl, "Effects of retinoic acid on proliferation, apoptosis, cytotoxicity, migration, and invasion of neuroblastoma cells," *Medical and Pediatric Oncology*, vol. 40, no. 4, pp. 205–213, 2003.
- [41] C. P. Reynolds, K. K. Matthay, J. G. Villablanca, and B. J. Maurer, "Retinoid therapy of high-risk neuroblastoma," *Cancer Letters*, vol. 197, no. 1-2, pp. 185–192, 2003.
- [42] A. Garaventa, R. Luksch, M. S. Lo Piccolo, et al., "Phase I trial and pharmacokinetics of fenretinide in children with neuroblastoma," *Clinical Cancer Research*, vol. 9, no. 6, pp. 2032–2039, 2003.
- [43] B. Desvergne and W. Wahli, "Peroxisome proliferatoractivated receptors: nuclear control of metabolism," *Endocrine Reviews*, vol. 20, no. 5, pp. 649–688, 1999.
- [44] M. J. Reginato, S. L. Krakow, S. T. Bailey, and M. A. Lazar, "Prostaglandins promote and block adipogenesis through opposing effects on peroxisome proliferator-activated receptor *y*," *Journal of Biological Chemistry*, vol. 23, no. 273, pp. 1855– 1858, 1998.

- [45] C. Juge-Aubry, A. Pernin, T. Favez, et al., "DNA binding properties of peroxisome proliferator-activated receptor subtypes on various natural peroxisome proliferator response elements: importance of the 5'-flanking region," *Journal of Biological Chemistry*, vol. 272, no. 40, pp. 25252–25259, 1997.
- [46] C. Grommes, G. E. Landreth, and M. T. Heneka, "Antineoplastic effects of peroxisome proliferator-activated receptor y agonists," *Lancet Oncology*, vol. 5, no. 7, pp. 419–429, 2004.
- [47] P. Tontonoz, S. Singer, B. M. Forman, et al., "Terminal differentiation of human liposarcoma cells induced by ligands for peroxisome proliferator-activated receptor *y* and the retinoid X receptor," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 1, pp. 237– 241, 1997.
- [48] M. Konopleva, E. Elstner, T. J. McQueen, et al., "Peroxisome proliferator-activated receptor y and retinoid X receptor ligands are potent inducers of differentiation and apoptosis in leukemias," *Molecular Cancer Therapeutics*, vol. 3, no. 10, pp. 1249–1262, 2004.
- [49] K. Yamazaki, M. Shimizu, M. Okuno, et al., "Synergistic effects of RXRα and PPARy ligands to inhibit growth in human colon cancer cells-phosphorylated RXRα is a critical target for colon cancer management," *Gut*, vol. 56, no. 11, pp. 1557–1563, 2007.
- [50] I. Avis, A. Martínez, J. Tauler, et al., "Inhibitors of the arachidonic acid pathway and peroxisome proliferator-activated receptor ligands have superadditive effects on lung cancer growth inhibition," *Cancer Research*, vol. 65, no. 10, pp. 4181– 4190, 2005.
- [51] D. L. Crowe and R. A. S. Chandraratna, "A retinoid X receptor (RXR)-selective retinoid reveals that RXR- α is potentially a therapeutic target in breast cancer cell lines, and that it potentiates antiproliferative and apoptotic responsive to peroxisome proliferator-activated receptor ligands," *Breast Cancer Research*, vol. 6, no. 5, pp. R546–R555, 2004.
- [52] G. C. Burdge, H. Rodway, J. A. Kohler, and K. A. Lillycrop, "Effect of fatty acid supplementation on growth and differentiation of human IMR-32 neuroblastoma cells in vitro," *Journal of Cellular Biochemistry*, vol. 80, no. 2, pp. 266–273, 2000.
- [53] S. Di Loreto, B. D'Angelo, M. A. D'Amico, et al., "PPARβ agonists trigger neuronal differentiation in the human neuroblastoma cell line SH-SY5Y," *Journal of Cellular Physiology*, vol. 211, no. 3, pp. 837–847, 2007.
- [54] S. W. Han, M. E. Greene, J. Pitts, R. K. Wada, and N. Sidell, "Novel expression and function of peroxisome proliferatoractivated receptor *y* (PPAR*y*) in human neuroblastoma cells," *Clinical Cancer Research*, vol. 7, no. 1, pp. 98–104, 2001.
- [55] Y. Sato, H. Sasaki, Y. Kobayashi, et al., "Expression of PPARgamma is correlated with the clinical course of neuroblastoma," *Journal of Pediatric Surgery*, vol. 38, no. 2, pp. 205–210, 2003.
- [56] A. Nakagawara, "Molecular basis of spontaneous regression of neuroblastoma: role of neurotrophic signals and genetic abnormalities," *Human Cell*, vol. 11, no. 3, pp. 115–124, 1998.
- [57] K. Yamamoto, R. Hanada, A. Kikuchi, et al., "Spontaneous regression of localized neuroblastoma detected by mass screening," *Journal of Clinical Oncology*, vol. 16, no. 4, pp. 1265–1269, 1998.
- [58] A. Yoneda, T. Oue, K. Imura, et al., "Observation of untreated patients with neuroblastoma dected by mass screening: a "wait and see" pilot study," *Medical and Pediatric Oncology*, vol. 36, no. 1, pp. 160–162, 2001.

- [59] M. Fukushima, "Biological activities and mechanisms of action of PGJ₂ and related compounds: an update," *Prostaglandins, Leukotrienes and Essential Fatty Acids*, vol. 47, no. 1, pp. 1–12, 1992.
- [60] T. T. Rohn, S. M. Wong, C. W. Cotman, and D. H. Cribbs, "15-deoxy-Δ^{12,14}-prostaglandin J₂, a specific ligand for peroxisome proliferator-activated receptor-*y*, induces neuronal apoptosis," *NeuroReport*, vol. 12, no. 4, pp. 839–843, 2001.
- [61] E. J. Kim, K. S. Park, S. Y. Chung, et al., "Peroxisome proliferator-activated receptor-γ activator 15-deoxy-Δ^{12,14}prostaglandin J₂ inhibits neuroblastoma cell growth through induction of apoptosis: association with extracellular signalregulated kinase signal pathway," *Journal of Pharmacology and Experimental Therapeutics*, vol. 307, no. 2, pp. 505–517, 2003.
- [62] H. A. Rodway, A. N. Hunt, J. A. Kohler, A. D. Postle, and K. A. Lillycrop, "Lysophosphatidic acid attenuates the cytotoxic effects and degree of peroxisome proliferator-activated receptor *γ* activation induced by 15-deoxyΔ^{12,14}-prostaglandin J₂ in neuroblastoma cells," *Biochemical Journal*, vol. 382, part 1, pp. 83–91, 2004.
- [63] T. Eichholtz, K. Jalink, I. Fahrenfort, and W. H. Moolenaar, "The bioactive phospholipid lysophosphatidic acid is released from activated platelets," *Biochemical Journal*, vol. 291, part 3, pp. 677–680, 1993.
- [64] Y. Yatomi, Y. Igarashi, L. Yang, et al., "Sphingosine 1phosphate, a bioactive sphingolipid abundantly stored in platelets, is a normal constituent of human plasma and serum," *Journal of Biochemistry*, vol. 121, no. 5, pp. 969–973, 1997.
- [65] K. M. Jung, K. S. Park, J. H. Oh, et al., "Activation of p38 mitogen-activated protein kinase and activator protein-1 during the promotion of neurite extension of PC-12 cells by 15-deoxy-Δ^{12,14}-prostaglandin J₂," *Molecular Pharmacology*, vol. 63, no. 3, pp. 607–616, 2003.
- [66] S. Han, R. K. Wada, and N. Sidell, "Differentiation of human neuroblastoma by phenylacetate is mediated by peroxisome proliferator-activated receptor *y*," *Cancer Research*, vol. 61, no. 10, pp. 3998–4002, 2001.
- [67] U. Valentiner, M. Carlsson, R. Erttmann, H. Hildebrandt, and U. Schumacher, "Ligands for the peroxisome proliferatoractivated receptor-y have inhibitory effects on growth of human neuroblastoma cells in vitro," *Toxicology*, vol. 213, no. 1-2, pp. 157–168, 2005.
- [68] V. C. Emmans, H. A. Rodway, A. N. Hunt, and K. A. Lillycrop, "Regulation of cellular processes by PPARy ligands in neuroblastoma cells is modulated by the level of retinoblastoma protein expression," *Biochemical Society Transactions*, vol. 32, part 5, pp. 840–842, 2004.
- [69] M. Kato, T. Nagaya, M. Fujieda, K. Saito, J. Yoshida, and H. Seo, "Expression of PPARy and its ligand-dependent growth inhibition in human brain tumor cell lines," *Japanese Journal of Cancer Research*, vol. 93, no. 6, pp. 660–666, 2002.
- [70] K. Schultze, B. Böck, A. Eckert, et al., "Troglitazone sensitizes tumor cells to TRAIL-induced apoptosis via down-regulation of FLIP and Survivin," *Apoptosis*, vol. 11, no. 9, pp. 1503–1512, 2006.
- [71] T. W. Jung, J. Y. Lee, W. S. Shim, et al., "Rosiglitazone protects human neuroblastoma SH-SY5Y cells against acetaldehydeinduced cytotoxicity," *Biochemical and Biophysical Research Communications*, vol. 340, no. 1, pp. 221–227, 2006.
- [72] T. W. Jung, J. Y. Lee, W. S. Shim, et al., "Rosiglitazone protects human neuroblastoma SH-SY5Y cells against MPP+ induced cytotoxicity via inhibition of mitochondrial dysfunction and

ROS production," *Journal of the Neurological Sciences*, vol. 253, no. 1-2, pp. 53–60, 2007.

- [73] T. Servidei, R. Morosetti, C. Ferlini, et al., "The cellular response to PPARy ligands is related to the phenotype of neuroblastoma cell lines," *Oncology Research*, vol. 14, no. 7-8, pp. 345–354, 2004.
- [74] I. Cellai, S. Benvenuti, P. Luciani, et al., "Antineoplastic effects of rosiglitazone and PPARy transactivation in neuroblastoma cells," *British Journal of Cancer*, vol. 95, no. 7, pp. 879–888, 2006.
- [75] Y. Sugiura, H. Shimada, R. C. Seeger, W. E. Laug, and Y. A. DeClerck, "Matrix metalloproteinases-2 and -9 are expressed in human neuroblastoma: contribution of stromal cells to their production and correlation with metastasis," *Cancer Research*, vol. 58, no. 10, pp. 2209–2216, 1998.
- [76] C. F. Chantrain, H. Shimada, S. Jodele, et al., "Stromal matrix metalloproteinase-9 regulates the vascular architecture in neuroblastoma by promoting pericyte recruitment," *Cancer Research*, vol. 64, no. 5, pp. 1675–1686, 2004.
- [77] S. Ogita, K. Tokiwa, N. Arizono, and T. Takahashi, "Neuroblastoma: incomplete differentiation on the way to maturation or morphological alteration resembling maturity?" *Oncology*, vol. 45, no. 3, pp. 148–152, 1988.
- [78] D. Shao and M. A. Lazar, "Modulating nuclear receptor function: may the phos be with you," *Journal of Clinical Investigation*, vol. 103, no. 12, pp. 1617–1618, 1999.