

CX3CL1/CX3CR1 Axis, as the Therapeutic Potential in Renal Diseases: Friend or Foe?



Quan Zhuang, Ke Cheng^{*} and Yingzi Ming^{*}

Transplantation Center of the 3rd Xiangya Hospital, Central South University, Changsha, Hunan 410013, China

ARTICLEHISTORY

Received: May 05, 2017 Revised: October 06, 2017 Accepted: January 14, 2018

DOI: 10.2174/1566523218666180214092536 **Abstract:** The fractalkine receptor chemokine (C-X3-C motif) receptor 1 (CX3CR1) and its highly selective ligand CX3CL1 mediate chemotaxis and adhesion of immune cells, which are involved in the pathogenesis and progression of numerous inflammatory disorders and malignancies. The CX3CL1/CX3CR1 axis has recently drawn attention as a potential therapeutic target because it is involved in the ontogeny, homeostatic migration, or colonization of renal phagocytes. We performed a Medline/PubMed search to detect recently published studies that explored the relationship between the CX3CL1/CX3CR1 axis and renal diseases and disorders, including diabetic nephropathy, renal allograft rejection, infectious renal diseases, IgA nephropathy, fibrotic kidney disease, lupus nephritis and glomerulonephritis, acute kidney injury and renal carcinoma. Most studies demonstrated its role in promoting renal pathopoiesis. Thus, the CX3CL1/CX3CR1 axis is now considered to be a double-edged sword that could provide novel perspectives into the pathogenesis and treatment of renal diseases and disorders.

Keywords: Fractalkine, CX3CL1, Chemokine receptor, CX3CR1, Renal disease, Kidney transplantation.

1. INTRODUCTION 1.1. CX3CL1/CX3CR1 Axis

Chemokine (C-X3-C motif) Receptor 1 (CX3CR1) is ubiquitously expressed in most tissues on mononuclear and circulatory lymphatic leucocytes [1]. Fractalkine, also known as CX3CL1, is the only ligand of CX3CR1, which is a sole chemokine that not only involves a chemoattractant function but also assists CX3CR1⁺ cells to adhere; thus, the CX3CL1/CX3CR1 axis is a new type of leukocyte-migration controller [2]. CX3CL1 is mainly produced by the endothelium and has two forms (membrane and soluble). Membrane CX3CL1 is an adhesion molecule, but soluble CX3CL1 is a chemoattractant for CX3CR1⁺ cells [3].

Leukocytes can tether and roll on endothelium in canonical pathway, by which leukocytes are exposed to the surface of the local endothelium, which produces chemokines by binding to glycosaminoglycans. When chemokines contact their homologous receptors, signal transduction activates integrins to form firm adhesions between rolling leukocytes and the endothelial surface. Finally, leukocytes change their shape and start diapedesis through the endothelium, arriving in the tissue [4]. It has been clearly demonstrated that CX3CR1 selectively exists on different lineages of cytotoxic leukocytes with high contents of intracellular perforin, granzyme B and death-signaling Fas ligand, which could help leukocytes undergo transendothial migration to infiltrate into inflamed tissues [5].

In humans, CX3CL1 is mainly expressed on the tubular epithelium, especially during inflammation. Concomitantly, CX3CR1-expressing monocytes and T cells are ubiquitously expressed in inflammatory renal tissues in patients [6, 7]. How does the inflammatory response link to the CX3CL1/CX3CR1 axis? When microbes, injuries and nonself-antigens stimulate the kidney, innate immune cells respond via special receptors, pattern recognition receptors (PRRs) [8]. Their ligands include Pathogen-Associated Molecular Patterns (PAMPs) [9], damage- or Danger-Associated Molecular Patterns (DAMPs) [10] and allogeneic non-self-associated patterns [11]. All of the above effects are able to initiate the inflammatory cascade and produce inflammatory cytokines. The CX3CL1/CX3CR1 axis is activated by inflammatory cytokines, such as interferon- γ (INF- γ), interleukin-1 (IL-1), tumor necrosis factor- α (TNF- α), IL-10, and IL-6 [12, 13]. CX3CL1 is mainly produced by glomerular endothelial cells and the tubular epithelium and can be detected in many other cells, such as podocytes, mesangial cells, renal tumor cells and stromal cells [14-16]. CX3CR1 is a G-protein coupled receptor that is expressed on CD8⁺ T lymphocytes [17], mast cells [18], Natural Killer (NK) cells [19], Dendritic Cells (DCs) [20],

^{*}Address correspondence to these authors at Transplantation Center, the 3rd Xiangya Hospital, Central South University, 138 Tongzipo Rd, Changsha 410013, China; Tel: +86-731-88618320; E-mails: chke1972@163.com; myz_china@aliyun.com

CX3CL1/CX3CR1 Axis, as the Therapeutic Potential in Renal Diseases

platelets [21], renal cancer cells [22], vascular smooth cells [23], tubular cells [24] mesenchymal cells [25], and monocytes/macrophages [26, 27]. After CX3CL1 conjugates with CX3CR1, the CX3CL1/CX3CR1 axis can initiate a cascade via several signaling pathways in the kidney, including ROS/MAPKS, Raf/MEK1/2-ERK1/2-Akt/PI3K, and nuclear factor of kappa light polypeptide gene enhancer in B cells 1 (NF-kB). CX3CL1/CX3CR1 axis directly up-regulates mesangial cell expansion via Reactive Oxygen Species (ROS) and Mitogen-Activated Protein Kinase (MAPK) diabetic nephropathy [28]. The CX3CL1/ CX3CR1 axis also activates vascular smooth muscle cell proliferation through Phosphatidylinositol-3 Kinase (PI3K), Akt and NF-kB [29]. According to Yao's observation, CX3CR1 was expressed in human clear cell Renal Cell Carcinoma (RCC) cell lines, and only membrane positive cells were responsible for CX3CL1-induced cell migration. Extracellular signal-Related Kinases (ERK1/2) and PI3K/Akt were triggered by soluble CX3CL1 dependent on time. [22]. These concepts are summarized schematically in Fig. (1).

2. CX3CL1/CX3CR1 AXIS IN NEPHROPATHIES

In many renal diseases, CX3CL1 expression and CX3CR1+ cells are evident in patients. A major observation regarding the role of CX3CL1/CX3CR1 axis was reported in renal disease in the context of ischemic Acute Renal Failure (ARF). Oh and colleagues recently reported the up-regulation of CX3CL1 in the renal endothelium and the protective effect of macrophage depletion during ischemic ARF in mice by blocking CX3CR1 on macrophages *via* specific blocking antibodies, thereby decreasing macrophage infiltration into the ischemic kidney. Similarly, decreased renal fibrosis after Ischemia-Reperfusion Injury (IRI) was achieved through CX3CR1-blocking antibodies [30]. Furthermore,

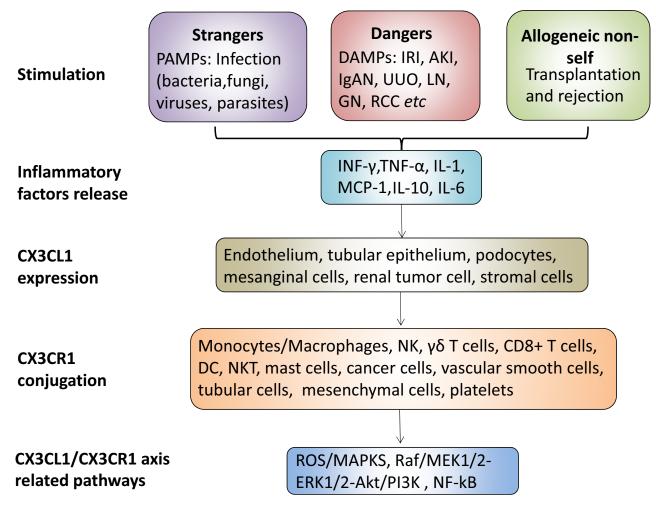


Fig. (1). The general process of CX3CL1/CX3CR1 axis cascade. (1) Stimulation process: stranger pattern (microbes), danger pattern (injuries) and allogeneic non-self pattern stimulate the kidney. (2) Inflammatory factors release process: INF- γ ,TNF- α , IL-1, MCP-1, IL-10, IL-6 are produced by (1). (3) CX3CL1 expression process: fractalkine is produced and increasingly expresses by endothelium, tubular epithelium, podocytes, mesangial cells, renal tumor cell, and stromal cells. (4) CX3CR1 conjugation process: fractalkine would conjugate with CX3CR1+ cells such as monocytes/macrophages, NK, $\gamma\delta$ T cells, CD8⁺ T cells, DC, NKT, mast cells, cancer cells, vascular smooth cells, tubular cells, mesenchymal cells, and platelets. (5) CX3CL1/CX3CR1 axis effect process: CX3CL1/CX3CR1 axis would produce effect *via* ROS/MAPKS, Raf/MEK1/2-ERK1/2-Akt/PI3K, NF-kB pathways.

CX3CL1 is known to play a critical role in glomerulonephritis by acting as a chemoattractant and adhesion molecule [31]. Indeed, the majority of leukocytes that infiltrate the kidney during glomerulonephritis or other nephropathies were shown to express CX3CR1. Moreover, Inoue et al. designed a potent CX3CL1 antagonist that delayed onset and progression of lupus nephritis in MRL/lpr mice, once again underlining the major therapeutic potential associated with CX3CR1 antagonism [32]. In a recent study, Lakkis et al. found the mainstream of recipient DCs which could replace donor DCs in heart and kidney grafts are non-conventional CD11b⁺CD11c⁺ DCs that produce IL-12 and originate from non-classical monocytes, which highly express CX3CR1 [33]. Nevertheless, Engel et al. revealed that CX3CR1 could reduce kidney fibrosis through preventing retention of profibrotic macrophages locally [27]. Chousterman et al. reported that inflammatory monocytes played a protective role in sepsis based on the CX3CR1 pathway [26]. Thus, the CX3CL1/CX3CR1 axis is considered to be a double-edged sword.

Pondering the relationship between the kidney disease therapy and the CX3CL1/CX3CR1 axis immediately brings to mind anti-inflammation, anti-fibrosis, antirejection and anti-cancer components (Fig. 2). We discuss the role of CX3CL1/CX3CR1 axis in kidney diseases and disorders in detail below to provide novel perspectives into the pathogenesis and treatment of renal diseases and disorders.

2.1. Diabetic Nephropathy

Diabetic nephropathy is the leading cause of end-stage renal disease (ESRD) worldwide [34]. CX3CL1/CX3CR1

axis in diabetic renal disease was firstly assessed by CX3CR1 Knockout (KO) mice twelve weeks after inducing diabetes. The function of the CX3CL1/CX3CR1 pathway was first investigated using a diabetic model, Streptozotocin (STZ)-injected CX3CR1 gene knockout mice. Macrophage aggression and renal Extracellular Matrix (ECM) accumulation were aggravated in diabetic mice [35]. CX3CL1 triggers vascular smooth muscle cell proliferation *via* PI3K/Akt and NF-kB [29] and stimulates the Janus kinase and Stat pathways in severe acute pancreatitis of rat [36]. ROS and MAPK were shown to be the signal molecules that were induced by CX3CL1, and blockade of ROS and MAPK successfully prevented synthesis of the mesangial ECM induced by CX3CL1 [28].

2.2. Allograft Function and Rejection

A small sample-size, half-quantified, retrospective study displayed that the CX3CL1 [15] and CX3CR1 [6] levels were higher in a transplanted kidney with a pathological diagnosis of acute cellular rejection than that with no sign of rejection. Hoffmann's work studied the expression of CX3CR1 in biopsies of 174 renal allografts. The results revealed that only a minor percentage of CX3CR1⁺ cells was detectable in normal kidney, while CX3CR1 was generally expressed in grafts that were pathologically diagnosed as acute tubulointerstitial and vascular allograft rejection. Additionally, in acute tubulointerstitial rejection, CX3CR1⁺ cells infiltrated the interstitial tissues and were mainly located around the peritubular capillaries, and they were also found in the glomeruli in a few patients. In acute vascular rejection, CX3CR1⁺ cells were ubiquitously detectable in the subendothelial regions of the arteries [37]. In fact, only a minor portion of CX3CR1⁺ cells could express CD4 and CD8 in allo-

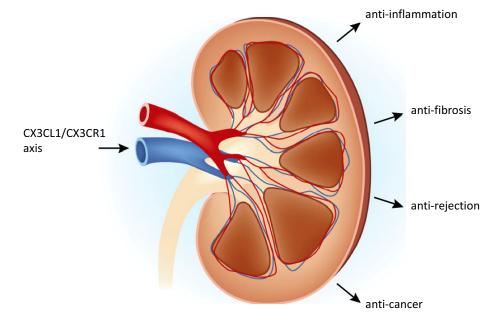


Fig. (2). The relationship between the kidney diseases therapy and the CX3CL1/CX3CR1 axis. Anti-inflammation, anti-fibrosis, anti-rejection and anti-cancer are the four main purposes to figure out CX3CL1/CX3CR1 axis. (The original kidney picture was copied and modi-fied from Yatim KM, Lakkis FG. A brief journey through the immune system. Clin J Am Soc Nephrol 2015; 10(7): 1274-1281.)

geneic renal transplantation tissue. However, the majority of CX3CR1⁺ cells were CD68⁺ mononuclear cells and CD209+ DCs. Because successive macrophages infiltrated both the glomeruli and interstitium of the grafts, which could cause fibrosis, fibrosis is also included in the pathogenesis of chronic kidney transplant rejection [38].

A recent work by our team showed that host DCs played a pivotal role in kidney allograft rejection, and most of the host DCs originated from non-classical monocytes (CX3CR1^{high}), which revealed that CX3CR1 might play a significant role in the formation of monocyte-derived DCs (mono-DCs) in kidney transplants [33]. In our paper, these CX3CR1^{high} mono-DCs formed stable, cognate interactions with effector T cells in the graft, as visualized by a multiphoton intravital microscope. We are presently investigating whether this cell-cell interaction can be attenuated by CX3CR1 blockade or gene knockout (unpublished data). Zhang et al. found that CX3CR1 and CX3CL1 were good predictors for diagnosing Acute Rejection (AR) in allogeneic renal transplant patients. They tested the CX3CL1 and CX3CR1 serum levels at day -1 and every other day (5 times in total) post-kidney transplantation. In the AR group with a pathological diagnosis, the serum levels of CX3CL1 and CX3CR1 were notably higher than those in the non-AR group. Moreover, in the AR group, high expression of serum CX3CL1 and CX3CR1 was positively associated with the onset time of AR. Moreover, in patients with increased CX3CL1 and CX3CR1 serum levels, 3 patients had pathological diagnoses of early AR but no sign of increased serum creatinine and proteinuria, which indicated that the serum CX3CL1 and CX3CR1 levels were more sensitive than increased serum creatinine at diagnosing early AR [39].

Delayed Graft Function (DGF), defined as a need for dialysis during the first 7 days after transplantation, is a common post-renal transplantation complication [40]. Dabrowska-Zamojcin *et al.* reported a relationship between the CX3CR1 gene V249I (rs3732379) Single-Nucleotide Polymorphism (SNP) and allograft function in the kidney. In their paper, DGF was diagnosed in 39.16% of individuals with the CC genotype, 22.73% with the CT genotype and 23.53% with the TT genotype. Therefore, the CC genotype was considered as an independent and substantial predictor of DGF [41].

As described in the studies mentioned above, in chronic renal allograft rejection, the levels of CX3CL1 and CX3CR1 were increased. Because microcapillary inflammation is a significant feature of chronic renal allograft rejection, Park *et al.* tested whether monocytes attached mesangial cells *via* the CX3CL1/CX3CR1 axis in a Lipopolysaccharide (LPS)-stimulated medium *ex vivo*. They found that siRNA against CX3CL1 or CX3CR1 effectively inhibited LPS-induced monocyte-mesangial cell attachment. In mesangial cells stimulated by LPS, the mRNA levels of CX3CL1 and CX3CR1 were improved and CX3CL1 protein synthesis was increased. In summary, it can be concluded that monocytes attach mesangial cells *via* the CX3CL1/CX3CR1 axis; moreover, the CX3CL1/CX3CR1 axis could provide the evidence to the development of inflammation, causing chronic renal allograft rejection [42].

2.3. Infection and Sepsis-induced Renal Diseases

As chemokines play key roles in regulating leukocyte migration, they are widely considered as possible therapeutic targets in sepsis. Raspé and his colleagues found increased levels of CX3CL1/CX3CR1 mRNA in sepsis models induced by Cecum Ligation and Puncture (CLP), and increased levels were also observed in plasma 24 and 48 hours after CLP. Furthermore, CLP-induced down-regulation of CX3CR1 in the kidney was reversed by pre-treatment with the selective NF-kB inhibitor Pyrrolidine Dithiocarbamate (PDTC). These results suggested that expression of the related ligands and receptors induced by CLP led to reversed regulation (up-regulation of CX3CL1 and down-regulation of CX3CR1), which might be regulated by the transcription factor NF-kB, probably through abridged release of inflammatory cytokines [43].

Chousterman et al. showed that inflammatory monocytes had protective effects on renal sepsis through a CX3CR1dependent adhesion mechanism. During sepsis with multiple pathogenic bacteria, inflammatory monocytes migrated from bone marrow, approached the renal cortex endothelial cells and stimulated monocytosis in several hours by increasing CX3CR1-related adhesion. Deficiency of CX3CR1 increased kidney damage and reduced mice mortality, which was related to the migration of monocytes. They also confirmed that the protective function of CX3CR1 was related to the reduction of inflammatory monocyte adhesion and IL-1ra secretion by Ly6C^{high} monocytes. In human diseases, CX3CR1 also took part in the pathogenesis of sepsis, and a study of CX3CR1 gene polymorphism showed the I249 CX3CR1 allele was correlated with more monocytes adhesion and less renal injury [26].

In acute systemic infection, Cytomegalovirus (CMV) spreads throughout the organism. Vascular endothelial cells are the main candidate host cells that promote virus transmission of the vascular pathway. CMV-infected endothelial cells are also believed to play a critical role in the inflammatory response in acute infection [44]. This is why CX3CL1⁺ endothelial cells are the critical site of CMV latency and reoccurrence. At the same time, latent CMV infected patients with ESRD is related to the expansion of circulation, late differentiation, and cytotoxicity of CD4+ T cells.

These effects are characterized by the lack of the costimulator marker CD28 from the cell surface (CD4+CD28^{null}) [45]. In Shabir's study, CD4⁺CD28^{null} T cells were mainly located in the patients with CMV infection and were amplified after transplantation. Therefore, improv-

ing CX3CR1⁺ CD4+CD27-CD28^{null} cells was of biological significance to CMV cellular immunity [46].

2.4. IgA Nephropathy

IgA nephropathy (IgAN) is the most common form of primary glomerulonephritis throughout the world and has a large impact on 20%-50% of patients with glomerulonephritis. Between 30%-40% of patients with IgAN develop ESRD in 2 decades [47]. Consequently, IgAN is considered as a foremost cause of ESRD in many countries. Kim and his colleagues recently found that mice treated with anti-APRIL (a proliferation-inducing ligand) antibodies decreased evolution of IgAN, serum IgA levels, and glomerular IgA accumulation, and also decreased inflammation mediated by CX3CL1/CX3CR1-related activation of monocytes [48].

Tonsillectomy has been reported to be one of the applicable therapies for IgAN patients [49]. In IgAN patients, an over-immune response of Tonsillar Mononuclear Cells (TMCs) to microbial DNA causes IFN- γ -mediated upregulation and enhanced production of B-cell-Activation Factor (BAFF), resulting in boosted production of IgA [50]. Otaka *et al.* recently presented an interesting study on the correlation between IgAN and tonsils. They analyzed the expression of CX3CR1 in tonsillar mononuclear cells of IgAN patients. Immunohistochemical analysis concluded that the distribution of CX3CR1⁺ cells in the inter-follicular region of the tonsillar area in IgAN patients was superior to that of non-IgAN patients.

Flow cytometry revealed that expression of CX3CR1⁺ CD8⁺ T cells in the tonsil was considerably increased in IgAN patients. Chemotaxis of CX3CL1 in the tonsillar mononuclear cells of IgAN patients was meaningfully increased. CX3CR1 had a highly significant effect on the expression of CD8⁺ T cells in IgAN patients. The toxicity of T cells after tonsillectomy was obviously decreased, and the hematuria was also relieved [17].

Cox *et al.* unveiled that high level of CX3CR1 on circulatory T cell subsets, for example, $\gamma\delta$ T cells, which adjusted IgA production in mucosa. CX3CR1 was also found to be up-regulated on NK cells, NKT cells, CD8+ T cells. *Ex vivo*, in Peripheral Blood Mononuclear Cells (PBMCs) of IgAN patients, but not from those with membranous or membranoproliferative glomerulonephritis, LPS-induced CX3CR1 expression was increased, implying an endogenous tendency of IgAN PBMCs expressing CX3CR1 [51, 52].

2.5. Fibrotic Kidney Disease

Some studies have shown that the CX3CL1/CX3CR1 axis might play a role in tubulointerstitial fibrosis, without considering the primary diseases. Koziolek *et al.* found that expression of CX3CR1 was higher in the kidneys of pa-

tients with renal fibrosis compared with those of patients with non-fibrotic, non-inflammatory nephropathies [53]. Peng and his colleagues showed that the CX3CL1-CX3CR1 pair acted as a destructive pathway in obstructed renal fibrosis, which appeared to operate by promoting Ly6C-CX3CR1^{high} macrophage accumulation in the kidney [54]. CX3CL1-CX3CR1 pair promoted Ly6C-CX3CR1^{high} macrophage survival and maintenance in the obstructed kidney. Nevertheless, these results also showed that the CX3CL1-CX3CR1 pair was able to replace monocyte trafficking or differentiation in the obstructed kidneys. Remarkably, a study by Engel et al. based on their previous study verified that CX3CR1 deficiency could attenuate renal fibrosis. However, through the DC-independent mice fibrotic model of Unilateral Ureteral Obstruction (UUO), renal fibrosis was unpredictably severer.

Macrophages were more plentiful in the nonappearance of CX3CR1 and formed more profibrotic mediators in the kidney *via* flow cytometry, such as Transforming Growth Factor- β (TGF- β) [27].

Chronic Kidney Disease (CKD) could accelerate the progress of organ impairment [55]. At stage 5 of CKD, basal activation of leukocytes has been associated with microinflammation and atherosclerosis. Uremic retention solutes and kidney-replacing treatment result in many indispensable alterations of leukocyte function, such as in ROS, apoptosis, chemotaxis and cytokine secretion, validating that leukocytes play a crucial role in vasculopathy of CKD [56]. CX3CR1⁺ CD16⁺ cells therefore favorably enter the endothelium and regions of inflammation by locally expressed CX3CL1, which has been shown to be up-regulated in chronic inflammatory situations [57]. Zaza *et al.* showed that the transcriptome profile of circulatory mononuclear cells (involving CX3CR1+ cells) was differentiated in patients of renal replacement therapy [58].

In the inflammatory response of the kidneys, CX3CL1 secreted from proximal tubular cells played a pivotal role in leukocyte enrollment and retaining to the interstitial tissue [59]. Additionally, these polymorphisms are also investigated in glucometabolic-, cardiovascular- and obesity-associated diseases [60, 61]. Shah *et al.* confirmed that CX3CR1 deficiency had a modest preventive impact on insulin resistance [62]. The II genotype and I allele frequencies of the CX3CR1 V249I polymorphism were discovered notably more frequently in Chronic Renal Failure (CRF), CRF with diabetes mellitus and atherosclerosis than control groups [63].

2.6. Lupus Nephritis and Glomerulonephritis

Systemic Lupus Erythematosus Erythema (SLE) is an autoimmune disease that mostly affects women at young ages and can affect a variety of organs and systems. If the kidney is involved, it is called Lupus Nephritis (LN), which is a serious complication of SLE, and is one of the highest causes of morbidity and mortality in SLE patients [64]. In

LN, CX3CL1 is mostly expressed on the glomerular endothelium, but mesangial cells and the interstitial microvasculature can also secrete it. CX3CR1 is expressed on macrophages and is essential for their migration [65].

In an inducible lupus model in which SCID mice are injected with hybridomas derived from MRL/lpr mice [66], diseased kidneys display up-regulated CX3CL1, which subsequently attracts macrophages. Cros *et al.* reported the occurrence of CD16⁺ monocytes within the glomerular blood vessels of LN patients, which correspond to the high CX3CR1⁺ expressing monocytes [67]. Taken together, this evidence indicates that targeting CX3CL1 signaling would be worthwhile for decreasing monocyte recruitment to the kidney, thus interfering with this pathway in LN pathogenesis.

DCs exist in almost all organs, playing a leading role in maintaining organ homeostasis and inducing the immune response against the invasion of pathogens [68, 69]. Hochheiser et al. determined that CX3CR1 could be defined as the specific "Homing Receptor" of the kidney DCs. DCs were notably abridged in CX3CR1 KO mice kidneys, which was not detected in other organs, excluding the small intestine. The symptoms of Glomerulonephritis (GN) were improved in the CX3CR1 KO mouse model due to the CX3CR1-dependent reduction of DCs, especially in the cortex of kidney. Conversely, immune defenses had no significant effect on the most frequent kidney infection, bacterial pyelonephritis, in CX3CR1 KO mice [70]. In summary, CX3CR1 is believe to be a therapeutic target for GN because CX3CR1 selectively affected the DCs in the kidneys and did not make mice more susceptible to bacterial renal infections.

2.7. Kidney Injury

Monocytes exist in two main forms, termed as, classical and non-classical. Evidence has shown that non-classical monocytes (CX3CR1^{high} CCR2- Ly6C-) perform a specialized form of immune homeostasis, as stimulators of acutely inflamed responses and making contribution to tissue remodeling [71, 72]. Finsterbusch et al. observed cell-cell contacts of CX3CR1^{high} monocytes and neutrophils in both noninflammatory and inflammatory glomeruli by multiphoton and confocal intravital microscopy. In the model of induced glomerular inflammation, the network of neutrophils and CX3CR1^{high} monocytes showed that the retention time of these cells in was prolonged in glomerular capillaries, and there was a tendency to produce ROS, which eventually resulted in kidney injury [73, 74]. In this study, inflammatory monocytes during the initial adhesion to the glomerular capillary required LFA-1 and CX3CR1. However, the neutrophils in the glomerular capillary were immediately arrested, which did not required the prior rolling process, but did require the function of P-selectin, a classical adhesion molecule, for leukocyte rolling [75]. Inflammation triggers TNF production of monocytes in the kidney, which can lead to

kidney damage by prolonging the dwell time of neutrophils and activating the release of ROS from neutrophils. TNF can also rapidly produce ROS by stimulating the activity of NAPDH oxidase in neutrophils [76]. Thus, in the acute inflammatory glomeruli, the deficit of CX3CR1 and consequent decrease in monocyte retention were correlated with decreased neutrophil activation.

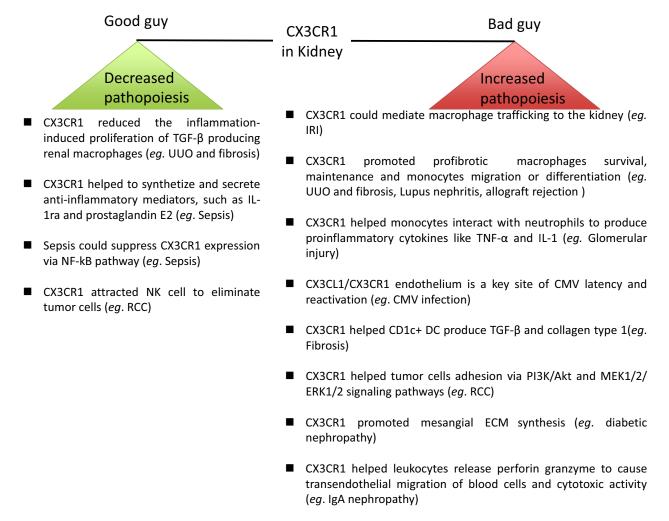
IRI is the main reason of acute and chronic renal dysfunction or failure. In renal IRI, invasive leukocytes and renal parenchyma cells, just like tubular epithelium, can produce chemokines [77]. In Stroo's IRI model, the expression level of CX3CR1 was appreciably improved a week after operation and the invasion of macrophages also reached its highest peak, which indicated that CX3CR1 mediates macrophage trafficking to the kidney [78]. Furuichi et al. [79] revealed elevated expression of CX3CR1 occurred at a later stage of renal IRI and that the absence of CX3CR1 could lead to a decrease in invasion of macrophage peak a week after operation. Nevertheless, Oh et al. [30] determined that early ischemic Acute renal Tubular Necrosis (ATN) was slightly correlated with a CX3CR1-dependent process, similar to serum creatinine. In the early stage of ischemia reperfusion, inhibition of CX3CR1 decreased the ATN score and macrophage infiltration.

2.8. Renal Tumor and Metastasis

In addition to their function in the immune system, chemokines and their receptors also play crucial roles in tumor initiation, progression, and metastasis [80]. Previous studies have found that tumor cells originated from diverse cancer express distinct chemokine receptors, and thus may cause diverse metastatic abilities [81]. Clear Cell RCC (CCRCC), as the main classification of RCC, is an aggressive and refractory cancer [82]. Consequently, it is imperative to identify the key proteins that regulate CCRCC metastasis. Yao et al. demonstrated that CX3CR1 could be expressed on a RCC cell line, and activation of CX3CL1 could contribute to cancer cell migration and activation. ERK1/2 and Akt were phosphorylated after CX3CL1 secretion and participated in CX3CL1-induced cell movement. In addition, high expression of CX3CR1 was associated with poor prognosis of CCRCC [22]. This research provides insights into that CX3CR1 promotes chemotaxis of tumor cells, and CX3CL1/CX3CR1 axis might play a certain role in tumor metastasis in the kidney.

Chemokine receptors contribute to cancer cell metastasis and must meet the following criteria. First, matched chemokines must be expressed on the host site depending on the metastatic profile of the targeting cancer [83]. Therefore, CX3CL1-expressing tissue can be the priority target of CX3CR1 expression in circulating renal cancer cells. Second, chemokines must be able to promote adhesion ability of tumor cells to endothelium and migration to target sites [84]. Membrane-CX3CL1 combined with CX3CR1 is rapid and resolute, which leads to captive and arrested leukocytes even in normal blood flow. Finally, the chemotaxis process requires expression of the tumor cell chemokine receptor [20].

Liu and his colleagues reported higher CX3CL1 expression in serum samples of patients with spinal metastases originated from kidney cancer. For one, it is reasonable to assume that endothelium distributed in bone marrow sinusoids can express membrane-associated CX3CL1. which could supply attachment sites for cancer cells expressing CX3CR1 in circulation. For another, membraneassociated CX3CL1 could be detached into the bone marrow and circulation. Thus, a concentration gradient of chemokines has been established, which could attract CX3CR1+ tumor cells from the peripheral circulation into the spine. However, immunohistochemical staining for expression of CX3CR1 in spinal metastases samples from the kidney showed negative staining [85]. The reason why CX3CR1 is not detectable might be attributed to Epithelial-Mesenchymal Transition (EMT) [86]. In the course of EMT, CX3CR1 is only expressed in mesenchymal cells because of changes in cell properties, and it tends to infiltrate and metastasize. Another conceivable explanation would be that tumor cells without expression or with little expression of CX3CR1 are more probable to spread to sites of metastasis rather than in situ. If neoplastic cells within the primary mass lose any part of the chemokine axis (ligand or receptor), they are less engaged and free to spread and metastasize [64]. Thus, higher CX3CL1 concentrations in bone marrow and plasma are needed to enroll lower CX3CR1-expressing tumor cells. In addition, the antitumor effects of the CX3CL1/CX3CR1 axis are mostly related to the attraction of cytotoxic CD8+ T lymphocytes and NK cells. CX3CR1 is usually expressed in cytotoxic lymphocytes of peripheral blood. These cells release intracellular perforin and granzyme B [5]. Hong et al. found that CX3CL1 secreted into the tumor environment directly recruits CX3CR1-expressing NK cells to tumor tissues, and then, these NK cells could lyse cancer cells. Removal of these NK cells result in the disappearance of the anti-tumor immune response, confirming the function of CX3CL1/ CX3CR1 axis in the NK cellmediated anti-tumor process. They also found that CD8⁺ T cells play an essential role in the CX3CL1-induced anti-



tumor effect as cance specific Cytotoxic T Lymphocytes (CTL). In addition, in tumor tissues, they observed that $CD4^+$ T cells can give rise into T helper 1 cells and enhance the ability of CTL to kill tumor cells through secretion of interferon gamma [87].

A malignant tumor after organ transplantation, such as post-transplantation lymphoproliferative disease (PTLD), is a foremost reason of recipient mortality without allograft dysfunction [88]. There are two ordinary SNPs situated in the coding sequence of the CX3CR1 gene, V249I and T280M, which were correlated with reduced numbers of CX3CL1 attached sites, attenuated cell adhesion, and reduced signaling and chemoattractant ability [89, 90]. In Courivaud's cohort study, they showed that 622 renal transplant recipients and demonstrated that I249M280 homozygotes had an independent increased risk of cancer. Thus, CX3CR1 gene polymorphisms were correlated with a higher rate of tumor occurrence in renal transplant recipients [14].

CONCLUDING REMARKS AND THERAPEUTIC PROSPECTS

CX3CR1 is usually considered to be a negative factor, increasing injury in kidney diseases. However, this view is more controversial now because CX3CR1 can be a positive factor under some circumstances. Regarding the mechanisms of increasing or decreasing renal pathopoiesis, several explanations are summarized as follows (Fig. 3). Increasing pathopoiesis mechanisms: 1) CX3CR1 could mediate macrophage trafficking; 2) CX3CR1 promoted profibrotic macrophage survival and maintenance, as well as monocyte migration or differentiation; 3) CX3CR1 helped monocytes interact with neutrophils to produce proinflammatory cytokines, such as TNF-a and IL-1; 4) the CX3CL1/CX3CR1 endothelium is a key site of CMV latency and reactivation; 5) CX3CR1 helped CD1c⁺ DC produce TGF- β and collagen type 1; 6) CX3CR1 helped tumor cell adhesion via the PI3K/Akt and MEK1/2/ERK1/2 signaling pathways; 7) CX3CR1 promoted mesangial ECM synthesis; and 8) CX3CR1 helped leukocytes release perforin and granzyme to cause transendothelial migration of blood cells and cytotoxic activity. Decreasing pathopoiesis mechanisms: 1) CX3CR1 reduced inflammation-induced proliferation of TGF-B producing renal macrophages; 2) CX3CR1 helped to synthesize and secrete anti-inflammatory mediators, such as IL-1ra and prostaglandin E2; 3) sepsis suppressed CX3CR1 expression via the NF-kB pathway; and 4) CX3CR1 attracted NK cells to eliminate tumor cells.

Although we discussed the dual function of the CX3CL1/CX3CR1 axis in the kidney diseases and disorders described above, many pathologies cannot be involved in one single disease. For instance, IRI and fibrosis could occur in the different stages of the kidney transplantation process. IRI mainly occurs in the stage of innate immunity of transplantation [91]. CX3CR1⁺ mononuclear cells, including DC, macrophages and CD4⁺ T cells, migrate into the kidney following IRI [92]. Human renal allograft chronic rejection is associated with endothelial cell damage, chronic vascular changes, and interstitial inflammation and fibrosis. The CX3CL1/CX3CR1 axis could contribute to the whole pathogenesis of renal allograft chronic rejection [93].

Therefore, we believe that both features should provide valuable opportunities in the future to inhibit renal injury in an accurate and safe manner. Despite potential future obstacles and disappointments, there is an urgent need to devise methods to manipulate the CX3CL1/ CX3CR1 axis due to its major and unique potential as a therapeutic target in inflammation, fibrosis, neoplasms, and transplantation rejection.

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

ACKNOWLEDGEMENTS

This study was supported by the New Xiangya Talent Project of the Third Xiangya Hospital of Central South University (JY201629).

REFERENCES

- Julia V, Staumont-Salle D, Dombrowicz D. Role of fractalkine/CX3CL1 and its receptor CX3CR1 in allergic diseases. Med Sci (Paris) 2016; 32: 260-6.
- [2] Stromberg A, Olsson K, Dijksterhuis JP, Rullman E, Schulte G, Gustafsson T. CX3CL1--a macrophage chemoattractant induced by a single bout of exercise in human skeletal muscle. Am J Physiol Regul Integr Comp Physiol 2016; 310: R297-304.
- [3] Liu W, Jiang L, Bian C, et al. Role of CX3CL1 in diseases. Arch Immunol Ther Exp (Warsz) 2016; 64: 371-83.
- [4] Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. Cell 1994; 76: 301-14.
- [5] Nishimura M, Umehara H, Nakayama T, et al. Dual functions of fractalkine/CX3C ligand 1 in trafficking of perforin+/granzyme B+ cytotoxic effector lymphocytes that are defined by CX3CR1 expression. J Immunol 2002; 168: 6173-80.
- [6] Segerer S, Hughes E, Hudkins KL, Mack M, Goodpaster T, Alpers CE. Expression of the fractalkine receptor (CX3CR1) in human kidney diseases. Kidney international 2002; 62: 488-95.
- [7] Yadav AK, Lal A, Jha V. Association of circulating fractalkine (CX3CL1) and CX3CR1(+)CD4(+) T cells with common carotid artery intima-media thickness in patients with chronic kidney disease. J Atheroscler Thromb 2011; 18: 958-65.
- [8] Oberbarnscheidt MH, Lakkis FG. Innate allorecognition. Immunol Rev 2014; 258: 145-9.
- [9] Beutler BA. TLRs and innate immunity. Blood 2009; 113: 1399-407.

- [10] Kono H, Rock KL. How dying cells alert the immune system to danger. Nat Rev Immunol 2008; 8: 279-89.
- [11] Zhuang Q, Lakkis FG. Dendritic cells and innate immunity in kidney transplantation. Kidney international 2015; 87: 712-8.
- [12] Fraticelli P, Sironi M, Bianchi G, et al. Fractalkine (CX3CL1) as an amplification circuit of polarized Th1 responses. J Clin Invest 2001; 107: 1173-81.
- [13] Wallquist C, Paulson JM, Hylander B, Lundahl J, Jacobson SH. Increased accumulation of CD16+ monocytes at local sites of inflammation in patients with chronic kidney disease. Scand J Immunol 2013; 78: 538-44.
- [14] Courivaud C, Bamoulid J, Loupy A, *et al.* Influence of fractalkine receptor gene polymorphisms V249I-T280M on cancer occurrence after renal transplantation. Transplantation 2013; 95: 728-32.
- [15] Cockwell P, Chakravorty SJ, Girdlestone J, Savage CO. Fractalkine expression in human renal inflammation. J Pathol 2002; 196: 85-90.
- [16] Kim KW, Vallon-Eberhard A, Zigmond E, et al. In vivo structure/function and expression analysis of the CX3C chemokine fractalkine. Blood 2011; 118: e156-67.
- [17] Otaka R, Takahara M, Ueda S, Nagato T, et al. Up-regulation of CX3CR1 on tonsillar CD8-positive cells in patients with IgA nephropathy. Hum Immunol 2017; 78: 375-83.
- [18] Papadopoulos EJ, Fitzhugh DJ, Tkaczyk C, et al. Mast cells migrate, but do not degranulate, in response to fractalkine, a membrane-bound chemokine expressed constitutively in diverse cells of the skin. Eur J Immunol 2000; 30: 2355-61.
- [19] Hamann I, Unterwalder N, Cardona AE, et al. Analyses of phenotypic and functional characteristics of CX3CR1-expressing natural killer cells. Immunology 2011; 133: 62-73.
- [20] Kitching AR. Dendritic cells in progressive renal disease: some answers, many questions. Nephrol Dial Transplant 2014; 29: 2185-93.
- [21] Schafer A, Schulz C, Eigenthaler M, *et al.* Novel role of the membrane-bound chemokine fractalkine in platelet activation and adhesion. Blood 2004; 103: 407-12.
- [22] Yao X, Qi L, Chen X, Du J, Zhang Z, Liu S. Expression of CX3CR1 associates with cellular migration, metastasis, and prognosis in human clear cell renal cell carcinoma. Urol Oncol 2014; 32: 162-70.
- [23] Martynowicz H, Janus A, Nowacki D, Mazur G. The role of chemokines in hypertension. Adv Clin Exp Med 2014; 23: 319-25.
- [24] Furuichi K, Kaneko S, Wada T. Chemokine/chemokine receptormediated inflammation regulates pathologic changes from acute kidney injury to chronic kidney disease. Clin Exp Nephrol 2009; 13: 9-14.
- [25] Ge J, Guo L, Wang S, et al. The size of mesenchymal stem cells is a significant cause of vascular obstructions and stroke. Stem Cell Rev 2014; 10: 295-303.
- [26] Chousterman BG, Boissonnas A, Poupel L, et al. Ly6Chigh Monocytes Protect against Kidney Damage during Sepsis via a CX3CR1-Dependent Adhesion Mechanism. J Am Soc Nephrol 2016; 27: 792-803.
- [27] Engel DR, Krause TA, Snelgrove SL, et al. CX3CR1 reduces kidney fibrosis by inhibiting local proliferation of profibrotic macrophages. J Immunol 2015; 194: 1628-38.
- [28] Park J, Song KH, Ha H. Fractalkine increases mesangial cell proliferation through reactive oxygen species and mitogen-activated protein kinases. Transplant Proc 2012; 44: 1026-8.
- [29] Chandrasekar B, Mummidi S, Perla RP, et al. Fractalkine (CX3CL1) stimulated by nuclear factor kappaB (NF-kappaB)dependent inflammatory signals induces aortic smooth muscle cell proliferation through an autocrine pathway. Biochem J 2003; 373: 547-58.

- [30] Oh DJ, Dursun B, He Z, et al. Fractalkine receptor (CX3CR1) inhibition is protective against ischemic acute renal failure in mice. Am J Physiol Renal Physiol 2008; 294: F264-71.
- [31] Ito Y, Kawachi H, Morioka Y, *et al.* Fractalkine expression and the recruitment of CX3CR1+ cells in the prolonged mesangial proliferative glomerulonephritis. Kidney international 2002; 61: 2044-57.
- [32] Inoue A, Hasegawa H, Kohno M, et al. Antagonist of fractalkine (CX3CL1) delays the initiation and ameliorates the progression of lupus nephritis in MRL/lpr mice. Arthritis Rheum 2005; 52: 1522-33.
- [33] Zhuang Q, Liu Q, Divito SJ, *et al.* Graft-infiltrating host dendritic cells play a key role in organ transplant rejection. Nat Commun 2016; 7: 12623.
- [34] Whiting DR, Guariguata L, Weil C, Shaw J. IDF diabetes atlas: global estimates of the prevalence of diabetes for 2011 and 2030. Diabetes Res Clin Pract 2011; 94: 311-21.
- [35] Song KH, Park J, Park JH, Natarajan R, Ha H. Fractalkine and its receptor mediate extracellular matrix accumulation in diabetic nephropathy in mice. Diabetologia 2013; 56: 1661-9.
- [36] Huang LY, Chen P, Xu LX, Zhou YF, Zhang YP, Yuan YZ. Fractalkine upregulates inflammation through CX3CR1 and the Jak-Stat pathway in severe acute pancreatitis rat model. Inflammation 2012; 35: 1023-30.
- [37] Hoffmann U, Bergler T, Segerer S, et al. Impact of chemokine receptor CX3CR1 in human renal allograft rejection. Transpl Immunol 2010; 23: 204-8.
- [38] Nankivell BJ, Borrows RJ, Fung CL, O'Connell PJ, Allen RD, Chapman JR. The natural history of chronic allograft nephropathy. N Engl J Med 2003; 349: 2326-33.
- [39] Zhang Q, Liu YF, Su ZX, Shi LP, Chen YH. Serum fractalkine and interferon-gamma inducible protein-10 concentrations are early detection markers for acute renal allograft rejection. Transplant Proc 2014; 46: 1420-5.
- [40] Domanski L, Kloda K, Kwiatkowska E, et al. Effect of delayed graft function, acute rejection and chronic allograft dysfunction on kidney allograft telomere length in patients after transplantation: a prospective cohort study. BMC Nephrol 2015; 16: 23.
- [41] Dabrowska-Zamojcin E, Dziedziejko V, Safranow K, Kurzawski M, Domanski L, Pawlik A. Association between the CX3CR1 gene V2491 polymorphism and delayed kidney allograft function. Transpl Immunol 2015; 32: 172-4.
- [42] Park J, Song KH, Ha H. Lipopolysaccharide increases monocyte binding to mesangial cells through fractalkine and its receptor. Transplant Proc 2012; 44: 1029-31.
- [43] Raspe C, Hocherl K, Rath S, Sauvant C, Bucher M. NF-kappaBmediated inverse regulation of fractalkine and CX3CR1 during CLP-induced sepsis. Cytokine 2013; 61: 97-103.
- [44] Adler B, Sinzger C. Endothelial cells in human cytomegalovirus infection: one host cell out of many or a crucial target for virus spread?. Thrombosis and haemostasis 2009; 102: 1057-63.
- [45] Betjes MG, Huisman M, Weimar W, Litjens NH. Expansion of cytolytic CD4+CD28- T cells in end-stage renal disease. Kidney international 2008; 74: 760-7.
- [46] Shabir S, Smith H, Kaul B, et al. Cytomegalovirus-associated CD4(+) CD28(null) Cells in NKG2D-dependent glomerular endothelial injury and kidney allograft dysfunction. Am J Transplant 2016; 16: 1113-28.
- [47] Manno C, Strippoli GFM, D'Altri C, Torres D, Rossini M, Schena FP. A novel simpler histological classification for renal survival in IgA nephropathy: A retrospective study. Am J Kidney Dis 2007; 49: 763-75.
- [48] Kim YG, Alvarez M, Suzuki H, et al. Pathogenic role of a proliferation-inducing ligand (APRIL) in murine IgA nephropathy. Plos One 2015; 10.

- [49] Liu LL, Wang LN, Jiang Y, et al. Tonsillectomy for IgA Nephropathy: A Meta-analysis. Am J Kidney Dis 2015; 65: 80-7.
- [50] Ramos MV, Fernandez GC, Brando RJF, et al. Interleukin-10 and interferon-gamma modulate surface expression of fractalkinereceptor (CX(3)CR1) via PI3K in monocytes. Immunology 2010; 129: 600-9.
- [51] Cox SN, Sallustio F, Serino G, et al. Activated innate immunity and the involvement of CX3CR1-fractalkine in promoting hematuria in patients with IgA nephropathy. Kidney international 2012; 82: 548-60.
- [52] Eitner F, Floege J. In search of a better understanding of IgA nephropathy-associated hematuria. Kidney Intl 2012; 82: 513-5.
- [53] Koziolek MJ, Vasko R, Bramlage C, Muller GA, Strutz F. The CX3C-chemokine fractalkine in kidney diseases. Mini-Rev Med Chem 2009; 9: 1215-28.
- [54] Peng XG, Zhang J, Xiao ZC, Dong YJ, Du J. CX3CL1-CX3CR1 interaction increases the population of Ly6C(-) CX3CR1(hi) macrophages contributing to unilateral ureteral obstruction-induced fibrosis. J Immunol 2015; 195: 2797-805.
- [55] Johansen KL, Lee C. Body composition in chronic kidney disease. Curr Opin Nephrol Hy 2015; 24: 268-75.
- [56] Brunet P, Gondouin B, Duval-Sabatier A, et al. Does uremia cause vascular dysfunction?. Kidney Blood Press R 2011; 34: 284-90.
- [57] Ruth JH, Volin MV, Haines GK, et al. Fractalkine, a novel chemokine in rheumatoid arthritis and in rat adjuvant-induced arthritis. Arthritis and Rheumatism 2001; 44: 1568-81.
- [58] Zaza G, Granata S, Rascio F, *et al.* A specific immune transcriptomic profile discriminates chronic kidney disease patients in predialysis from hemodialyzed patients. Bmc Med Genom 2013; 6: 17.
- [59] Chakravorty SJ, Cockwell P, Girdlestone J, Brooks CJ, Savage COS. Fractalkine expression on human renal tubular epithelial cells: potential role in mononuclear cell adhesion. Clin Exp Immunol 2002; 129: 150-9.
- [60] Sirois-Gagnon D, Chamberland A, Perron S, Brisson D, Gaudet D, Laprise C. Association of common polymorphisms in the fractalkine receptor (cx3cr1) with obesity. Obesity 2011; 19: 222-7.
- [61] Ting KH, Ueng KC, Chiang WL, Chou YE, Yang SF, Wang PH. Relationship of genetic polymorphisms of the chemokine, ccl5, and its receptor, ccr5, with coronary artery disease in taiwan. Evid-Based Compl Alt 2015.
- [62] Shah R, O'Neill SM, Hinkle C, et al. Metabolic effects of cx3cr1 deficiency in diet-induced obese mice. Plos One 2015; 10.
- [63] Bagci B, Bagci G, Huzmeli C, Sezgin I, Ozdemir O. Associations of fractalkine receptor (CX3CR1) and CCR5 gene variants with hypertension, diabetes and atherosclerosis in chronic renal failure patients undergoing hemodialysis. Int Urol Nephrol 2016; 48: 1163-70.
- [64] Mohan C, Putterman C. Genetics and pathogenesis of systemic lupus erythematosus and lupus nephritis. Nat Rev Nephrol 2015; 11: 329-41.
- [65] Yona S, Kim KW, Wolf Y, *et al.* Fate mapping reveals origins and dynamics of monocytes and tissue macrophages under homeostasi. Immunity 2013; 38: 1073-9.
- [66] Nakatani K, Yoshimoto S, Iwano M, et al. Fractalkine expression and CD16(+) monocyte accumulation in glomerular lesions: association with their severity and diversity in lupus models. Am J Physiol-Renal 2010; 299: F207-F16.
- [67] Cros J, Cagnard N, Woollard K, et al. Human CD14(dim) monocytes patrol and sense nucleic acids and Viruses via TLR7 and TLR8 Receptors. Immunity 2010; 33: 375-86.
- [68] Riedel JH, Paust HJ, Turner JE, et al. Immature renal dendritic cells recruit regulatory CXCR6(+) invariant natural killer t cells

to attenuate crescentic GN. J Am Soc Nephrol 2012; 23: 1987-2000.

- [69] Heymann F, Meyer-Schwesinger C, Hamilton-Williams EE, et al. Kidney dendritic cell activation is required for progression of renal disease in a mouse model of glomerular injury (vol 119, pg 1286, 2009). J Clini Invest 2009; 119: 2114-.
- [70] Hochheiser K, Heuser C, Krause TA, et al. Exclusive CX(3)CR1 dependence of kidney DCs impacts glomerulonephritis progression. J Clini Invest 2013; 123: 4242-54.
- [71] Nahrendorf M, Swirski FK, Aikawa E, et al. The healing myocardium sequentially mobilizes two monocyte subsets with divergent and complementary functions. J Exp Med 2007; 204: 3037-47.
- [72] Geissmann F, Jung S, Littman DR. Blood monocytes consist of two principal subsets with distinct migratory properties. Immunity 2003; 19: 71-82.
- [73] Devi S, Li AQ, Westhorpe CLV, *et al.* Multiphoton imaging reveals a new leukocyte recruitment paradigm in the glomerulus. Nat Med 2013; 19: 107-12.
- [74] Finsterbusch M, Hall P, Li AQ, et al. Patrolling monocytes promote intravascular neutrophil activation and glomerular injury in the acutely inflamed glomerulus. P Natl Acad Sci USA 2016; 113: E5172-E81.
- [75] Kuligowski MP, Kitching AR, Hickey MJ. Leukocyte recruitment to the inflamed glomerulus: a critical role for plateletderived P-selectin in the absence of rolling. J Immunol 2006; 176: 6991-9.
- [76] Morgan MJ, Kim YS, Liu ZG. TNFalpha and reactive oxygen species in necrotic cell death. Cell research 2008; 18: 343-9.
- [77] Segerer S, Nelson PJ, Schlondorff D. Chemokines, chemokine receptors, and renal disease: from basic science to pathophysiologic and therapeutic studies. J Am Soc Nephrol 2000; 11: 152-76.
- [78] Stroo I, Stokman G, Teske GJD, et al. Chemokine expression in renal ischemia/reperfusion injury is most profound during the reparative phase. Int Immunol 2010; 22: 433-42.
- [79] Furuichi K, Gao JL, Murphy PM. Chemokine receptor CX3CR1 regulates renal interstitial fibrosis after ischemia-reperfusion injury. Am J Pathol 2006; 169: 372-87.
- [80] Strieter RM, Belperio JA, Phillips RJ, Keane MP. CXC chemokines in angiogenesis of cancer. Seminars in cancer biology 2004; 14: 195-200.
- [81] Raman D, Baugher PJ, Thu YM, Richmond A. Role of chemokines in tumor growth. Cancer Lett 2007; 256: 137-65.
- [82] Jemal A, Siegel R, Xu JQ, Ward E. Cancer Statistics, 2010. Ca-Cancer J Clin 2010; 60: 277-300.
- [83] Ahn SY, Cho CH, Park KG, et al. Tumor necrosis factor-alpha induces fractalkine expression preferentially in arterial endothelial cells and mithramycin A suppresses TNF-alpha-induced fractalkine expression. Am J Pathol 2004; 164: 1663-72.
- [84] Fong AM, Robinson LA, Steeber DA, et al. Fractalkine and CX3CR1 mediate a novel mechanism of leukocyte capture, firm adhesion, and activation under physiologic flow. J Exp Med 1998; 188: 1413-9.
- [85] Liu WM, Bian C, Liang Y, Jiang LB, Qian C, Dong J. CX3CL1: a potential chemokine widely involved in the process spinal metastases. Oncotarget 2017; 8: 15213-9.
- [86] Aparicio LA, Blanco M, Castosa R, et al. Clinical implications of epithelial cell plasticity in cancer progression. Cancer Lett 2015; 366: 1-10.
- [87] Xin H, Kikuchi T, Andarini S, et al. Antitumor immune response by CX3CL1 fractalkine gene transfer depends on both NK and T cells. Eur J Immunol 2005; 35: 1371-80.
- [88] Webster AC, Craig JC, Simpson JM, Jones MP, Chapman JR. Identifying high risk groups and quantifying absolute risk of cancer

after kidney transplantation: A cohort study of 15183 recipients. Am J Transplant 2007; 7: 2140-51.

- [89] Faure S, Meyer L, Costagliola D, et al. Rapid progression to AIDS in HIV+ individuals with a structural variant of the chemokine receptor CX(3)CR1. Science 2000; 287: 2274-7.
- [90] Yu YRA, Fong AM, Combadiere C, Gao JL, Murphy PM, Patel DD. Defective antitumor responses in CX3CR1-deficient mice. Int J Cancer 2007; 121: 316-22.
- [91] Kezic A, Stajic N, Thaiss F. Innate immune response in kidney ischemia/reperfusion injury: potential target for therapy. J Immunol Res 2017; 2017: 6305439.
- [92] Rogers NM, Matthews TJ, Kausman JY, Kitching AR, Coates PT. Review article: Kidney dendritic cells: their role in homeostasis, inflammation and transplantation. Nephrology 2009; 14: 625-35.
- [93] Cao G, Lu Y, Gao R, et al. Expression of fractalkine, CX3CR1, and vascular endothelial growth factor in human chronic renal allograft rejection. Transplant Proc 2006; 38: 1998-2000.