

LETTER

Contemporary evolution of an invasive grass in response to elevated atmospheric CO₂ at a Mojave Desert FACE site

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Abstract

Elevated atmospheric CO₂ has been shown to rapidly alter plant physiology and ecosystem productivity, but contemporary evolutionary responses to increased CO₂ have yet to be demonstrated in the field. At a Mojave Desert FACE (free-air CO₂ enrichment) facility, we tested whether an annual grass weed (*Bromus madritensis* ssp. *rubens*) has evolved in response to elevated atmospheric CO₂. Within 7 years, field populations exposed to elevated CO₂ evolved lower rates of leaf stomatal conductance; a physiological adaptation known to conserve water in other desert or water-limited ecosystems. Evolution of lower conductance was accompanied by reduced plasticity in upregulating conductance when CO₂ was more limiting; this reduction in conductance plasticity suggests that genetic assimilation may be ongoing. Reproductive fitness costs associated with this reduction in phenotypic plasticity were demonstrated under ambient levels of CO₂. Our findings suggest that contemporary evolution may facilitate this invasive species' spread in this desert ecosystem.

Keywords

Bromus rubens, contemporary evolution, desert ecosystem, elevated atmospheric CO₂, genetic assimilation, invasive species, norms of reaction, phenotypic plasticity, stomatal conductance.

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INTRODUCTION

Human-induced changes to the global environment are pervasive (IPCC 2013) and predictions of long-term ecological impacts of these changes may depend on whether organisms can evolve in response to rapid changes in climate (Bradshaw & McNeilly 1991; Geber & Griffen 2003; Franks *et al.* 2007). The success of invasive species, another important agent of global change, may be due in part to their ability to adapt rapidly to changing environmental conditions (Sakai *et al.* 2001; Lee 2002). Kingsolver (1996) suggested that demographic characteristics of many invasive species, such as short generation time and large local population size, should facilitate their rapid evolutionary response to global climate change. Rising levels of atmospheric CO₂ represent an important component of climate change and, despite the potential for invasive plants to evolve in response to elevated CO₂, we have little information on the synergistic interaction between these two major components of global change. Numerous studies have documented pronounced ecological and physiological phenotypic responses of plants to increased CO₂ (Ainsworth & Long 2005; Franks *et al.* 2013) and some studies have demonstrated heritable variation within populations in physiological and growth responses to elevated CO₂ (Schmid *et al.* 1996; Thomas & Jasienski 1996; Case *et al.* 1998). Nonetheless, there still appears to be little evidence for con-

temporary evolution in response to elevated atmospheric CO₂ (Leakey & Lau 2012). Even in controlled artificial selection experiments under varying CO₂ levels, evidence for adaptive evolutionary response is variable and somewhat limited (Tousignant & Potvin 1996; Ward *et al.* 2000; Collins & Bell 2004; Wieneke *et al.* 2004; Frenck *et al.* 2013). In particular, there is a lack of information on contemporary evolutionary responses of plant species to elevated CO₂ under realistic field conditions that include complex networks of both abiotic and biotic stressors (Leakey & Lau 2012).

The worldwide network of free-air CO₂ enrichment (FACE) facilities represents replicated, large-scale manipulations of atmospheric CO₂ under natural field conditions. FACE experiments have provided important information on ecological and physiological responses to elevated atmospheric CO₂ (Long *et al.* 2004). Given rising atmospheric CO₂, numerous studies and reviews have indicated that rates of leaf stomatal conductance should decrease, primarily as an adaptive response to conserve water (Tyree & Alexander 1993; Drake *et al.* 1997; Buckley & Schymanski 2014). Plants typically exhibit reduced stomatal conductance in elevated CO₂ conditions because the exaggerated concentration gradient facilitates stomatal uptake of CO₂. Lower stomatal conductance results in reduced water loss from transpiration, and C₃ plants generally exhibit a greater response to enriched CO₂ compared to C₄ plants. This differential response suggests

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that elevated atmospheric CO₂ may contribute to changes in community assemblages, although other environmental variables may also affect any relative advantages between C3 and C4 plants as CO₂ increases (Bowes 1993). The adaptive importance of reduced stomatal conductance under elevated atmospheric CO₂ is further supported by a comprehensive review of physiological results from FACE sites that indicated widespread and consistent phenotypically plastic reductions in stomatal conductance under increased CO₂ (Ainsworth & Long 2005). Despite this strong evidence for the adaptive significance of plastic reductions in conductance, none of the studies reported in this review examined potential evolutionary changes in stomatal conductance in response to elevated CO₂.

Norms of reaction in phenotypic response or the capacity to acclimate to variation in atmospheric CO₂ can also evolve if there is genetic variation in phenotypic plasticity itself (Scheiner 1993). As noted above, a meta-analysis of studies conducted at 12 large-scale FACE sites over 15 years indicated widespread acclimation in several physiological properties (Ainsworth & Long 2005). Although there is some evidence that there may be genetic variance for plasticity in stomatal response (Case *et al.* 1998), it remains unknown whether any genetic-based capacity for phenotypic plasticity in conductance may change under elevated CO₂ levels. A comparison of conductance plasticity between ambient and elevated CO₂ populations at a FACE site facilitates the study of possible canalisation in phenotypic response, whereby genetic assimilation (Waddington 1961) may act to “fix” lower conductance in populations exposed to elevated CO₂. Demonstration of genetic assimilation in the field has been difficult, but it has been argued that the loss of plasticity and the evolution of a canalised phenotype may occur relatively rapidly and thus may be difficult to detect (Pigliucci *et al.* 2006). The experimental manipulations in atmospheric CO₂ available at FACE sites offer a unique opportunity to detect potentially transient genetic assimilation and the evolution of reduced plasticity in stomatal conductance.

To test if invasive plants can evolve very rapidly (i.e. within 7 years) in response to elevated atmospheric CO₂ under natural field conditions, we examined the evolutionary response of an introduced grass species *Bromus madritensis* ssp. *rubens* (L.) Husn. (hereafter, referred to as *Bromus*) to elevated CO₂ treatments at the Nevada Desert FACE Facility (NDFF) (Jordan *et al.* 1999). *Bromus* is an invasive annual C3 grass that is spreading widely within the south-western United States (Hunter 1991). Previous work on *Bromus* at NDFF indicates that elevated CO₂ elicits plastic changes in physiology, reproductive allocation and seed production (Huxman *et al.* 1999; Smith *et al.* 2000; Huxman *et al.* 2001). These strong phenotypic responses in traits related to fitness suggested that selection for reduced stomatal conductance may be strong and so we examined both evolutionary and plastic responses in *Bromus* stomatal conductance to elevated atmospheric CO₂. For *Bromus* populations collected from elevated and ambient CO₂ FACE rings at the NDFF, we measured stomatal conductance in plants grown under ambient and high CO₂ levels in growth chambers. Our central hypothesis was that *Bromus* populations from elevated CO₂ FACE rings should evolve

lower conductance rates to better conserve water in the field. By growing plants from these populations under both ambient and high CO₂ conditions in growth chambers, we were also able to test whether plasticity in stomatal response varied between populations. In particular, a reduced capacity for conductance plasticity in *Bromus* populations from elevated CO₂ FACE rings would suggest that genetic assimilation for conductance has occurred in these populations. To explore potential adaptive consequences of both genetic and plastic changes in response to elevated CO₂, we also measured reproductive output from plants in the growth chambers. If there is a fitness cost to the evolution of reduced conductance or conductance plasticity when CO₂ is more limiting, we would expect that elevated CO₂ field populations should have a lower reproductive output than ambient CO₂ field populations when both populations are grown under ambient CO₂. Given previous studies that indicate a plastic growth and reproductive response in *Bromus* to elevated CO₂ (Smith *et al.* 2000; Huxman *et al.* 1999), we also expected a plastic adaptive response to high CO₂ growth conditions such that, regardless of field population origin, plants growing under high CO₂ should exhibit higher reproductive output.

MATERIALS AND METHODS

The NDFF is located on the Nevada Test Site (36°49' N, 115°55' W). On April 28, 1997, the NDFF began exposing circular plots of the Mojave Desert (23 m diameter rings) to ambient (blower controls) or elevated (*c.* 550 μmol mol⁻¹ CO₂) atmospheric CO₂ treatments. On 6 May 2004, we collected a minimum of 30 *Bromus* maternal family seed samples from each of the three ambient and three elevated CO₂ FACE rings that had been under continuous treatment for 7 years. Previous research on *Bromus* has demonstrated some effects of maternal growth environment on phenotypic expression in offspring (Huxman *et al.* 1998). Thus, before testing for genetic differences between *Bromus* populations from ambient and elevated CO₂ treatments, we controlled for maternal environmental effects by growing out the seeds we collected from the NDFF in an outdoor common garden at the University of California, Davis, California (38°32' 41" N, 121°44' 25" W). In December 2004, we planted this outdoor common garden using 164 mL conetainers (Stuewe and Sons, Tangent, OR, USA) filled with Yolo loam. Plants were arranged in a randomised complete block design and irrigated with tap water. Overall plant mortality was less than 5% and no maternal families were lost. We then used seed produced from this outdoor common garden to plant experimental populations that replicated maternal families in ambient CO₂ (360 μmol mol⁻¹) and high CO₂ (700 μmol mol⁻¹) growth chambers (Model PGR15; Conviron, Pembina, ND, USA). Plants in growth chambers were grown in trays with 20.7 cm³ planting cells (ITML, Ontario, Canada). Each planting cell was filled with 15.5 cm³ of Turface MV™ and *c.* 5 cm³ of Turface QuickDry™ (Profile, Buffalo Grove, IL, USA), coarse- vs. fine-grained forms of calcined montmorillonite clay respectively. The QuickDry™ substrate was amended with solid, slow-release fertilisers (Simplot, Boise, ID, USA) in the forms of polymer-coated potassium nitrate (13-0-43) and tri-

ple superphosphate (0-45-0) at the rate of 0.8 g N L⁻¹, 2 g K L⁻¹ and 0.4 g P L⁻¹. Tap water was applied to the plants as needed so water was not limiting. Temperature in the growth chambers was maintained at 22°C and all growth chamber lights (four metal halide, four high-pressure sodium and four incandescent lights) were applied for a 14 h day length including gradual one-hour transitions between light and dark periods.

We repeated the growth chamber trials and switched the CO₂ levels in the chambers to avoid confounding chamber effects with treatment effects. We measured stomatal conductance for plants in both trials using a steady-state leaf porometer (Model SC-1; Decagon Devices, Pullman, WA, USA). Inflorescence weights were obtained from the first growth chamber trial where plants were grown to flowering and seed production. Inflorescence weight was used as an index of reproductive fitness because it is highly correlated with viable seed number ($r = 0.93$; $P < 0.0001$; $n = 45$). Although inflorescence weight data were collected only from the first growth chamber trial, we feel that chamber effects were minimal because environmental conditions were very similar between the two chambers. For example, average temperatures (\pm SD) in high-CO₂ vs. low-CO₂ chambers were $22.0 \pm 0.04^\circ\text{C}$ and $22.0 \pm 0.08^\circ\text{C}$, respectively, while per cent relative humidity was 81.4 ± 2.2 and 81.9 ± 3.0 respectively. In addition, fertilisation and watering regimes were identical. Initial individual seed weight for each maternal plant was used as a covariate in the analysis of both conductance and inflorescence weight to further account for maternal effects. Averaging of the conductance and reproductive data across maternal families within each FACE ring resulted in a total sample size of 24 replicates (from both growth chamber trials) for conductance and 12 replicates (from the first growth chamber trial) for inflorescence weight. Stomatal conductance data from both growth chamber trials were analysed with chamber effect as a random factor. Analyses of variance of both conductance and reproductive data were conducted using JMP v.10 software (SAS Institute, Inc., Cary, NC, USA).

Although the number of maternal families in our experiment was relatively low, we initially planned to use family-level data to estimate broad-sense heritability for both conductance and inflorescence weight. However, an initial analysis of the data revealed that variability around estimated parameters was so large as to render the estimated values unreliable and so these data and analyses are not presented. To estimate rates of evolutionary change in conductance, we used formula found in Bone & Farres (2001) to calculate the rate of evolution in haldanes (change in mean per generation, in phenotypic standard deviation units) between FACE seed sources for populations from both growth chamber CO₂ environments.

RESULTS

Under ambient (360 $\mu\text{mol mol}^{-1}$) CO₂ conditions in the growth chamber, *Bromus* plants from elevated CO₂ field populations exhibited significantly lower stomatal conductance than ambient CO₂ field populations (contrast $F = 14.28$, $P = 0.0014$; Fig. 1 and Table 1A). Under high-CO₂ growth

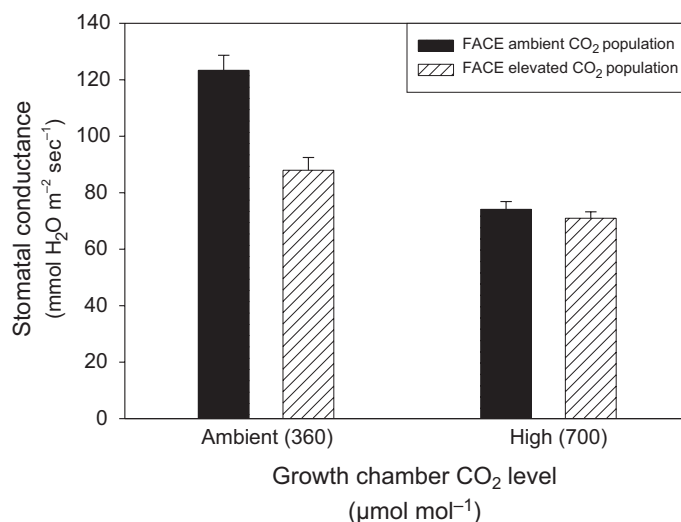


Figure 1 Evolution of reduced plasticity in stomatal conductance. Genetic differences in plastic response are indicated by interactive effects of field population source and growth chamber CO₂ level on stomatal conductance in *Bromus* (mean \pm SE).

Table 1 Analyses of variance for the two-way factorial design examining the main and interactive effects of FACE field population source and growth chamber CO₂ levels on (A) stomatal conductance, (B) inflorescence weight, (C) concentration of leaf nitrogen (microgrammes of N per gramme of leaf) and (D) number of days from planting to first flower.

Source	d.f.	Mean square	<i>F</i> ratio	<i>P</i> value
A. Stomatal conductance				
FACE population source	1	1766.12	7.53	0.0134
Growth chamber CO ₂ level	1	6396.19	27.26	<0.0001
Population \times CO ₂ level	1	1593.12	6.79	0.0179
Average maternal seed weight	1	87.67	0.37	0.5486
Chamber identity (random factor)	1	1507.44		0.3940
Error	18	234.61		
B. Inflorescence weight				
FACE population source	1	0.6274	10.40	0.0145
Growth chamber CO ₂ level	1	0.7349	12.19	0.0101
Population \times CO ₂ level	1	0.2925	4.85	0.0635
Average maternal seed weight	1	0.2012	3.34	0.1105
Error	7	0.0603		
C. Leaf [N]				
FACE population source	1	0.0352	0.5319	0.4866
Growth chamber CO ₂ level	1	4.0420	61.1618	<0.0001
Population \times CO ₂ level	1	0.0415	0.6276	0.4511
Error	8	0.0661		
D. Days to first flower				
FACE population source	1	3.485	0.0272	0.8732
Growth chamber CO ₂ level	1	94.454	0.7360	0.4159
Population \times CO ₂ level	1	28.779	0.2242	0.6485
Error	8	128.340		

chamber conditions, the lack of a significant population source effect (contrast $F = 0.01$, $P = 0.9132$; Table 1A) suggests a pronounced reduction or canalisation of phenotypic differences in conductance between field populations (Fig. 1). Averaged across field source populations, plants grown in

high-CO₂ growth chambers exhibited significantly lower stomatal conductance than plants grown in ambient-CO₂ growth chambers ($F = 27.26$, $P < 0.0001$; Table 1A). This main effect of growth chamber CO₂ level indicates a phenotypically plastic reduction in conductance for both field source populations in response to higher CO₂. The interaction between population source and growth chamber CO₂ level was also significant ($F = 6.79$, $P = 0.0179$; Fig. 1 and Table 1A), suggesting that elevated CO₂ field populations have a reduced capacity for phenotypically plastic conductance adjustments in response to lower levels of CO₂. Chamber effects on conductance were not significant ($F = 0.87$, $P = 0.3940$; Table 1A). The rates of evolutionary divergence between FACE seed sources in the ambient- vs. high-CO₂ growth chamber environments were -0.243 and -0.105 haldanes, respectively.

When grown in the ambient-CO₂ growth chamber, *Bromus* plants descendent from elevated CO₂ field populations produced significantly smaller inflorescences than ambient CO₂ field populations (contrast $F = 14.65$, $P = 0.0065$; Table 1B and Fig. 2). Under high-CO₂ growth chamber conditions, there was not a significant difference in reproduction between FACE field population sources (contrast $F = 0.66$, $P = 0.44$; Table 1B and Fig. 2).

Leaf tissue quality as measured by leaf tissue nitrogen concentration (microgrammes of N per gramme of leaf) was affected by the growth chamber CO₂ environment, but not by FACE seed source. There was no significant difference in leaf tissue nitrogen concentration between ambient versus elevated CO₂ FACE seed sources (Table 1C), but leaf tissue nitrogen concentration was significantly greater in ambient- (4.2 ± 0.1 SE) vs. high (3.0 ± 0.1 SE)-CO₂ growth chamber environments. There were no significant interactions between FACE seed sources and growth chamber CO₂ levels for leaf tissue nitrogen concentrations. There were no significant differences in phenology between FACE seed sources or growth

chamber CO₂ levels, as measured by time to first flower in the first trial of the growth chamber common gardens (Table 1D).

DISCUSSION

Physiological responses in *Bromus* to elevated CO₂ treatments (i.e. changes in stomatal conductance) are consistent with an evolutionary response to FACE CO₂ treatments in the field as well as a reduction in the capacity for phenotypic plasticity in conductance. We also detected a plastic reproductive fitness response to growth chamber CO₂ treatments that differed between FACE CO₂ populations. The FACE CO₂ population source effect indicates that *Bromus* populations under elevated CO₂ treatments evolved lower conductance in the field within a 7-year period. Because water was not limiting in the growth chambers, we could not experimentally test for the adaptive value of reduced conductance when water is in short supply. However, water is a primary limiting resource in this desert environment (Smith 1997) and a large body of literature on adaptation to xeric habitats suggests that this evolution of reduced conductance represents an adaptive response to conserve water and thus increase water-use efficiency (Bowes 1993; Tyree & Alexander 1993; Drake *et al.* 1997; Buckley & Schymanski 2014). Despite the low replication of CO₂ treatments at the FACE site, the highly significant differences between FACE CO₂ treatment populations suggest a robust evolutionary response in stomatal conductance. Our results indicating an evolutionary basis for reduced conductance under elevated atmospheric CO₂ are consistent with research in several FACE experiments that demonstrated plastic decreases in stomatal conductance that resulted in increased water-use efficiency (Ainsworth & Long 2005).

A reduced capacity for conductance plasticity was found for the elevated CO₂ FACE populations, suggesting that changes in conductance reaction norms may have occurred rapidly in these populations. It has been suggested that genetic assimilation may be more common than generally recognised, in part, because the process of genetic assimilation may occur relatively rapidly and thus escape detection (Pigliucci *et al.* 2006). We believe that this reduced capacity for plasticity in conductance likely represents selection on standing genetic variance in the norms of reaction associated with conductance plasticity that existed before elevated CO₂ FACE treatments began. An examination of conductance plasticity differences among maternal families within the NDFP ambient control treatment provides evidence for the existence of this initial standing genetic variation in reaction norms. For example, two maternal families from separate populations (i.e. two different FACE rings) of the NDFP ambient control treatment exhibited conductance rates lower than the average for elevated sources when exposed to the ambient-CO₂ growth chamber treatment. This suggests that genotypes with lower conductance and a reduced capacity for plastic response previously existed in the unselected populations. Empirical evidence from a long history of quantitative genetics (Roff 2007) as well as more recent theoretical developments (Barrett & Schluter 2007) indicate that adaptive responses based on standing genetic variation, rather than new mutations, are more likely to foster the type of rapid evolutionary change in conductance

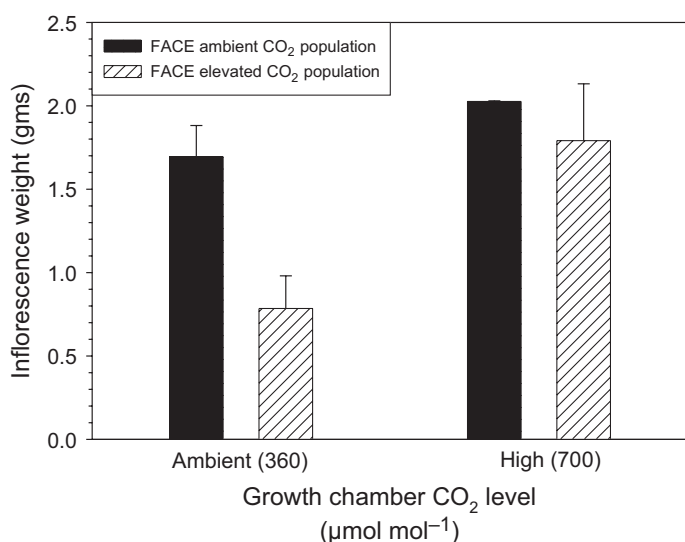


Figure 2 Fitness consequences of CO₂ level and population source. Interactive effects of genetic and growth chamber CO₂ level on reproductive fitness in *Bromus* as measured by inflorescence weight (mean \pm SE).

reaction norms that we observed in *Bromus*. In support of this idea, Case *et al.* (1998) reported differences among paternal families of wild radish in both the direction and magnitude of stomatal index responses to changing CO₂; results suggesting that standing genetic variation in norms of reaction related to stomatal function exists in other weedy species.

A comparison of reproductive output between ambient and elevated CO₂ FACE populations growing under ambient CO₂ levels suggests a fitness cost associated with reduced capacity in these populations for plastic increases in conductance when CO₂ is limiting (Fig. 2). Although we did not test for costs of plasticity directly, this reduction in conductance plasticity is at least consistent with the hypothesis that there are fitness costs associated with maintaining this plasticity when CO₂ is less limiting (Pigliucci *et al.* 2006). Conversely, because the evolution of reduced conductance in the elevated CO₂ populations is likely a selective response to conserve water and given that water was not limiting in the growth chambers, we did not expect to observe a fitness advantage associated with reduced conductance.

As an annual grass with little persistent seed bank development (Jurand *et al.* 2013), *Bromus* has a short generation time that may facilitate rapid evolutionary response to elevated atmospheric CO₂. In contrast, long-lived perennial plants or annual plants with persistent seed banks (Templeton & Levin 1979) in this desert plant community have effectively longer generation times and thus may be expected to respond more slowly to selective pressures. In addition, variation in breeding systems among different plant species may significantly affect the likelihood of rapid evolutionary response. In particular, outcrossing can significantly increase gene flow that, in turn, reduces the effectiveness of local selection to evolve differences in adaptive traits. The highly selfing nature of *Bromus* may reduce gene flow among the FACE experimental rings and thus promote the local evolution of decreased stomatal conductance under elevated CO₂.

Although we found evidence for rapid evolution in stomatal conductance in *Bromus*, information is lacking on whether other species in this desert environment have evolved physiological adaptations to rising atmospheric CO₂. A field study at NDFP comparing growth, phenology and physiology between *Bromus* and a native short-lived perennial (*Eriogonum inflatum* Torr. & Frem.) indicated that *E. inflatum* exhibited a pronounced, season-long reduction in stomatal conductance under elevated CO₂ (Huxman & Smith 2001). In fact, phenotypic reduction in conductance in *E. inflatum* plants exposed to elevated CO₂ was stronger and more prolonged than in *Bromus*. Whether there is a genetic basis for reduced conductance in *E. inflatum* remains unknown, but the magnitude and extended duration of the response suggest that selection for reduced conductance in this native perennial might be strong under elevated CO₂. If there is selection for reduced conductance in *E. inflatum*, this desert ephemeral species would be a prime candidate for testing whether this phenotypic response in conductance has a genetic basis. In contrast, a recent study by Newingham *et al.* (2014) on the response of the perennial plant community to elevated CO₂ at the NDFP site suggests that evolutionary response in long-lived perennials may be unlikely. Despite a decade of exposure to elevated CO₂, they

found very little change in total cover in the woody and herbaceous perennial species examined. The authors suggest that slow growth and sporadic recruitment may constrain species responses at a community level; these growth and demographic characteristics might also reduce any evolutionary response within species.

Studies focused on potentially complex traits, such as plant morphology, growth and phenology have failed to detect significant evolutionary responses to elevated CO₂ (Lau *et al.* 2007; Leakey & Lau 2012). It is possible that the genetic basis for variation in stomatal conductance may be simpler than the genetic structure underlying phenotypic variation in these more downstream traits. Recent genomic research in wheat indicates that multiple traits related to stomatal conductance and water-use efficiency may be controlled by relatively few genes and thus may respond fairly rapidly to selection (Panio *et al.* 2013). Similar research is not yet available for *Bromus*, but these results at least provide a possible genetic basis for more rapid evolutionary responses in physiological traits such as stomatal conductance. In general, differences among plant traits in the potential for evolutionary response are common and may reflect both environmental effects on the phenotypic expression of genetic variability (e.g. heritability) as well as genetic constraints (e.g. negative genetic correlations among traits) (Ackerly *et al.* 2000; Geber & Griffen 2003).

In addition, the natural field conditions within the FACE rings may have promoted an evolutionary response in conductance by providing a complex environment, whereby the effects of elevated CO₂ on interspecific interactions might alter both phenotypic expression of genetic variation and the efficacy of the selection in causing evolutionary change (Lau *et al.* 2014). The effects of competition on evolutionary responses in *Bromus* at NDFP remain unknown. However, research on potential responses of a winter annual community in the Sonoran Desert to environmental variation and climate change suggest that interspecific interactions and temporal variability in rainfall may act to maintain trade-offs between water-use efficiency and relative growth rate (Huxman *et al.* 2013). This research focused on among-species differences, but it is not unreasonable to suggest that these interactions between biotic and abiotic stressors might also influence the capacity of selection to alter within-species variation in traits associated with water-use efficiency (e.g. stomatal conductance). At an ecosystem level, the effects of elevated CO₂ on conductance, assimilation and water-use efficiency can be especially important within a desert environment. Within xeric communities, changes in conductance can influence a number of ecosystem properties ranging from increased soil water to evapotranspiration dynamics (Morgan *et al.* 2004).

A logistic factor that may contribute to a general lack of evidence for evolutionary responses to elevated CO₂ is that the level of replication of CO₂ treatments at a FACE site is typically low. It is possible that concerns over low statistical power at FACE sites have discouraged exploration of evolutionary response to elevated CO₂ especially if the initial assumption is that evolutionary responses will be weak (i.e. small treatment effect sizes). Our results indicate that the magnitude of effects associated with evolutionary response to elevated CO₂ can be substantial and so further research focused

on detecting genetic shifts might reveal that rapid evolutionary responses to FACE treatments are more common than currently supposed.

Our findings corroborate previous research (Huxman *et al.* 1998; Long *et al.* 2004; Ainsworth & Long 2005) that plants respond phenotypically to elevated CO₂ by expressing reduced tissue quality (i.e. nitrogen concentration) and thus an increased carbon-to-nitrogen (C:N) ratio of leaf tissue. If NO₃ is the primary source of nitrogen, as it was in our study, then a reduction in leaf nitrogen may reflect a reduced capacity for NO₃ uptake under elevated CO₂ because of a reduction in photorespiration (Bloom *et al.* 2012). Reduced tissue quality may negatively impact *Bromus* populations in particular because of an associated reduction in seed quality and seedling vigour (Huxman *et al.* 1998, 2001). Compared to seeds produced under ambient CO₂, seeds from *Bromus* plants grown under elevated (700 µmol mol⁻¹) CO₂ had higher C:N ratios and reduced seed reserves. Upon germination, seedlings from *Bromus* plants grown under elevated CO₂ also grew more slowly and produced fewer leaves (Huxman *et al.* 1998). As noted by Huxman *et al.* (1998), this reduction in seed quality and seedling vigour could have a substantial impact on the future demographic dynamics and invasive success of *Bromus* as atmospheric CO₂ increases. Thus, there may be a significant fitness trade-off in that higher seed production under elevated CO₂ may be offset by a reduction in seedling survival and vigour that, in turn, might constrain *Bromus* population growth and invasive spread.

Rates of evolutionary divergence in stomatal conductance between FACE seed sources grown in the ambient- vs. high-CO₂ growth chamber environments (−0.243 and −0.105 haldanes, respectively) are near the high end of the studies reviewed by Bone & Farres (2001). These relatively high rates of evolutionary change in stomatal conductance support their suggestion that physiological traits may evolve more rapidly than morphological traits. Although the same FACE populations were used in high- and ambient-CO₂ growth chambers, estimates of evolutionary rates were much higher in the ambient-CO₂ growth environment. This environmental effect on the phenotypic expression of evolutionary change lends support to the observation that estimates of many evolutionary parameters, including local adaptation and heritability, may vary with environmental conditions (Williams *et al.* 2008).

Evolutionary responses in this invasive species to rising CO₂ could have broad ecosystem impacts because of its effect on fire regimes. As *Bromus* populations spread throughout the Mojave Desert ecosystem, they provide a continuous fine fuel source that increases the frequency and size of fires. This change in fire regime, in turn, threatens native plant ecosystems that are not adapted to frequent, widespread fires (Smith *et al.* 2000). Previous investigators have suggested that the long-term success and dominance of *Bromus* in this arid ecosystem will be promoted by phenotypically plastic increases in biomass and seed production in response to elevated CO₂ (Smith *et al.* 2000). Our evidence for contemporary *in situ* evolution of *Bromus* populations suggests that genetic shifts in response to elevated CO₂ may also promote the persistence and dominance of this desert weed in the Mojave Desert ecosystem. In general, the current wave of biological invasions

may be due, in part, to the ability of non-native species to evolve rapidly in response to human-induced environmental change (Lee 2002; Rice & Emery 2003).

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AUTHORSHIP

J.G. and K.R. designed experiment, J.G. collected the data and J.G. and K.R. analysed the data. J.G. wrote the first draft of the manuscript and both authors contributed substantially to revisions.

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