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Identification of a regulation network in response to cadmium toxicity using blood clam *Tegillarca granosa* as model

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Clam, a filter-feeding lamellibranch mollusk, is capable to accumulate high levels of trace metals and has therefore become a model for investigation the mechanism of heavy metal toxicification. In this study, the effects of cadmium were characterized in the gills of *Tegillarca granosa* during a 96-hour exposure course using integrated metabolomic and proteomic approaches. Neurotoxicity and disturbances in energy metabolism were implicated according to the metabolic responses after Cd exposure, and eventually affected the osmotic function of gill tissue. Proteomic analysis showed that oxidative stress, calcium-binding and sulfur-compound metabolism proteins were key factors responding to Cd challenge. A knowledge-based network regulation model was constructed with both metabolic and proteomic data. The model suggests that Cd stimulation mainly inhibits a core regulation network that is associated with histone function, ribosome processing and tight junctions, with the hub proteins actin, gamma 1 and Calmodulin 1. Moreover, myosin complex inhibition causes abnormal tight junctions and is linked to the irregular synthesis of amino acids. For the first time, this study provides insight into the proteomic and metabolomic changes caused by Cd in the blood clam *T. granosa* and suggests a potential toxicological pathway for Cd.

One of the most important concern in marine and coastal environments is the heavy metal due to their toxic effects on marine organisms^{1,2}. Among them, cadmium (Cd), a non-essential metal element required for many of living organisms, can easily enter the environment as a pollutant from many anthropogenic activities³. The development and industrialization of coastal zones has contributed greatly to Cd pollution in marine environments⁴. As a result, Cd is one of seven most common released heavy metals for the environment (Cd, Cr, Cu, Hg, Ni, Pb, and Zn)⁵ and affects many invertebrates in marine, such as gastropod mollusks and bivalve⁶. In the East China Sea, especially along the coasts in Zhejiang Province, Cd has been determined as the most abundant heavy-metal pollutant in *Tegillarca granosa*⁷. Therefore, bivalve responses to Cd contamination represent an important research topic in environmental toxicology, public health and costal conservation⁸.

Molluscs are known and used worldwide as healthy indicators of oceans, because of their ability of metal concentration accumulation and integration⁹. Despite many studies on Cd-induced toxicological effects at the physiological level, the mechanisms underlying the observed responses are still poorly understood. Previous studies have found some proteins, such as metallothionein (MT), as bioindicators of response to induced Cd exposure^{10–12} or superoxide dismutase (SOD) activity as an index to estimate Cd induction level¹³. However, change in MT or SOD level is only one of many underlying mechanisms related to Cd resistance in animals.

Metabolomics is a high-throughputs technology for detecting small metabolites and molecular products of metabolism^{14,15}. ¹H NMR spectroscopy is often used to detect a large range of endogenous metabolites in an organism. Rich quantitative and structural information by ¹H NMR allows analyzing many metabolites

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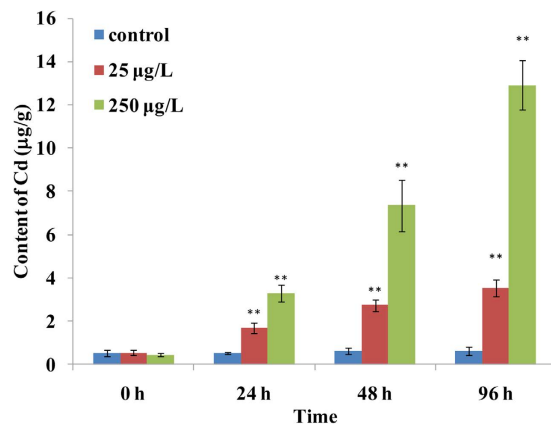


Figure 1. The accumulated concentrations ($\mu\text{g/g}$) of cadmium (Cd) in the gills from blood clams *Tegillarca granosa* after exposure to Cd for 24, 48 and 96 hours. The cadmium concentrations were presented as the mean \pm standard deviation. The significance of the difference (** $p < 0.01$) between control and Cd treated-groups was tested by a one-way analysis of variance.

simultaneously. However, reports of the application of NMR-based toxicology studies in marine invertebrates have been limited to date^{16–19}. High applicability and reliability of this technology has been demonstrated by applying it to investigate the toxicological mechanisms of cadmium and copper on green mussels²⁰. Blaise *et al.*²¹ applied ¹H NMR to reveal the latent phenotype associated to superoxide dismutase and catalase *Caenorhabditis elegans* mutations, and other studies also used ¹H NMR to obtain metabolic profiles of *C. elegans* responses to cadmium²².

Proteomics is a large-scale technology in study of protein structure and function. It has shown its potential to enhance understanding of stress response mechanisms at molecular level²³. iTRAQ, isobaric tags for relative and absolute quantification, uses labeled peptides for protein identification and quantification. Because the labeled peptides can be identified by sensitive mass spectrometers, it makes iTRAQ the one of currently widely used proteomic approach²⁴. A number of studies have characterized the metabolomics or proteomic responses to several stressors in mollusks, including heavy metal exposure, arsenic exposure, and pathogen infection^{3,25–28}. For instance, Stress responses in the gills of the pacific oyster *Crassostrea gigas* were conducted by iTRAQ proteomic analysis²⁸. iTRAQ-based proteomics also revealed novel proteins involved in pathogen challenge in Sea Cucumber *Apostichopus japonicus*⁸. However, to our knowledge, no iTRAQ-based proteomic and NMR-based metabolomic combination studies have been conducted to understand the responses of clams to Cd. Perturbation in metabolites and proteins that are involved in same metabolic pathways can be detected by proteomics and metabolomics. Therefore, their combination can lead to a better understanding of toxicological effects of contaminants in bioindicator species^{29,30}. For example proteomic and metabolomic analysis revealed gender-specific responses of mussel *Mytilus galloprovincialis* to tetrabromodiphenyl ether³⁰, responses of clam *R. philippinarum* to arsenic exposure under different salinities³¹, and responses of Pacific oyster *C. gigas* to elevated pCO₂ exposure³².

Our work integrated iTRAQ and H NMR approach to detect biomarker response to Cd in gill and to elucidate the toxicological effects of Cd (25 $\mu\text{g/L}$ 250 $\mu\text{g/L}$) in the blood of the clam *T. granosa* at three (24, 48 and 96 h) exposure time points. There were no obvious histopathological changes in gill at selected 250 $\mu\text{g/L}$ for Cd exposures compared with the control within 96 h according to pre-experiment. Here our study aimed to address the following points: (1) Which metabolite and proteins or functional groups of proteins alter their expressions significantly in response to the three time points previously mentioned? (2) Are these responses specific or common to the three different time points? (3) Construction of a regulation network to better understand the relationship between the proteins, affected pathways and downstream metabolic compounds.

Results and Discussion

Cadmium bioaccumulation in the gills of blood clams. Figure 1 shows the accumulation of cadmium in blood clam gill tissues after 24, 48 and 96 hours of exposure. The average concentration of cadmium in gill tissues that were exposed to a low dose of cadmium (25 $\mu\text{g/L}$) was approximately 3 times higher than that of the control group after 24 hours of exposure ($p < 0.01$), while the cadmium from the high dose of cadmium (250 $\mu\text{g/L}$)-exposed group was approximately 6-fold ($p < 0.01$) higher than that of the control group. The concentrations of cadmium in blood clam gill tissue were continuously elevated with the increased exposure time for both low and high cadmium-dosed groups. The Cd concentrations in 25 and 250 $\mu\text{g/L}$ cadmium-exposed blood clam samples were approximately 5 and 16 ($p < 0.01$) times higher than those of the control group after Cd exposure for 48 hours, respectively. At 96 h post exposure, the gill tissue Cd levels for the low and high Cd exposures were 6.4 and 28 times higher, respectively, than that of the control group. This finding indicates that this marine bivalve has a high accumulation of Cd and is a good bioindicator for Cd monitoring and ecotoxicology. Previous study also showed that of the five most common molluscs along Zhejiang coast in China, *T. granosa* had greater aptitude for bioaccumulation of Cd⁷.

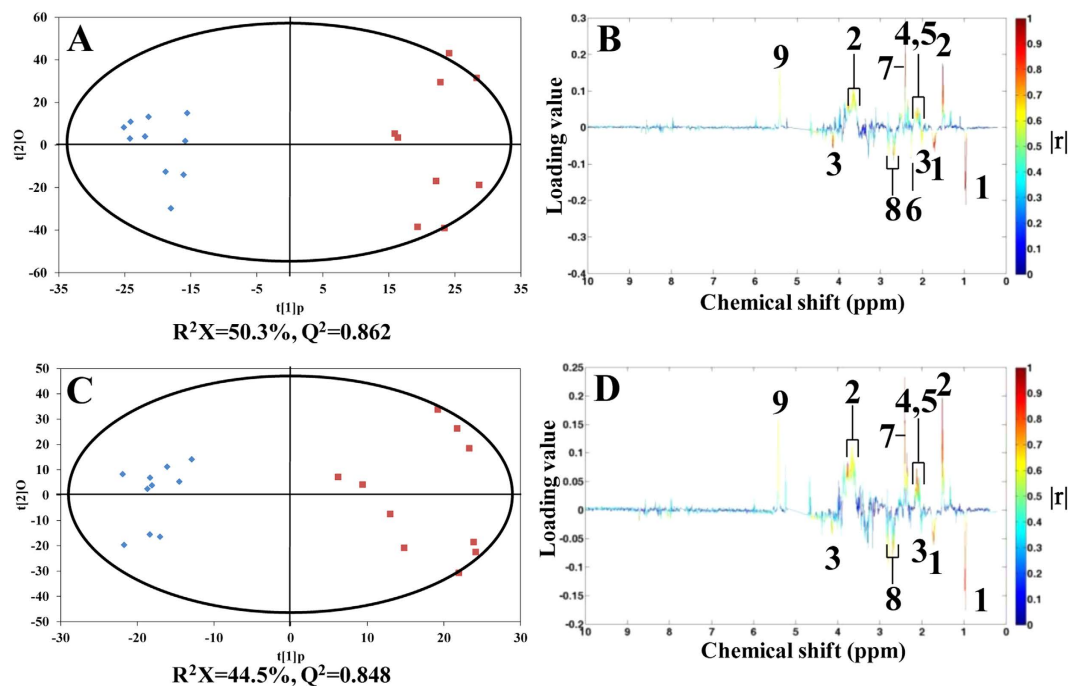


Figure 2. OPLS-DA score plots derived from ^1H NMR spectra of gill tissue extracts from the control (\blacklozenge) and exposed (\blacksquare) blood clam groups with (A) 25 and (C) 250 $\mu\text{g/L}$ Cd, respectively, after exposure for 24 hours and corresponding coefficient plots (B,D). The most significant metabolic variations between Cd exposure and control group are shown in the coefficient plots: positive directed peaks for up-regulated metabolites in Cd exposure group; negative directed peaks for down-regulated metabolites. Key: (1) leucine, (2) alanine, (3) proline, (4) glutamate, (5) glutamine, (6) acetoacetate, (7) succinate, (8) aspartate and (9) glycogen.

NMR-based metabolomics of Cd challenging in blood clams. Supplementary Fig. S1 displays a typical 1-D ^1H NMR spectrum of gill tissue extracts from a control blood clam. Several different classes of metabolites were identified by the Chemomx software, including amino acids, energy storage compounds, organic osmolytes, and intermediates of the tricarboxylic acid cycle.

OPLS-DA analysis was performed on the NMR spectral data of blood clam gill tissue extracts to seek metabolic biomarkers related to Cd exposure after 24, 48 and 96 h. Metabolite assignments in the 1-dimensional 500-MHz ^1H NMR spectra of tissue extracts from the gills of *T. granosa* are shown in Supplementary Table S1. After Cd exposure for 24 h, the gill samples from the control and both doses (25 and 250 $\mu\text{g/L}$) of Cd-exposed groups were clearly separated in OPLS-DA score plots (Fig. 2A,C), with reliable Q^2 values greater than 0.8. Based on the loading plot of OPLS-DA (Fig. 2B,D), both doses (25 and 250 $\mu\text{g/L}$) of Cd induced identical metabolic responses in blood clam gill tissues. Although the Cd concentration differed by 10 times in water between the 2 exposure groups, the difference was 1.9 times in tissues between them. This result could explain the identical metabolic responses that were induced by both doses of Cd in blood clam gills at 24 hpe. For those blood clam samples that were sampled at 48 hpe (Supplementary Fig. S2), the low dose of Cd (25 $\mu\text{g/L}$) increased the levels of alanine, glutamine and succinate and decreased the levels of leucine, aspartate, adenine and proline. Although the high dose of Cd (250 $\mu\text{g/L}$) induced some similar metabolic responses, including increased alanine, glutamine and succinate and decreased leucine, aspartate and proline, glycogen was uniquely elevated, and adenine was not altered in this dose of Cd-exposed blood clam gill tissues. For those blood clam samples that were sampled at 96 hpe (Supplementary Fig. S3), the results indicate that the low dose of Cd caused increases in alanine, glutamate, succinate and glycogen and decreases in leucine, β -alanine, hypotaurine, proline and adenine, whereas the high dose of Cd induced similar metabolic biomarkers, including increased alanine, glutamate, succinate and glycogen and decreased leucine, β -alanine, hypotaurine and proline.

Overall, the dose- and time-dependent toxicological effects of Cd exposure included neurotoxicity and disturbances in energy metabolism after exposure for 24 and 48 hours, as indicated by elevated glutamate, glutamine, alanine, succinate and decreased leucine, aspartate and proline^{20,33}. After exposure for 96 hours, however, both Cd exposures induced disturbance in the osmotic regulation in blood clam gill tissues, as displayed by decreased hypotaurine and β -alanine. Interestingly, both Cd exposures induced identical metabolic biomarkers at 24 h post-treatment and no significant difference between the two doses (25 and 250 $\mu\text{g/L}$) of Cd-exposed groups. This result probably can be explained by the similar levels of accumulated Cd in blood clam gill tissues. Thus, we selected the high dose 250 $\mu\text{g/L}$ Cd-exposed group for further proteomic analysis.

GO and KEGG analysis of the differential expression proteins at three time points. Only proteins that contained at least two peptides and that were detected in both replicates were used for quantification; we described global proteome changes in the *T. granosa* gill during the short-term course of Cd exposure. We

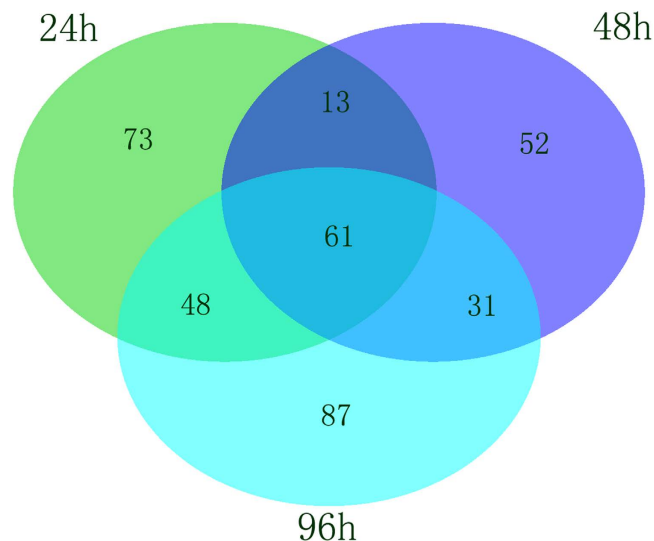


Figure 3. Changed proteome distribution between different time points. Venn diagram showing unique and shared proteins between time points.

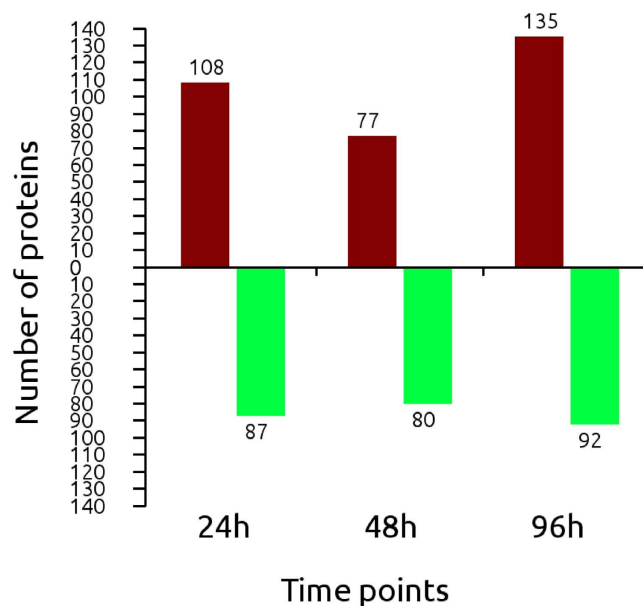


Figure 4. Number of differential expressed proteins detected at 24, 48 and 96 h.

identified 2329 distinct proteins, of which 2318 were identified and quantified reliably at a global false discovery rate (FDR) of 1%. 365 identified proteins had significant changes in expression at different time points, compared with the control group, and 61 proteins were differentially expressed at all of the examined time points (Fig. 3, Supplementary Table S2). Among these differentially expressed proteins, 195, 157 and 227 up- or down-regulated proteins were identified in Cd-challenged *T. granosa* at 24, 48 and 96 h, respectively (Fig. 4).

For an overview of the function of all of the detected differential expression proteins (DEPs) and the potential linkage between them, a GO enrichment analysis was performed for DEPs detected at each time point. The DEPs were classified into the Biological process (BP), Cellular component (CC) and Molecular function (MF) categories. The DEPs were predominantly binding proteins that were involved in cellular component organization or biogenesis and in the response to inorganic substances 24 h post Cd treatment. At 48 hpe, DEPs were also primarily binding proteins that were involved in the cellular aldehyde metabolic and organic acid metabolic processes, and at 96 hpe, DEPs were predominantly proteins that were related to cellular component assembly and cytoskeleton organization (Fig. 5). Further KEGG pathway enrichment revealed that these proteins were mainly involved in glycolysis/gluconeogenesis, ribosomes, tight junctions and proximal tubule bicarbonate reclamation at 24 hpe; glycolysis/gluconeogenesis and the HIF-1 signaling pathway at 48 hpe; and glycolysis/gluconeogenesis, ribosomes, ECM-receptor interaction and tight junctions at 96 hpe (Fig. 6). The common pathways from three

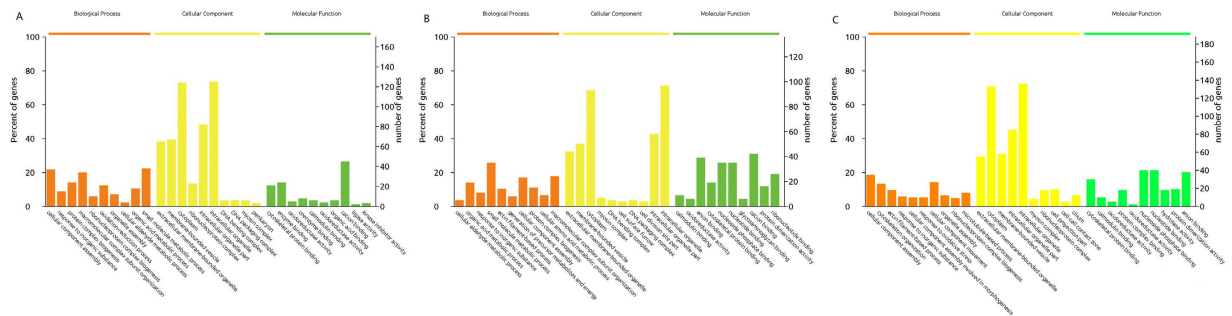


Figure 5. Most significant enriched GO terms of differentially expressed proteins at each time point. (A) 24 h; (B) 48 h; and (C) 96 h.

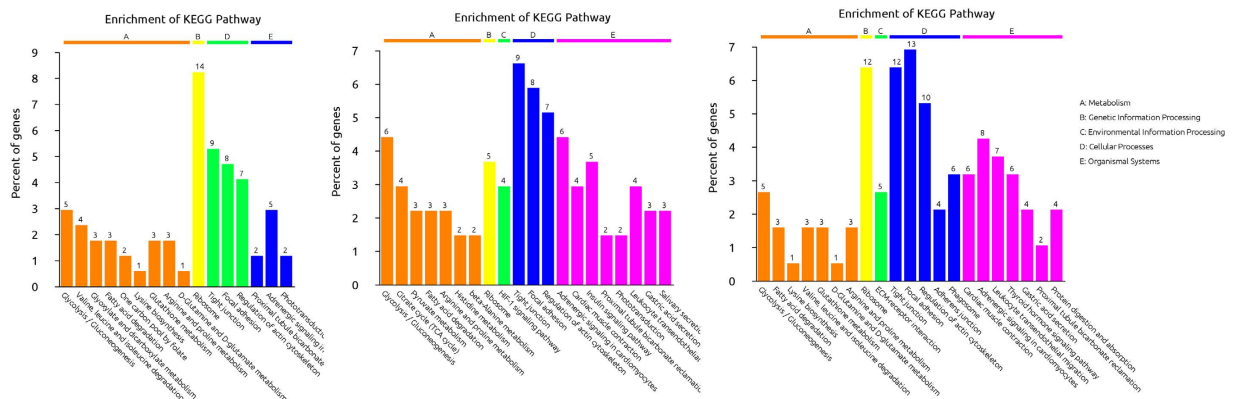


Figure 6. Most significant associated KEGG pathways of differentially expressed proteins at each time point. (A) 24 h; (B) 48 h; and (C) 96 h.

time points are glycolysis/gluconeogenesis, tight junctions and ribosomes, which should be closely related to Cd toxicity and metabolism.

In humans, Cd induces changes in carbohydrate metabolism and gluconeogenesis through various effects on the liver, kidneys, adrenal glands and pancreas. Laboratory-controlled injection and ingestive intake of Cd produce an increase in gluconeogenesis and blood glucose³⁴. Cadmium induced gluconeogenic changes in rat kidney and liver have been reported³⁵. Tight junctions mediate cell-cell adhesion and can regulate protein transport through the extracellular matrix³⁶. *In vivo*, disrupted tight-junction associated microfilaments in rat sertoli cells were detected after Cd exposure³⁷. It suggests that tight junction, i.e., intracellular adhesion and the interaction of the actomyosin cytoskeleton with the plasma membrane were likely associated with Cd in blood clams³⁸. It is likely that Cd in blood clams could destroy the function of tight junctions, i.e., intracellular adhesion and the interaction of the actomyosin cytoskeleton with the plasma membrane³⁸. Ribosomal genes have been reported to be early targets of cadmium-induced toxicity in *Chironomus riparius* larvae, indicating that cadmium can interfere cellular survival via interaction with ribosomal genes that led to extreme impairment of nucleolus activity³⁹. Significant quantities of cadmium have been found in purified ribosomes and from rat liver, people have detected significant quantities of Cd, also suggesting the attachment of Cd with ribosomes⁴⁰.

GO and KEGG analysis of common DEPs of three time points. Among all DEPs, 61 proteins were detected in all three time points. A GO and KEGG analysis of 61 common DEPs showed that the functions of its DEPs are distributed in cytoskeletal protein binding, calmodulin binding and oxidoreductase activity, which are involved in the glycolysis/gluconeogenesis, tight junction and adrenergic signaling pathways (Supplementary Figs S4 and S5).

Moreover, altered glycolysis was also seen in our NMR-based metabolomics study, with four metabolites, glutamine, alanine, glucose and glycogen, which related to glycolysis/gluconeogenesis process⁴¹, all up-regulated after Cd exposure. The results of proteomics and metabolomics coincide, showing that up-regulation of glycolysis/gluconeogenesis pathways are important for Cd metabolism in *T. granosa*. Similar responses were found in plant tomato roots after Cd treatment⁴². The breakdown of tight junction is a sign of cadmium treatment reported in previous studies^{37,43}, and our results also indicate that cadmium affects the integrity of tight junctions in blood clam.

Proteome characterization induced by Cd exposure. Some well-documented stress- or immunity-related, calcium-binding and sulfur-compound metabolism proteins were found with large fold-changes at three time points (Table 1). The universal stress protein A-like protein was significantly

Biological function	Protein description	E-value	Protein accession no.	Fold change		
				24 h	48 h	96 h
Stress response	universal stress protein A-like protein	1e-53	XP_011429497	17.39	4.09	24.88
	core histone macro-H2A	0.0	XP_011454400	0.19	0.39	0.33
	histone H2A isoform 2	3e-79	ACJ12611	0.22	0.39	0.53
Antioxidant defense	Cu/Zn superoxide dismutase	4e-40	AFQ32467	10.29	10.43	11.81
	glutathione S-transferase	6e-83	AGN03945	—	1.90	3.16
Calcium-binding protein	neuronal calcium sensor (EF-hand motif)	4e-05	P36608	6.36	11.47	2.20
	sarcoplasmic calcium-binding protein	2e-80	BAA89417	5.43	5.30	3.27
	calmodulin	4e-18	XP_010966045	6.36	11.47	2.20
Sulfur compounds metabolism	sulfotransferase	4e-92	XP_004633325	6.39	7.40	5.06
	protein disulfide-isomerase	8e-49	ERE68284	—	2.03	2.72
Cell structure	tenascin-X	0.24	XP_006537513	3.95	4.23	3.00
	mucin-17-like	2e-136	XP_011425282	3.25	4.12	4.91
	tropomyosin	3e-28	XP_011437747	3.20	4.14	3.41
Metabolic process	retinal dehydrogenase	0.0	XP_006743256	4.00	6.23	4.29
	isocitrate dehydrogenase	0.0	EKC40376	3.37	2.08	3.05
	phosphohistidine phosphatase	8e-47	XP_004848882	3.14	2.93	2.55
	annexin	5e-137	XP_004561320	3.07	2.96	5.47
	protein mab-21	9e-58	XP_004549366	3.00	4.62	3.66
	1,4-alpha-glucan-branching enzyme-like	0.0	XP_011434285	2.85	3.20	3.30
	glyceraldehyde-3-phosphate dehydrogenase	0.0	XP_010609934	2.51	2.45	3.06
	hemoglobin I	3e-37	AGA03854	2.45	2.23	2.21
	thioredoxin	6e-33	XP_002504076	2.41	4.63	3.85
	dipeptidase 1-like	3e-110	XP_011438699	2.31	2.38	2.51
	enolase-like	5e-115	XP_011436227	2.10	2.05	2.33
	pyruvate carboxylase	0.0	EKC30574	2.05	2.89	2.22
	complement C1q tumor necrosis factor-related protein	2e-14	XP_011436150	0.09	0.43	0.28
	collagen alpha-1	1e-08	XP_006792865	0.21	0.32	0.29

Table 1. Summary of differential expressed protein after 96 h Cd exposure (fold change >2 or <0.5 in all three time points).

up-regulated at all three time points and sharply increased to 24.88-fold compared to that of the control group after 96 h of exposure. The universal stress proteins (USPs) appear to play an active role in the abiotic stress response in bacteria and plants^{44,45}, but their functions remain largely unknown in animals. Cu/Zn superoxide dismutase was also highly expressed, increasing by more than 10-fold compared to that in the control at all three time points. It is generally agreed that oxidative stress plays important roles in acute Cd poisoning⁴⁶; Cd-generated superoxide anion, hydrogen peroxide, and hydroxyl radicals have been detected *in vivo* by electron spin resonance spectra⁴⁷. Cu/ZnSOD is an important antioxidant enzyme that catalyzes the conversion of superoxide to oxygen and hydrogen peroxide in aerobic organisms. The study also shows that the metal-response element (MRE) in the proximal 5' flanking sequence of Cu/ZnSOD is required for the induction of the gene by heavy metals⁴⁸. Therefore, it is very important that Cu/ZnSOD contributes to detoxifying Cd poisoning and maintaining the dynamic balance of ROS. Increasing evidence suggests that the toxicity of Cd is mediated by oxidative stress-induced cell death. The important antioxidant defense enzyme glutathione S-transferase (GST) was also up-regulated; other studies have reported that GST plays a critical role in the detoxification of Cd⁴⁹. Cd can disrupt Ca²⁺ homeostasis, causing apoptosis in a variety of cells⁵⁰. In the present study, we found that some Ca²⁺ related proteins were significantly up-regulated after Cd exposure. Three proteins, including neuronal calcium sensor (Ncs1), sarcoplasmic calcium-binding protein (Hrc) and calmodulin (Calm1), were up-regulated more than 5-fold compared with the control group. The biophysicochemical similarities of Cd²⁺ and Ca²⁺ allow Cd²⁺ to displace Ca²⁺ in some Ca²⁺-binding proteins and to disrupt Ca-mediated signaling pathways⁵¹. Further determination of the short-term and long-term factors that are affected by spatial and molecular interactions of Cd²⁺ and Ca²⁺ will provide additional insight into toxic mechanisms.

In plants, many studies have shown that Cd can induce sulfate uptake and that sulfur metabolism is associated with the detoxification of Cd⁵²⁻⁵⁶. However, few studies have reported the Cd and sulfur compound relationship in animals. In our study, we found that sulfotransferase (Sult1b1) was significantly up-regulated after Cd challenge. This result suggests that sulfur-related metabolism might also be involved in the Cd-induced response and the detoxification of Cd in animals.

Heavy metals can change nucleosome structure by binding to its core component and this may have a potential function in toxicity⁵⁷. In our study, the key nucleosome components H2, H3 and H4 significantly decreased after Cd exposure. It might implicate Cd-induced DNA damage and apoptosis⁵⁸. Another study showed that

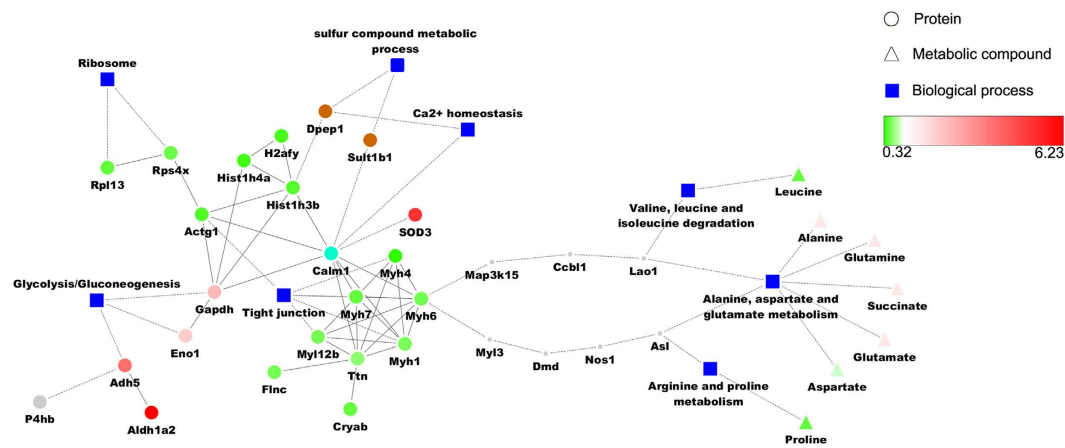


Figure 7. Key regulation model constructed using proteomic and metabolomic data. Circle nodes indicate proteins and triangle nodes indicate metabolic compounds. Filled color represents expression fold change using gradient color from blue to red. Blue colored rectangle indicates KEGG pathways. A solid line between two proteins indicates a known interaction annotated in the STRING database; a dashed line between proteins indicates indirect interaction. Gray-colored dots are extended proteins from the database that built up a potential path from the proteomic to the metabolomic layer.

heavy metal exposure results truncation of H2A and H2B, and the elimination of some functional modification sites⁵⁹. Similar metabolism progress might also appear in the cell after Cd treatment, probably resulting in the truncation of histones.

Regulation model construction. A regulation network was constructed to better understand the relationship between the proteins, affected pathways and downstream metabolic compounds. As shown in the model (Fig. 7), Rpl13 and Rps4x, two ribosome proteins, were significantly down-regulated at all three time points; the expression of the glycolysis-related proteins, Adh5, Eno1 and Gapdh notably increased, suggesting an activation of glycolysis in Cd stimulation; and Myh7, Actg1, Myh6, Myl12b, Myh4, and Myh1, which are involved in tight junctions, were suppressed. Histone proteins, such as H2afy, Hist1h3b and Hist1h4a were also markedly down-regulated; these findings are consistent with those of previous studies⁵⁸. Moreover, the model demonstrates a potential crosstalk between these pathways: protein Actg1 (cytoplasmic actin) is a connector of ribosomes, histones and tight junctions; and Gapdh (glyceraldehyde-3-phosphate dehydrogenase) is overexpressed, linking glycolysis and histones together. Gapdh also interacts with Actg1, suggesting the probable inhibitory function of Gapdh in the Cd response. Calm1, a Calmodulin, mediates the control of a large number of Ca^{2+} -driven enzymes, kinases and phosphatases involved in cytokinesis; it is a connector of tight junctions and histones. Interestingly, Calm1 was first overexpressed by more than 1.9-fold at 24 h and then inhibited at 48 h and 96 h. Calm4 (not shown in the model), another calcium-binding protein, shows a similar profile as that of Calm1, first being overexpressed by 2.2-fold at 24 h and 48 h and then down-regulated at 96 h. These results suggest that the level of calcium-binding proteins is crucial in protection against Cd. Associated with Calm1 is SOD3 (extracellular Cu/ZnSOD), an important anti-toxic protein protecting the extracellular space in biological systems. The expression of this protein increased more than 10-fold at all 3 time points, and the expression of SOD1 (not shown in model) increased more than 10-fold only at 96 h, suggesting that SOD1 might function as a backup enhancer that activates under long-term Cd exposure. The complementary expression profile of SOD1 and Calm4, SOD3 and Calm1 might suggest vital novel regulation signaling controlled by the SOD and Calm family.

In tight junctions, members of the Myosin-II family are inhibited by Cd, including Myh1, 4, 6, 7 and 12b⁶⁰; these proteins are key components in the assembly and contraction of muscle⁶¹. The model also simulates a connection possibility of the key differentially expressed proteins with significantly-expressed metabolic amino acids. This model shows 2 possible paths linking amino acid metabolism pathways, including valine, leucine and isoleucine degradation; alanine, aspartate and glutamate metabolism; and arginine and proline metabolism. One possible path is associated with map3k15 (mitogen-activated protein kinase kinase kinase 15 Gene), ccbl1 (cysteine conjugate-beta lyase 1), and Lao1 (L-amino acid oxidase 1); these proteins might be involved in oxidative stress processes^{62,63}. The other pathway is through myl3 (myosin, light polypeptide 3), Dmd (Dystrophin), Nos1 (nitric oxide synthase 1), and Asl (argininosuccinate lyase), an important player in the arginine and proline metabolic pathway, as indicated by the KEGG database. This pathway might be related to NO regulation, a messenger of neurotransmitter signals in the nervous system^{64–67}.

In summary, the model illustrates a possible regulation network consisting of important proteome and metabolomic changes upon Cd stimulation. It suggests that the ribosome was likely down-regulated, glycolysis was likely activated and tight junctions were likely suppressed. These pathways were interlinked with down-regulated histone complex. Compromising tight junctions is likely to interfere with oxidative stress and NO regulation, and in turn be responsible for up-regulated calcium-binding related proteins and downstream up-regulated amino acid metabolism. Noteworthy, some previous studies have reported response of Cd in evolutionally distant

species rather than clam^{20,51}. Although we cannot directly compare the changes at molecular level, genes in clam do not necessarily have homologous in distant related species, and Cd does not necessarily effect same genes in different species. But we have found relationship at the biological process level, such as tight junction system and glycolysis system of both tomato and clam were significantly affected by Cd exposure. This study provides a potential regulation network model consistent of these biological processes based on mammalian interactomes, further investigations should be done to validate the key molecules and interactions.

Conclusion

Cd toxicity is a multigenetic, complex process. The use of proteomics and metabolomics has provided insight into the comprehensive landscape of change at the molecular level. A total of 365 proteins and 12 metabolites were significantly changed after Cd exposure. Interestingly, the metabolic biomarkers were consistent in the time series analysis, suggesting a conserved mechanism of Cd toxicity. To explore the core mechanism, we focused on differentially expressed proteins, independent of the exposure time. Bioinformatic analysis identified a core connected regulatory network consisting of 15 down-regulated proteins; these proteins were functionally involved in histone function, ribosomal process and tight junctions. This result might suggest that histones and ribosomes are the primary targets of Cd. Glycolysis, sulfur metabolic process and Ca^{2+} homeostasis were also detected in our study, and these processes were connected with the core network, suggesting the mechanisms of Cd toxicity were interlinked. In contrast, the myosin complex was significantly suppressed, leading to the abnormal function of the tight junction, which could be a potential mechanism to disturb the neural and osmotic function of tissue. This work is the first multi-omics study of the basic mechanism of Cd toxicity and the results provide a list of potential key regulators of Cd toxicity in blood clam *T. granosa*. The continuous functional investigation of these proteins could eventually solve the key targets of Cd toxicity.

Methods

Clam, exposure conditions, and sample collection. Adult blood clams *T. granosa* (shell length: 2.8–3.2 cm) were collected from a local culturing farm in Ningbo, China. These clams were allowed to acclimate in aerated seawater (20 °C, 30‰ salinity, collected from a pristine environment) in the laboratory for 2 weeks and fed with *Platymonas subcordiformis* at 2% of the tissue dry weight per day. After acclimatization, all the blood clams were divided into three 50-L tanks (three cadmium exposure levels 0, 25, 250 µg/L), each containing sixty blood clams. For Cd exposure, the blood clams were exposed to dissolved 25 and 250 µg/L Cd^{2+} , respectively. Cadmium solution (12.5 g/L) were prepared from $\text{Cd}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ (analytical grade), 0.1 L and 1 L cadmium solution were added to 49.9 L and 49 L sea water for final concentrations of 25 and 250 µg/L Cd^{2+} , respectively. The low experimental concentration (25 µg/L) of Cd^{2+} can be found in heavily polluted seawaters along the coast of China⁵¹. At each sampling time point (24, 48 and 96 hpe), fifteen blood clams from the control and Cd-exposed groups were immediately dissected for the gill tissues (~100 mg). In total, 180 clams representing five biological replicates and four time points (0 h as control) of three experiments were used for the following cadmium concentration determination, proteomic and metabolomics analysis. All of the gill tissues were flash frozen in liquid nitrogen and then stored at –80 °C.

Cadmium concentration determination. Five gill samples were dried and digested according to Wu *et al.* lab protocol²⁰. PE Analyst-800 was used to quantify the Cd concentration. For internal quality control, we have used the tissue of GBW08571 marine muscle as certified reference⁶⁸. Three individual spiking experiments were performed to recovery the target elements. Restriction level for Cd was set to 95.6–104.3%. One-way ANOVA followed by an unpaired student t-test was applied. *P*-value < 0.05 was considered as significant. SPSS19 software was used for the statistical analysis. The blood clams (*T. granosa*) here are commercially cultured animals, and all the experiments were conducted in accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. The study protocol was approved by the Experimental Animal Ethics Committee of Zhejiang Wanli University, China.

Metabolomics analysis. Modified extraction protocol⁶⁹ was used to obtain polar metabolites from gills. Briefly, 100 mg gill tissue was homogenized (Percellys 24, Bertin Technologies, France) and mixed with 4 mL/g methanol, 0.85 mL/g water, and 2 mL/g chloroform, then shaken and centrifuged at 4 °C, 3000 g, for 5 minutes. The supernatant substance was then mixed with 2 mL/g chloroform and 2 mL/g water, vortexed and centrifuged for 10 minutes, at 3000 g, 4 °C. The methanol/water layer was dried, re-suspended in 600 µL of 100 mM phosphate buffer (Na_2HPO_4 and NaH_2PO_4 with 0.5 mM TSP (sodium 3-trimethylsilyl [2, 2, 3, 3-²D₄] propionate) as the internal standard for the chemical shift, pH 7.0) in D_2O , vortexed and centrifuged for 5 minutes at 3000 g, 4 °C. The supernatant substance was used for NMR analysis.

One-dimensional ¹H NMR spectra of gill extracts were acquired on a Bruker AV 500 NMR spectrometer performing at 500.13 MHz at 298 K³⁰. All the ¹H NMR spectra were acquired using water suppressed NOESYPR1D pulse sequence (recycle delay-90°-*t*₁-90°-*t*_m-90°-acquisition). Sixteen K data points of the NMR signal were acquired with a 6009.6 Hz spectral width, an acquisition time of 2.7 s, and a recycle delay time of 3.0 s using 128 scans. The water suppression was achieved with the selective irradiation on the water peak during a fixed interval *t*₁ of 4 µs and the mixing time *t*_m of 100 ms. Then, the datasets were zero-filled to 32,768 points, and exponential line-broadenings of 0.3 Hz were applied before Fourier transformation. All ¹H NMR spectra were phase- and baseline corrected and chemical shift was calibrated to TSP (δ 0.00) manually using TopSpin (version 2.1, Bruker). Then, each spectrum was segmented into 0.005 ppm bins between 0.2 and 9.1 ppm with bins from 4.72 to 4.96 ppm (water) excluded from all the NMR spectra. The area for each segmented region was calculated and normalized to the total integrated area of the spectra. All the NMR spectra were generalized by log transformation to stabilize the variance across the spectral bins and to increase the weightings of the less intense peaks.

Partial least squares discriminant analysis (PLS-DA) and orthogonal projection to latent structure with discriminant analysis (OPLS-DA) were performed on the NMR data of clam groups at 24 h, 48 h, 96 h against 0 h, respectively⁷⁰. The quality of the constructed PLS-DA models was assessed using cross-validation with five-way split Venetian blinds. A Q^2 score of >0.08 indicates that the PLS-DA model is significantly better than chance, while a score between 0.7 and 1.0 indicates that the model is highly robust and the corresponding Latent Variable weight plots of OPLS-DA can usually present the contributive NMR bins (peaks) of metabolites for the separation between control and exposed groups⁷¹. The validity of each constructed OPLS-DA model was evaluated with the cross validation-ANOVA approach ($p < 0.05$). The OPLS-DA results were visualized in score plots to show the classifications and in corresponding loading plots to show the NMR spectral variables contributing to the classifications. Score plots were used for classification visualization and corresponding loading plots were used to show variables contribution. Then, MATLAB (V7.0, MathWorks Inc., Natwick, USA) was used to perform coefficient-coded loadings plots with an in-house-developed program and were color-coded with the absolute value of the correlation coefficients (r). The metabolites with absolute value of r above 0.602 were considered to be statistically significant ($p < 0.05$). Chemomx (Evaluation Version, Chemomx Inc., Edmonton, Alberta, Canada) was used for metabolites assignments.

Proteomics analysis. Five biological replicates were mixed, then divided into two experimental replicates of each time point of the 250 $\mu\text{g/L}$ Cd^{2+} group were prepared. Total protein of each gill sample was powdered with liquid nitrogen. Mixture of the powder and 9 M urea were centrifuged at 15,000 g for 15 min at 4 °C. Bradford method was used for supernatant quantification⁷². The samples were then centrifuged at 15,000 g for 15 min at 4 °C. The supernatant was collected and quantified by the Bradford method. After applying 20- μg samples to a 12% SDS-PAGE gel, the gel was visualized by CBB stain according Candiano's protocol for checking the protein⁷³.

100 mg of protein for each sample were mixed with a dissolution buffer of AB sciex (Foster City, CA, USA), digested with trypsin and labeled with iTRAQ reagents 8-plex kit (AB sciex). Samples and independent biological replicates were labeled 113–117, 114–118, 115–119, and 116–121 Da, respectively. Labeled samples were then pooled and dried in an Eppendorf vacuum concentrator.

Agilent 1200 HPLC was used for sample purify, Exigent nanoLC-Ultra 2D system (AB SCIEX) was used for sample separation. Triple TOF 5600 mass spectrometer and a Nanospray III source (AB SCIEX) was used to perform Mass spectrometer data acquisition.

Protein Pilot (v4.5) was used for raw data processing against the *T. granosa* transcription and *C. gigas* genome database⁷⁴. Protein identification was performed with the search option of emphasis on biological modifications. The database search parameters were as follows: the instrument was TripleTOF 5600, iTRAQ quantification, cysteine modified with iodoacetamide; and biological modifications were selected as ID focus and trypsin digestion. Peptides with global false discovery rate (FDR) $< 1\%$ were used for further protein annotation. Average ration of 114/118, 113/117 at 24 h, 115/119 and 113/117 at 48 h, and 116/121 and 113/117 at 96 h were assigned as fold change value. Proteins with a corrected fold change of > 1.50 or < 0.67 in any duplicate were considered to be significantly differentially expressed.

Bioinformatics Analysis. Blastp was used against the *Mus musculus* proteome database at uniprot (<http://www.uniprot.org/>). The Protein with best identity to query protein among the best 10 hits with $e\text{-value} < 1e^{-10}$ was considered as homologous of the gill proteins in mouse. DAVID toolkit was used for gene ontology enrichment analysis. Associated KEGG signaling pathways were calculated using fish's exact tests. The STRING database (version 9.1)⁷⁵ was used to obtain protein-protein interaction information with default score setting. Potential paths of differentially expressed proteins and metabolic compounds were searched in the knowledge-driven database with a shortest-path algorithm. Cytoscape (<http://cytoscape.org/>) version 3.1 was used for regulation network construction.

References

- Bargagli, R. Trace metals in Antarctica related to climate change and increasing human impact. *Rev. Environ. Contam. Toxicol.* **166**, 129–174 (2000).
- Zhang, G., Liu, D., Wu, H., Chen, L. & Han, Q. Heavy metal contamination in the marine organisms in Yantai coast, northern Yellow Sea of China. *Ecotoxicology* **21**, 1726–1733 (2012).
- Chora, S. *et al.* Effect of cadmium in the clam *Ruditapes decussatus* assessed by proteomic analysis. *Aquat. Toxicol.* **94**, 300–308 (2009).
- Isani, G. *et al.* Cadmium accumulation and biochemical responses in *Sparus aurata* following sub-lethal Cd exposure. *Ecotoxicol. Environ. Saf.* **72**, 224–230 (2009).
- Han, F. X. *et al.* Industrial age anthropogenic inputs of heavy metals into the pedosphere. *Naturwissenschaften* **89**, 497–504 (2002).
- Shi, W. *et al.* Ocean acidification increases cadmium accumulation in marine bivalves: a potential threat to seafood safety. *Sci. Rep.* **6**, 20197 (2016).
- Huang, H., Wu, J. Y. & Wu, J. H. Heavy metal monitoring using bivalve shellfish from Zhejiang coastal waters, East China Sea. *Environ. Monit. Assess.* **129**, 315–320 (2006).
- Zhang, P. *et al.* iTRAQ-Based proteomics reveals novel members involved in pathogen challenge in sea cucumber *Apostichopus japonicus*. *PLoS One* **9**, e100492 (2014).
- Bebianno, M. J. & Serafim, M. A. Comparison of metallothionein induction in response to cadmium in the gills of the bivalve molluscs *Mytilus galloprovincialis* and *Ruditapes decussatus*. *Sci. Total. Environ.* **214**, 123–131 (1998).
- Roméo, M. & Gnassia-Barelli, M. Metal distribution in different tissues and in subcellular fractions of the Mediterranean clam *Ruditapes decussatus* treated with cadmium, copper, or zinc. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* **111**, 457–463 (1995).
- Bebianno, M., Nott, J. & Langston, W. Cadmium metabolism in the clam *Ruditapes decussata*: the role of metallothioneins. *Aquat. Toxicol.* **27**, 315–333 (1993).
- Roesijadi, G., Brubacher, L., Unger, M. & Anderson, R. Metallothionein mRNA induction and generation of reactive oxygen species in molluscan hemocytes exposed to cadmium *in vitro*. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* **118**, 171–176 (1997).
- Geret, F., Serafim, A., Barreira, L. & Bebianno, M. J. Effect of cadmium on antioxidant enzyme activities and lipid peroxidation in the gills of the clam *Ruditapes decussatus*. *Biomarkers* **7**, 242–256 (2002).

14. Jones, O. A., Dondero, F., Viarengo, A. & Griffin, J. L. Metabolic profiling of *Mytilus galloprovincialis* and its potential applications for pollution assessment. *Mar. Ecol. Prog. Ser.* **369**, 169–179 (2008).
15. Viant, M. R. *et al.* International NMR-based environmental metabolomics intercomparison exercise. *Environ. Sci. Technol.* **43**, 219–225 (2008).
16. Tuffnail, W., Mills, G. A., Cary, P. & Greenwood, R. An environmental ¹H NMR metabolomic study of the exposure of the marine mussel *Mytilus edulis* to atrazine, lindane, hypoxia and starvation. *Metabolomics* **5**, 33–43 (2009).
17. Zhang, L. *et al.* Metabolic responses in gills of Manila clam *Ruditapes philippinarum* exposed to copper using NMR-based metabolomics. *Mar. Environ. Res.* **72**, 33–39 (2011).
18. Liu, X. *et al.* Differential toxicological effects induced by mercury in gills from three pedigrees of Manila clam *Ruditapes philippinarum* by NMR-based metabolomics. *Ecotoxicology* **20**, 177–186 (2011).
19. Aru, V., Sarais, G., Savorani, F., Engelsens, S. B. & Marincola, F. C. Metabolic responses of clams, *Ruditapes decussatus* and *Ruditapes philippinarum*, to short-term exposure to lead and zinc. *Mar. Pollut. Bull.* **107**, 292–299 (2016).
20. Wu, H. & Wang, W. X. NMR-based metabolomic studies on the toxicological effects of cadmium and copper on green mussels *Perna viridis*. *Aquat. Toxicol.* **100**, 339–345 (2010).
21. Blaise, B. J. *et al.* Metabotyping of *Caenorhabditis elegans* reveals latent phenotypes. *Proc Natl. Acad. Sci. USA* **104**, 19808–19812 (2007).
22. Hughes, S. L., Bundy, J. G., Want, E. J., Kille, P. & Stürzenbaum, S. R. The metabolomic responses of *Caenorhabditis elegans* to cadmium are largely independent of metallothionein status, but dominated by changes in cystathionine and phytochelatins. *J. Proteome Res.* **8**, 3512–3519 (2009).
23. Buckley, B. A., Gracey, A. Y. & Somero, G. N. The cellular response to heat stress in the goby *Gillichthys mirabilis*: a cDNA microarray and protein-level analysis. *J. Exp. Biol.* **209**, 2660–2677 (2006).
24. Evans, C. *et al.* An insight into iTRAQ: where do we stand now? *Anal. Bioanal. Chem.* **404**, 1011–1027 (2012).
25. Leung, P. T., Wang, Y., Mak, S. S., Ng, W. C. & Leung, K. M. Differential proteomic responses in hepatopancreas and adductor muscles of the green-lipped mussel *Perna viridis* to stresses induced by cadmium and hydrogen peroxide. *Aquat. Toxicol.* **105**, 49–61 (2011).
26. Thompson, E. L. *et al.* A proteomic analysis of the effects of metal contamination on Sydney Rock Oyster (*Saccostrea glomerata*) haemolymph. *Aquat. Toxicol.* **103**, 241–249 (2011).
27. Corporeau, C., Tamayo, D., Pernet, F., Quéré, C. & Madec, S. Proteomic signatures of the oyster metabolic response to herpesvirus OsHV-1 μ Var infection. *J. Proteomics* **109**, 176–187 (2014).
28. Zhang, Y. *et al.* Proteomic basis of stress responses in the gills of the pacific oyster *Crassostrea gigas*. *J. Proteome Res.* **14**, 304–317 (2015).
29. Marrocco, C., Zolla, V. & Zolla, L. Meat quality of the longissimus lumborum muscle of Casertana and Large White pigs: metabolomics and proteomics intertwined. *J. Proteomics* **75**, 610–627 (2011).
30. Ji, C., Wu, H., Wei, L., Zhao, J. & Yu, J. Proteomic and metabolomic analysis reveal gender-specific responses of mussel *Mytilus galloprovincialis* to 2,2',4,4'-tetrabromodiphenyl ether BDE 47. *Aquat. Toxicol.* **140–141**, 449–457 (2013).
31. Wu, H. *et al.* Proteomic and metabolomic responses of clam *Ruditapes philippinarum* to arsenic exposure under different salinities. *Aquat. Toxicol.* **136**, 91–100 (2013).
32. Wei, L., Wang, Q., Wu, H., Ji, C. & Zhao, J. Proteomic and metabolomic responses of Pacific oyster *Crassostrea gigas* to elevated pCO₂ exposure. *J. Proteomics* **112C**, 83–94 (2014).
33. Kelly, A. & Stanley, C. A. Disorders of glutamate metabolism. *Ment. Retard. Dev. Disabil. Res. Rev.* **7**, 287–295 (2001).
34. Di Giulio, R. T. & Scanlon, P. F. Effects of cadmium ingestion and food restriction on energy metabolism and tissue metal concentrations in mallard ducks *Anas platyrhynchos*. *Environ. Res.* **37**, 433–444 (1985).
35. Chapatwala, K. D., Rajanna, E. & Desai, D. Cadmium induced changes in gluconeogenic enzymes in rat kidney and liver. *Drug Chem. Toxicol.* **3**, 407–420 (1980).
36. Yu, Q. H. & Yang, Q. Diversity of tight junctions TJs. between gastrointestinal epithelial cells and their function in maintaining the mucosal barrier. *Cell. Biol. Int.* **33**, 78–82 (2009).
37. Hew, K. W., Heath, G. L., Jiwa, A. H. & Welsh, M. J. Cadmium *in vivo* causes disruption of tight junction-associated microfilaments in rat Sertoli cells. *Biol. Reprod.* **49**, 840–849 (1993).
38. Shin, K., Fogg, V. C. & Margolis, B. Tight junctions and cell polarity. *Annu. Rev. Cell Dev. Biol.* **22**, 207–235 (2006).
39. Planelló, R., Martínez-Guitarte, J. L. & Morcillo, G. Ribosomal genes as early targets of cadmium-induced toxicity in *Chironomus riparius* larvae. *Sci. Total. Environ.* **373**, 113–121 (2007).
40. Norton, K. B. & Kench, J. E. Effects of cadmium on ribosomal protein synthesis in rat liver. *Environ. Res.* **13**, 102–110 (1977).
41. Gerich, J. E., Meyer, C., Woerle, H. J. & Stumvoll, M. Renal gluconeogenesis: its importance in human glucose homeostasis. *Diabetes Care* **24**, 382–391 (2001).
42. Rodríguez-Celma, J., Rellán-Alvarez, R., Abadía, A., Abadía, J. & Lopez-Millán, A. F. Changes induced by two levels of cadmium toxicity in the 2-DE protein profile of tomato roots. *J. Proteomics* **73**, 1694–1706 (2010).
43. Chung, N. P. & Cheng, C. Y. Is cadmium chloride-induced inter-Sertoli tight junction permeability barrier disruption a suitable *in vitro* model to study the events of junction disassembly during spermatogenesis in the rat testis? *Endocrinology* **142**, 1878–1888 (2001).
44. Kvint, K., Nachin, L., Diez, A. & Nyström, T. The bacterial universal stress protein: function and regulation. *Curr. Opin. Microbiol.* **6**, 140–145 (2003).
45. Loukehaich, R. *et al.* SpUSP, an annexin-interacting universal stress protein, enhances drought tolerance in tomato. *J. Exp. Bot.* **63**, 5593–5606 (2012).
46. Pathak, N. & Khandelwal, S. Oxidative stress and apoptotic changes in murine splenocytes exposed to cadmium. *Toxicology* **220**, 26–36 (2006).
47. Liu, J., Qu, W. & Kadiiska, M. B. Role of oxidative stress in cadmium toxicity and carcinogenesis. *Toxicol. Appl. Pharmacol.* **238**, 209–214 (2009).
48. Yoo, H. Y., Chang, M. S. & Rho, H. M. Heavy metal-mediated activation of the rat Cu/Zn superoxide dismutase gene via a metal-responsive element. *Mol. Gen. Genet.* **262**, 310–313 (1999).
49. Kim, J. H. *et al.* Expression profiles of seven glutathione S-transferase (GST) genes in cadmium-exposed river pufferfish (*Takifugu obscurus*). *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* **151**, 99–106 (2010).
50. Xu, B. *et al.* Calcium signaling is involved in cadmium-induced neuronal apoptosis via induction of reactive oxygen species and activation of MAPK/mTOR network. *PLoS ONE* **6**, e19052 (2011).
51. Choong, G., Liu, Y. & Templeton, D. M. Interplay of calcium and cadmium in mediating cadmium toxicity. *Chem. Biol. Interact.* **211**, 54–65 (2014).
52. Nussbaum, S., Schmutz, D. & Brunold, C. Regulation of assimilatory sulfate reduction by cadmium in *Zea mays* L. *Plant. Biol.* **88**, 1407–1410 (1988).
53. Nocito, F. F., Pirovano, L., Cocucci, M. & Sacchi, G. A. Cadmium-induced sulfate uptake in maize roots. *Plant. Physiol.* **129**, 1872–1879 (2002).
54. Astolfi, S., Zuchi, S. & Passera, C. Role of sulphur availability on cadmium-induced changes of nitrogen and sulphur metabolism in maize (*Zea mays* L.) leaves. *J. Plant. Physiol.* **161**, 795–802 (2004).

55. Clemens, S. Toxic metal accumulation, responses to exposure and mechanisms of tolerance in plants. *Biochimie* **88**, 1707–1719 (2006).
56. Scheerer, U. *et al.* Sulphur flux through the sulphate assimilation pathway is differently controlled by adenosine 5'-phosphosulphate reductase under stress and in transgenic poplar plants overexpressing γ -ECS, SO, or APR. *J. Exp. Bot.* **61**, 609–622 (2010).
57. Mohideen, K., Muhammad, R. & Davey, C. A. Perturbations in nucleosome structure from heavy metal association. *Nucleic Acids Res.* **38**, 6301–6311 (2010).
58. Pawlak, S. & Deckert, J. Histone modifications under environmental stress. *Biol. Lett.* **44**, 65–73 (2007).
59. Karaczyn, A. A., Golebiowski, F. & Kasprzak, K. S. Truncation, deamidation, and oxidation of histone H2B in cells cultured with nickel II. *Chem. Res. Toxicol.* **18**, 1934–1942 (2005).
60. Le Goff, X., Motegi, F., Salimova, E., Mabuchi, I. & Simanis, V. The *S. pombe* rlc1 gene encodes a putative myosin regulatory light chain that binds the type II myosins myo3p and myo2p. *J. Cell. Sci.* **113**, 4157–4163 (2000).
61. Saez, L. & Leinwand, L. A. Characterization of diverse forms of myosin heavy chain expressed in adult human skeletal muscle. *Nucleic Acids Res.* **14**, 2951–2969 (1986).
62. Suematsu, N. *et al.* Hyperhomocysteinemia alters cardiac substrate metabolism by impairing nitric oxide bioavailability through oxidative stress. *Circulation* **115**, 255–262 (2007).
63. Lee, J. I. *et al.* α -Keto acid metabolites of naturally occurring organoselenium compounds as inhibitors of histone deacetylase in human prostate cancer cells. *Cancer Prev. Res.* **2**, 683–693 (2009).
64. Brenman, J. E., Chao, D. S., Xia, H., Aldape, K. & Bredt, D. S. Nitric oxide synthase complexed with dystrophin and absent from skeletal muscle sarcolemma in Duchenne muscular dystrophy. *Cell* **82**, 743–752 (1995).
65. Milacic, M. *et al.* Annotating cancer variants and anti-cancer therapeutics in reactome. *Cancers* **4**, 1180–1211 (2012).
66. Croft, D. *et al.* The Reactome pathway knowledgebase. *Nucleic Acids Res.* **42**, D472–D477 (2014).
67. Kanehisa, M. *et al.* Data, information, knowledge and principle: back to metabolism in KEGG. *Nucleic Acids Res.* **42**, D199–D205 (2014).
68. Zhang, X. Investigation of pollution of Pb, Cd, Hg, As in sea water and deposit of Bohai Sea area. *Heilongjiang. Environ. J.* **25**, 87–90 (2001).
69. Li, L. *et al.* Uptake pathways and subcellular fractionation of Cd in the polychaete *Nereis diversicolor*. *Ecotoxicology* **2012**, **21**, 104–110.
70. Wu, H., Southam, A. D., Hines, A. & Viant, M. R. High-throughput tissue extraction protocol for NMR- and MS-based metabolomics. *Anal. Biochem.* **372**, 204–212 (2008).
71. Wu, H., Liu, X., Zhang, X., Ji, C., Zhao, J. & Yu, J. Proteomic and metabolomic responses of clam *Ruditapes philippinarum* to arsenic exposure under different salinities. *Aquat. Toxicol.* **136**, 91–100 (2013).
72. Lindon, J. C., Nicholson, J. K. & Everett, J. R. NMR spectroscopy of biofluid. *Ann. Rep. NMR Spectrosc.* **38**, 1–88 (1999).
73. Bradford, M. M. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* **72**, 248–254 (1976).
74. Candiano, G. *et al.* Blue silver: a very sensitive colloidal Coomassie G-250 staining for proteome analysis. *Electrophoresis* **25**, 1327–1333 (2004).
75. Shilov, I. V. *et al.* The Paragon Algorithm, a next generation search engine that uses sequence temperature values and feature probabilities to identify peptides from tandem mass spectra. *Mol. Cell Proteomics* **6**, 1638–1655 (2007).
76. Szklarczyk, D. *et al.* STRING v10: protein-protein interaction networks, integrated over the tree of life. *Nucleic Acids Res.* **43**, D447–D452 (2015).

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Author Contributions

Y.B. performed the whole experiment and wrote the manuscript. Z.L. and C.L. designed the experiments, wrote and revised the manuscript, and contributed reagents. J.C. and W.L. contributed reagents/materials/analysis tools. X.L. and W.Z. discussed the project and analyzed data. All authors contributed to the editing of the manuscript.

Additional Information

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