

Mitochondrial content is central to nuclear gene expression: Profound implications for human health

Rebecca Muir, Alan Diot and Joanna Poulton*

We review a recent paper in *Genome Research* by Guantes et al. showing that nuclear gene expression is influenced by the bioenergetic status of the mitochondria. The amount of energy that mitochondria make available for gene expression varies considerably. It depends on: the energetic demands of the tissue; the mitochondrial DNA (mtDNA) mutant load; the number of mitochondria; stressors present in the cell. Hence, when failing mitochondria place the cell in energy crisis there are major effects on gene expression affecting the risk of degenerative diseases, cancer and ageing. In 2015 the UK parliament approved a change in the regulation of IVF techniques, allowing Mitochondrial replacement therapy to become a reproductive choice for women at risk of transmitting mitochondrial disease to their children. This is the first time that this technique will be available. Therefore understanding the interaction between mitochondria and the nucleus has never been more important.

Keywords:

ageing; gene expression; gene regulation; mitochondria; mitochondrial biology; mtDNA; single cell analysis

DOI 10.1002/bies.201500105

Nuffield Department of Obstetrics and Gynaecology, University of Oxford, John Radcliffe Hospital, Oxford, UK

*Corresponding author:

Joanna Poulton
E-mail: joanna.poulton@obs-gyn.ox.ac.uk

Abbreviations:

AS, alternative splicing; GWA, genome wide association; GWAS, genome wide association studies; HIF, hypoxia inducible factor; OXPHOS, oxidative phosphorylation; SDH, succinate dehydrogenase; SNPs, single nucleotide polymorphisms; TCA cycle, the citric acid cycle.

Introduction

Guantes et al. [1] begin by observing that gene expression is noisy in a population of genetically identical cells, and that this has consequences for cell behavior [2]. Something else must be affecting expression, either downstream or upstream. Reasoning that gene expression uses a high proportion of cellular energy (indeed, 75% in *E. coli* [3]), the authors demonstrate that the cellular content of mitochondrial proteins seems to exert far-reaching effects on nuclear gene expression. They previously showed that heterogeneity in the volume of mitochondria in individual cells is a source of variability in transcription rate [2], which in turn is highly sensitive to the concentration of ATP. This links transcription to mitochondrial function [2]. In the current study, Guantes et al. attribute the cause of variability in global gene expression to energetic modulation by the mitochondrial network [1]. They start by showing that mitochondrial mass, measured as the integrated signal of a mitochondrial dye called CMXRos, is proportional to the abundance of a number of mitochondrial proteins involved in energy production, referring to this as “mitochondrial content”. They measure the mitochondrial content of individual cells together with either (a) the protein content, (b) the RNA polymerase II content/activity, or (c) the chromatin modification. They then investigate the variation independent of the mitochondria, and the co-variation with mitochondria; finally they calculate the mitochondrial contribution to the variability of (a), (b), and (c).

Their results show that mitochondrial content accounts for half of the protein variability in a cell population. This genome-wide effect is due to an effect at four different levels of the gene expression: 1) on chromatin activation via the acetylation of histones; 2) on transcription activity, 3) on alternative splicing (AS), 4) on protein synthesis. This is illustrated in Fig. 1.

All of these effects are likely to be due to the ATP levels linked to the mitochondrial content and function of the cell. Although it seems counterintuitive to think that a global

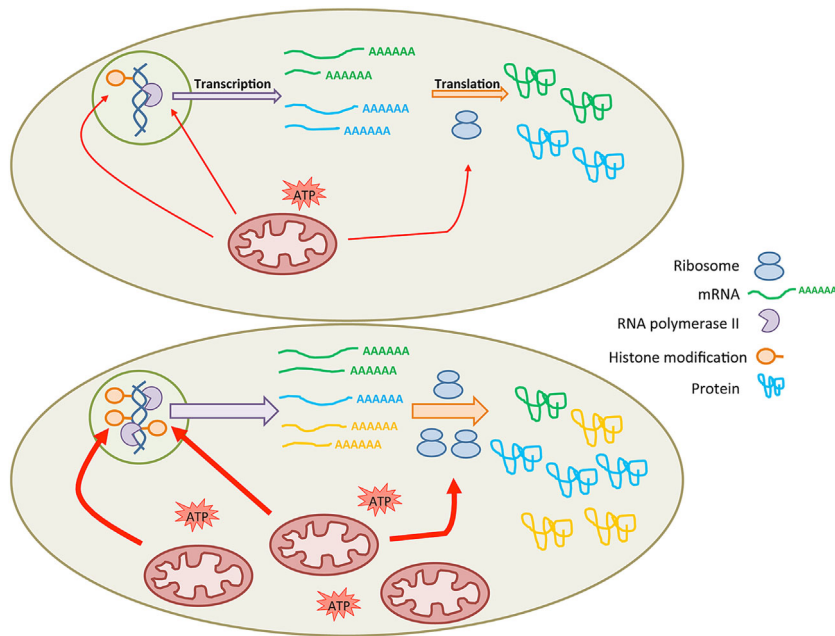


Figure 1. Guantes et al. have shown that mitochondria are an important factor influencing the cell's protein content variability. Mitochondria affect the histone profile, the transcription level, the alternative splicing, and the mRNA translation into proteins (upper panel). An increase in mitochondrial content, i.e. ATP content, of the cell will lead to an activation of the chromatin by histones acetylation and methylation, a greater number of genes transcribed by the RNA Polymerase 2, a different pattern of alternative splicing and a higher number of ribosomes for protein translation (lower panel). This results in different sets of proteins between cells with a high content of mitochondria and cells with a lower content of mitochondria.

constraint such as energy can be responsible for the substantial individual variation between cells, it is apparent that mitochondria are heterogeneous in structure, number, and function across a population of cells, especially in pathological states. It follows that energy production will be heterogeneous. Gene expression has multiple steps with very different energy demands along the way. Mitochondrial variation is likely to play a central role in diseases because of its striking effects on the transcriptional profile of cells when different levels of mutant mtDNA are present [4].

ATP levels are often used to assess mitochondrial function, but this is problematic because homeostatic mechanisms obscure changes. ATP flux is a better reflection of changes in mitochondrial function, but it is difficult to measure accurately. This limitation has masked the central role of mitochondrial variation. Most of the data implicating mitochondria in disease arise from genetic studies of mitochondrial DNA (mtDNA). Hence, the methodology of Guantes et al. is of particular note because of the primary focus on mitochondrial mass and gene expression, rather than on ATP/mtDNA.

This paper is also one of the first to describe non-genetic variability in human cell lines that exhibit characteristics of human cancer and disease. We hypothesize that the differences in gene expression have important implications for multifactorial disease and cancer. Additionally, we believe that understanding the mitochondrial influence on gene expression adds

another dimension to the social and ethical issues associated with recent advances in nuclear transfer techniques.

Mitochondria are responsible for nuclear genome complexity

The nuclear and mitochondrial genomes in eukaryotes have always been co-dependent, essentially guiding each other's evolution through deep time as a result of the original endosymbiosis [5]. In the eukaryotic cell, mitochondria produce energy via oxidative phosphorylation (OXPHOS), a chain of chemical reactions resulting in ATP production. Unlike chromosomal genes, mitochondrial genes are packed into a tiny circle of DNA containing 13 protein reading frames encoding OXPHOS-related proteins. OXPHOS is hypothesized to be integral to the evolution of multicellular life, because cells with a tiny energy budget have to restrict their genome size and complexity [3]. From the point of view of a cell, the expression of genes is enormously expensive, and yet it is both greater and more diverse in eukaryotes compared to prokaryotes.

The reciprocal interaction between mitochondria and the nuclear genome is archaic and highly evolved [3], leading Guantes et al.

to explore the mitochondrial influence on nuclear gene expression, characterizing the impact that mitochondria have on nuclear genes from a quantitative perspective.

Differences in genetically identical cell populations [6] are found even when grown in identical environments [7]. Previous research has shown that gene and protein expression levels can vary significantly among the same cell population with different functional results [8, 9], but Guantes et al. have gone a step further by using single-cell analysis with a focus on gene expression and mitochondrial content. Single-cell analysis is now a crucial tool for understanding the complexity behind variations in cell populations. Genetic and phenotypic heterogeneity among cells is the rule, not the exception [10]. Much of the literature has focused on microbes rather than human cells. Therefore this paper is an important step towards identifying the causes of noise in human disease [11].

Mitochondria have been implicated in cancer and a wide range of metabolic and degenerative diseases, including ageing itself. In many cases the links are apparent but incomplete. The new findings presented by Guantes et al. have the potential to fill gaps across the board because of their focus on several levels of gene regulation [1]. First we will highlight how these regulatory mechanisms are influenced by the mitochondria. Then we will discuss how mitochondria are implicated in cancer, multifactorial disease, and neurodegenerative disease, offering our interpretation of how the research of Guantes et al. can be applied to these situations.

Mitochondria drive alternative splicing

AS is crucial to Iborra and co-workers main finding that mitochondria contribute over half of protein diversity in the cell. AS allows the economical storage of genetic information, and can result in more transcripts than the number of genes in a human's entire collective genome [12]. Although this mechanism allows tissue specificity and evolutionary flexibility, the high potential for protein diversity brings with it the potential for malignancy. A common theme in the "hallmarks of cancer" is a switch in splicing patterns associated with a more aggressive, invasive cancer phenotype. Misregulation of an angiogenesis splice form has been found, as have splice forms that contribute to resistance to apoptosis [13]. Diseases associated with AS are well-documented [14, 15]. Guantes et al. postulate that mitochondria will affect AS either through impacting the energy supply of the AS molecular machinery, or indirectly through mechanisms that are highly energy dependent and play a role in AS, such as the epigenetic mechanisms of chromatin remodeling [16].

Mitochondria influence epigenetic chromatin modifications

Epigenetic modification is one of the regulatory steps in gene expression, and its mechanisms fall broadly into three categories: methylation, histone modification, and RNA-associated silencing [17]. Chinnery et al. speculated that the varying severity seen in patients with mitochondrial disease might be partly explained by the downstream effects caused by modification of nuclear DNA [18]. Guantes et al. provide evidence that mitochondria play a role in epigenetic modification: they discovered three histone modifications that co-vary with mitochondrial content and are linked with chromatin activation, namely H4K16 [19], H3K4me3, and H3K36me2 [20]. However, they did not find links with histone modifications involved in chromatin repression. Such acquired epigenetic changes [21] may link environmental risk factors to important diseases including human cancer. Guantes et al. also comment that the mitochondria provide metabolites needed for epigenetic modification, and therefore that a lack of mitochondrial metabolites may be rate-limiting and affect the frequency of nuclear gene modification [22]. Their findings add substance to the notion of a "mitocheckpoint" in which damage to mitochondria affects the availability of S-adenosyl methionine, thus modulating methylation of the nuclear genome [23]. Although this field is in its infancy, mitochondrial epigenetics may contribute to the effect that variance in mitochondrial number and functionality has on the nucleus, affecting penetrance of disease [13].

Mitochondria regulate expression of cancer genes

After Otto Warburg observed that cancer cells rely on glycolysis instead of OXPHOS for energy production [24], mitochondria have been thought to be implicated in carcinogenesis. Later discoveries continued to place mitochondria at the forefront of

cancer research as it was found that nuclear-mitochondrial and mitochondria-to-nucleus signal integration, control of apoptosis, and various metabolic pathways were under mitochondrial control. Notably, up-regulation of hypoxia inducible factor (HIF) and of HIF-responsive genes is a feature of many cancers. Rarely, mutations in genes encoding TCA cycle enzymes [25], such as the mitochondrial complex succinate dehydrogenase (SDH) [26], are central to neoplastic transformation. MtDNA analysis reveals that most or all cancers accumulate somatic mitochondrial as well as nuclear genome mutations [27]. The origins and impact of cancer-associated mutations in mtDNA were unclear, and some investigators therefore suggested that abnormal mitochondrial function might cause oncogenic transformation. If this were widely applicable, therapeutic approaches should aim to augment and not inhibit mitochondrial function [28].

However, recent sequencing that extensively explores the somatic alterations in the mitochondrial genomes of cancers suggests that the reverse is true for the majority of cancers. Ju et al. found that most mitochondrial genome mutations have no discernible effect [29]. The apparent high frequency of "passenger" mtDNA mutations in tumors could arise from their stochastic accumulation. Furthermore, DNA mutations that damage normal mitochondrial activity were less likely to be maintained in cancer cells. While there are undoubtedly additional metabolic alterations and dependencies of cancer cells that may be exploited to improve anticancer therapy, it is clear that most cancers need their mitochondria for proliferation, maintenance and invasive behavior. The increased risk conferred by impaired SDH activity is therefore the exception and not the rule [26]. Hence for most tumor types, these new findings suggest that drugs that impair mitochondrial function may slow down cancer progression by influencing nuclear gene transcription. Studying the mitochondrial activity and transcriptional profile of primary and cancer cell lines will help clarify the relationship between mitochondria and cancer cell behavior. Further work is needed to determine how this relates to cancers that arise from dysregulated stem cells.

Mitochondria regulate genes in multifactorial disease

Chromosomes segregate in accordance with Mendel's laws, and in this process, genes are subject to rigorous selection pressures, leading to disease-causing genes being eliminated from the population. Mitochondrial genes work differently. There are thousands of copies of mtDNA in every nucleated cell, and these have a high mutation rate, and a maternal mode of inheritance. Furthermore, healthy and mutant mtDNA are able to co-exist in the same cell, in a state called heteroplasmy. The "Mendelian disease" paradigm led to Genome-Wide Association Studies (GWAS) gaining immense popularity within genetics research. While this generated useful results for some diseases, many other diseases remain relatively intractable [30]. This may be because the mechanism behind many diseases is actually bioenergetic, given the strong links between mtDNA and some types of disease [31]. The supposedly "genome-wide" GWAS techniques typically

did not even consider mitochondrial DNA. To overcome this failing, GWAS investigators then set out to explore the effects of mtDNA single nucleotide polymorphisms (SNPs) in multifactorial diseases. However, several seemingly well-designed studies failed to detect associations between disease-associated variants located in the mitochondrial genome and diabetes [32–34].

This is because mtDNA poses several problems. Firstly, these studies could have missed heteroplasmic mtDNA mutants in blood, which often do not reflect the load in tissues, such as the insulin-secreting β cells of the pancreas. In β cells they may reach levels sufficient to change transcriptional profile while blood levels may decline to undetectable levels [35]. Secondly, a reproducible association may be overlooked if SNPs are considered as independent entities, the default position for GWAS studies. There are several examples where the context of mtDNA variants may affect the risk of disease that they confer [36, 37]. Independence of SNPs is less appropriate for mitochondrial than for nuclear SNPs, because intermolecular recombination is negligible in the context of maternal inheritance. In some [36, 37] but not all [38] such examples, the group of specific mtDNA SNPs implicated in conferring risk of disease have been identified. One way to probe groups of SNPs, which co-segregate because they are ancient associations, is to consider mtDNA haplogroups (in which people of different geographical regions have mtDNA variants which are population-specific) [39]. However, mtDNA haplogroups may be less helpful [40, 41] for rapidly mutating susceptibility variants [42] such as the OriB variant [43]. This variant is situated within an origin of replication (OriB) [44]. It lies within a homopolymeric C tract in the D-loop region of the mitochondrial genome. It is unique because SNPs in the local mtDNA sequence that affect the risk conferred interrupt the homopolymeric tract and compromise its secondary structure. Intuitively such changes should affect the function of the origin of replication, potentially being mildly deleterious [45]. We demonstrated a significant association between sequence variation around OriB and type 2 diabetes, initially in a UK population [40]. Despite being missed in the large GWAS studies [32] the association was confirmed in a large meta-analysis of Europid [43] and other populations [45] that took the local sequence into account. This variant predisposes to thinness, both at birth [46] and in young adults [47], helping to explain the link between low birth weight and diabetes.

Hudson et al considered haplotypes when they re-investigated mtDNA SNPs in 38,638 individuals with 11 major late-onset diseases and 17,483 controls in a GWAS study [39]. Their data suggests that deleterious variants that are associated with one or more common diseases are escaping selection. Like the OriB variant (which was not investigated in their study), many of these mutations emerged recently. They postulated that these recent mutations interact with nuclear loci and modify the risk of developing multiple common diseases. In line with Iborra's findings, these apparently detrimental variants could impair global nuclear gene expression, potentially contributing significantly to the pathogenesis. The next step will be to investigate single cell transcripts and protein profiles relative to heteroplasmic variants, and the protein profile in the OriB variant compared to controls.

Mitochondria regulate genes associated with degenerative diseases and ageing

Mitochondria are thought to play an important role in neurodegeneration and ageing. Significantly, Attardi showed that fibroblast respiratory chain function declines with age [48]. Several authors reported accumulation of low levels of mtDNA rearrangements (the “common deletion”) in Alzheimers [49], Parkinson's disease [50] and ageing [51], in failing and ageing hearts [52, 53], and even in human oocytes [54]. This led to the “vicious circle” theory of mitochondrial ageing, in which somatic mutation of mtDNA engenders respiratory chain dysfunction, enhancing the production of DNA-damaging oxygen radicals and hence mtDNA damage [55]. However, the levels of mutant mtDNA detected were often extremely low (<1%) compared with the levels found in bonafide mitochondrial disease [56]. Such levels are more likely to reflect an underlying problem than to be damaging in themselves. More recent work on neurodegeneration implies disturbances in cellular and organellar quality control mechanisms and in mitochondrial dynamics (mitochondrial shape, size, distribution, movement, and anchorage). Mitochondrial dynamics are central to mitophagy (the type of quality control among several that is most likely to maintain mtDNA), and genes required for mitophagy are involved in familial Parkinson's disease [57]. We recently showed that fibroblast mitophagy declines with the age of the donor [58]. This again implicates declining mitochondrial function as important in the ageing process, and potentially explains the accumulation of mtDNA deletions in human ageing. There are additional reasons for thinking that energy metabolism might be important in neurodegeneration: for instance, more gene products implicated in neurodegeneration are targeted to the mitochondria than expected by mere chance alone (23% compared with 8%) [59]. Hence mitochondrial defects may be important in neurodegenerative disorders.

The most convincing recent evidence that mitochondria are involved in ageing came from studies of mtDNA maintenance. Larsson and co-workers generated a mouse with a proofreading defect in polymerase gamma, a protein that is essential for all mitochondrial DNA replication. Unexpectedly the main phenotype of this mouse was premature ageing [60], and this positioned the accuracy of mitochondrial DNA replication at the center of the ageing debate. Subsequent studies, however, did not support a simplistic interpretation of the data largely because of the inadequacies of the hypotheses being tested [61]. Even if mitochondrial damage were important, how did this translate into effects on ageing? Reactive oxygen species, an essential component to the vicious circle of ageing did not accumulate [62]. Neither did the common deletion, widely considered to be a hallmark of mitochondrial DNA damage in humans [63]. Iborra's hypothesis that mitochondria are central to gene expression provides a first step. But if a decline in oxidative phosphorylation were sufficient to cause premature ageing, should it not be a common feature of mitochondrial disease? Rather, the between-cell variability in mitochondrial function present in this mouse, which is already known to have profound effects [64], is the more

plausible answer. Again, use of the impressive range of single cell techniques that are now available to explore these questions will ultimately confirm or refute this claim. Indeed, investigating the activity level of telomerase in cells with a high content of mitochondria or ATP and conversely in cells with less energy could bring together the role of mitochondria and telomerase in ageing. Given our results showing a decrease in mitochondrial quality control with age, the older cells may have an altered energetic status and thus differ in net protein activities, among them telomerase.

Social and ethical insights

In this essay we have seen that mitochondria alter the nuclear gene expression profile of cells. This concept takes its lead from the knowledge that a high amount of mtDNA sequence variance exists in the human population [65] and that over evolutionary time, mitochondrial-nuclear interactions become highly specific [66]. These findings are important for understanding the ethical ramifications of the UK Parliament approval of “mitochondrial donation” or “mitochondrial replacement therapy” [67–69]. These terms describe an in vitro fertilization-based technique for reducing the dose of mutant mtDNA in the pre-implantation embryo. Successful experiments have been carried out in mice [70] and monkeys [71] and are technically possible in human embryos [72]. Concerns have been expressed that nuclear gene expression may need to be compatible with the mtDNA of the donor [73], and that the mtDNA of the donor should be closely related to that of the recipient [68, 74]. However, the concern raised by the findings of Guantes et al., that the mitochondria themselves may be important regulators of nuclear genes, has barely been considered. The view that “all you need is enough energy” and that mitochondrial transfer is equivalent to “changing a laptop battery” is inadequate [67]. Human development involves a carefully choreographed sequence of expressed genes, and in the case of the respiratory chain, these are encoded by both nuclear and mtDNA. As Guantes et al show, ATP is a key regulator of gene expression. Fundamentally altering the energetics is likely to affect the balance of the complex processes underlying health and personality. The UK parliament was briefed with the information that “Mitochondria have their own separate DNA, which carries just a few genes. All of these genes are involved in energy production but determine no other characteristics. And so, any faults in these genes lead only to problems in energy production... all available scientific evidence indicates that the genes contributing to personal characteristics and traits come solely from the nuclear DNA” (http://www.hfea.gov.uk/docs/2014-10-01_Mitochondrial_donation_an_introduutory_briefing_note_-_final.pdf). However, the data presented by Guantes et al suggest that mitochondria are determinants of important traits including multifactorial disease, cancer, and ageing. Mitochondrial replacement therapy may well be a viable strategy for fixing the energy deficits in mitochondrial disease, but the donated mitochondria are likely to influence other characteristics. In future, regulators would be well advised to ensure that the mitochondrial and nuclear genomes of donor and recipient

are sufficiently matched, as this may be critical to the success of mitochondrial replacement therapy [68, 73, 74].

Conclusions and outlook

Guantes et al. have begun to explain the genetic mechanisms by which ATP alter the cell, namely epigenetics and alternative splicing, and we have shown that their central tenet can be applied to established theories in cancer metabolism and ageing. This research has revealed that the mitochondria, the ancient organelles once seen simply as generators of ATP, affect cellular longevity and health by affecting nuclear transcription and translation. Future work should focus on verifying the links between the energetic status of the cell and its individual portfolio of expressed genes in health and disease. Outcomes of such studies might open promising new therapeutic strategies. Moreover, they will improve the safety of mitochondrial replacement therapy by helping to clarify the cross-talk between mitochondria and the nucleus.

Acknowledgments

The authors thank Karl Morten and Iain McLean for ideas and helpful feedback, and Stephen Kennedy for support. This work was supported in part by the MRC (MR/J010448/1), the Wellcome Trust (0948685/Z/10/Z), the NewLife Foundation (SG/14-15/11), the Angus Memorial Mitochondrial Fund, and the NHS Specialized Services Rare Mitochondrial Disorders Service.

The authors have declared no conflict of interest.

References

1. Guantes R, Rastrojo A, Neves R, Lima A, et al. 2015. Global variability in gene expression and alternative splicing is modulated by mitochondrial content. *Genome Res* **25**: 633–44.
2. das Neves RP, Jones NS, Andreu L, Gupta R, et al. 2010. Connecting variability in global transcription rate to mitochondrial variability. *PLoS Biol* **8**: 1000560.
3. Lane N, Martin W. 2010. The energetics of genome complexity. *Nature* **467**: 929–34.
4. Picard M, Zhang J, Hancock S, Derbeneva O, et al. 2014. Progressive increase in mtDNA 3243A>G heteroplasmy causes abrupt transcriptional reprogramming. *Proc Natl Acad Sci USA* **E42**.
5. Greiner S, Bock R. 2013. Tuning a ménage à trois: co-evolution and co-adaptation of nuclear and organellar genomes in plants. *BioEssays* **35**: 354–65.
6. Raj A, van Oudenaarden A. 2008. Nature, nurture, or chance: stochastic gene expression and its consequences. *Cell* **135**: 216–26.
7. Raser JM, O'Shea EK. 2005. Noise in gene expression: origins, consequences, and control. *Science* **309**: 2010–3.
8. Cohen AA, Geva-Zatorsky N, Eden E, Frenkel-Morgenstern M, et al. 2008. Dynamic proteomics of individual cancer cells in response to a drug. *Science* **322**: 1511–6.
9. Snijder B, Pelkmans L. 2011. Origins of regulated cell-to-cell variability. *Nat Rev Mol Cell Biol* **12**: 119–25.
10. Kalisky T, Blainey P, Quake SR. 2011. Genomic analysis at the single-cell level. *Annu Rev Genet* **45**: 431–45.
11. Maheshri N, O'Shea EK. 2007. Living with noisy genes: how cells function reliably with inherent variability in gene expression. *Annu Rev Biophys Biomol Struct* **36**: 413–34.
12. Graveley BR. 2001. Alternative splicing: increasing diversity in the proteomic world. *Trends Genet* **17**: 100–7.

13. Venables JP. 2006. Unbalanced alternative splicing and its significance in cancer. *BioEssays* **28**: 378–86.
14. Tazi J, Bakkour N, Stamm S. 2009. Alternative splicing and disease. *Biochim Biophys Acta* **1792**: 14–26.
15. Lara-Pezzi E, Gómez-Salineró J, Gatto A, García-Pavía P. 2013. The alternative heart: Impact of alternative splicing in heart disease. *J Cardiovasc Trans Res* **6**: 945–55.
16. Modrek B, Lee C. 2002. A genomic view of alternative splicing. *Nat Genet* **30**: 13–9.
17. Simmons D. 2008. Epigenetic influence and disease. *Nat Educ* **1**: 6.
18. Chinnery PF, Elliott HR, Hudson G, Samuels DC, et al. 2012. Epigenetics, epidemiology and mitochondrial DNA diseases. *Int J Epidemiol* **41**: 177–87.
19. Canals-Hamann AZ, das Neves RP, Reittie JE, Iniguez C, et al. 2013. A biophysical model for transcription factories. *BMC Biophys* **6**: 2.
20. Henikoff S, Shilatifard A. 2011. Histone modification: cause or cog? *Trends Genet* **27**: 389–96.
21. Fraga MF, Ballestar E, Paz MF, Ropero S, et al. 2005. Epigenetic differences arise during the lifetime of monozygotic twins. *Proc Natl Acad Sci USA* **102**: 10604–9.
22. Shaughnessy DT, McAllister K, Worth L, Haugen AC, et al. 2014. Mitochondria, energetics, epigenetics, and cellular responses to stress. *Environ Health Perspect* **122**: 1271–8.
23. Minocherhomji S, Tollefsbol TO, Singh KK. 2012. Mitochondrial regulation of epigenetics and its role in human diseases. *Epigenetics* **7**: 326–34.
24. Vander Heiden MG, Cantley LC, Thompson CB. 2009. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science* **324**: 1029–33.
25. Pollard PJ, Wortham NC, Tomlinson IP. 2003. The TCA cycle and tumorigenesis: the examples of fumarate hydratase and succinate dehydrogenase. *Ann Med* **35**: 632–9.
26. Baysal BE, Ferrell RE, Willett-Brozick JE, Lawrence EC, et al. 2000. Mutations in SDHD, a mitochondrial complex II gene, in hereditary paraganglioma. *Science* **287**: 848–51.
27. Polyak K, Li Y, Zhu H, Lengauer C, et al. 1998. Somatic mutations of the mitochondrial genome in human colorectal tumours. *Nat Genet* **20**: 291–3.
28. Bonnet S, Archer SL, Allalunis-Turner J, Haromy A, et al. 2007. A mitochondrial-K⁺ channel axis is suppressed in cancer and its normalization promotes apoptosis and inhibits cancer growth. *Cancer Cell* **11**: 37–51.
29. Ju YS, Alexandrov LB, Gerstung M, Martincorena I, et al. 2014. Origins and functional consequences of somatic mitochondrial DNA mutations in human cancer. *eLife* **3**: e02935.
30. Visscher Peter M, Brown Matthew A, McCarthy Mark I, Yang J. 2012. Five years of GWAS discovery. *Am J Hum Genet* **90**: 7–24.
31. Wallace DC. 2011. Bioenergetic origins of complexity and disease. *Cold Spring Harb Symp Quant Biol* **76**: 1–6.
32. Saxena R, de Bakker PI, Singer K, Mootha V, et al. 2006. Comprehensive association testing of common mitochondrial DNA variation in metabolic disease. *Am J Hum Genet* **79**: 54–61.
33. Mohlke KL, Jackson AU, Scott LJ, Peck EC, et al. 2005. Mitochondrial polymorphisms and susceptibility to type 2 diabetes-related traits in Finns. *Hum Genet* **118**: 245–54.
34. Andrews R, Reginiotes T, Turnbull DM, Birch M, et al. 2006. The role of mitochondrial haplogroups in primary open angle glaucoma. *Br J Ophthalmol* **90**: 488–90.
35. Rahman S, Poulton J, Marchington D, Suomalainen A. 2001. Decrease of 3243 A>G mtDNA mutation from blood in MELAS syndrome: a longitudinal study. *Am J Hum Genet* **68**: 238–40.
36. Bhat A, Koul A, Sharma S, Rai E, et al. 2007. The possible role of 10398A and 16189C mtDNA variants in providing susceptibility to T2DM in two North Indian populations: a replicative study. *Hum Genet* **120**: 821–6.
37. Niemi AK, Moilanen JS, Tanaka M, Hervonen A, et al. 2005. A combination of three common inherited mitochondrial DNA polymorphisms promotes longevity in Finnish and Japanese subjects. *Eur J Hum Genet* **13**: 166–70.
38. Brown MD, Sun F, Wallace DC. 1997. Clustering of Caucasian Leber hereditary optic neuropathy patients containing the 11778 or 14484 mutations on an mtDNA lineage. *Am J Hum Genet* **60**: 381–7.
39. Hudson G, Gomez-Duran A, Wilson IJ, Chinnery PF. 2014. Recent mitochondrial DNA mutations increase the risk of developing common late-onset human diseases. *PLoS Genet* **10**: e1004369.
40. Poulton J, Luan J, Macaulay V, Hennings S, et al. 2002. Type 2 diabetes is associated with a common mitochondrial variant: evidence from a population-based case-control study. *Hum Mol Genet* **11**: 1581–3.
41. Barrett T, Scott-Brown M, Seller A, Smith P, et al. 1998. Mitochondrial studies in Wolfram Syndrome. *Dev Med Child Neurol* **77**: 3.
42. Pritchard J. 2001. Are rare variants responsible for susceptibility to complex diseases? *Am J Hum Genet* **69**: 124–37.
43. Ye Z, Gillson C, Sims M, Khaw KT, et al. 2013. The association of the mitochondrial DNA OriB variant (16184–16193 polycytosine tract) with type 2 diabetes in Europan populations. *Diabetologia* **56**: 1907–13.
44. Yakusawa T, Yang M, Jacobs H, Holt I. 2005. A bidirectional origin of replication maps to the major non-coding region of human mitochondrial DNA. *Mol Cell* **18**: 651–62.
45. Park KS, Chan JC, Chuang LM, Suzuki S, et al. 2008. A mitochondrial DNA variant at position 16189 is associated with type 2 diabetes mellitus in Asians. *Diabetologia* **51**: 602–8.
46. Casteels K, Ong K, Phillips D, Bendall H, et al. 1999. Mitochondrial DNA variant, thinness at birth, and type-2 diabetes. ALSFAC study team. Avon Longitudinal Study of Pregnancy and Childhood. *Lancet* **353**: 1499–500.
47. Parker E, Phillips D, Cockington R, Cull C, et al. 2005. A common mitochondrial DNA variant is associated with thinness in mothers and their 20 year old offspring. *Am J Physiol Endocrinol Metab* **289**: 1110–4.
48. Laderman KA, Penny JR, Mazzucchelli F, Bresolin N, et al. 1996. Aging-dependent functional alterations of mitochondrial DNA (mtDNA) from human fibroblasts transferred into mtDNA-less cells. *J Biol Chem* **271**: 15891–7.
49. Coskun PE, Beal MF, Wallace DC. 2004. Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proc Natl Acad Sci USA* **101**: 10726–31.
50. Bender A, Krishnan KJ, Morris CM, Taylor GA, et al. 2006. High levels of mitochondrial DNA deletions in substantia nigra neurons in aging and Parkinson disease. *Nat Genet* **38**: 515–7.
51. Melov S, Schneider JA, Coskun PE, Bennett DA, et al. 1999. Mitochondrial DNA rearrangements in aging human brain and in situ PCR of mtDNA. *Neurobiol Aging* **20**: 565–71.
52. Cortopassi GA, Arnheim N. 1990. Detection of a specific mitochondrial DNA deletion in tissues of older humans. *Nucleic Acids Res* **18**: 6927–33.
53. Marin-Garcia J, Pi Y, Goldenthal MJ. 2006. Mitochondrial-nuclear cross-talk in the aging and failing heart. *Cardiovasc Drugs Ther* **20**: 477–91.
54. Chen X, Prosser R, Simonetti S, Sadlock J, et al. 1995. Rearranged mitochondrial genomes are present in human oocytes. *Am J Hum Genet* **57**: 239–47.
55. Jacobs HT. 2003. The mitochondrial theory of aging: dead or alive? *Aging Cell* **2**: 11–7.
56. Cortopassi GA, Shibata D, Soong NW, Arnheim N. 1992. A pattern of accumulation of a somatic deletion of mitochondrial DNA in aging human tissues. *Proc Natl Acad Sci USA* **89**: 7370–4.
57. Narendra D, Tanaka A, Suen DF, Youle RJ. 2008. Parkin is recruited selectively to impaired mitochondria and promotes their autophagy. *J Cell Biol* **183**: 795–803.
58. Diot A, Hinks-Roberts A, Lodge T, Liao C, et al. 2015. A novel quantitative assay of mitophagy: combining high content analysis fluorescence with mitochondrial DNA mutant load to identify novel pharmacological modulators of mitophagy. *Pharmacol Res* **100**: 24–35.
59. Schon EA, Przedborski S. 2011. Mitochondria: the next (neurode) generation. *Neuron* **70**: 1033–53.
60. Trifunovic A, Wredenberg A, Falkenberg M, Spelbrink JN, et al. 2004. Premature ageing in mice expressing defective mitochondrial DNA polymerase. *Nature* **429**: 417–23.
61. Kirkwood TB, Kowald A. 2012. The free-radical theory of ageing-older, wiser and still alive: modelling positional effects of the primary targets of ROS reveals new support. *BioEssays* **34**: 692–700.
62. Trifunovic A, Hansson A, Wredenberg A, Rovio AT, et al. 2005. Somatic mtDNA mutations cause aging phenotypes without affecting reactive oxygen species production. *Proc Natl Acad Sci USA* **102**: 17993–8.
63. Kukat A, Trifunovic A. 2009. Somatic mtDNA mutations and aging-facts and fancies. *Exp Gerontol* **44**: 101–5.
64. Ross JM, Stewart JB, Hagstrom E, Brene S, et al. 2013. Germline mitochondrial DNA mutations aggravate ageing and can impair brain development. *Nature* **501**: 412–5.
65. Wallace DC. 1994. Mitochondrial DNA sequence variation in human evolution and disease. *Proc Natl Acad Sci USA* **91**: 8739–46.
66. Reinhardt K, Dowling DK, Morrow EH. 2013. Mitochondrial replacement, evolution, and the clinic. *Science* **341**: 1345–6.

67. **Gemmell N, Wolff JN.** 2015. Mitochondrial replacement therapy: cautiously replace the master manipulator. *BioEssays* **37**: 584–5.
68. **Burgstaller JP, Johnston IG, Poulton J.** 2015. Mitochondrial DNA disease and developmental implications for reproductive strategies. *Mol Hum Reprod* **21**: 11–22.
69. **Poulton J, Kennedy S, Oakeshott P, Wells D.** 2009. Preventing transmission of maternally inherited mitochondrial DNA diseases. *BMJ* **338**: 345–9.
70. **Sato A, Kono T, Nakada K, Ishikawa K,** et al. 2005. Gene therapy for progeny of mito-mice carrying pathogenic mtDNA by nuclear transplantation. *Proc Natl Acad Sci USA* **102**: 16765–70.
71. **Tachibana M, Sparman M, Sritanaudomchai H, Ma H,** et al. 2009. Mitochondrial gene replacement in primate offspring and embryonic stem cells. *Nature* **461**: 367–72.
72. **Craven L, Tuppen HA, Greggains GD, Harbottle SJ,** et al. 2010. Pronuclear transfer in human embryos to prevent transmission of mitochondrial DNA disease. *Nature* **465**: 82–5.
73. **Dunham-Snary KJ, Ballinger SW.** 2015. GENETICS. Mitochondrial-nuclear DNA mismatch matters. *Science* **349**: 1449–50.
74. **Burgstaller J, Johnston I, Jones N, Albrechtová J,** et al. 2014. MtDNA segregation in heteroplasmic tissues is common in vivo and modulated by haplotype differences and developmental stage. *Cell Rep* **7**: 2031–41.