

Detection of Multiple Viral and Bacterial Infections in Acute Exacerbation of Chronic Obstructive Pulmonary Disease: A Pilot Prospective Study

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Few studies have evaluated the contribution of multiple virus and bacterial infections in acute exacerbation of chronic obstructive pulmonary disease. This study estimated the burden of multiple viral and bacterial respiratory infections in moderate to very severe chronic obstructive pulmonary disease patients that were prospectively followed-up during a 12-month pilot study. Clinical data were collected monthly and sputum was collected at the time of each acute exacerbation event. Classical culture techniques for bacteria and multiplex polymerase chain reaction (PCR) and microarray detection assays were performed to identify viral and atypical bacterial pathogens in the sputum. Overall, 51 patients were included and 45 acute exacerbation events were investigated clinically and microbiologically. Among the 45 acute exacerbation events, 44% had evidence of viral infection involving human rhinovirus (HRV) and metapneumovirus (hMPV) in 20% and 18%, respectively. Intracellular bacteria were not found in sputum by PCR. Common bacterial pathogens were identified in 42% of acute exacerbation patients, most frequently *Branhamella catarrhalis*, *Streptococcus pneumoniae* and *Haemophilus influenzae*. Viral or virus and bacteria co-infections were detected in 27% of acute exacerbation events ($n = 12$) with HRV and hMPV involved in 92% of cases. Patients with co-infections did not present greater clinical severity scores at exacerbation and more recurrence of acute exacerbation events at 3 and 6 months than those with single infections ($P > 0.4$). These results suggest that HRV and hMPV may be contributors or cofactors of AECOPD. These findings indicate that viral or virus and bacterial co-infections do not impact significantly on the clinical severity of acute

exacerbation of chronic obstructive pulmonary disease and recurrence at 3 and 6 months.

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INTRODUCTION

Acute exacerbation of chronic obstructive pulmonary disease is an important cause of morbidity and mortality. This disease is associated with worsening quality of life and decline in lung function [Seemungal et al., 1998; Donaldson et al., 2002]. Acute exacerbation of chronic obstructive pulmonary disease is triggered by bacterial or viral infection of the airways but the pathophysiological importance of these infectious agents is not fully understood [Melbye et al., 1992;

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Falsey et al., 1995; Dowell et al., 1996; Soler et al., 1998]. Classical bacteria are usually isolated in 40–60% of sputum sampled at the time of acute exacerbation of chronic obstructive pulmonary disease but 20–40% of patients with stable chronic obstructive pulmonary disease have also positive sputum cultures suggesting that bacteria may sometimes be innocent bystanders [Sethi et al., 2007]. The role of viruses has been largely investigated with recent reports describing viral infection in 29–44% of acute exacerbation of chronic obstructive pulmonary disease events. Using PCR techniques, the most common detected viruses in upper respiratory airway samples of acute exacerbation of chronic obstructive pulmonary disease patients were rhinoviruses, respiratory syncytial viruses, influenza A and B viruses, parainfluenza viruses, coronaviruses, and adenoviruses [Mohan et al., 2010]. Viral infections can also be detected in stable chronic obstructive pulmonary disease patients suggesting that some respiratory viruses may cause persistent low-grade infection contributing to the pathogenesis of the disease [Seemungal et al., 2001; Papi et al., 2006; Mohan et al., 2010]. Only few recent studies investigated the presence of mixed viral and classical and intracellular bacterial infections in sputum samples taken at the time of acute exacerbation event [Soler et al., 1998; Bandi et al., 2003; Papi et al., 2006; Wilkinson et al., 2006a; De Serres et al., 2009]. In the present time, the burden of multiple viral and bacterial respiratory infections as contributors of acute exacerbation of chronic obstructive pulmonary disease remains to be investigated.

In this prospective 1-year follow-up study, the burden of multiple viral and bacterial respiratory pathogens was assessed in moderate-to-severe chronic obstructive pulmonary disease patients and its relationship with the clinical severity criteria of acute exacerbation of chronic obstructive pulmonary disease and the frequency of recurrence at 3 and 6 months was assessed.

PATIENTS AND METHODS

Patients

Patients with chronic obstructive pulmonary disease were consecutively recruited from the Department of Respiratory Medicine of Reims (France) from January 2006 to December 2008. Chronic obstructive pulmonary disease was clinically defined in accordance with the Global Initiative for Chronic Obstructive Lung Disease criteria (GOLD) [Rabe et al., 2007] with post-bronchodilator $FEV_1 < 80\%$ and $FEV_1/FVC < 70\%$. At inclusion, all patients were stable with no acute exacerbation of chronic obstructive pulmonary disease for 4 weeks. Patients with asthma, bronchiectasis, cancer, or other respiratory diseases were excluded. The Hospital Ethics Committee approved the study (UCH Reims, Champagne Ardenne; number 2006-05, France) and a written informed consent was obtained from the patients.

Study Design and Clinical Follow-Up

Clinical characteristics of the patients were assessed at the time of inclusion, monthly and at the time of each acute exacerbation of chronic obstructive pulmonary disease event during 1 year. A diary card was given to each patient to determine a daily clinical score based upon self-reported symptoms: dyspnoea, sputum production, and purulence. Symptoms were noted as 1 (as usual), 2 (worse than usual), or 3 (much worse than usual). Acute exacerbation was defined as a minor worsening (+1) of at least two symptoms or a major worsening (+2) of at least one symptom for at least 2 days [Seemungal et al., 2001; Sethi et al., 2002]. Patients remained on their regular treatment for the duration of the study. Patients were consulting monthly to record changes in symptoms; medication and lung function and they were asked to contact the study team at each acute exacerbation of chronic obstructive pulmonary disease. For each patient a physical exam was performed to assess acute exacerbation of chronic obstructive pulmonary disease severity. Arterial blood gases, chest X-ray, and pulmonary functional tests were performed. Induced sputum was obtained prior to initiation of AECOPD treatment. One week after acute exacerbation of chronic obstructive pulmonary disease event, recovery was clinically assessed and monthly follow-up was resumed. Pulmonary functional tests were performed using a Body box 5500 plethysmograph (Medisoft, Sorinnes, Belgium). Sputum samples collected at the time of acute exacerbation were induced with hypertonic saline at 3%, 4% then 5% concentration delivered by ultrasonic nebuliser as previously described [Papi et al., 2006].

Microbiological Assays

Bacteriological assays. Samples were homogenized as described previously [Sethi et al., 2002]. Serial dilutions were placed on blood, chocolate, and MacConkey agar plates. Bacterial identification was performed by standard techniques. Sputum isolates were classified as potential pathogens microorganisms or normal flora. Potential pathogens microorganisms were: *Haemophilus influenzae*, *Moraxella catarrhalis*, *Streptococcus pneumoniae*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, and other gram-negative rods [Sethi et al., 2002]. Colony forming units (cfu)/ml were then calculated.

Classical virological assays. Immunofluorescence assays for detection of respiratory syncytial viruses A and B, influenza viruses, parainfluenza viruses, and adenoviruses antigens were performed as described previously [Bouscambert-Duchamp et al., 2005]. Virus isolates were typed by the standard method of virus neutralization onto cell culture for enteroviruses and by classical immunofluorescence antigen detection assays onto infected cell monolayers [Bouscambert-Duchamp et al., 2005; Jacques et al., 2006].

Multiplex PCR assays detection of viruses and atypical bacteria. Total nucleic acid extraction (DNA/RNA) was performed using the NucliSens easy-MAG instrument (BioMérieux, Lyon, France). Nucleic acids were eluted in a final volume of 50 μ l and then stored at -80°C [Boom et al., 1990; Loens et al., 2007]. CLART[®] PneumoVir and CLART[®] FluAVir kits (Genomica, Madrid, Spain) were performed according to the manufacturer's instructions allowing a simultaneous detection and identification of 20 different types and subtypes of human respiratory viruses (Influenza A (seasonal A/H1N1 and A/H3N2 and new influenza A/H1N1v/2009 strains), influenza B and C, parainfluenza (PIV 1, 2, 3, 4A, 4B), respiratory syncytial virus (RSV A and RSV B), human rhinovirus (HRV), adenovirus, human enteroviruses, human bocaviruses, human coronavirus E-229, human metapneumovirus (hMPV-A and hMPV-B) [Renois et al., 2010]. RT-PCR inhibitors were not detected in the analyzed samples as demonstrated by the positive detection of the internal control [Renois et al., 2010]. Each sputum sample was also tested for the presence of intracellular bacteria using a multiplex PCR and microplate assay (Chlamydege assay, Argene, Varhiles, France) allowing the detection of *Legionella pneumophila*, *Chlamydia pneumoniae*, and *Mycoplasma pneumoniae* [Ginevra et al., 2005].

HRV genotyping. In cases of positive HRV genomic RNA detection, the VP4 region and part of the VP2 genome region (VP4-VP2) were amplified with specific primers as described previously [Savolainen-Kopra et al., 2009; Wisdom et al., 2009]. To determine the rhinovirus type, the obtained consensus sequences of VP4-VP2 were compared to all corresponding HRV sequences available in GenBank for each region by using BLAST software. Sequence alignments were

performed using the Clustal W program (version 1.81). Phylogenetic and molecular evolutionary analyses were conducted using MEGA (version 4.0) software and allowed us to perform reliable HRV genotyping identification [Savolainen-Kopra et al., 2009; Wisdom et al., 2009]. The sequences generated in the present study have been assigned to the following GenBank accession numbers: JQ918189–JQ918195.

Statistical analyses. Clinical or biological data were expressed as mean values \pm standard deviation. Chi-squared test, Mc Nemar Chi-squared test, Fisher's exact test or Wilcoxon rank-sums test was carried out when necessary using the SAS software, version 9.1.3 (SAS Institute, Cary, NC). A *P*-value <0.05 was considered significant.

RESULTS

Patients' Characteristics

Fifty-one chronic obstructive pulmonary disease patients were included consecutively from January 2006 to December 2008. According to GOLD classification, 10 patients (20%) were classified as GOLD II (moderate), 32 (63%) GOLD III (severe), and 9 (17%) were GOLD IV (very severe). Twenty-five patients (49%) reported at least one acute exacerbation of chronic obstructive pulmonary disease during the 12-month follow-up (mean: 1.8 ± 0.8 acute exacerbation event per year; Table I). Eleven patients (22%) reported one acute exacerbation of chronic obstructive pulmonary disease, nine patients (18%) reported two acute exacerbations, four patients (7%) reported three acute exacerbations, and only one patient (2%) reported four acute exacerbations. A total of 45 acute exacerbations of chronic obstructive pulmonary

TABLE I. Clinical and Functional Baseline Characteristics of 51 Prospectively Studied Patients

	No reported AECOPD (n = 26)	1 reported AECOPD (n = 11)	>2 reported AECOPD (n = 14)
Age, year	66 \pm 8	60 \pm 9	60 \pm 8*
Male gender, %	84	73	86
Current smoker, %	23	55	29
Smoking history, pack-year	43 \pm 20	57 \pm 24	41 \pm 15
Chronic bronchitis, %	54	82	79
Influenza vaccination, %	68	56	89
Pneumococcus vaccination, %	47	40	50
FEV1, % predicted	45 \pm 8	44 \pm 13	41 \pm 16
FVC, % predicted	75 \pm 14	69 \pm 9	72 \pm 15
FEV1/FVC, %	47 \pm 9	49 \pm 14	45 \pm 14
BMI, kg/m ²	28 \pm 9	27 \pm 6	27 \pm 9
PaO ₂ , mmHg	74 \pm 9	74 \pm 6	74 \pm 13
PaCO ₂ , mmHg	41 \pm 5	41 \pm 5	42 \pm 5
pH	7.41 \pm 0.04	7.42 \pm 0.02	7.41 \pm 0.04
Inhaled corticosteroid, %	88	81	93
Long-acting β 2-agonist, %	96	91	100
Short-acting bronchodilator, %	50	91	71
Long term oxygen therapy, %	4	9	0

AECOPD, acute exacerbation of chronic obstructive pulmonary disease.

Data are presented as mean \pm SD.

**P* < 0.05 versus no AECOPD.

disease were reported and analyzed in the study. Nine acute exacerbations events (20%) required hospitalization. One patient died from acute respiratory failure. The retrospective analysis of diary cards showed that 22 acute exacerbation events were not reported. Comparison between patients with no reported acute exacerbation of chronic obstructive pulmonary disease, one acute exacerbation of chronic obstructive pulmonary disease, and >2 acute exacerbations of chronic obstructive pulmonary disease did not show any differences, except a slight difference for age (66 ± 8 years for no acute exacerbation vs. 60 ± 8 years for >2 acute exacerbation events, $P < 0.05$; Table I).

Microbiological Findings in Acute Exacerbation of Chronic Obstructive Pulmonary Disease

Sputum samples were analyzed in 45 acute exacerbations of chronic obstructive pulmonary disease (Table IIA). No pathogen was found in 16 acute exacerbations of chronic obstructive pulmonary disease (35%). Viruses were detected by PCR in 20 (44%) acute exacerbations of chronic obstructive pulmonary disease cases: one virus in eight cases (18%), two viruses in two cases (4%), and co-infection virus and potential pathogen microorganisms in 10 cases (22%; Table IIB). HRV or hMPV represented 72% of detected viruses. As expected, viral detection rates by

TABLE II. Microbiological Findings Obtained at Exacerbation of Chronic Obstructive Pulmonary Disease A Global Findings Obtained Using Classical Culture Techniques for Bacteria and Viruses Associated to Multiplex Polymerase Chain Reaction (PCR) Assays for the Detection of Viral and Atypical Bacterial Pathogens in Sputum Specimens

	Number of samples positive by ^a			
	PCR microarray techniques		Culture assays	
	n	%	n	%
Respiratory viruses ^b	20	44	3	6
HRV	9	20	0	0
HMPV	8	18	0	0
Influenza A virus	2	4	1	2
Influenza B virus	1	2	0	0
RSV	1	2	1	2
Parainfluenza virus	1	2	1	2
Respiratory bacteria			19	42
Classical PPM				
<i>Moraxella catarrhalis</i>	ND	ND	6	13
<i>Haemophilus influenzae</i>	ND	ND	8	17
<i>Streptococcus pneumoniae</i>	ND	ND	3	7
Streptococcus species	ND	ND	1	2
<i>Pasteurella multocida</i>	ND	ND	ND	2
<i>Citrobacter freundii</i>	ND	ND	ND	2
<i>Haemophilus parainfluenzae</i>	ND	ND	1	2
<i>Proteus vulgaris</i>	ND	ND	1	2
Atypical respiratory pathogens				
<i>Legionella pneumophila</i>	0	0		
<i>Mycoplasma pneumoniae</i>	0	0		
<i>Chlamydia pneumoniae</i>	0	0		

B-Viral and bacterial co-infections at exacerbation of chronic obstructive pulmonary disease

Coinfections		
Pathogen 1	Pathogen 2	n = 12
HMPV	Influenza A virus	6
	HRV	1
	<i>H. influenzae</i>	1
	<i>M. catarrhalis</i>	1
	<i>S. pneumoniae</i>	1
	<i>H. parainfluenzae</i>	1
HRV	HMPV	6
	<i>H. influenzae</i>	1
	<i>M. catarrhalis</i>	4
RSV	<i>M. catarrhalis</i>	1
	<i>H. influenzae</i>	1

^aMore than one pathogen possible.

^bTwenty viral pathogens were detected by microarray system; only positive detected viral types were notified in this table.

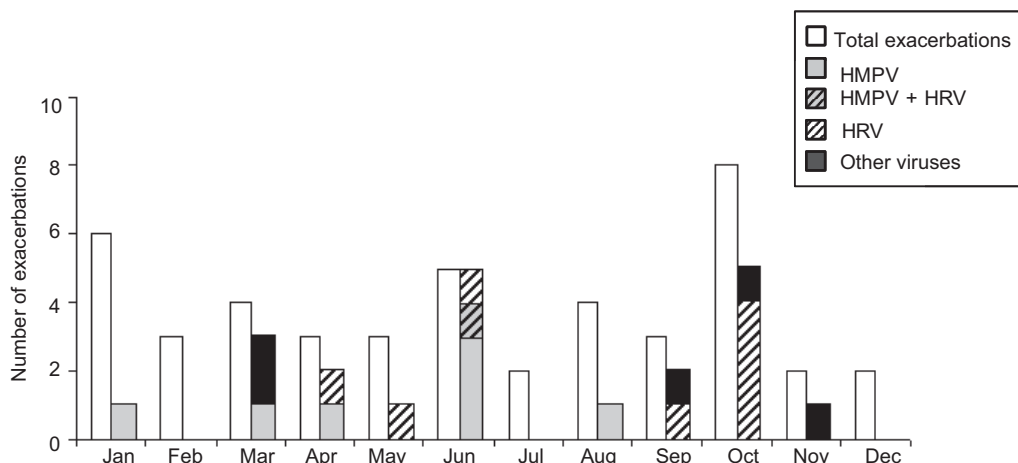


Fig. 1. Monthly distribution of viruses detected by PCR-microarray assays at acute exacerbation of chronic obstructive pulmonary disease.

PCR were significantly greater than those observed by classical viral culture assays (44% vs. 7%, respectively, $P < 0.001$). hMPV was mainly detected in spring whereas HRV was predominant in autumn and spring (Fig. 1). In case of positive HRV detection at time exacerbation ($n = 4$), HRV genotyping assay on VP4/VP2 capsid viral genes was performed as previously described [Wisdom et al., 2009]. These genotyping results identified several HRV phylogenetic groups corresponding to the common geno-groups A (serotypes 14 and 18) and B (serotype 44) and interestingly to the newly described geno-group C (not shown).

Potential pathogen microorganisms were detected in 19 acute exacerbations of chronic obstructive pulmonary disease (42%): one in 17 cases, two in one case, and three in one case (Table IIA). *M. catarrhalis*, *H. influenzae*, or *Streptococcus pneumoniae* were found in 77% of potential pathogen microorganism detection. The bacterial load in positive samples was 10^6 cfu/ml or greater. Intracellular bacteria (*Legionella pneumophila*, *Mycoplasma pneumoniae*, and *Chlamydia pneumoniae*) were not found in sputum samples at time of acute exacerbation of chronic obstructive pulmonary disease by PCR. Viral or virus and bacteria co-infections were detected in

27% of acute exacerbations of chronic obstructive pulmonary disease ($n = 12$). HRV and HMPV were involved in 11/12 co-infections (92%; Table IIB). HRV was associated with *H. influenzae* in four co-infection cases (33%).

Clinical Findings and Outcome

The influence of the viral infection and the impact of multiple infections on severity of acute exacerbation of chronic obstructive pulmonary disease were analyzed. Interestingly, viral detection by PCR at time of acute exacerbation of chronic obstructive pulmonary disease was not associated statistically with the severity of exacerbation in term of hospitalization rate and clinical score, FEV₁ and PaO₂ variations from baseline ($P > 0.2$; Table III). In addition, co-infections cases (virus/bacteria or virus/virus) did not present greater clinical severity scores at exacerbation than those with single infections ($P = 0.45$; Table III). Next, we analyzed the potential statistical relationships between the frequency of acute exacerbation recurrence and multiple infections. We observed no significant statistical differences in the frequency of recurrence after acute exacerbation of chronic obstructive pulmonary disease between patients with

TABLE III. Clinical Severity Criteria of Acute Exacerbation of Chronic Obstructive Pulmonary Disease According to the Microbiological Findings

	Total (n = 45)	Virus ^a (n = 8)	Co-infection ^a (n = 12)	Bacteria ^a (n = 9)	No pathogen ^a (n = 16)
Clinical score	6.1 ± 1.2	6.4 ± 1.2	6.5 ± 1.4	6.3 ± 1.2	5.6 ± 1.2
Δ clinical score	3.1 ± 1.2	3.4 ± 1.2	3.5 ± 1.4	3.1 ± 1.2	2.6 ± 1.1
Hospitalization, %	17	20	10	33	19
Δ FEV ₁ , % predicted	1 ± 10	6 ± 7	3 ± 7	0.3 ± 10	2 ± 12
Δ PaO ₂	-0.9 ± 10	-0.7 ± 6.8	4 ± 11.6	-5.7 ± 9	-2.5 ± 10.4

Data are presented as mean ± SD.

Δ, difference from baseline.

^aNo significant differences between all groups.

and without multiple viral or bacterial infections at 3 months (17% vs. 33%, respectively, $P = 0.46$) and at 6 months (42% vs. 52%, respectively, $P = 0.74$; data not shown). Multiple infection rates were not significantly different in cases of frequent (>2 per year) or infrequent acute exacerbation of chronic obstructive pulmonary disease (<2 per year) (29% vs. 18% of exacerbations, respectively, $P = 0.70$). Finally, co-infected patients did not present greater clinical severity scores at exacerbation and more recurrences of exacerbation at 3 and 6 months than those with single infections ($P > 0.4$; Table III).

DISCUSSION

Several studies have measured the burden of viral or bacterial infections, but only few have assessed the importance of both pathogens and the presence of mixed viral or virus and bacteria multiple infections [Soler et al., 1998; De Serres et al., 2009]. In this prospective 1-year follow-up study, 51 patients were included and 45 acute exacerbation events were clinically and microbiologically investigated. Of the 45 acute exacerbation events studied, 44% had evidence of viral infections, 42% demonstrated classical bacterial pathogens, and 27% had evidence of multiple viral or virus and bacteria co-infections (Table II). The detection rates of classical bacterial pathogens and of respiratory viruses by RT-PCR techniques were in agreement with those previously published in recent reports [Melbye et al., 1992; Falsey et al., 1995; Dowell et al., 1996; Soler et al., 1998; De Serres et al., 2009]. However the distribution of type of respiratory viruses appeared to be different from previously published studies with identification of HRV and HMPV as the primary contributors to acute exacerbation while RSV and influenza were infrequently detected during this 1-year pilot study. Potential pathogen microorganisms were detected in 42% of acute exacerbation events demonstrating as previously described the predominance of *M. catarrhalis*, *Haemophilus influenzae*, and *Streptococcus pneumoniae* that represented 77% of classical bacteria. No intracellular bacteria were detected at time of acute exacerbation events in patient with chronic obstructive disease (Table II).

In the present study, HMPV was identified as one of potential primary contributors of acute exacerbation events whereas this respiratory virus has been rarely detected at time of exacerbation in previous studies ranging from 0% to 4.7% with a weighted mean prevalence of 0.7% in a recent review (Table II) [Beckham et al., 2005; Rohde et al., 2005; Mohan et al., 2010]. These discrepant results could be explained by the sensitivity of the used techniques and by the design of the study or the temporal epidemiological characteristics. HRV was previously identified as a one of the major contributor of acute exacerbation of chronic obstructive pulmonary disease and interestingly our genotyping analyses showed

that the geno-group C could trigger acute exacerbation events. Because HRV group C strains are not easily detectable by classical culture techniques; these results indicate the need to use PCR techniques to explore the relationship between virus infections and acute exacerbation of chronic obstructive pulmonary disease. In the present study, viral respiratory pathogens were detected by use of a multiplex PCR-DNA and microarray detection assay allowing a rapid and accurate detection of conventional and newly discovered viral respiratory pathogens. Respiratory viruses were detected in 44% of acute exacerbations of chronic obstructive pulmonary disease by multiplex PCR-DNA microarray system, a rate 6.7 higher than those obtained by classical viral detection techniques. However, this qualitative molecular approach was not able to distinguish an ongoing infection from a chronic or persistent viral shedding [Seemungal et al., 2001; Kuypers et al., 2006; De Serres et al., 2009]. Further prospective clinical studies testing the viral load levels at time of acute exacerbation versus stable state are now necessary to differentiate ongoing infection from past or beginning viral infections in acute exacerbation of chronic obstructive pulmonary disease cases.

One of the main objectives of this study was to evaluate the potential relationships between multiple bacterial and viral infections and the severity and risk of recurrence of acute exacerbation of chronic obstructive pulmonary disease. In the present study, multiple infections were not related to demographic characteristics, smoking history, lung function or inhaled treatment, in accordance with previous studies [Seemungal et al., 2001; Wilkinson et al., 2006b]. Finally, clinical outcomes of acute exacerbation of chronic obstructive pulmonary disease that were depending on multiple infections were analyzed. Mixed infections (virus-virus or virus-bacteria) were not found to be associated with the severity of acute exacerbation of chronic obstructive pulmonary disease or with an increased risk of recurrence at 3 and 6 months ($P > 0.4$). A recent study using PCR detection methods for nasal aspirate samples described that acute exacerbation of chronic obstructive pulmonary disease cases of mixed virus-bacterial infections do not appear to be more severe among virus-infected patients [De Serres et al., 2009]. The present study provides additional data on the impact of mixed virus or bacterial infections onto the clinical severity and the risk of recurrence of acute exacerbation that are critical elements in chronic obstructive pulmonary disease management. However, it must be pointed out that the present monocentric study was conducted in a relatively low number of chronic obstructive pulmonary disease patients and acute exacerbation of chronic obstructive pulmonary disease events. Further multicentric studies involving larger population are needed to come to definitive conclusions regarding the clinical impact of multiple infections in acute exacerbation of chronic obstructive pulmonary disease.

The present results demonstrated chronic obstructive pulmonary disease patients infected by two respiratory viruses therefore suggesting a potential intrinsic susceptibility to particular triggers of acute exacerbation of chronic obstructive pulmonary disease [Seemungal et al., 2001]. This susceptibility could be related to polymorphism of genes encoding CCL1, a chemotactic factor for leukocytes [Takabatake et al., 2006], mannose binding lectin (MBL2) [Yang et al., 2003] or surfactant protein B [Foreman et al., 2008], altering host defense factors against pathogens. Recently, an experimental human model of HRV infection clearly showed the role of HRV as triggering factor in acute exacerbation of chronic obstructive pulmonary disease [Mallia et al., 2011]. A deficient induction of type III interferon- λ by HRV in asthmatic primary bronchial cells and alveolar macrophages was correlated with the severity of HRV related asthma exacerbation [Contoli et al., 2006]. The hypothesis of a lower immunological response against viral infections in acute exacerbation of chronic obstructive pulmonary disease remains to be investigated in further clinical prospective studies.

In conclusion, these results suggest that HRV and hMPV may be contributors or cofactors of acute exacerbation of chronic obstructive pulmonary disease. These findings indicate that viral or virus and bacterial co-infections do not impact significantly on the clinical severity of acute exacerbation of chronic obstructive pulmonary disease and recurrence at 3 and 6 months.

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