

Development of Nanomaterials to Target Articular Cartilage for Osteoarthritis Therapy

Chenyu Rao and Sirong Shi*

State Key Laboratory of Oral Diseases, West China Hospital of Stomatology, Sichuan University, Chengdu, China

Osteoarthritis (OA) is an obstinate, degradative, and complicated disease that has drawn much attention worldwide. Characterized by its stubborn symptoms and various sequela, OA causes much financial burden on both patients and the health system. What's more, conventional systematic therapy is not effective enough and causes multiple side effects. There's much evidence that nanoparticles have unique properties such as high penetration, biostability, and large specific surface area. Thus, it is urgent to exploit novel medications for OA. Nanomaterials have been sufficiently studied, exploiting diverse nano-drug delivery systems (DDSs) and targeted nano therapeutical molecules. The nanomaterials are primarily intra-articular injected under the advantages of high topical concentration and low dosage. After administration, the DDS and targeted nano therapeutical molecules can specifically react with the components, including cartilage and synovium of a joint in OA, furthermore attenuate the chondrocyte apoptosis, matrix degradation, and macrophage recruitment. Thus, arthritis would be alleviated. The DDSs could load with conventional anti-inflammatory drugs, antibodies, RNA, and so on, targeting chondrocytes, synovium, or extracellular matrix (ECM) and releasing the molecules sequentially. The targeted nano therapeutical molecules could directly get to the targeted tissue, alleviating the inflammation and promoting tissue healing. This review will comprehensively collect and evaluate the targeted nanomaterials to articular cartilage in OA.

OPEN ACCESS

Edited by:

Jianhua Li, Shandong University, China

Reviewed by:

Hongyu Zhang, Tsinghua University, China Lei Sui, Tianjin Medical University, China Jingang Xiao, Southwest Medical University, China

> *Correspondence: Sirong Shi sirongshi@scu.edu.cn

Specialty section:

This article was submitted to Nanobiotechnology, a section of the journal Frontiers in Molecular Biosciences

> Received: 23 March 2022 Accepted: 09 May 2022 Published: 11 August 2022

Citation:

Rao C and Shi S (2022) Development of Nanomaterials to Target Articular Cartilage for Osteoarthritis Therapy. Front. Mol. Biosci. 9:900344. doi: 10.3389/fmolb.2022.900344 Keywords: osteoarthritis, drug delivery system, targeted nanomaterials, articular cartilage, synovium

INTRODUCTION

Epidemiologically, the outbreak of osteoarthritis (OA) shows a prejudice of females in gender and the age more than 65 (Felson et al., 1987; Xia et al., 2014; Szilagyi et al., 2022). Clinical symptoms vary among patients. Knee damage displays principally, followed by hip and hand joints (Ummarino et al., 2020). Evidence reveals that obesity, trauma, and genetic factors are related to the appearance and development of OA (Vina and Kwoh, 2018; Liu et al., 2021). Obesity could not only metabolically impact OA with a higher level of inflammatory factors but also increase the burden of the knee and hip joint, resulting in chronic mechanical damage, which means further progress of OA (Vina and Kwoh, 2018; Ummarino et al., 2020). Joint trauma caused by intense sports, accidents, and surgeons could also assist the OA pathology (Antoni et al., 2021). Congenital deficiency, including joint dysplasia and deformity, shows a susceptible trend towards the disease, attributed to abnormal joint stress and secondary mechanical damage (Vina and Kwoh, 2018).

Conventionally, the administration of OA is classified into two categories, including medication and surgeon (Abramoff and Caldera, 2020; Ummarino et al., 2020). The current clinical drug treatment mainly refers to non-steroidal antiinflammatory drugs and corticosteroids (Abramoff and Caldera, 2020). However, non-steroidal anti-inflammatory drugs have obvious adverse reactions involving the gastrointestinal tract, cardiovascular and cerebrovascular (Pelletier et al., 2016). While long-term use of corticosteroids may cause osteoporosis, hypertension, diabetes, etc., (Wernecke et al., 2015). Furthermore when the above two drugs are taken orally, due to the clearance of the drugs by the liver and plasma, higher doses are required to achieve effective blood drug concentrations, which further increases the possibility of adverse reactions (Wernecke et al., 2015; Pelletier et al., 2016; Abramoff and Caldera, 2020; Ummarino et al., 2020).

To overcome the defects above, we need to figure out a novel drug administration approach based on higher local concentration and lower systemic dosage. Thus, the efficacy would be enhanced while the adverse effects would be alleviated. Nanotechnology has been widely exploited, introducing various nanomaterials with unique properties such as high permeate, specifical and long endurance *in vivo* or *in vitro* (Brown et al., 2019; Zhang T. et al., 2020; Ma et al., 2022). Plenty of evidence shows that nanomaterials could be therapeutic molecules combining with targeted tissue directly and be drug delivery systems (DDSs) transporting the drug molecules specifically (Brown et al., 2019; Ummarino et al., 2020). In addition, previous studies have demonstrated that nanomaterials have multiple bio-functions (Zhang et al., 2021) [anti-tumor (Ma et al., 2022), anti-inflammation (Wang et al., 2022; Zhang et al., 2022), differentiation promotion (Li S et al., 2021), immune modulation (Qin et al., 2022), and neuroprotection (Zhou et al., 2021; Zhu et al., 2022)]

This manuscript comprehensively summarizes the current OA-targeted nanodrugs, classified by their targeted tissues, including chondrocytes, the cartilage extracellular matrix (ECM), synovium matrix, and synovial cells. We also distinguish the nanodrugs into nano-therapeutic molecules that directly react with their target and DDSs that specifically transport the drug molecules embedded in them.

STRUCTURE OF ARTICULAR CARTILAGE AND SYNOVIUM

Chondrocytes

As is known to all, the joint undertakes the burden of sports with a series of physiological bases such as articular cartilage, synovium, and ligaments. Articular cartilage consists of chondrocyte and ECM, being the major portion of degradation caused by OA, meanwhile the important target of the therapeutic molecules.

Generally, chondrocytes participate in osteogenesis via the process of proliferation and terminal hypertrophic alternation. Previous investigation has proven that chondrocytes are derived from mesenchymal stem cells (MSCs) at the embryonic stage, followed by hypertrophic change (Kronenberg, 2003). The hypertrophic chondrocytes are endowed with various functions, including osteogenesis and bone mineralization. The spatial distribution of chondrocytes is characterized by its regular occurrence in cartilage lacuna and elimination at the chondroosseous junction. Several scholars have studied that the postmature chondrocytes are likely to differentiate into osteoblasts and keep the promising potency by differentiating into multiple lineages (Yang et al., 2014).

The exclusive presence of chondrocytes in the ECM makes articular cartilage devoid of vascular vessels and nerves. The fluid nourishes chondrocytes from synovium and subchondral bone (**Figure 1D**) (Armiento et al., 2018; Messina et al., 2019). With the stagnation of the premature stage, chondrocytes intervene in the perpetual balance of articular cartilage vitally (Komori, 2020). Chondrocytes synthesize and secrete collagen, polysaccharides, and their derivatives to construct ECM.

ECM

ECM is the essential part of articular cartilage to accommodate chondrocytes, composed of fibers, proteoglycans, and glycoproteins (Carballo et al., 2017). The spatial distribution of cartilage ECM varies overtly, which means five zones of ECM with diverse components arranged respectively from surface to depth (Brown et al., 2019).

Collagen is the domaining fiber constituting the ECM, classified as collagen II (the predominant one), III, IV, VI, IX, X, and XI (COL-II, III, IV, VI, IX, X, XI), endowing the mechanical properties of cartilage such as tenacity and flexibility (Wilusz et al., 2014; Carballo et al., 2017). Previous studies show that COL-II takes up about 90% volume of fibrils and interacts with COL-IX and XI, and further becomes coarser and cross-linked with the help of leucine-rich proteoglycans and cartilage oligomeric matrix protein (Brown et al., 2019). COL-IV constructs micro-fibrils distributed around chondrocytes, endowing the ECM with the elastic mechanical property. The COL-IV and X don't form macromolecular fibrous structures, mainly located at tangential and calcified zones (van der Kraan et al., 2001; Foldager et al., 2014). Chondrocytes merely take up about 2% volume of the articular cartilage, involving the formation of ECM with the regular releasement of inflammatory and growth factors while maintaining the cartilage homeostasis to keep it healthy (Carballo et al., 2017).

Proteoglycans and glycoproteins are also the main components of cartilage ECM. Glycoproteins constitute proteoglycans via combining glycosaminoglycans (GAGs) on their sidechains no less than three. GAGs are bioactive and comprise chondroitin sulfate (ChS), dermatan sulfate (DS), heparan sulfate (HS), keratan sulfate (KS), and hyaluronic acid (HA) (Brown et al., 2019). Aggrecans have the largest quantity in proteoglycans of ECM, followed by perlecan and laminin (Maroudas, 1976; Brown et al., 2019). Aggrecans can form aggregates as core protein via conjugating with HA molecules, thereby exerting their bio-function in the reticulated structure of collagens (Schneevoigt et al., 2017; Han et al., 2019). What's more, aggrecans can also endow cartilage with the property of elasticity through the associated water molecules (Schulz and Bader, 2007).



FIGURE 1 | Pathology of OA Joint and Spatial Structure of Cartilage. The spatial structure of cartilage is divided into three parts. The collagens in superficial zone are aligned parallel to the tissue surface (A). At the same time, they are randomly orientated in the middle zone (B) and are perpendicular to the interface of cartilage and subchondral bone in deep zone (C). Normal articular cartilage contains chondrocytes and ECM full of collagens, proteoglycans, and glycosaminoglycans (D). In contrast, the OA cartilage is recognized by the apoptosis of chondrocytes and ECM fragments (E). Normal synovium consists of synovial cells and synovial ECM, which is similar to cartilage (F). At the same time, the OA synovium is characterized by infiltration of macrophages, multinucleated giant cells, fibrosis, and angiogenesis (G).

Synovium

The physiological functions of synovium comprising joint cavity formation, fluid dynamic accommodation, and chondrocytes' nutritional transport in an articulation mustn't be ignored. Synovium is also a vital component of a joint (**Figure 1F**) (Mathiessen and Conaghan, 2017). There are two layers constituting normal synovium. The outer one is called subintima, characterized by the vascular and lymphatic vessels within it, while the inner one is identified by its macrophages and fibroblasts (Smith, 2011; Mathiessen and Conaghan, 2017). Taking burdens of immune reactions, macrophages widely exist in the synovium. In other words, macrophages phagocytose and polarize, thus proceed an inflammatory process. As a cyst-like structure formed with synovium, the synovial cavity contains a small amount of synovial fluid abundant with HA and water.

Spatial Structure of Cartilage

The spatial structure of articular cartilage is complicated and orderly, divided into three parts from superficial to deep (**Figures 1A–C**): the superficial zone where the collagen fibrils are aligned parallel to the tissue surface, the middle zone where the collagen fibrils orientation is random, and the deep zone that is adjacent to subchondral bone in which the fibrils are perpendicular to the interface (Carballo et al., 2017). Previous studies have shown that the number of molecules contained in articular cartilage varies from region to region. The middle zone has the largest charge because it contains the most negatively charged proteoglycan. In contrast, the collagen fibers have the highest content in the superficial zone, giving it a higher function of shear resistance (Mow and Guo, 2002; Carballo et al., 2017).

PATHOLOGY OF OSTEOARTHRITIS

OA is an obstinate disease characterized by degradation and various disorders such as joint pain, impaired mobility, and inflammation, with much attention to public health authorities and governments worldwide (Reddi et al., 2011; Bottini et al., 2016; Shah et al., 2020). It is reported that 151 million individuals have been affected by the disease, leading to a fiscal burden of \$185.5 billion annually in the United States alone (Brown et al., 2019). OA has become a socioeconomic problem impacting the residents worldwide due to its increasing morbidity (Hootman and Helmick, 2006). It has been widely explored that the risk factors of OA mostly ascribe to family inheritance, senescence, overweight and joint trauma, whereas the pathogenesis of OA is still largely vague (Xia et al., 2014). Currently, it is widely known that OA is a complicated process in which severe degradation of articular cartilage is the most notable. The whole joint is influenced during OA, impairing the synovium, joint ligaments, and subchondral bone (Abramoff and Caldera, 2020).

ECM Degradation

Degradation of the ECM might appear first during the progression of OA (Goldring and Goldring, 2010; Mobasheri and Batt, 2016) (**Figure 1E**). First, the various metalloproteinases and aggrecanases in ECM will increase abnormally, and increase the catabolism of ECM, and cause the gradual degradation of macromolecular substances such as collagen fibers that maintain the stable structure of ECM, triggering the destruction of structure and function of ECM (Bottini et al., 2016).

Furthermore, ECM is damaged by the inflammatory cytokines (IL-1, IL-6, and TNF), reactive oxygen species (ROS), and peroxynitrite released by various cells at the lesion (Bottini et al., 2016). Matrix metalloproteinases (MMPs) are a family (Winer et al., 2018), relying on Ca^{2+} , Zn^+ which are the cofactors to exert their function of matrix resolution (Wang and Khalil, 2018; Kou et al., 2021). Scholars have classified MMPs into 26 categories, numbered MMP1~26. The substrates of MMP are almost all the protein components of ECM, so scientists divide it into collagenase, gelatinase, matrix-degrading, and so on according to the different substrates (Cui et al., 2017). ROS is a one-electron reduction product of oxygen, including superoxide anion, hydrogen peroxide (H2O2), and hydroxyl radicals (Turrens, 2003; Brieger et al., 2012). ROS are detrimental to the components of ECM via amino acid modification, peptide breakage, protein oxidative allostery, and undermining the structure of macromolecules (Jakubczyk et al., 2020).

Chondrocytes Alteration

The enhanced catabolism of ECM leads to an increase in the content of matrix fragments and chemokines, which in turn causes a series of changes in the phenotype of chondrocytes (Bottini et al., 2016). Chondrocytes have various manifestations of hypertrophy, terminal differentiation, and apoptosis (Mobasheri and Batt, 2016). Generally, they show a tendency to repair the damage.

In the second stage of OA, chondrocytes are stimulated by arthritis (**Figure 1E**). Chondrocytes upregulate the secretion of catabolic enzymes and mediums, enhancing cartilage degradation, and further promoting the apoptosis of chondrocytes themselves. Some evidence shows that the alternation of ECM in OA precedes the changes of osmolality and ionic microenvironment, leading to dynamic adjust of chondrocytes (Jeremiasse et al., 2020).

Synovium Inflammation

OA can cause synovium inflammation (Figure 1G). Some scholars reckon that synovial membranes and fluid are the keys between systematic inflammation and OA. Topical arthritis is mainly brought by synovium and synovial fluid via synthesizing and releasing inflammatory factors (IL-1, IL-6, TNF) and MMPs. Leading to catabolic loop and cartilage damage, synovial fluid can also be a biochemical pathway to unravel the complicated molecular mechanism of OA (Bottini et al., 2016; Ingale et al., 2021). Many of the synovial cells are macrophages, the innate immune cells derived from mononuclear leucocytes (Falconer et al., 2018). Macrophages exert the inflammatory response via the polarization toward pro-inflammatory (M1) and anti-inflammatory (M2) phenotypes (Muñoz et al., 2020). In the context of early-stage arthritis, the macrophages in the synovium are stimulated by inflammatory factors such as LPS, IFN- γ , and TNF- α , thus polarizing towards the M1 phenotype (Cook et al., 2003). The M1 cells would release even more inflammatory factors, including IL-6, IL-12, IL-23, and TNF-a. Not only that, ROS, NO, MMP-1,3,13 secreted by M1 cells can harm articular cartilage directly (Nagai et al., 2003; Haltmayer et al., 2019; Jakubczyk et al., 2020).

The inflammatory and damage factors mentioned above will recruit more leucocytes, including mononuclear cells and macrophages, sequentially promote their proliferation and differentiation towards M1 phenotype macrophages, creating a vicious cycle (Culemann et al., 2019).

THERAPEUTIC ADMINISTRATION

Route of Administration

Drug molecules need to reach the lesion to perform the corresponding pharmacological functions, thereby alleviating symptoms and curing diseases. There are many ways to administer medicine in the medical profession. The common ones are oral, subcutaneous injection, intravenous injection, sublingual administration, etc. As for the drug treatment of OA, the routes can be divided into two major groups, including systematic treatment and topical treatment.

Systematic treatment of OA consists of oral and injection administration with conventional anti-inflammatory medicine such as NSAIDs, capsaicin, weak opioids, and narcotic analgesics. It is more conventional but less effective and targeting (Abramoff and Caldera, 2020).

The topical treatment of OA is intra-articular injection. Compared with systemic administration, intra-articular injection administration has the advantages of small systemic side effects, high local drug concentration, and long drug retention time. Local injection of hormones and antiinflammatory drugs can stabilize cell membranes, reduce inflammatory exudation and relieve symptoms (Altman and Barthel, 2011; Brown et al., 2019). Notwithstanding the universal application of intra-articular injection with the drugs mentioned above, the blemishes such as rapid drug diminishment and apparent off-target effects of the drugs are still unignorable (Brown et al., 2019). In recent years, intra-articular injection of nano-medicine has gradually entered the public eye. Many scholars have tried to use nano-medicine. Many nano-DDSs and nano-medicine have been exploited because of their strong penetration, high retention, and targeting properties.

Conventional Medicine Has Obvious Defects

Destruction of the articulation is the most obvious feature during the OA process, resulting in multiple clinical symptoms, including inflammation, pain, and restricted movement of joints (Abramoff and Caldera, 2020). To relieve symptoms and eliminate the disease, scholars have recommended various treatment methods of which nonpharmacologic, pharmacologic, and surgical therapy (Taruc-Uy and Lynch, 2013). Paracetamol, other non-steroidal anti-inflammatory drugs (NSAIDs), and capsaicin are topically utilized to treat OA, leading to a wide range of adverse events (AEs) such as hepatotoxicity, renal damage, and cardiovascular injury. Meanwhile, a relatively minimal therapeutic effect (Altman and Barthel, 2011; Taruc-Uy and Lynch, 2013). Paracetamol is a universally used analgesic-antipyretic, leading to hepatotoxicity

at a dose of 3.25 g/day, which merely causes little therapeutic effects. NSAIDs have extensive AEs on digestive, cardiovascular, and urinary systems, relying on the dose, duration, and age factors (Altman and Barthel, 2011). Capsaicin can also have AEs such as irritation, nerve degeneration, and occasional coughing (Mason et al., 2004; Gibbons et al., 2010; Altman and Barthel, 2011). Thus, conventional pharmacological treatment emerges with its defective aspect of high risks of heterogeneous side effects and low safe dose. Therefore, we need to find new drugs that can target therapy to increase the local drug concentration while reducing the dose, achieving the goal of increasing efficacy and reducing systemic reactions. Therapeutic nanoparticles (also called nanodrugs) have gradually been recognized by the medical field, with unsurpassed targeting, membrane penetrability, thus could lower the risk brought by high plasma concentration (Brown et al., 2019).

Nanoparticles Have Diverse Unique Properties

The size of particles significantly affects their physical, chemical, and biological properties. The size of drug molecules can even directly affect its pharmacological properties, enhancing its therapeutic effects, such as anti-tumor, anti-angiogenic, antiinflammatory, and proliferation promoting functions. Scholars endow the medicine with a lower dosage and adverse effects rate at the nanoscale. Nanodrugs and nano-DDSs possess specific traits likely to be attributed to their properties of nanoparticles elaborated below.

The particles' size and specific surface area affect their biological properties, including the particles' penetration, distribution, and clearance after entering the body. The smaller the particle size and the larger the specific surface area, the more likely it will cause adverse reactions. Nanoparticles may damage normal tissue by generating free radicals, an effect that intensifies as the size of the nanoparticles decreases (Gatoo et al., 2014). Some studies have also suggested that nanoparticles smaller than 50 nm have strong penetration and can be distributed in almost all tissues, resulting in enhanced side effects (De Jong et al., 2008; Gatoo et al., 2014).

In vitro experiments have shown that nanoparticles with a size of about 100 nm are most easily endocytosed by cells (Xu et al., 2012; Liu et al., 2017). This might be attributed to a better combination and absorption of surface cluster receptors resulting from nanoparticles with a size of 100 nm. Thus the particles trigger invagination of the cell membrane to form vesicles and mediate endocytosis (Liu et al., 2017). In addition, studies have shown that nanoparticles *in vivo* tend to adsorb various molecules on the surface to form protein coronas (PCs), thus the size of the particles interacting with the cell membrane is often larger than the size of the nanoparticles just composited (Walkey et al., 2012; Sun et al., 2013). There is evidence that the optimal nanoparticle size for mediating endocytosis *in vivo* is less than 34 nm (Choi et al., 2010).

Nanoparticles have different morphologies after synthesis, including spherical, rod-shaped, tubular, and tetrahedral



(Gatoo et al., 2014). Different shapes of nanoparticles have different effects on triggering endocytosis (Liu et al., 2017). Previous studies have confirmed that spherical nanoparticles better mediate endocytosis and are less toxic (Champion and Mitragotri, 2006; Lee et al., 2007). At the same time, the higher the aspect ratio and the more elongated morphology of nanoparticles, the greater their potential toxicity, which may be because the longer fibers are not easily cleared by macrophages, causing inflammation in the tissue (Fubini et al., 2011; Gatoo et al., 2014).

Free radicals, metal ions, etc. on the surface of nanoparticles can lead to the generation of ROS, triggering different degrees of toxic reactions, while specific modifications on the surface of nanoparticles (such as hydrophilic polyethylene glycol and other surface-active copolymers) can reduce the side effects and enhance the stability of nanoparticles (Liu et al., 2017).

The charge on the surface of nanoparticles can also affect their function. For example, positively charged nanoparticles have higher clearance rates but also have a better affinity for certain negatively charged biological structures (Gatoo et al., 2014; Liu et al., 2017; He et al., 2020).

The hydrophobicity of the surface of nanoparticles can also affect the efficiency of their cellular uptake. Hydrophobic molecules are more likely to enter the cell membrane and be endocytosed by cells; nanoparticles with a rough surface have a stronger interaction with cells, making it easier to enter cells and mediate endocytosis (Gatoo et al., 2014).

CARTILAGE TARGETED NANODRUGS

Chondrocytes Targeted Nanodrugs

Chondrocytes undertake the burden of the generation of ECM and renewal of its components, attributed to their unique existence in ECM (Figure 1D). Researchers have investigated a variety of nanodrugs to treat OA to ease the symptoms, eliminate the cause of the disease, and improve the quality of daily life (Figure 2).

Various therapeutic molecules could decline the inflammatory reaction and ease the symptoms of OA. Still, they are restricted by their off-target effects and low concentration in joint. Previous studies have proved that nanoparticles could be designed as a medication delivery system (**Table 1**). Cationic polyethylenimine

Name	Туре	Mechanism	Citation
CAP-PEI	Drug delivery system	Utilize the nanoparticles' inherent properties of high penetration, stability and compatibility to deliver therapeutic molecules	Bottini et al. (2016)
PKDi-Nano CL@miR-140 Lipo@TGF-β1 HGdPDW MSC-exosome TFNAs@woqonin		Specifically modified DDSs could obtain targeting	Cho et al. (2019) He K et al. (2021) Velot et al. (2022) Ouyang et al. (2019) Kim et al. (2020) Sirong et al. (2020)
IGF-1	Therapeutic molecule	Promote chondrocytes to synthesize ECM components	(Loffredo et al., 2014; Geiger et al., 2018)
ChS		Enhance chondrocytes proliferation and migration	Hsu et al. (2022)
SeCS		Reduce apoptosis of chondrocytes	Wang et al. (2020)
3'-SL		Inhibit apoptosis, enhance synthesis of chondrocytes	Baek et al. (2021)
RXRα modulator K- 80003		Target chondrocyte nuclear receptor $RXR\alpha$	Li S et al. (2021)
CircSERPINE2		Target miR-1271-5p and ERG	Shen et al. (2019)
Circ_0020093		Inhibit miR-23b	Feng et al. (2021)
CircPDE4B		Regulate p-38/MAPK signaling pathway	Shen et al. (2021)
CYTOR		The knock of CYTOR could reverse anti-OA drug effects, indicating CYTOR to be a promising target drug	Wang et al. (2021)

TABLE 1	Chondrocyte	targeted	nanomaterials.
---------	-------------	----------	----------------

(PEI) is a vital approach to delivering drugs to the lesion site. Scholars modified a targeting ligand (Chondrocyte-homing peptide, CAP) on the PEI to reduce the toxicity and dose meanwhile assemble the pharmaceutical molecule (Bottini et al., 2016). Lipid-based carriers are also recognized as a conventional nano-delivery tool to load hydrophobic therapeutic molecules, while polymeric nanoparticles can encapsulate multiple medicines targeting chondrocytes (de Silva et al., 1979). For instance, in 2019, Cho, H. et al. successfully loaded protein kinase D inhibitor (PKDi) onto nanosomes, creating PKDi-Nano. PKDi can specifically ease the inflammatory reaction primarily caused by PKD, while the clinical application is restricted by its off-target effects and low cellular internalization. PKDi-Nano can significantly decline the defects above and reduce the inflammation damage via the nuclear factor kappa-light-chain enhancer of the activated B cells (NF-κB) pathway. The lipid-based nanosome endows PKDi with stability, leading to a relatively long-time NF-KB pathway activation. Thus, the restoration of chondrocytes would be enhanced (Cho et al., 2019). Some scholars synthesized cationic liposomes via film dispersion and loaded microRNA-140, demonstrating that CL@miR-140 could successfully lead to chondrocyte remedy. After the nanomaterial gets to the chondrocytes, the DDS could release and transport the miR-140 into the cells. Thus, the mRNA could specifically upregulate the COL2A1 mRNA, modulating COL-II synthesis (He K et al., 2021). In 2022, Velot É et al. synthesized agro-based rapeseed liposomes carried transforming growth factor (TGF)-β1 (Lipo@TGF-β1) (Velot et al., 2022). The liposomes encapsulated the TGF-\beta1, endowing it with biostability, a long half-life, and good penetration in cartilage. Then the TGF-B1 would bind with chondrocytes receptors to mediate several signaling pathways (ERK/p-38 MAPK/Smad), which retains the chondrocytes' articular phenotype. Other nanoparticles can also be used to target chondrocytes. In 2019,

Ouyang, Z. et al. (Ouyang et al., 2019) successfully synthesized Gd₂(CO₃)₃ core-based nanoparticles, then anchored a cartilagetargeting peptide and loaded hesperetin (Hes) into the nanoparticles, forming a chondrocyte-targeted drug delivery system called Hes-Gd₂(CO₃)₃@PDA-PEG-DWpeptide (HGdPDW). The scholars demonstrated that HGdPDW could specifically inhibit chondrocyte TLR-2 to alleviate the degeneration via TLR-2/NF-κB/Akt signaling pathway. Exosomes have been exploited for OA therapy as well. In 2017, Cosenza, S. et al. successfully synthesized mesenchymal stem cells derived exosomes and confirmed the protective function of OA cartilage (Cosenza et al., 2017). In 2020, Kim, Y. et al. (Kim et al., 2020) demonstrated that exosomes from mesenchymal stem cells (MSC-exosomes) could be used as a targeting drug delivery system for chondrocytes. The MSCexosomes could specifically protect chondrocytes via multiple methods, such as downregulating the inflammatory factors secretion, declining the expression of prostaglandin E2 (PGE2), or reducing the binding affinity of transcription factor c-jun activating protein-1 (AP-1) and NF-κB. In 2020, Sirong, S. et al. (Sirong et al., 2020) verified that the tetrahedral framework nucleic acids loaded with wogonin (TFNAs@wogonin) could restrain the inflammation with chondrocytes apoptosis attenuation as well as chondrogenic marker expression enhancement.

Several scholars have also shown that nanoparticles can be used directly as therapeutic molecules or as carriers and effectors simultaneously (**Table 1**). The previous investigation revealed that insulin-like growth factor 1 (IGF-1) could upregulate the synthesis of ChS-rich aggregating proteoglycans even at the relatively high concentration of inflammatory factors such as IL-1 and TNF- α , indicating that IGF-1 has the potential to ease arthritis (McQuillan et al., 1986; Tyler, 1989). In 2014, the heparin-binding (HpB) domain of human EGF was assembled on IGF-1, creating HpB-IGF-1 nanomaterial by Loffredo et al. (Loffredo et al., 2014) The HpB-IGF-1 obtained much more stability than single IGF-1. Geiger, BC et al. (Geiger et al., 2018) also modified the IGF-1 with PAMAM dendrimer, endowing the nanoparticles with higher levels of residence time. In 2014 Jain A et al. (Jain et al., 2014) composited diacerein-loaded liposomes and then spliced them onto ChS, thus creating a nanoparticle with anti-inflammation and targeting functions. Further, in 2022, Hsc, HC. et al. (Hsu et al., 2022) demonstrated that ChS could enhance chondrocytes proliferation and migration via inhibiting AKT/ NF-κB pathway and inducing β-catenin. Taken together, ChS is a promising chondrocyte-targeted nanodrug. Wang, L. et al. (Wang et al., 2020) cultured Kashin-Beck disease (KBD) chondrocytes with nano-Se (SeCS) and consequently found that SeCS could reduce the apoptosis of KBD chondrocytes with the upregulation of carbohydrate sulfotransferase 12 and 15 (CHST-12, 15) uronyl 2-O-sulfotransferase (UST) on protein and mRNA level. The scholars firstly demonstrated that the concentration of Se in RA, OA, and KBD is significantly lower than that in control, indicating that the deficiency of Se may be a cause of RA, OA, and KBD. They treated KBD chondrocytes with SeCS and showed that the viability and ultrastructure were improved. The Western blot and q-PCR showed that CHST-12, 15 and UST were obviously upregulated on both protein and mRNA levels. It shows SeCS may recue the KBD/OA chondrocytes via modulating the expression of CHST-12, 15 and UST in them. D'Atri, D. et al. (D'Atri et al., 2021) proved that NGs (nanoghosts) could be both a multifunctional DDS and a targeting medication towards inflammatory chondrocytes in OA via a proof-of-concept experiment. A recent research studied by Baek, A. et al. (Baek et al., 2021) has shown that 3'-Sialyllactose (3'-SL) could reduce arthritis induced by IL-1β via various pathways, including reducing the level of ROS, inhibiting apoptosis of chondrocytes genetically, promoting chondrocytes synthesizing and secreting matrix components. It has been previously studied that a nuclear receptor, Retinoid X receptor a (RXRa), is widely expressed in chondrocytes (Collins-Racie et al., 2009; Ratneswaran et al., 2017). Thus, some scholars demonstrated that RXRa modulator K-80003 could alleviate the degradation of cartilage and synovium inflammation utilizing the property. (Li H et al., 2021).

Recently, the inherent properties of targeting, biocompatibility, and specific bio-functions of nucleic acid are drawing attention in the field. Current studies have proposed using circular RNAs (circRNAs) as targeted drugs that can inhibit the genes associated with the process of OA. Shen, S et al. (Shen et al., 2019) demonstrated that the circRNA (CircSERPINE2) could target the OA chondrocytes via targeting miR-1271-5p and E26 transformation-specific-related genes (ERG). The scholars firstly confirmed that CircSERPINE2 was relatively low in OA cartilage, then they treated the cells with CircSERPINE2. Eventually, they concluded that the drug-treated group downregulated MMPs expression, which was related to the degradation during the OA process. Feng, M. et al. (Feng et al., 2021) found that circ_0020093 could ease the degradation and apoptosis brought by IL-1 β , a kev inflammatory factor. The scholars found that

circ_0020093 and SPRY1 expressions declined in IL-1βinduced OA chondrocytes. Further, they demonstrated that circ 0020093 could upregulate the SPRY1 expression via targeting miR-23b inhibition, which could alleviate the apoptosis induced by IL-1β. In a word, circ_0020093 can target the miR-23b/SPRY1 axis in chondrocytes, leading to the prevention of OA apoptosis. Shen, S. et al. (Shen et al., 2021) concluded that circPDE4B could lower the degradation and upregulate the repairment of articular cartilage via a series of experiments. Like the study mentioned above, circPDE4B was also downregulated in OA chondrocytes. The researchers further demonstrated that circPDE4B could promote RIC8 guaninenucleotide exchange factor A (RIC8A) degradation, both of them participated in OA process. Eventually, the scholars confirmed that circPDE4B/RIC8A could regulate p-38/MAPK signaling pathway in chondrocytes. The overexpression of circPDE4B would inhibit the pathway, resulting in the decline of MMP and enhancement of cell viability. Long non-coding RNA (lncRNA) induced by icariin is also confirmed to promote proliferation and repress the loss of chondrocytes in an inflammatory microenvironment. Cytoskeleton regulator RNA (CYTOR) is a kind of lncRNA, Wang, G. et al. (Wang et al., 2021) demonstrated that the CYTOR knockdown could reverse the anti-OA drug's protective effect in chondrocytes, indicating CYTOR to be a promising chondrocyte-targeted nanodrug for OA.

ECM Targeted Nanodrugs

ECM is secreted by chondrocytes which are the only kind of cells showing presence in articular cartilage and is mainly composed of fibers (collagen), proteoglycans, and glycoproteins (Carballo et al., 2017). Those macromolecules could be the specific ligands for targeting nanodrugs. In this section, we will primarily describe the proteoglycan and collagen-targeted nanodrugs (**Table 2**).

Collagen Targeted Nanodrugs

Collagen is the major fiber in the ECM of cartilage. More precisely, collagen II (COL-II) is the predominant one. Thus, various targeted drugs could bind with COL-II specifically. Previous studies have confirmed that a six amino acid peptide (sequence WYRGRL) has the properties of binding COL-II and conjugating with therapeutic molecules, thus fabricating an ECM-targeted nanodrug. The scholars utilized phage display of peptide library to select appropriate ligands of the ECM. Eventually, they obtained the purpose via the selection of COL-II targeted WYRGRL with five rounds of bioplanning. In addition, they sequentially demonstrated that the peptide could significantly enhance the target effect for OA cartilage (Rothenfluh et al., 2008). Antibodies to COL-II can also enhance the reactivity of cartilage-targeted nanodrugs with ECM. Cho, H. and his colleagues have deeply investigated the area. The team conjugated COL-II antibody with liposomal nanomaterial to diagnose and rehabilitate OA in 2013. They utilized nanosomes to encapsulate the fluorescence and conjugate COL-II antibody to detect the OA at an early stage (Cho et al., 2014; Cho et al., 2015). The above researches performed by Cho,

Name	Туре	Mechanism	Citation
WYRGRL	Drug delivery system	Specifically interact with COL-II	Rothenfluh et al. (2008)
COL-II antibody			(Cho et al., 2014; Cho et al., 2015)
mAV		Target the negative charge of the ECM	He et al. (2020)
RH-SLNs			Ebada et al. (2022)
RAPA@Lipo@HMs			Lei et al. (2022)
PLL		Stimulated by OA environment	Lan et al. (2020)
PPNP			Li H et al. (2021)

H. et al. indicate that the nanomaterials can promisingly deliver therapeutic molecules towards COL-II, thus targeting the OA lesion.

Polysaccharide and Its Derivatives Targeted Nanodrugs

Glycosaminoglycan (GAG) is widely distributed in ECM of cartilage, including hyaluronic acid (HA), chondroitin sulfate (ChS), and so on. Negatively charged glycosaminoglycans are enriched in the cartilage ECM and form cross-linked proteoglycans to form a negatively charged network structure (Shapiro et al., 2002; Palmer et al., 2006). Some previous studies have demonstrated that the negative charge of ECM is a promising targeting lesion for OA treatment. Researchers targeted the negatively charged proteoglycans in ECM by modulating different nanomaterials' zeta potential (Brown et al., 2019).

He, T. et al. successfully confirmed that nano-Avidin (mAv) covalently conjugating drugs can lead to fast penetration and long maintain time in cartilage via reversibly integrating with the aggrecans with a negative charge (He et al., 2020). What's more, Ebada, HM. et al. and Lei, Y. et al. composited two cationic liposomes named rhein hydrophobic ion pairing integrated solid lipid nanoparticles (RH-SLNs) and rapamycin-liposome-incorporating hyaluronic acid-based HMs (RAPA@ Lipo@HMs) as DDSs to target negatively charged ECM of cartilage (Ebada et al., 2022; Lei et al., 2022).

Other Multiple Components Targeted Nanodrugs

The microenvironment of arthritis cartilage is different from the normal. For instance, some enzyme and inflammatory factors may be upregulated, and the pH may be lower. Therefore, the scholars can synthesize new nanoparticles which have a stronger response to the arthritis cartilage. Lan, Q. et al. produced a novel nano-DDS including two specific motifs that target the OA lesion, a conventional anti-inflammatory drug, and a biomaterial scaffold that releases the drug continuously. The scholars utilized poly (2-ethyl-2-oxazoline)-poly (e-caprolactone) (PLL) to endow the nanomaterial with pH-response. Then they conjugated a specific peptide substrate of MMP-13 enzyme to obtain MMP-response. Taken together, the nanomaterial could specifically release its loaded therapeutic molecules in OA lesions (Lan et al., 2020). An ROS-responsive drug release system, also called boronate-stabilized polyphenol-poloxamer (PPNP) assembled dexamethasone (DEX) nanodrug, was exploited by Li, X. et al., in 2021. PPNP was reported to obtain limited ROS-



response. It can deliver drug molecules to OA lesions, then the relatively high level of ROS could break the PPNP delivery system, leading to a drug release. The scholars demonstrated that the nanomaterial was highly sensitive in a 37° C, 50×10^{-6} M H₂O₂ environment (high ROS microenvironment *in vitro*), leading to 85% releasement of DEX (Li X et al., 2021).

SYNOVIUM TARGETED NANODRUGS

Synovium Cells Targeted Nanodrugs

Synovium consists of two layers which are divided into inner and outer ones. The inner one contains synovial cells, while the outer one gathers macrophages, fibroblasts, and capillaries (Mathiessen and Conaghan, 2017). Thus, nanomaterials can target the synovium in arthritis by exploiting the cells mentioned above (**Figure 3**) (**Table 3**).

Plenty of DDSs have been explored to improve the drug treatment effect on synovium, such as polymers, polysaccharides, carbon-nanotubes, micelles, liposomes, and lipids (Brown et al., 2019). Macrophages play an important role in arthritis and are widely distributed in the synovium, mediating phagocytosis and polarization. Thus, macrophages are involved in the occurrence, development, and outcome of synovial inflammation in OA. In 2000, Barrera, P. et al. (Barrera et al., 2000) took advantage of the feature induced by macrophages, compositing nano-liposomal preparations that could be phagocytosed by macrophages and induce apoptosis via the NF- κ B pathway. The researchers found that the injection of the clodronate liposomes results in the depletion of synovial

TABLE 3 | Synovium targeted nanomaterials.

Name	Туре	Mechanism	Citation
Nano-liposomal preparation	Drug delivery system	Phagocytosed by macrophages and induce apoptosis via the NF- κB pathway	Barrera et al. (2000)
Nano-thiolated glycol chitosan packaged siRNA		Influence the polarization of macrophages via Notch 1 pathway	Kim et al. (2015)
miR365 antagomir/NPs-YCWP		Phagocytosed by macrophages, downregulated inflammatory factors and upregulated Nr1D2	Zhang L et al. (2020)
SOD-NPs		Endocytosed by synovial cells	Gui et al. (2022)
IL-Ra modified nanoparticle	Therapeutic	Target synoviocytes via interacting with IL-1 receptors	Whitmire et al. (2012)
Vitronectin	molecule	Interact with FLSs via integrin- $\alpha\beta$	(Zhou et al., 2009; Ciregia et al., 2021)
Rh-PLGA-NPs@NH	Drug delivery	Stimulated by OA environment	Hu et al. (2020)
PAA-MSNs system			He M et al. (2021)

macrophages. Then they discovered that the expression of intercellular adhesion molecule 1 (ICAM-1) and vascular cell adhesion molecule 1 (VCAM-1), two factors in synovitis, were downregulated after the clodronate liposomes treatment, further confirming the anti-inflammation effect via macrophage depletion. In 2015, Kim, MJ. et al. (Kim et al., 2015) used nano-thiolated glycol chitosan to package the siRNA, specifically modulating the Notch 1 pathway. The scholars demonstrated that the LPS-induced macrophages internalized the nanoparticles, the siRNA then specifically inhibited Notch 1 pathway, reducing the level of relevant mRNA (detected by realtime PCR). Thus, the activation of macrophages in the synovium is inhibited. In the same year, Jain, S. et al. (Jain et al., 2015) exploited alginate-decorated IL-10 plasmid nanoparticles which specifically bind macrophages in the synovium via Fc and neuropilin-1 receptors. The alginate-based nanoparticles could target macrophages, and then the plasmid could induce M2 polarization, alleviating arthritis. In 2020, Zhang, L. et al. (Zhang L et al., 2020) exploited a nano-tube delivery system mediated by yeast cell wall particles (YCWP) and loaded with miR365 antagomir, which could be specifically recognized and phagocytosed by macrophages after oral administration. In vitro experiment showed that macrophages successfully engulfed the miR365 antagomir/NPs-YCWP, sequentially downregulated inflammatory factors and upregulated Nr1D2. They also demonstrated that the nanoparticle could alleviate the OA of mice. Taken together, we can confirm that miR365 antagomir/ NPs-YCWP could target synovial macrophages for OA therapy. In 2022, Gui, T. et al. exploited a superoxide dismutase-loaded porous polymersomes (SOD-NPs) that mainly accumulated in synovium tissue and reduced ROS production, further preventing the catabolism. The scholars firstly discovered that SOD-NPs were mainly distributed in synovial tissue in vivo. Then they demonstrated that synovial cells endocytosed SOD-NPs, leading to declining ROS, MMP, and other inflammatory factors. Thus, they confirmed that SOD-NPs could target OA synovial cells (Gui et al., 2022).

Nanoparticles can also directly react with targeting tissue, thus unleashing its treatment function. In 2012, Whitmire, RE. et al. (Whitmire et al., 2012) fabricated novel self-assembled nanoparticles, forming a submicron-scale structure that can bind interleukin-1 receptor antagonist (IL-Ra). The IL-Ra is the natural protein inhibitor of IL-1, thus could be a therapeutic molecule for OA. The scholars demonstrated that IL-Ra could specifically bind with IL-1 receptor on synoviocytes, endowing the nanoparticles with synoviocyte targeting and arthritis easement capacity. Inflamed synovium may show a higher presence of angiogenic endothelial cells, thus improving the level of integrin- $\alpha_v\beta_3$. Early in 2009, Zhou, H. et al. (Zhou et al., 2009) demonstrated that nanoparticles modified with peptidomimetic vitronectin antagonists complementary with integrin- $\alpha_v\beta_3$. While in 2021, Ciregia, F. et al. (Ciregia et al., 2021) confirmed that integrin- α_β was expressed on fibroblast-like synoviocytes (FLSs) as a receptor, allowing its ligand, vitronectin, to bind with itself specifically, thus mediating an arthritis-eliminating process.

Synovial Fluid Targeted Nanodrugs

Synovial fluid is mainly secreted by synovial cells, consists of water, hyaluronic acid (HA), complement, polysaccharides, and cytokines. Multiple targeted nanomaterials have been endowed with the properties of high specificity towards synovial fluid and penetration through synovium layers (**Table 3**).

Synovial fluid contains a lot of HAs, an acidic mucopolysaccharide, lubricating joints and reducing the friction between articular cartilage. A review published by Altman, R et al. (Altman et al., 2019) has already shown that HA could specifically bind with its surface receptors, including CD44, toll-like receptor (TLR), ICAM-1, and layilin (LAYN). Murakami, T et al. (Murakami et al., 2019) previously corroborated that HA potentially reduces the apoptosis of chondrocytes via CD44, indicating CD44 could be an ideal targeted molecule for HA. Ragni, E. et al. (Ragni et al., 2019) demonstrated that CD44, a kind of receptor of HA, has the property of recruitment of extracellular vesicles (EVs) with CD44 markers. The statistics indicated that at the concentration of 2 mg/L of HA, the uptake of the EVs mediating by synoviocytes could be significantly improved. The research further explored the recruitment brought by the interaction between HA and CD44.

Like cartilage ECM, the composition of synovial fluid during OA changes, which leads to a lower pH and higher level of inflammatory factors (IL-1, IL-6, TNF, etc.). Hu, B. et al. (Hu et al., 2020) synthesized lactic-co-glycolic acid (PLGA)

nanoparticles (NPs) loaded with rhein (Rh) and NHHCO (NH) (Rh-PLGA-NPs@NH). The Rh-PLGA-NPs@NH releases more therapeutic molecules in low pH synovial fluid environment which often shows in OA. He, M. et al. (He M et al., 2021) also designed a novel DDS consisting of pH-responsive polyacrylic acid (PAA) and mesoporous silica nanoparticles (MSNs). Therefore, the DDS could be more active in the acidic environment in OA, releasing more therapeutic molecules and improving the retention level of the medicine.

CONCLUSION AND DISCUSSION

The exploitation of targeted nano-medicine for OA has been for several years. In these years, many researchers have synthesized different nanoparticles. These nanoparticles are used to construct DDSs for the targeted conveyance of medication or directly fabricate targeted nanodrugs. The above-mentioned two methods achieve the same goal through different approaches eventually increasing the concentration of the drug in the local lesion, the retention time of the drug, and reducing the side effects and the dosage of the drugs.

For targeted tissues, targeted nano-medicine for chondrocytes and cartilage matrix has been developed in large quantities. In contrast, the corresponding targeted nano-medicine for synovial matrix, synovial fluid, and synovial cells is still in urgent need of development. The main targets of ECM-targeted nanodrugs are

REFERENCES

- Abramoff, B., and Caldera, F. E. (2020). Osteoarthritis: Pathology, Diagnosis, and Treatment Options. *Med. Clin. North Am.* 104, 293–311. doi:10.1016/j.mcna. 2019.10.007
- Altman, R., Bedi, A., Manjoo, A., Niazi, F., Shaw, P., Mease, P. J. C., et al. (2019). Anti-Inflammatory Effects of Intra-articular Hyaluronic Acid: A Systematic Review. *Cartilage* 10, 43–52. doi:10.1177/1947603517749919
- Altman, R. D., and Barthel, H. R. (2011). Topical Therapies for Osteoarthritis. Drugs 71, 1259–1279. doi:10.2165/11592550-000000000-00000
- Antoni, M., Ginot, G., Mereb, T., Clement, X., Eichler, D., Kempf, J., et al. (2021). Post-Traumatic Elbow Osteoarthritis after Radial Head Arthroplasty: Prevalence and Risk Factors. Orthop. Traumatol. Surg. Res. 107, 102814. doi:10.1016/j.otsr.2021.102814
- Armiento, A. R., Stoddart, M. J., Alini, M., and Eglin, D. (2018). Biomaterials for Articular Cartilage Tissue Engineering: Learning from Biology. Acta Biomater. 65, 1–20. doi:10.1016/j.actbio.2017.11.021
- Baek, A., Jung, S., Pyo, S., Kim, S., Jo, S., Kim, L., et al. (2021). 3'-Sialyllactose Protects SW1353 Chondrocytic Cells From Interleukin-1β-Induced Oxidative Stress and Inflammation. *Front. Pharmacol.* 12, 609817. doi:10.3389/fphar. 2021.609817
- Barrera, P., Blom, A., van Lent, P. L., van Bloois, L., Beijnen, J. H., van Rooijen, N., et al. (2000). Synovial Macrophage Depletion with Clodronate-Containing Liposomes in Rheumatoid Arthritis. Arthritis Rheum. 43, 1951–1959. doi:10.1002/1529-0131(200009)43:9<1951::AID-ANR5>3.0. CO;2-K
- Bottini, M., Bhattacharya, K., Fadeel, B., Magrini, A., Bottini, N., Rosato, N., et al. (2016). Nanodrugs to Target Articular Cartilage: An Emerging Platform for Osteoarthritis Therapy. *Nanomedicine* 12, 255–268. doi:10.1016/j.nano.2015. 09.013
- Brieger, K., Schiavone, S., Miller, F. J., JR., and Krause, K. H. (2012). Reactive Oxygen Species: from Health to Disease. *Swiss Med. Wkly.* 142, w13659. doi:10. 4414/smw.2012.13659

collagen fibers and glycosaminoglycans, and collagen fibers are mainly COL-II. However, ECM contains a large amount of other collagen, so the development of these collagen fibers targeted nanodrugs still needs improvement.

When reviewing the literature in related fields, the author found the following problems. ①The synovial matrix and cartilage matrix are similar in composition. Can the targeted nano-drugs targeting the cartilage matrix also target the synovial matrix? ②The pH of synovial fluid and ECM can be lower in OA at the same time. Can the targeting of the drugs be carried out towards both sites?

AUTHOR CONTRIBUTIONS

All authors contributed to the study conception and design. The first draft of the manuscript was written by CR and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

FUNDING

This work was supported by Sichuan University Postdoctoral Interdisciplinary Innovation Fund, the Fundamental Research Funds for the Central Universities, Postdoctoral Science Foundation of China (Grant 2021M692271).

- Brown, S., Kumar, S., and Sharma, B. (2019). Intra-articular Targeting of Nanomaterials for the Treatment of Osteoarthritis. *Acta Biomater*. 93, 239–257. doi:10.1016/j.actbio.2019.03.010
- Carballo, C. B., Nakagawa, Y., Sekiya, I., and Rodeo, S. A. (2017). Basic Science of Articular Cartilage. *Clin. Sports Med.* 36, 413–425. doi:10.1016/j.csm.2017. 02.001
- Champion, J. A., and Mitragotri, S. (2006). Role of Target Geometry in Phagocytosis. Proc. Natl. Acad. Sci. U. S. A. 103, 4930–4934. doi:10.1073/ pnas.0600997103
- Cho, H., Bhatti, F., Hasty, K., and Yi, A. J. I. J. O. N. (2019). Nanosome-Mediated Delivery of Protein Kinase D Inhibitor Protects Chondrocytes From Interleukin-1β-Induced Stress and Apoptotic Death. *Int. J. Nanomedicine* 14, 8835–8846. doi:10.2147/IJN.S218901
- Cho, H., Pinkhassik, E., David, V., Stuart, J. M., and Hasty, K. A. (2015). Detection of Early Cartilage Damage Using Targeted Nanosomes in a Post-traumatic Osteoarthritis Mouse Model. *Nanomedicine* 11, 939–946. doi:10.1016/j.nano. 2015.01.011
- Cho, H., Stuart, J. M., Magid, R., Danila, D. C., Hunsaker, T., Pinkhassik, E., et al. (2014). Theranostic Immunoliposomes for Osteoarthritis. *Nanomedicine* 10, 619–627. doi:10.1016/j.nano.2013.09.004
- Choi, H. S., Ashitate, Y., Lee, J. H., Kim, S. H., Matsui, A., Insin, N., et al. (2010). Rapid Translocation of Nanoparticles from the Lung Airspaces to the Body. *Nat. Biotechnol.* 28, 1300–1303. doi:10.1038/nbt.1696
- Ciregia, F., Deroyer, C., Cobraiville, G., Plener, Z., Malaise, O., Gillet, P., et al. (2021). Modulation of $\alpha_V \beta_6$ Integrin in Osteoarthritis-Related Synovitis and the Interaction with VTN_(381-397 a.a.) Competing for TGF- β 1 Activation.. *Exp. Mol. Med.* 53, 210–222. doi:10.1038/s12276-021-00558-2
- Collins-Racie, L. A., Yang, Z., Arai, M., Li, N., Majumdar, M. K., Nagpal, S., et al. (2009). Global Analysis of Nuclear Receptor Expression and Dysregulation in Human Osteoarthritic Articular Cartilage: Reduced LXR Signaling Contributes to Catabolic Metabolism Typical of Osteoarthritis. Osteoarthr. Cartil. 17, 832–842. doi:10.1016/j.joca.2008.12.011
- Cook, A. D., Braine, E. L., and Hamilton, J. A. (2003). The Phenotype of Inflammatory Macrophages Is Stimulus Dependent: Implications for the

Nature of the Inflammatory Response. J. Immunol. 171, 4816–4823. doi:10. 4049/jimmunol.171.9.4816

- Cosenza, S., Ruiz, M., Toupet, K., Jorgensen, C., and Noël, D. (2017). Mesenchymal Stem Cells Derived Exosomes and Microparticles Protect Cartilage and Bone from Degradation in Osteoarthritis. *Sci. Rep.* 7, 16214. doi:10.1038/s41598-017-15376-8
- Cui, N., Hu, M., and Khalil, R. A. (2017). Biochemical and Biological Attributes of Matrix Metalloproteinases. *Prog. Mol. Biol. Transl. Sci.* 147, 1–73. doi:10.1016/ bs.pmbts.2017.02.005
- Culemann, S., Grüneboom, A., Nicolás-Ávila, J., Weidner, D., Lämmle, K. F., Rothe, T., et al. (2019). Locally Renewing Resident Synovial Macrophages Provide a Protective Barrier for the Joint. *Nature* 572, 670–675. doi:10.1038/ s41586-019-1471-1
- De Jong, W. H., Hagens, W. I., Krystek, P., Burger, M. C., Sips, A. J., Geertsma, R. E., et al. (2008). Particle Size-dependent Organ Distribution of Gold Nanoparticles after Intravenous Administration. *Biomaterials* 29, 1912–1919. doi:10.1016/j.biomaterials.2007.12.037
- D'Atri, D., Zerrillo, L., Garcia, J., Oieni, J., Lupu-Haber, Y., Schomann, T., et al. (2021). Nanoghosts: Mesenchymal Stem Cells Derived Nanoparticles as a Unique Approach for Cartilage Regeneration. *J. Control Release* 337, 472–481. doi:10.1016/j.jconrel.2021.05.015
- de Silva, M., Hazleman, B. L., Thomas, D. P., and Wraight, P. (1979). Liposomes in Arthritis: a New Approach. Lancet 1, 1320–1322. doi:10.1016/s0140-6736(79) 91951-2
- Ebada, H. M., Nasra, M. M., Nasra, R. A., Solaiman, A. A., and Abdallah, O. Y. (2022). Cationic Nanocarrier of Rhein Based on Hydrophobic Ion Pairing Approach as Intra-articular Targeted Regenerative Therapy for Osteoarthritis. *Colloids Surf. B Biointerfaces* 211, 112285. doi:10.1016/j.colsurfb.2021.112285
- Falconer, J., Murphy, A. N., Young, S. P., Clark, A. R., Tiziani, S., Guma, M., et al. (2018). Review: Synovial Cell Metabolism and Chronic Inflammation in Rheumatoid Arthritis. *Arthritis Rheumatol.* 70, 984–999. doi:10.1002/art.40504
- Felson, D. T., Naimark, A., Anderson, J., Kazis, L., Castelli, W., and Meenan, R. F. (1987). The Prevalence of Knee Osteoarthritis in the Elderly. The Framingham Osteoarthritis Study. *Arthritis Rheum.* 30, 914–918. doi:10.1002/art. 1780300811
- Feng, M., Jing, L., Cheng, J., An, S., Huang, J., Yan, Q. J. M., et al. (2021). Circ_0020093 Ameliorates IL-1β-induced Apoptosis and Extracellular Matrix Degradation of Human Chondrocytes by Upregulating SPRY1 via Targeting miR-23b. *Mol. Cell.*. *Biochem.* 476, 3623–3633. doi:10.1007/s11010-021-04186-2
- Foldager, C. B., Toh, W. S., Gomoll, A. H., Olsen, B. R., and Spector, M. (2014). Distribution of Basement Membrane Molecules, Laminin and Collagen Type IV, in Normal and Degenerated Cartilage Tissues. *Cartilage* 5, 123–132. doi:10. 1177/1947603513518217
- Fubini, B., Fenoglio, I., Tomatis, M., and Turci, F. (2011). Effect of Chemical Composition and State of the Surface on the Toxic Response to High Aspect Ratio Nanomaterials. *Nanomedicine (Lond)* 6, 899–920. doi:10.2217/nnm.11.80
- Gatoo, M. A., Naseem, S., Arfat, M. Y., Dar, A. M., Qasim, K., Zubair, S., et al. (2014). Physicochemical Properties of Nanomaterials: Implication in Associated Toxic Manifestations. *Biomed. Res. Int.* 2014, 498420. doi:10. 1155/2014/498420
- Geiger, B., Wang, S., Padera, R., Grodzinsky, A., and Hammond, P. (2018). Cartilage-penetrating Nanocarriers Improve Delivery and Efficacy of Growth Factor Treatment of Osteoarthritis. *Sci. Transl. Med.* 10, eaat8800. doi:10.1126/scitranslmed.aat8800
- Gibbons, C. H., Wang, N., and Freeman, R. (2010). Capsaicin Induces Degeneration of Cutaneous Autonomic Nerve Fibers. Ann. Neurol. 68, 888–898. doi:10.1002/ana.22126
- Goldring, M. B., and Goldring, S. R. (2010). Articular Cartilage and Subchondral Bone in the Pathogenesis of Osteoarthritis. *Ann. N. Y. Acad. Sci.* 1192, 230–237. doi:10.1111/j.1749-6632.2009.05240.x
- Gui, T., Luo, L., Chhay, B., Zhong, L., Wei, Y., Yao, L., et al. (2022). Superoxide Dismutase-Loaded Porous Polymersomes as Highly Efficient Antioxidant Nanoparticles Targeting Synovium for Osteoarthritis Therapy. *Biomaterials* 283, 121437. doi:10.1016/j.biomaterials.2022.121437
- Haltmayer, E., Ribitsch, I., Gabner, S., Rosser, J., Gueltekin, S., Peham, J., et al. (2019). Co-culture of Osteochondral Explants and Synovial Membrane as *In*

Vitro Model for Osteoarthritis. PLoS One 14, e0214709. doi:10.1371/journal. pone.0214709

- Han, B., Li, Q., Wang, C., Patel, P., Adams, S. M., Doyran, B., et al. (2019). Decorin Regulates the Aggrecan Network Integrity and Biomechanical Functions of Cartilage Extracellular Matrix.. ACS Nano 13, 11320–11333. doi:10.1021/ acsnano.9b04477
- He, K., Huang, X., Shan, R., Yang, X., Song, R., Xie, F., et al. (2021). Intra-articular Injection of Lornoxicam and MicroRNA-140 Co-loaded Cationic Liposomes Enhanced the Therapeutic Treatment of Experimental Osteoarthritis. AAPS PharmSciTech 23, 9. doi:10.1208/s12249-021-02149-w
- He, M., Qin, Z., Liang, X., He, X., Zhu, B., Lu, Z., et al. (2021). A pH-Responsive Mesoporous Silica Nanoparticles-Based Drug Delivery System with Controlled Release of Andrographolide for OA Treatment. *Regen. Biomater.* 8, rbab020. doi:10.1093/rb/rbab020
- He, T., Zhang, C., Vedadghavami, A., Mehta, S., Clark, H. A., Porter, R. M., et al. (2020). Multi-arm Avidin Nano-Construct for Intra-cartilage Delivery of Small Molecule Drugs. *J. Control. Release* 318, 109–123. doi:10.1016/j.jconrel.2019. 12.020
- Hootman, J. M., and Helmick, C. G. (2006). Projections of US Prevalence of Arthritis and Associated Activity Limitations. *Arthritis Rheum.* 54, 226–229. doi:10.1002/art.21562
- Hsu, H., Ke, Y., Lai, Y., Hsieh, W., Lin, C., Huang, S., et al. (2022). Chondroitin Sulfate Enhances Proliferation and Migration via Inducing β-Catenin and Intracellular ROS as Well as Suppressing Metalloproteinases through Akt/ NF-κB Pathway Inhibition in Human Chondrocytes. J. Nutr. Health Aging 26, 307–313. doi:10.1007/s12603-022-1752-5
- Hu, B., Gao, F., Li, C., Zhang, B., An, M., Lu, M., et al. (2020). Rhein Laden pH-Responsive Polymeric Nanoparticles for Treatment of Osteoarthritis. *Amb. Express* 10, 158. doi:10.1186/s13568-020-01095-3
- Ingale, D., Kulkarni, P., Electricwala, A., Moghe, A., Kamyab, S., Jagtap, S., et al. (2021). Synovium-Synovial Fluid Axis in Osteoarthritis Pathology: A Key Regulator of the Cartilage Degradation Process. *Genes. (Basel)* 12, 989. doi:10.3390/genes12070989
- Jain, A., Mishra, S. K., Vuddanda, P. R., Singh, S. K., Singh, R., Singh, S., et al. (2014). Targeting of Diacerein Loaded Lipid Nanoparticles to Intra-articular Cartilage Using Chondroitin Sulfate as Homing Carrier for Treatment of Osteoarthritis in Rats. *Nanomedicine* 10, 1031–1040. doi:10.1016/j.nano. 2014.01.008
- Jain, S., Tran, T. H., and Amiji, M. (2015). Macrophage Repolarization with Targeted Alginate Nanoparticles Containing IL-10 Plasmid DNA for the Treatment of Experimental Arthritis. *Biomaterials* 61, 162–177. doi:10.1016/ j.biomaterials.2015.05.028
- Jakubczyk, K., Dec, K., Kałduńska, J., Kawczuga, D., Kochman, J., Janda, K., et al. (2020). Reactive Oxygen Species - Sources, Functions, Oxidative Damage. *Pol. Merkur. Lek.* 48, 124–127.
- Jeremiasse, B., Matta, C., Fellows, C., Boocock, D., Smith, J., Liddell, S., et al. (2020). Alterations in the Chondrocyte Surfaceome in Response to Pro-inflammatory Cytokines. BMC Mol. Cell. Biol. 21, 47. doi:10.1186/s12860-020-00288-9
- Kim, M. J., Park, J. S., Lee, S. J., Jang, J., Park, J. S., Back, S. H., et al. (2015). Notch1 Targeting siRNA Delivery Nanoparticles for Rheumatoid Arthritis Therapy. J. Control. Release 216, 140–148. doi:10.1016/j.jconrel.2015.08.025
- Kim, Y., Choi, J., and Kim, K. J. B. J. (2020). Mesenchymal Stem Cell-Derived Exosomes for Effective Cartilage Tissue Repair and Treatment of Osteoarthritis. *Biotechnol. J.* 15, e2000082. doi:10.1002/biot.202000082
- Komori, T. (2020). Molecular Processes in Chondrocyte Biology. Int. J. Mol. Sci. 21, E4161. doi:10.3390/ijms21114161
- Kou, L., Jiang, X., Lin, X., Huang, H., Wang, J., Yao, Q., et al. (2021). Matrix Metalloproteinase Inspired Therapeutic Strategies for Bone Diseases. *Curr. Pharm. Biotechnol.* 22, 451–467. doi:10.2174/1389201021666200630140735
- Kronenberg, H. M. (2003). Developmental Regulation of the Growth Plate. Nature 423, 332–336. doi:10.1038/nature01657
- Lan, Q., Lu, R., Chen, H., Pang, Y., Xiong, F., Shen, C., et al. (2020). MMP-13 Enzyme and pH Responsive Theranostic Nanoplatform for Osteoarthritis. J. Nanobiotechnology 18, 117. doi:10.1186/s12951-020-00666-7
- Lee, M. K., Lim, S. J., and Kim, C. K. (2007). Preparation, Characterization and In Vitro Cytotoxicity of Paclitaxel-Loaded Sterically Stabilized Solid Lipid Nanoparticles. *Biomaterials* 28, 2137–2146. doi:10.1016/j.biomaterials.2007.01.014

- Lei, Y., Wang, Y., Shen, J., Cai, Z., Zhao, C., Chen, H., et al. (2022). Injectable Hydrogel Microspheres with Self-Renewable Hydration Layers Alleviate Osteoarthritis. Sci. Adv. 8, eabl6449. doi:10.1126/sciadv.abl6449
- Li, H., Li, X., Yang, B., Su, J., Cai, S., Huang, J., et al. (2021). The Retinoid X Receptor α Modulator K-80003 Suppresses Inflammatory and Catabolic Responses in a Rat Model of Osteoarthritis. *Sci. Rep.* 11, 16956. doi:10.1038/ s41598-021-96517-y
- Li, S., Liu, Y., Tian, T., Zhang, T., Lin, S., Zhou, M., et al. (2021). Bioswitchable Delivery of microRNA by Framework Nucleic Acids: Application to Bone Regeneration. Small 17, e2104359. doi:10.1002/smll.202104359
- Li, X., Wang, X., Liu, Q., Yan, J., Pan, D., Wang, L., et al. (2021). ROS-responsive Boronate-Stabilized Polyphenol-Poloxamer 188 Assembled Dexamethasone Nanodrug for Macrophage Repolarization in Osteoarthritis Treatment. Adv. Healthc. Mat. 10, e2100883. doi:10.1002/adhm.202100883
- Liu, K., Cai, F., Liu, Y., Abulaiti, A., Ren, P., Yusufu, A. J. B. M. D., et al. (2021). Risk Factors of Ankle Osteoarthritis in the Treatment of Critical Bone Defects Using Ilizarov Technique. *BMC Musculoskelet. Disord.* 22, 339. doi:10.1186/s12891-021-04214-8
- Liu, Y., Workalemahu, B., and Jiang, X. (2017). The Effects of Physicochemical Properties of Nanomaterials on Their Cellular Uptake In Vitro and In Vivo. Small 13, 1701815. doi:10.1002/smll.201701815
- Loffredo, F. S., Pancoast, J. R., Cai, L., Vannelli, T., Dong, J. Z., Lee, R. T., et al. (2014). Targeted Delivery to Cartilage Is Critical for *In Vivo* Efficacy of Insulinlike Growth Factor 1 in a Rat Model of Osteoarthritis. *Arthritis Rheumatol.* 66, 1247–1255. doi:10.1002/art.38357
- Ma, W., Yang, Y., Zhu, J., Jia, W., Zhang, T., Liu, Z., et al. (2022). Biomimetic Nanoerythrosome-Coated Aptamer-DNA Tetrahedron/Maytansine Conjugates: pH-Responsive and Targeted Cytotoxicity for HER2-Positive Breast Cancer. Adv. Mat., e2109609. doi:10.1002/adma.202109609 [online ahead of print].
- Maroudas, A. I. (1976). Balance between Swelling Pressure and Collagen Tension in Normal and Degenerate Cartilage. *Nature* 260, 808–809. doi:10.1038/ 260808a0
- Mason, L., Moore, R. A., Derry, S., Edwards, J. E., and Mcquay, H. J. (2004). Systematic Review of Topical Capsaicin for the Treatment of Chronic Pain. *Bmj* 328, 991. doi:10.1136/bmj.38042.506748.EE
- Mathiessen, A., and Conaghan, P. G. (2017). Synovitis in Osteoarthritis: Current Understanding with Therapeutic Implications. *Arthritis Res. Ther.* 19, 18. doi:10.1186/s13075-017-1229-9
- McQuillan, D. J., Handley, C. J., Campbell, M. A., Bolis, S., Milway, V. E., Herington, A. C., et al. (1986). Stimulation of Proteoglycan Biosynthesis by Serum and Insulin-like Growth Factor-I in Cultured Bovine Articular Cartilage. *Biochem. J.* 240, 423–430. doi:10.1042/bj2400423
- Messina, O. D., Vidal Wilman, M., and Vidal Neira, L. F. (2019). Nutrition, Osteoarthritis and Cartilage Metabolism. *Aging Clin. Exp. Res.* 31, 807–813. doi:10.1007/s40520-019-01191-w
- Mobasheri, A., and Batt, M. (2016). An Update on the Pathophysiology of Osteoarthritis. Ann. Phys. Rehabil. Med. 59, 333–339. doi:10.1016/j.rehab. 2016.07.004
- Mow, V. C., and Guo, X. E. (2002). Mechano-electrochemical Properties of Articular Cartilage: Their Inhomogeneities and Anisotropies. Annu. Rev. Biomed. Eng. 4, 175–209. doi:10.1146/annurev.bioeng.4.110701.120309
- Muñoz, J., Akhavan, N. S., Mullins, A. P., and Arjmandi, B. H. (2020). Macrophage Polarization and Osteoporosis: A Review. *Nutrients* 12, E2999. doi:10.3390/ nu12102999
- Murakami, T., Otsuki, S., Okamoto, Y., Nakagawa, K., Wakama, H., Okuno, N., et al. (2019). Hyaluronic Acid Promotes Proliferation and Migration of Human Meniscus Cells via a CD44-dependent Mechanism. *Connect. Tissue Res.* 60, 117–127. doi:10.1080/03008207.2018.1465053
- Nagai, H., Kumamoto, H., Fukuda, M., and Takahashi, T. (2003). Inducible Nitric Oxide Synthase and Apoptosis-Related Factors in the Synovial Tissues of Temporomandibular Joints with Internal Derangement and Osteoarthritis. J. Oral Maxillofac. Surg. 61, 801–807. doi:10.1016/s0278-2391(03)00155-1
- Ouyang, Z., Tan, T., Liu, C., Duan, J., Wang, W., Guo, X., et al. (2019). Targeted Delivery of Hesperetin to Cartilage Attenuates Osteoarthritis by Bimodal Imaging with Gd2(CO3)3@PDA Nanoparticles via TLR-2/NF-κB/Akt Signaling.. *Biomaterials* 205, 50–63. doi:10.1016/j.biomaterials.2019.03.018

- Palmer, A. W., Guldberg, R. E., and Levenston, M. E. (2006). Analysis of Cartilage Matrix Fixed Charge Density and Three-Dimensional Morphology via Contrast-Enhanced Microcomputed Tomography. *Proc. Natl. Acad. Sci. U.* S. A. 103, 19255–19260. doi:10.1073/pnas.0606406103
- Pelletier, J. P., Martel-Pelletier, J., Rannou, F., and Cooper, C. (2016). Efficacy and Safety of Oral NSAIDs and Analgesics in the Management of Osteoarthritis: Evidence from Real-Life Setting Trials and Surveys. *Semin. Arthritis Rheum.* 45, S22–S27. doi:10.1016/j.semarthrit.2015.11.009
- Qin, X., Xiao, L., Li, N., Hou, C., Li, W., Li, J., et al. (2022). Tetrahedral Framework Nucleic Acids-Based Delivery of microRNA-155 Inhibits Choroidal Neovascularization by Regulating the Polarization of Macrophages. *Bioact. Mat.* 14, 134–144. doi:10.1016/j.bioactmat.2021.11.031
- Ragni, E., Perucca Orfei, C., de Luca, P., Lugano, G., Viganò, M., Colombini, A., et al. (2019). Interaction with Hyaluronan Matrix and miRNA Cargo as Contributors for *In Vitro* Potential of Mesenchymal Stem Cell-Derived Extracellular Vesicles in a Model of Human Osteoarthritic Synoviocytes. *Stem Cell. Res. Ther.* 10, 109. doi:10.1186/s13287-019-1215-z
- Ratneswaran, A., Sun, M., Dupuis, H., Sawyez, C., Borradaile, N., Beier, F., et al. (2017). Nuclear Receptors Regulate Lipid Metabolism and Oxidative Stress Markers in Chondrocytes. J. Mol. Med. 95, 431–444. doi:10.1007/s00109-016-1501-5
- Reddi, A. H., Becerra, J., and Andrades, J. A. (2011). Nanomaterials and Hydrogel Scaffolds for Articular Cartilage Regeneration. *Tissue Eng. Part B Rev.* 17, 301–305. doi:10.1089/ten.TEB.2011.0141
- Rothenfluh, D. A., Bermudez, H., O'Neil, C. P., and Hubbell, J. A. (2008). Biofunctional Polymer Nanoparticles for Intra-articular Targeting and Retention in Cartilage. *Nat. Mat.* 7, 248–254. doi:10.1038/nmat2116
- Schneevoigt, J., Fabian, C., Leovsky, C., Seeger, J., and Bahramsoltani, M. (2017). In Vitro Expression of the Extracellular Matrix Components Aggrecan, Collagen Types I and II by Articular Cartilage-Derived Chondrocytes. Anat. Histol. Embryol. 46, 43–50. doi:10.1111/ahe.12230
- Schulz, R. M., and Bader, A. (2007). Cartilage Tissue Engineering and Bioreactor Systems for the Cultivation and Stimulation of Chondrocytes. *Eur. Biophys. J.* 36, 539–568. doi:10.1007/s00249-007-0139-1
- Shah, K., Yang, X., Lane, J., Collins, G., Arden, N., Furniss, D., et al. (2020). Risk Factors for the Progression of Finger Interphalangeal Joint Osteoarthritis: a Systematic Review. *Rheumatol. Int.* 40, 1781–1792. doi:10.1007/s00296-020-04687-1
- Shapiro, E. M., Borthakur, A., Gougoutas, A., and Reddy, R. (2002). 23Na MRI Accurately Measures Fixed Charge Density in Articular Cartilage. *Magn. Reson. Med.* 47, 284–291. doi:10.1002/mrm.10054
- Shen, S., Wu, Y., Chen, J., Xie, Z., Huang, K., Wang, G., et al. (2019). CircSERPINE2 Protects against Osteoarthritis by Targeting miR-1271 and ETS-Related Gene. Ann. Rheum. Dis. 78, 826–836. doi:10.1136/ annrheumdis-2018-214786
- Shen, S., Yang, Y., Shen, P., Ma, J., Fang, B., Wang, Q., et al. (2021). circPDE4B Prevents Articular Cartilage Degeneration and Promotes Repair by Acting as a Scaffold for RIC8A and MID1. Ann. Rheum. Dis. 80, 1209–1219. doi:10.1136/ annrheumdis-2021-219969
- Sirong, S., Yang, C., Taoran, T., Songhang, L., Shiyu, L., Yuxin, Z., et al. (2020). Effects of Tetrahedral Framework Nucleic Acid/wogonin Complexes on Osteoarthritis. *Bone Res.* 8, 6. doi:10.1038/s41413-019-0077-4
- Smith, M. D. (2011). The Normal Synovium. Open Rheumatol. J. 5, 100–106. doi:10.2174/1874312901105010100
- Sun, J., Xianyu, Y., Li, M., Liu, W., Zhang, L., Liu, D., et al. (2013). A Microfluidic Origami Chip for Synthesis of Functionalized Polymeric Nanoparticles. *Nanoscale* 5, 5262–5265. doi:10.1039/c3nr01289a
- Szilagyi, I., Waarsing, J., Schiphof, D., van Meurs, J., and Bierma-Zeinstra, S. J. R. (2022). Towards Sex-specific Osteoarthritis Risk Models: Evaluation of Risk Factors for Knee Osteoarthritis in Males and Females. *Rheumatol. Oxf.* 61, 648–657. doi:10.1093/rheumatology/keab378
- Taruc-Uy, R. L., and Lynch, S. A. (2013). Diagnosis and Treatment of Osteoarthritis. Prim. Care 40, 821–836. doi:10.1016/j.pop.2013.08.003
- Turrens, J. F. (2003). Mitochondrial Formation of Reactive Oxygen Species. J. Physiol. 552, 335–344. doi:10.1113/jphysiol.2003.049478
- Tyler, J. A. (1989). Insulin-like Growth Factor 1 Can Decrease Degradation and Promote Synthesis of Proteoglycan in Cartilage Exposed to Cytokines. *Biochem. J.* 260, 543–548. doi:10.1042/bj2600543

- Ummarino, A., Gambaro, F. M., Kon, E., and Torres Andón, F. (2020)., 10. Nanomaterials (Basel), E1562. doi:10.3390/nano10081562Therapeutic Manipulation of Macrophages Using Nanotechnological Approaches for the Treatment of OsteoarthritisNanomaterials
- van der kraan, P. M., Stoop, R., Meijers, T. H., Poole, A. R., and van Den Berg, W. B. (2001). Expression of Type X Collagen in Young and Old C57Bl/6 and Balb/c Mice. Relation with Articular Cartilage Degeneration. Osteoarthr. Cartil. 9, 92–100. doi:10.1053/joca.2000.0364
- Velot, É., Elkhoury, K., Kahn, C., Kempf, H., Linder, M., Arab-Tehrany, E., et al. (2022). Efficient TGF-β1 Delivery to Articular Chondrocytes *In Vitro* Using Agro-Based Liposomes. *Int. J. Mol. Sci.* 23, 2864. doi:10.3390/ijms23052864
- Vina, E. R., and Kwoh, C. K. (2018). Epidemiology of Osteoarthritis: Literature Update. Curr. Opin. Rheumatol. 30, 160–167. doi:10.1097/BOR.000000000000479
- Walkey, C. D., Olsen, J. B., Guo, H., Emili, A., and Chan, W. C. (2012). Nanoparticle Size and Surface Chemistry Determine Serum Protein Adsorption and Macrophage Uptake. J. Am. Chem. Soc. 134, 2139–2147. doi:10.1021/ja2084338
- Wang, G., Zhang, L., Shen, H., Hao, Q., Fu, S., Liu, X. J. B. C. M., et al. (2021). Upregulation of Long Non-coding RNA CYTOR Induced by Icariin Promotes the Viability and Inhibits the Apoptosis of Chondrocytes. *BMC Complement. Med. Ther.* 21, 152. doi:10.1186/s12906-021-03322-1
- Wang, L., Yin, J., Yang, B., Qu, C., Lei, J., Han, J., et al. (2020). Serious Selenium Deficiency in the Serum of Patients with Kashin-Beck Disease and the Effect of Nano-Selenium on Their Chondrocytes. *Biol. Trace Elem. Res.* 194, 96–104. doi:10.1007/s12011-019-01759-7
- Wang, X., and Khalil, R. A. (2018). Matrix Metalloproteinases, Vascular Remodeling, and Vascular Disease. Adv. Pharmacol. 81, 241–330. doi:10. 1016/bs.apha.2017.08.002
- Wang, Y., Li, Y., Gao, S., Yu, X., Chen, Y., Lin, Y., et al. (2022). Tetrahedral Framework Nucleic Acids Can Alleviate Taurocholate-Induced Severe Acute Pancreatitis and its Subsequent Multiorgan Injury in Mice. *Nano Lett.* 22, 1759–1768. doi:10.1021/acs.nanolett.1c05003
- Wernecke, C., Braun, H. J., and Dragoo, J. L. (2015). The Effect of Intra-articular Corticosteroids on Articular Cartilage: A Systematic Review. Orthop. J. Sports Med. 3, 2325967115581163. doi:10.1177/2325967115581163
- Whitmire, R., Wilson, D., Singh, A., Levenston, M., Murthy, N., García, A. J. B., et al. (2012). Self-assembling Nanoparticles for Intra-articular Delivery of Antiinflammatory Proteins. *Biomaterials* 33, 7665–7675. doi:10.1016/j.biomaterials. 2012.06.101
- Wilusz, R. E., Sanchez-Adams, J., and Guilak, F. (2014). The Structure and Function of the Pericellular Matrix of Articular Cartilage. *Matrix Biol.* 39, 25–32. doi:10.1016/j.matbio.2014.08.009
- Winer, A., Adams, S., and Mignatti, P. (2018). Matrix Metalloproteinase Inhibitors in Cancer Therapy: Turning Past Failures into Future Successes. *Mol. Cancer Ther.* 17, 1147–1155. doi:10.1158/1535-7163.MCT-17-0646
- Xia, B., di, C., Zhang, J., Hu, S., Jin, H., Tong, P., et al. (2014). Osteoarthritis Pathogenesis: a Review of Molecular Mechanisms. *Calcif. Tissue Int.* 95, 495–505. doi:10.1007/s00223-014-9917-9
- Xu, A., Yao, M., Xu, G., Ying, J., Ma, W., Li, B., et al. (2012). A Physical Model for the Size-dependent Cellular Uptake of Nanoparticles Modified with

Cationic Surfactants. Int. J. Nanomedicine 7, 3547–3554. doi:10.2147/IJN. S32188

- Yang, G., Zhu, L., Hou, N., Lan, Y., Wu, X. M., Zhou, B., et al. (2014). Osteogenic Fate of Hypertrophic Chondrocytes. *Cell. Res.* 24, 1266–1269. doi:10.1038/cr. 2014.111
- Zhang, L., Peng, H., Zhang, W., Li, Y., Liu, L., Leng, T., et al. (2020). Yeast Cell Wall Particle Mediated Nanotube-RNA Delivery System Loaded with miR365 Antagomir for Post-traumatic Osteoarthritis Therapy via Oral Route. *Theranostics* 10, 8479–8493. doi:10.7150/thno.46761
- Zhang, M., Zhang, X., Tian, T., Zhang, Q., Wen, Y., Zhu, J., et al. (2022). Antiinflammatory Activity of Curcumin-Loaded Tetrahedral Framework Nucleic Acids on Acute Gouty Arthritis. *Bioact. Mat.* 8, 368–380. doi:10.1016/j. bioactmat.2021.06.003
- Zhang, T., Tian, T., and Lin, Y. (2021). Functionalizing Framework Nucleic-Acid-Based Nanostructures for Biomedical Application. Adv. Mat., e2107820. doi:10. 1002/adma.202107820
- Zhang, T., Tian, T., Zhou, R., Li, S., Ma, W., Zhang, Y., et al. (2020). Design, Fabrication and Applications of Tetrahedral DNA Nanostructure-Based Multifunctional Complexes in Drug Delivery and Biomedical Treatment. *Nat. Protoc.* 15 (8), 2728–2757. doi:10.1038/s41596-020-0355-z
- Zhou, H. F., Chan, H. W., Wickline, S. A., Lanza, G. M., and Pham, C. T. (2009). Alphavbeta3-targeted Nanotherapy Suppresses Inflammatory Arthritis in Mice. *Faseb J.* 23, 2978–2985. doi:10.1096/fj.09-129874
- Zhou, M., Zhang, T., Zhang, B., Zhang, X., Gao, S., Zhang, T., et al. (2021). A DNA Nanostructure-Based Neuroprotectant against Neuronal Apoptosis via Inhibiting Toll-like Receptor 2 Signaling Pathway in Acute Ischemic Stroke. ACS Nano 16, 1456–1470. doi:10.1021/acsnano.1c09626
- Zhu, J., Yang, Y., Ma, W., Wang, Y., Chen, L., Xiong, H., et al. (2022). Antiepilepticus Effects of Tetrahedral Framework Nucleic Acid via Inhibition of Gliosis-Induced Downregulation of Glutamine Synthetase and Increased AMPAR Internalization in the Postsynaptic Membrane. *Nano Lett.* 22, 2381–2390. doi:10.1021/acs.nanolett.2c00025

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Rao and Shi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.