



Review

Multidimensional excavation of the current status and trends of mechanobiology in cardiovascular homeostasis and remodeling within 20 years

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ABSTRACT

Mechanobiology is essential for cardiovascular structure and function and regulates the normal physiological and pathological processes of the cardiovascular system. Cells in the cardiovascular system are extremely sensitive to their mechanical environment, and once mechanical stimulation is abnormal, the homeostasis mechanism is damaged or lost, leading to the occurrence of pathological remodeling diseases. In the past 20 years, many articles concerning the mechanobiology of cardiovascular homeostasis and remodeling have been published. To better understand the current development status, research hotspots and future development trends in the field, this paper uses CiteSpace software for bibliometric analysis, quantifies and visualizes the articles published in this field in the past 20 years, and reviews the research hotspots and emerging trends. The regulatory effects of mechanical stimulation on the biological behavior of endothelial cells, smooth muscle cells and the extracellular matrix, as well as the mechanical-related remodeling mechanism in heart failure, have always been research hotspots in this field. This paper reviews the research advances of these research hotspots in detail. This paper also introduces the research status of emerging hotspots, such as those related to cardiac fibrosis, homeostasis, mechanosensitive transcription factors and mechanosensitive ion channels. We hope to provide a systematic framework and new ideas for follow-up research on mechanobiology in the field of cardiovascular homeostasis and remodeling and promote the discovery of more therapeutic targets and novel markers of mechanobiology in the cardiovascular system.

1. Introduction

Mechanobiology focuses on how mechanical stimulation interacts with chemical signals to regulate cell and tissue functions and their molecular mechanisms. With the stress-growth law proposed by Y.C.

Fung in 1990, which mentioned that the remodeling of blood vessels involving the growth or resorption of cells and extracellular materials is linked to mechanical stresses,¹ researchers have gradually turned their attention to the mechanism of the mechanical regulation of life processes. With the accelerated development of molecular biology and cell biology,

Abbreviations: AS, Atherosclerosis; CFD, Computational fluid dynamics; ECM, Extracellular matrix; ECs, Endothelial cells; EndMT, Endothelial-mesenchymal transformation; GAGs, Glycosaminoglycans; HF, Heart failure; LAE, Left atrial enlargement; LSS, Laminar shear stress; mTORC1, mammalian target protein complex 1; NETs, Neutrophil extracellular traps; OSS, Oscillatory shear stress; PDE5, Phosphodiesterase 5; TEAD, Tea-domain transcription factor; VSMCs, Vascular smooth muscle cells; WWTR1, WW domain-containing transcription regulator 1; YAP, Yes associated protein.

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a new interdisciplinary field, mechanobiology, has gradually emerged. Mechanobiology studies not only reveal the biomechanical mechanisms of normal growth, development and aging but also clarify the role of mechanical stimulation in the pathogenesis of diseases.² Although the vast majority of cells in the body are subjected to mechanical stimulation, most current research on mechanobiology focuses on the cardiovascular system.^{3–5} The cardiovascular system, which is centered on the heart, pumps blood out of the heart to the entire body through the heart's mechanical pumping action, achieving energy transfer and material exchange. This process involves many mechanical effects, including the mechanical force generated by the beating of the heart and the mechanical stimulation of blood pressure and flow through blood vessels. Mechanical stimulation is a mechanical signal that is the main regulator of cell structure and function. It causes disturbances in the position and morphology of cells and the vascular microenvironment. Mechanosensitive sensors on cell membranes can sense these mechanical changes and transduce them into chemical signals, activating multiple signaling pathways and responding by remodeling, growth, and repair.⁶

Cardiovascular homeostasis and remodeling have always been research hotspots in the field of cardiovascular diseases. Cardiovascular homeostasis is a physiological state in which cardiovascular physiological functions as well as internal and external environmental compositions and properties are maintained in a relatively dynamic balance, whereas cardiovascular remodeling is a dynamic pathological process.⁷ Mechanical stimulation is a key regulator of cardiovascular homeostasis and pathological remodeling.⁸ Under physiological conditions, the cardiovascular system senses mechanical stimulation and then maintains the relative balance of cardiovascular composition, structure and function through various regulatory mechanisms to achieve cardiovascular homeostasis.⁹ When mechanical stimulation is abnormal or the cardiovascular mechanosensation or mechanotransduction capacity is impaired, pathological cardiovascular remodeling can occur.^{9–11} Different cardiovascular cells and components respond differently to abnormal mechanical stimulation, causing different cardiovascular remodeling events. Pathological remodeling changes in the cardiovascular system are manifested in both structure and function. Structural changes include intimal hyperplasia, thickening or thinning of the media, adventitial fibrosis, and ventricular concentric hypertrophy,^{10,12} which in turn affect functional changes manifested as changes in vascular compliance and vascular regulation disorders.^{9,13} The biological processes involved include the synthesis, degradation, and reorganization of the extracellular matrix (ECM), as well as the excessive proliferation, migration, and apoptosis of endothelial cells (ECs) and vascular smooth muscle cells (VSMCs).^{14–16} Pathological mechanical stimulation or mechanotransduction induces cardiovascular remodeling leading to a range of cardiovascular diseases, including hypertension, atherosclerosis (AS), heart failure (HF), and restenosis after vein grafting.^{17–19} Therefore, in cardiovascular homeostasis and remodeling, a deep understanding of the biological effects of mechanical stimulation and how to produce biological effects is of great theoretical and clinical importance for identifying potential mechanics-related drug targets and new biomarkers and laying a mechanobiological foundation for the prevention and treatment of pathological cardiovascular remodeling diseases.

Currently, with the continuous development of mechanobiology in cardiovascular homeostasis and remodeling, the number of related articles has increased rapidly. Therefore, it is necessary to conduct a retrospective analysis of published articles in this field to better understand the status of past research and provide guidance for future research. Bibliometrics is a branch of informatics. It quantitatively analyzes relevant information in the scientific literature to understand the knowledge structure and research trends in a certain research field. It is an effective tool for objectively studying the current status of a discipline and reflecting its development.²⁰ Recently, many reviews related to bibliometrics have been published.^{21–23} By using bibliometric analysis software to construct a knowledge graph, a comprehensive literature review of the proposed research dimension is conducted. It usually takes scientific publications as input, quickly organizes many published articles,

generates interactive visualization maps of complex structures, visualizes the structure and dynamics of a certain research field, and then analyzes the results to determine the main research topics and frontiers.²⁴ CiteSpace is a bibliometric software for visual analysis of scientific literature. It presents the structure, laws and distribution of scientific knowledge by drawing a series of visual maps, helping researchers to better understand the development context of the research field.²⁵ This article uses CiteSpace bibliometric analysis software to conduct a bibliometric analysis of the field of mechanobiology in cardiovascular homeostasis and remodeling, comprehensively analyzes the development status and trends of this field, better elucidates the current research hotspots and emerging trends in this field, and promotes the development of this field.

2. Bibliometric methods

2.1. Data collection and retrieval strategy

The Web of Science is the world's largest information resource platform, covering the fields of natural sciences, engineering technology, biomedicine, social sciences, arts and humanities. It also includes the world's highest-level authoritative academic journals, monographs, conferences, abstracts and conference proceedings with powerful citation indexing and academic influence evaluation functions. In this work, the Web of Science Core Collection (WoSCC) is used as the data source, and the data are searched from 2004 to 2024 November as the time period. The search terms used were ALL=((cardiac OR vascular OR artery OR arterial OR vessel OR cardiovascular) AND (remodel* OR homeostasis)) AND (mechanobiology OR mechanics). A total of 2872 studies were retrieved, and 2796 studies were analyzed via bibliometrics after the duplicate articles were deleted. Standard bibliometric indicators were used to analyze the number of published articles, countries, institutions, authors, cited references, and keywords in CiteSpace 6.4.R1.

2.2. Bibliometric analysis tool - CiteSpace

This study selected 6.4.R1 from CiteSpace and used 2796 articles retrieved from WoSCC as the data source. This analysis included 2212 articles, 354 reviews, 95 proceedings papers, etc (Fig. 1). We Select January 2004 to November 2024 for time slicing, with each slice set to 1 year, the selection standard Top N set to 50, and the node types selected as author, institution, country, keyword, and reference to visualize the obtained valid literature data. The analysis results include annual publication analysis, national and institutional cooperation analysis, author analysis and citation explosion keywords. The relevant content is visualized in an information graph, and the key nodes and their connections in different graphs are analyzed. The higher the frequency of occurrence in the graph is, the larger the node. The color and thickness of the inner circle of a node represent the frequency of occurrence at different times, whereas the purple outer circle of the node indicates its high betweenness centrality, expressing the intermediary connection role of the node in the collinear network, which can be considered a turning point and plays a key role in the information graph. The connecting lines between nodes are represented as co-occurrence or co-citation relationships, and the thickness of the connecting lines indicates the strength of co-occurrence. The more connections there are and the thicker the connecting lines between nodes are, the closer the relationship is. Finally, the research hotspots are visually displayed through an information graph, and research trends are predicted.

3. Bibliometric analysis

3.1. Annual publication volume analysis

Since 2004, there have been a total of 2796 research articles in this field, increasing from 30 in 2004 to over 240, showing an overall upward trend. In 2021, more than 200 papers were published, reaching its peak.

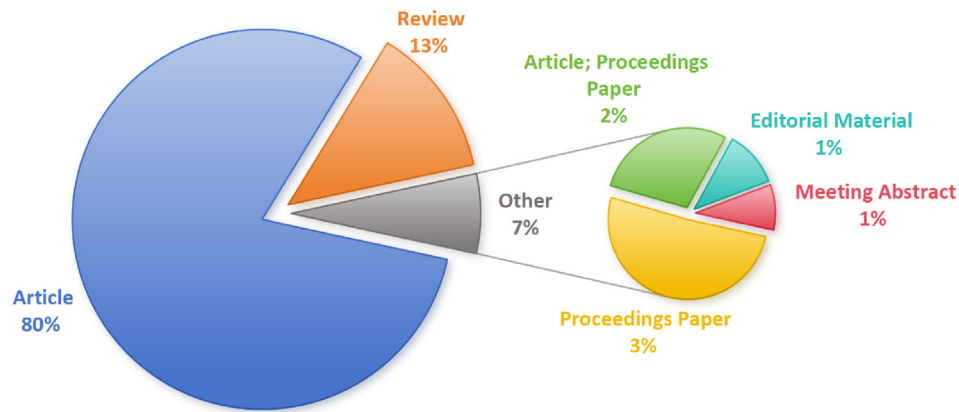


Fig. 1. Types of publications in the field of mechanobiology in cardiovascular homeostasis and remodeling from 2004 to 2024.

Within two years, the number of published articles remained at approximately 240. As of the search date, the number of papers published in 2024 was 197. The regression curves predict that the number of publications in this field will be 245 in 2024 (Fig. 2). With increasing social attention given to cardiovascular disease and the continuous development and improvement of strategies for disease modeling and treatment, this field has also received widespread attention. The stable upward trend of published articles indicates that this field has become a hot research topic in recent years and has attracted widespread attention.

3.2. Collaborative relationship analysis

3.2.1. Country/Regional cooperation network

A total of 82 nodes and 893 connections of the country cooperation network map were obtained after visualized analysis with "country" as the node. Each node represents a country. The larger the node is, the more articles published by that country. The top five countries are the United States, China, Italy, the United Kingdom, and Germany (Fig. 3). The United States has the highest number of publications, with a total of 1449 articles (Fig. 4). Through CiteSpace visualization, it is possible to analyze the cooperative relationships between countries (Fig. 3), where the multiple and dense connections between countries indicate close cooperation between them. The cooperation between countries is as follows. The United States has close ties with China, the United Kingdom, Italy, Germany, Canada, France, and many of the countries shown

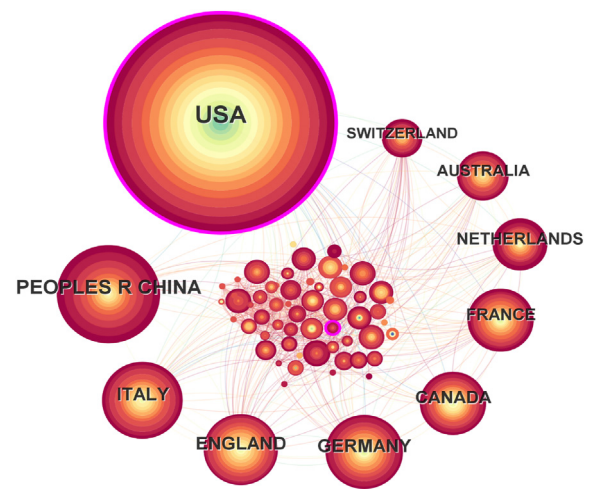


Fig. 3. Network diagram of the effects of national cooperation in the field of mechanobiology in cardiovascular homeostasis and remodeling from 2004 to 2024.

(Fig. 3); China has close ties with South Africa, Britain, Germany, Canada, France and other European and American countries, as well as Japan, South Korea and other Asian countries; Italy has close ties with

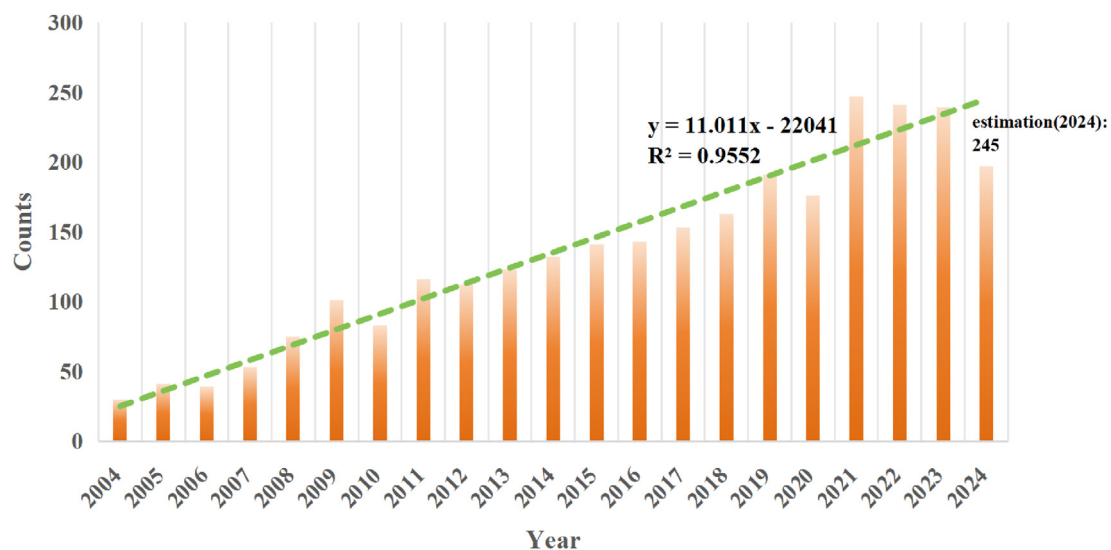


Fig. 2. Number of publications in the field of mechanobiology in cardiovascular homeostasis and remodeling from 2004 to 2024.

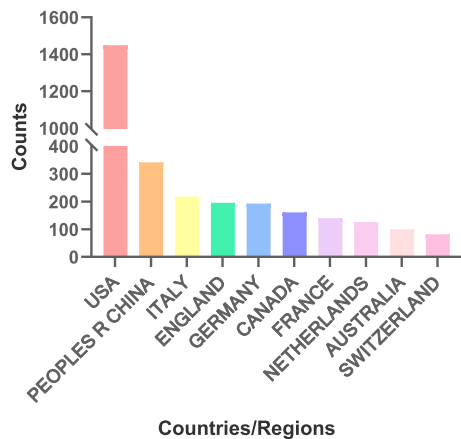


Fig. 4. Top 10 countries in the field of mechanobiology in cardiovascular homeostasis and remodeling in terms of publication volume.

France, Greece, Canada, the Netherlands and Australia; Britain has close ties with Switzerland, Singapore, Greece, Spain and India; and Germany has close ties with Belgium, Ireland, South Korea, Japan and South Africa. The top five countries in terms of centrality are the United States (0.35), South Africa (0.11), England (0.07), Egypt (0.07), and Slovenia (0.06). The centrality of America and South Africa is greater than 0.1, and the high centrality in the national cooperation network also indicates that these 2 countries have a certain influence on cardiovascular homeostasis and remodeling in the field of mechanobiology.

3.2.2. Institutional cooperation network

The information graph of articles published by institutions in various countries from 2004 to 2024 includes 441 nodes and 2906 connections. According to the institutional cooperation network (Fig. 5), the top ten institutions with the greatest number of publications can be obtained, among which the top five institutions are the University of California System, Harvard University, Yale University, Pennsylvania Commonwealth System of Higher Education (PCSHE), and University System of Georgia (Table 1). In the network diagram of institutional cooperation (Fig. 5), University of California System and University of California Davis, University of California Berkeley, University of California Irvine,

Table 1
Top 9 institutions in terms of publication volume from 2004 to 2024.

Institution	Counts	Countries
University of California System	159	USA
Harvard University	118	USA
Yale University	98	USA
Pennsylvania Commonwealth System of Higher Education (PCSHE)	93	USA
University System of Georgia	92	USA
University of Texas System	86	USA
Georgia Institute of Technology	85	USA
Institut National de la Sante et de la Recherche Medicale (Inserm)	84	FRANCE
Stanford University	76	USA

Austral University, University of Illinois System, University of California San Francisco, University of Illinois Chicago Hospital have close ties in the field of mechanobiology in cardiovascular homeostasis and remodeling. Harvard University works closely with Beth Israel Deaconess Medical Center, Harvard Medical School, and the Broad Institute. Yale University and Texas A&M University College Station, Texas A&M University System, Universitat Politcnica de Valencia and Universite de Montpellier have close links in this field. Pennsylvania Commonwealth System of Higher Education (PCSHE) and the University of Pittsburgh, the Mayo Clinic, the University of Pittsburgh Palermo, Boston Children's Hospital, the University of Limerick, Pennsylvania State University, Pennsylvania State University and Duke University cooperate and exchange. There are close ties between the University System of Georgia and the Georgia Institute of Technology, the US Department of Veterans Affairs, the Veterans Health Administration (VHA), Augusta University, the Atlanta VA Medical Center, and the Atlanta VA Health Care System.

The CiteSpace visualization of institutional collaboration network graphs reveals that the nodes of institutions such as University of Maryland Baltimore (0.27), CNRS - National Institute for Biology (INSB) (0.24), Johns Hopkins University (0.22), Brigham & Women's Hospital (0.2), National Institutes of Health (NIH) - USA (0.2), University of Toronto (0.19), University of California System (0.18), and Centre National de la Recherche Scientifique (CNRS) (0.17) all have purple outer circles, indicating that these institutions have high centrality and play important roles in research and development in this field (Fig. 5). Currently, the main research in this field is focused on schools, with few

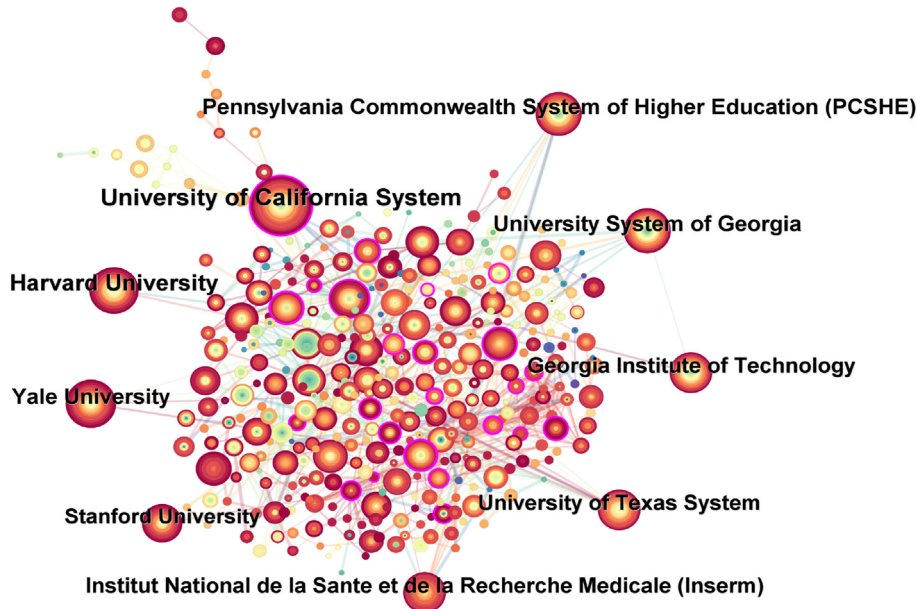


Fig. 5. Network diagram of institutional cooperation in the field of mechanobiology in cardiovascular homeostasis and remodeling from 2004 to 2024.

hospitals conducting related research. Approximately ninety percent of the top nine institutions in terms of publication volume are from the United States, which has a certain leading position in research in this field.

3.2.3. Author collaboration network

Through the analysis of the number of articles published by authors in the field of mechanobiology in cardiovascular homeostasis and remodeling, as well as the collaboration network, the collaboration network diagram includes 744 nodes and 1144 connections. The size of the nodes in the collaboration diagram represents the number of articles published by the authors, and the thickness of the connection lines reflects the strength of their collaboration relationship. According to the institutional cooperation network (Fig. 6), the top ten authors with the highest number of publications can be obtained, among which the top five authors are Jay D. Humphrey, and Marijana. Tadic, Dao Wen. Wang, Cesare. Cuspidi, Rudolph L. Gleason. Nearly half of the top 10 authors in the past 20 years are from the United States (Table 2).

The cooperative network diagram closely related to national and institutional cooperation is different. The cooperative relationships among scholars are relatively scattered, and scholars are closely connected on a small scale (Fig. 6). David A. Vorp and Jay D. Humphrey jointly published information about abdominal aortic aneurysms and biomechanical behavior research.²⁶ Marijana Tadic and Cesare. Cuspidi closely collaborated in the field of vascular research. Together, they have participated in a number of studies in the areas of hypertension, cardiac mechanics, and cardiology. Specifically in patients with hypertension, they collaborated to investigate the relationship between left atrial enlargement (LAE) and changes in heart structure and function, as well as the impact of these changes on prognosis.²⁷ Moreover, the prevalence of LAE and its association with left ventricular hypertrophy are also discussed in untreated and treated hypertension patients.²⁸ These collaborative results not only improve the understanding of vascular disease mechanisms, but also provide valuable insights for clinical practice.

The collaboration between Seungik Vorp. Baek and David A., in the field of vascular research, are reflected mainly in the joint research projects and published academic papers. They have collaborated on a number of occasions, including a special issue of Annals of Biomedical Engineering exploring cardiovascular biomechanics and biofluids, in particular, modeling cardiovascular structures.²⁹ In addition, they collaborated on the publication of research on multiscale modeling of blood vessels via a fluid–solid growth framework, which involves advances in medical imaging, vascular biology, and biomechanics and is expected to play an increasingly important role in clinical

Table 2
Top 10 authors in terms of publication volume from 2004 to 2024.

Authors	Counts	Affiliations	Countries
Jay D. Humphrey	61	Yale University	USA
Marijana. Tadic	25	Tadic, Marijana	GERMANY
Dao Wen. Wang	24	Union Hospital, Tongji Medical College, Huazhong University of Science and Technology	PEOPLES R CHINA
Cesare. Cuspidi	21	Università Milano-Bicocca	ITALY
Rudolph L. Gleason	20	Georgia Institute of Technology	USA
Hai-Chao. Han	17	University of Texas at San Antonio	USA
Seungik. Baek	16	Michigan State University	USA
Chen. Chen	16	Xidian University	PEOPLES R CHINA
David A. Vorp	15	University of Pittsburgh Swanson School of Engineering	USA
Gerhard A. Holzapfel	15	Graz University of Technology	AUSTRIA

decision-making processes. Their collaboration highlights the importance of applying biomechanics and biofluidics to the progression of vascular disease and shows how to analyze aneurysm enlargement or wall stress through computational models. These collaborative results not only promote an understanding of the biomechanics of vascular diseases but also provide a new perspective and method for clinical treatment. The cooperative work of Chen, Chen and Wang, Dao Wen focused mainly on the mechanical pathogenic mechanism involved in the occurrence and development of HF and revealed a series of mechanically regulated signaling pathways.^{30–32} The roles of miR-665, mammalian target protein complex 1 (mTORC1) and phosphodiesterase 5 (PDE5) in heart failure are closely related to the biomechanical properties of the heart. MiR-665 affects cardiac function by affecting phosphatase and tensin cognates, mTORC1 affects vascular tone and blood pressure via Yes-associated proteins, and PDE5 inhibition prevents cardiac fibrosis by affecting Smad signaling, all of which are important components of cardiac biomechanics research. Understanding how these molecules and signaling pathways regulate the mechanical properties of the heart could provide new strategies for the treatment of heart disease.

3.3. Research hotspot analysis

High-frequency keywords often reflect research hotspots in the field. To further study the current research hotspots and trends of mechanobiology in cardiovascular homeostasis and remodeling, a keyword co-occurrence network diagram (Fig. 7) was generated through CiteSpace, which contains 597 nodes and 6016 connections. The top 10 most frequently occurring keywords included "extracellular matrix", "shear stress", "smooth muscle cells", "heart failure", "endothelial cells", "blood pressure", "blood flow", "computational fluid dynamics", "myocardial infarction", and "hypertension", indicating that these are the most studied directions by researchers in the past 20 years. The research hotspots in the field of cardiovascular homeostasis and remodeling in mechanobiology influence each other. The fluid shear stress generated by blood flow mainly acts on ECs, and the mechanical load caused by blood pressure mainly acts on SMCs and cardiomyocytes. These cardiovascular cells regulate the remodeling process of the ECM in a paracrine manner. Cardiovascular cells can also influence each other in a paracrine manner. These findings indicate that cardiovascular remodeling and homeostasis constitute a complex overall process that is inseparable from the division of labor and cooperation among various cardiovascular components. On the basis of the top five most frequent keywords in the keyword co-occurrence network map, the following focuses on the role of the biological behavior of ECs, smooth muscle cells and the ECM under

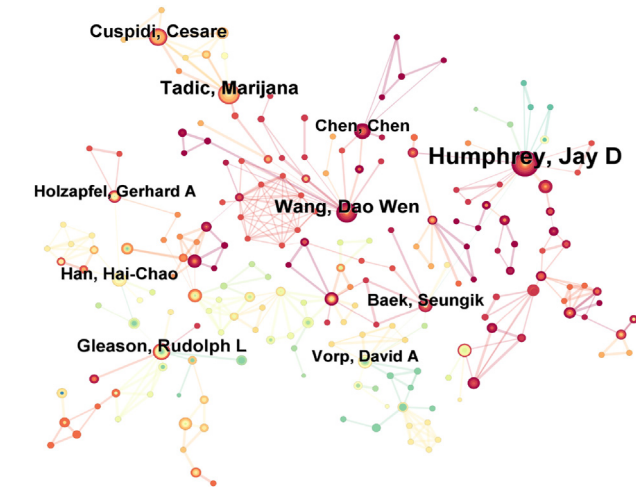


Fig. 6. Author collaboration network in the field of mechanobiology in cardiovascular homeostasis and remodeling from 2004 to 2024.

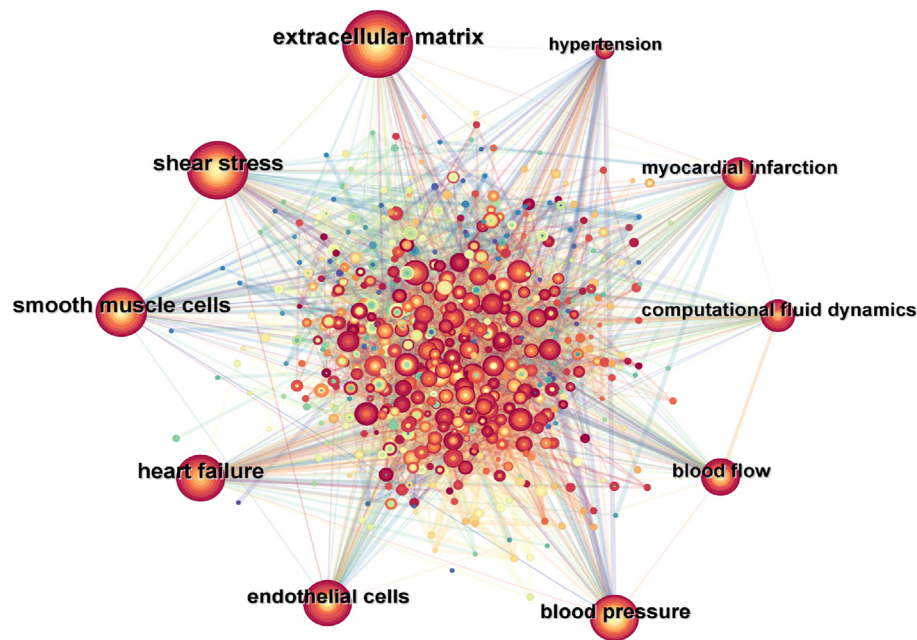


Fig. 7. Cooccurrence of key words in the field of mechanobiology in cardiovascular homeostasis and remodeling from 2004 to 2024.

mechanical stimulation in cardiovascular remodeling and homeostasis, as well as the mechanical regulatory mechanism in the occurrence and development of HF.

3.3.1. Mechanobiology of the endothelial cell response to shear stress

ECs, as the cells in direct contact with blood in vessels, are continuously exposed to hemodynamics and play an important role in the regulation of cardiovascular homeostasis and remodeling by mechanical stimulation, which has become the focus of researchers. Shear stress caused by blood flow is the main mechanical force sensed by ECs.^{17,33} ECs have excellent mechanosensitive properties and mechanotransduction ability and can continuously sense the magnitude, pulsation and direction of shear stress.^{8,34,35} ECs convert mechanical signals into intracellular biochemical signals through specific mechanosensitive sensors, such as ion channels, integrins, the glycocalyx and adhesion molecules.^{33,36} This signal transduction process causes changes in the morphology, structure, function and gene expression of ECs and subsequently affects cardiovascular homeostasis or remodeling.^{37–39} The hemodynamics of the cardiovascular system is complex, and shear stress manifests itself in many forms. In the vertical part of the cardiovascular system, blood flow is in a single direction, and shear stress manifests itself as laminar shear stress (LSS),⁴⁰ whereas in arterial bifurcations, bends, or valves, where blood flow is weaker and direction changes are complex, shear stress manifests itself as low oscillatory shear stress (OSS).^{41,42} Different forms of shear stress stimulate ECs and cause different biological processes. Many studies have reported the protective effects of physiological LSS on endothelial function, including promoting the formation of tight cell junctions in ECs and exerting normal barrier functions.^{43,44} It also regulates the balance of vasodilator and vasomotor substances secreted by ECs to maintain normal vascular tension.^{45,46} Many studies have also investigated the role of LSS in maintaining the anti-proliferative, anti-inflammatory and anti-thrombotic phenotypes of ECs.^{47–50} Recent studies have also shown that LSS promotes EC polarization and directional migration by regulating the remodeling of adhesion junctions between ECs and plays an important role in repairing and maintaining vascular integrity after vascular injury.⁵¹

Sustained stimulation by pathological shear stress induces endothelial dysfunction leading to cardiovascular pathological remodeling diseases, of which atherosclerosis is the most common and studied disease.^{17,38,52–54} Shear stress is the main cause of the focal distribution

of atherosclerotic lesions.^{55,56} In AS-susceptible areas, ECs are exposed to OSS and low shear stress, increasing the expression of proinflammatory factors and cell adhesion factors^{57–59} and activating the oxidative pathways that cause EC damage and the subsequent formation and development of AS lesions.^{58,60–62} In addition, OSS or low shear stress disrupts the barrier function of ECs⁶³ and promotes the deposition of lipids and inflammatory cells under the endothelium, leading to inward remodeling of the vascular wall and causing the occurrence of atherosclerotic stenosis.^{64,65} Similarly, pathological shear stress is also a major cause of heart valve disease,^{66–68} and there is a spatial correlation between the location of the lesion and hemodynamics. Studies have shown that lipid deposition and calcification of the aortic valve are more likely to occur on the aortic side,^{69–71} and shear stress manifests as low OSS, which may be related to the fact that low OSS promotes EC damage.^{72,73} Moreover, abnormal shear stress in heart valve ECs promotes endothelial-mesenchymal transformation (EndMT), causing pathological remodeling of the aortic valve and leading to valvular dysfunction.⁷⁴ Low OSS plays an important role in the pathological remodeling process of aortic aneurysms.^{75–77} Aortic aneurysms are in a disturbed blood flow environment. ECs are chronically stimulated by low OSS, and the pathological phenotype is activated, which manifests as increased oxidative stress, EndMT, and increased EC permeability.^{78,79} In addition, ECs of the pathological phenotype secrete a variety of key growth factors and matrix-regulating proteins that regulate ECM remodeling and SMC phenotypic transformation,⁸⁰ and these processes are associated with aortic aneurysm growth and rupture.^{80,81} Hemodynamic disturbances are also a major cause of poor prognosis in some surgical procedures such as angioplasty and stent implantation, with altered hemodynamics in the postprocedural region manifesting as low OSS, which promotes neoplastic endothelial proliferation by inducing EC injury.^{82–85} Abnormal shear stress-induced EC dysfunction is also the primary cause of vascular stenosis after arterial bypass grafting.⁸⁶ When veins are suddenly exposed to the shear stress in the arterial system, and the magnitude of the shear stress is much greater than the shear stress previously carried by venous ECs, EC inflammation and apoptosis are included, promoting subsequent angiogenesis and intimal hyperplasia.^{87–89} In summary, the effects of hemodynamics on EC physiology and pathology have been extensively studied and will continue to be a major focus in the future. Elucidating the role of blood flow patterns in EC dysfunction can provide insights into the pathogenesis of lesions in

specific regions of the cardiovascular system and help develop therapeutic strategies based on hemodynamics.

3.3.2. *Mechanobiology in the extracellular matrix*

The ECM is a three-dimensional structural entity within the cardiovascular system that mainly includes elastin, collagen, glycosaminoglycans (GAGs) and related proteoglycans. It is the main component of the cardiovascular microenvironment,⁹⁰ providing mechanical support for cells and maintaining the normal structure–function relationship of the cardiovascular system.^{91,92} It is also the main conductor of mechanical force within the cardiovascular system, transmitting key signals to vascular cells, cardiomyocytes and interstitial cells.^{93–95} Under physiological conditions, the ECM remains relatively stable to maintain normal cardiovascular function.⁹⁶ In pathological vascular remodeling events caused by abnormal mechanical forces, ECM remodeling is a key link, which manifests as changes in the composition of matrix proteins, matrix metalloproteinase-mediated matrix protein degradation, and changes in the topological characteristics of the ECM.^{97–99} When blood vessels are exposed to LSS, the ECM composition is altered to exhibit pro-inflammatory and pro-tissue repair phenotypes, promoting the generation of an intravascular inflammatory microenvironment.^{100,101} Increased circumferential stress can also trigger changes in the ECM components of the vascular wall, manifested as the breakage of elastic fibers and excessive deposition of collagen,^{102–105} while also affecting the rearrangement of matrix proteins and proteoglycans.^{105,106} The topological structure of the ECM can also change due to pressure overload.^{105,107} The pathological remodeling of the ECM can lead to changes in vascular structure and function, manifested as thickening of the arterial wall, increased vascular stiffness, decreased compliance, and increased peripheral resistance.^{90,106,108} A study revealed that the speed of the blood pressure pulse wave also affects the composition of the ECM, indicating that, in hypertensive patients, the relative content of collagen is positively correlated with the speed of the pulse wave, whereas the relative content of elastin is negatively correlated with it.¹⁰⁹ The occurrence and development of aortic aneurysms are related to changes in the ECM.^{110–112} Pathological remodeling of the ECM in aortic aneurysms can affect the strength of the vascular wall. When the wall stress exceeds the wall strength, aortic dissection or rupture occurs.¹¹³ Mechanical stimulation can also affect the ECM remodeling of heart valves. OSS increases the production of collagen and GAG.^{66,114} Both cyclic stretching and pressure overload lead to remodeling of the ECM of the aortic valve, causing degenerative aortic valve disease.^{115,116} Changes in the composition and structure of the ECM can also affect the response of cardiovascular cells to mechanical stimulation. Fibronectin can increase the mechanical sensitivity of cells to high shear stress, whereas the presence of collagen types I and IV and laminin can improve the mechanical sensitivity of cells to low shear stress.¹¹⁷ In addition, the topological characteristics of the ECM help to resist the inflammatory and proliferative activation of ECs by turbulent flow.¹⁰¹ In Marfan syndrome mice, the ECM structure of the heart valves is destroyed, and the proper structure–function relationship of the heart cannot be maintained under mechanical stimulation, resulting in pathological remodeling of the heart valves and myxomatous valvular disease.⁹² In summary, ECM remodeling under mechanical stimulation has been a popular topic that researchers have explored over the past 20 years. The discovery of new mechanisms and perspectives in this field is a continuous improvement in the understanding of the pathogenesis of cardiovascular pathological remodeling diseases.

3.3.3. *Mechanobiology in vascular smooth muscle cells*

VSMCs are the main cellular components of the tunica media of the vascular wall and are mainly subjected to mechanical stimulation generated by pulsatile blood pressure in the form of cyclical stretch.¹¹⁸ VSMCs are also important participants in vascular homeostasis and remodeling under mechanical stimulation.^{119–121} VSMCs have a high degree of plasticity. Under vascular homeostasis, VSMCs exhibit a

contractile phenotype to maintain normal vascular tension and function.^{14,122} However, in a hemodynamic environment that promotes pathological vascular remodeling, VSMCs undergo phenotypic switching and differentiate into a more migratory, secretory, and proliferative synthetic phenotype, promoting the formation of vascular lesions.^{123–125} An increasing number of studies have shown that mechanical stimulation regulates VSMC phenotypic switching through various mechanisms.¹²⁶ In the event of an acute increase in blood pressure, smooth muscle cells contract rapidly to maintain normal wall stress.¹²⁷ In arterial remodeling induced by chronically high blood pressure, the contribution of vascular smooth muscle is most pronounced, including chronic vascular constriction and luminal narrowing due to dysfunction and aberrant proliferation of SMCs, as well as vascular fibrosis resulting from the dedifferentiation of SMCs into myofibroblasts.^{126,128,129} In addition, vascular smooth muscle is a key regulator of ECM remodeling after increased circumferential stress. It participates in vascular remodeling by secreting a variety of matrix metalloproteinases to regulate the remodeling of the ECM,^{130,131} causing pathological remodeling of conduction arteries and resistance arteries.¹³⁰ The abnormal adaptation of SMCs to increased circumferential stress is also the main cause of pathological remodeling in aortic aneurysms.^{132,133} In addition, VSMCs respond to mechanical stimulation of increased blood pressure by altering cell adhesion and the expression of ECM molecules.^{134,135} In a healthy state, VSMCs are not directly exposed to shear stress. When atherosclerotic lesions rupture or invasive techniques such as angioplasty cause damage to ECs, VSMCs are directly exposed to blood flow, and receiving shear stress stimulation, leading to corresponding changes in intracellular molecular signaling and function. Physiological LSS inhibits the proliferation of VSMCs by increasing the expression of nitric oxide synthase and AMP-activated protein kinase phosphorylation.¹³⁶ However, low shear stress and OSS promote the proliferation and migration of VSMCs to the intima, resulting in an EC phenotype, with a significant increase in the expression of EC markers,¹³⁷ which contributes to the formation of focal neointima and promotes the formation of atherosclerotic stenosis lesions.^{138–140} After endarterectomy, the exposure of VSMCs to high LSS leads to increased apoptosis, which helps reduce restenosis after vascular injury.¹⁴¹ Similarly, in vascular grafts, high shear stress can inhibit smooth muscle cell proliferation and reduce neointima formation.¹²⁴ In summary, the phenotypic switching of VSMCs under mechanical stimulation has always been a hot area of research for researchers, and its mechanism of action may be a potential therapeutic target in the future.

3.3.4. *Mechanobiology in heart failure*

HF is a clinical syndrome and the endpoint of remodeling in most cardiovascular diseases.¹⁴² Clinical manifestations include impaired cardiac pumping function and the inability of cardiac output to meet the metabolic needs of systemic tissues. Pathological features include ventricular enlargement and relative thinning of the ventricular wall.¹⁴³ In the early stage of mechanical overload, the heart is able to maintain cardiac output and cardiac systolic and diastolic function to maintain cardiovascular homeostasis through compensatory mechanisms of myocardial hypertrophy. However, as mechanical overload persists, cardiac remodeling becomes decompensated hypertrophy, accompanied by changes in structure, molecules, metabolism, and the ECM, leading to cardiac dysfunction and eventually HF.¹⁴⁴ Both pressure and volume overload are important pathogenic factors of HF,¹⁴⁵ leading to progressive remodeling of ventricular geometry and myocardial structure. These remodeling processes affect the mechanical properties of the heart, which in turn aggravates the cardiac remodeling process and eventually results in HF. Excessive ECM deposition is observed in HF induced by pressure overload, contributing to ventricular wall thickening,^{146,147} whereas increased ECM degradation contributes to ventricular wall thinning and ventricular cavity dilation in HF induced by volume overload.¹⁴⁸ In the context of myocardial hypertrophy and dysfunction caused by pressure overload, many studies have shown that inflammation is an important pathogenic mechanism that causes cardiac

dysfunction by inducing cardiomyocyte death and damage.^{149–152} In addition, mitochondrial dysfunction is a key pathogenic factor. Pressure overload promotes the development of HF by downregulating mitochondrial autophagy and causing mitochondrial dysfunction.¹⁵³ Similarly, the activation of the cardiac immune response mechanism is also involved in adverse cardiac remodeling and HF caused by mechanical overload.¹⁵⁴ Studies have shown that under the action of hemodynamic mechanical forces, metabolic remodeling of the heart precedes structural remodeling. Pressure overload reduces the expression of cardiac pyruvate kinase muscle isoenzymes, reduces the utilization of glucose by cardiomyocytes, and subsequently leads to cardiomyocyte death, aggravating myocardial dysfunction and fibrosis and ultimately causing HF.¹⁵⁵ Recent studies have shown that hypoxia can protect against the development of cardiac hypertrophy and HF caused by pressure overload by increasing the expression of hypoxia-inducible factor-1 α and reducing the expression of genes related to pathological remodeling.¹⁵⁶ Many studies have also been conducted on the pathological remodeling mechanisms of HF volume overload. During volume overload, the left ventricle undergoes extensive remodeling, manifested as chamber dilation, increased myocardial wall stress, and progressive myocardial cell contractile dysfunction. The mechanisms involved include myocardial and EC apoptosis, altered cardiac fibroblast function, and extensive ECM remodeling.¹⁴⁸ Volume overload also leads to the upregulation of metabolic genes related to myocardial stress and glycolysis in the ventricle.¹⁵⁷ Hypertension and adverse ventricular remodeling after myocardial infarction can lead to HF. Hemodynamic changes after acute myocardial infarction cause pathological remodeling of cardiomyocytes, leading to left ventricular systolic and diastolic dysfunction.¹⁵⁸ In hypertension, enhanced cyclic stretch induces a hypertrophic response in cardiomyocytes by inducing NF- κ B activation and increasing VEGF secretion, promoting pathological myocardial remodeling.¹⁵⁹ Research on the mechanobiology of heart failure has focused mainly on the influence of mechanical regulatory mechanisms under volume overload and pressure overload on the biological behavior of cardiac cells and ECM, which will continue to be a direction of interest in the future.

3.4. Emerging trends analysis

Keywords with the strongest citation bursts (Fig. 8) can indicate emerging research hotspots and provide a basis for exploring future research trends and cutting-edge topics. In the research on mechanobiology in cardiovascular homeostasis and remodeling, the keywords whose citation bursts have continued to the present are responses, homeostasis, fibrosis, and mechanisms, indicating that these four keywords may represent future research trends. In recent years, the mechanical regulatory mechanisms of cardiovascular remodeling and homeostasis have been continuously discovered, among which mechanosensitive transcription factors and mechanosensitive ion channel piezo1 have received the most attention. In the past, researchers focused their attention on the effects of mechanical stimulation on cardiovascular remodeling. In recent years, researchers have gradually focused on the role of mechanical regulation in cardiovascular homeostasis. In cardiac remodeling, cardiac fibrosis has recently attracted increasing attention from researchers, and mechanical-related regulatory mechanisms have been continuously discovered. The following is a detailed introduction to the research progress of these emerging research hotspots in the mechanobiology of cardiovascular remodeling and homeostasis.

3.4.1. Mechanotransduction transcription factors

A core mechanism by which mechanical force affects cell behavior involves changes in gene expression, and various mechanosensitive transcription factors play key roles in this process. Mechanical stimulation regulates gene expression by activating different mechanosensitive transcription factors, thereby affecting the function of cardiovascular cells.^{160,161} These include Krüppel-like factor 2, Krüppel-like factor 2, NF- κ B and Yes associated protein (YAP)/WW domain-containing transcription regulator 1 (TAZ, also known as WWTR1)/TEA-domain transcription factor (TEAD).^{162–165} In recent years, researchers have focused on the effect of mechanical stimulation on YAP/TAZ/TEAD and its role in cardiovascular homeostasis and remodeling and have obtained many valuable findings. YAP/TAZ, mechanosensitive transcriptional

Top 20 Keywords with the Strongest Citation Bursts

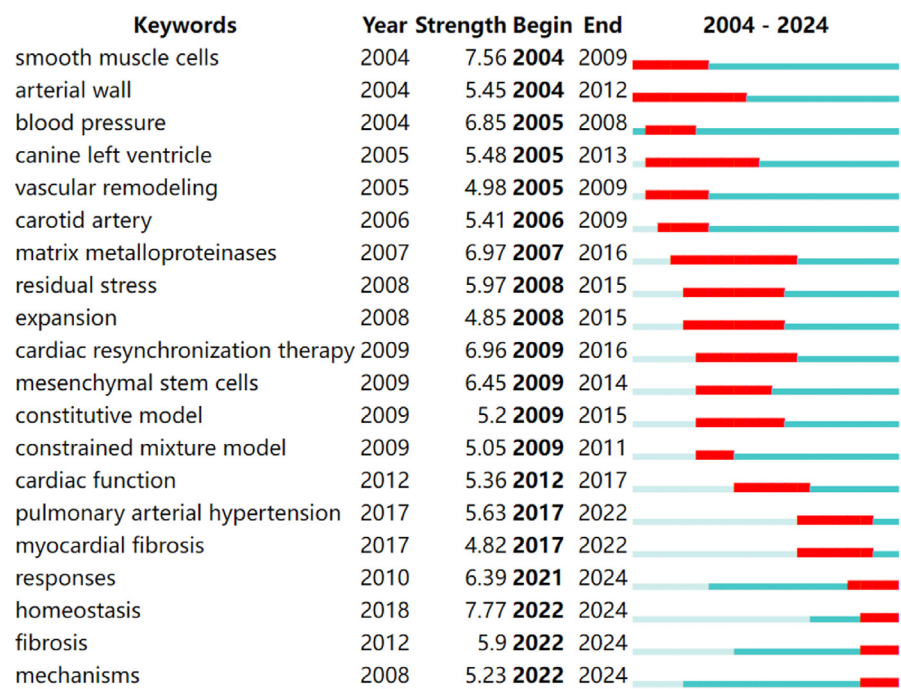


Fig. 8. Top 20 keywords with the strongest citation bursts in the field of mechanobiology in cardiovascular homeostasis and remodeling from 2004 to 2024.

coactivators that regulate the TEAD family of transcription factors, are the final effector molecules downstream of Hippo signaling.¹⁶⁶ It plays an integral role in the development of the cardiovascular system and the maintenance of cardiovascular homeostasis.^{167–169} Many studies have shown that it is the most important mechanosensitive transcription factor activated by disturbance,^{170–173} and can shuttle between the cytoplasm and the nucleus to respond to mechanotransduction.¹⁷² Changes in cell morphology, rigidity and topology of the ECM, shear stress and stretching can affect the distribution of YAP within the cell and the corresponding biological effects.¹⁷⁴ The role of YAP in the mechanical conduction of ECs has been continuously studied and has attracted increasing attention. Studies have shown that LSS can inhibit YAP activity in ECs, thereby inhibiting inflammation and playing an anti-atherosclerotic role, whereas OSS induces YAP activation and translocation into the nucleus of ECs, leading to increased EC proliferation and monocyte adhesion factor expression, resulting in the occurrence of key pathogenic events in atherosclerosis, such as intimal thickening and foam cell formation.^{173,175} These studies have laid a theoretical foundation for the use of YAP as a therapeutic target for mechanically stimulated atherosclerotic diseases. In myocardial ischemia–reperfusion injury, LSS exerts a protective effect on microvascular ECs by activating YAP and alleviating myocardial injury.¹⁷⁶ In hypertension, blood pressure overload increases YAP nuclear translocation in vascular ECs, promoting oxidative stress and inhibiting NO bioavailability, leading to endothelial dysfunction.¹⁷⁷ In heart valves, YAP activation is also affected by mechanical forces, affecting valve growth and remodeling.⁶⁷ In addition to ECs, YAP/TAZ also play important roles in the mechanotransduction of smooth muscle cells. Proper YAP/TAZ mechanotransduction in VSMCs is a key factor in maintaining the contractile phenotype and function of vascular smooth muscle and is essential for maintaining vascular homeostasis. In pressure overload, SMC-specific YAP/TAZ loss leads to decreased vasocontractility, impaired myogenic response, and increased compliance.^{178,179} In addition, YAP/TAZ loss also affects the regulation of ECM remodeling by SMCs under pressure overload, which manifests as reduced assembly of elastic fibers and increased degradation of elastin, leading to the formation of aneurysms.^{180,181} Pathological mechanical stretch downregulates enhancer of zeste homolog 2 via activation of the YAP-TEAD1 pathway, thereby exacerbating VSMC apoptosis and vascular remodeling.¹⁸² These studies suggest that the balance of YAP/TAZ activation represents the balance of vascular cell function and that the deletion or increased nuclear translocation of YAP/TAZ causes the activation of downstream pathogenic signaling pathways, leading to the occurrence of vascular pathological remodeling. At present, the role of YAP/TAZ in cardiovascular homeostasis and remodeling is still in the stage of continuous in-depth study. In the future, many studies are needed to elucidate its specific mechanism of action and explore the regulatory factors affecting its activation/inactivation balance to provide a new perspective for the treatment of cardiovascular remodeling diseases.

3.4.2. Piezo1 in mechanobiology

Piezo1 is a mechanosensitive cationic channel on the cell membrane that responds to different mechanical forces and converts them into intracellular signals, participating in the occurrence and development of cardiovascular remodeling diseases.^{183–185} In recent years, its pathogenic mechanism in cardiovascular remodeling induced by mechanical stimulation has been continuously explored and enriched. In the heart, piezo1 is a key mechanical sensor that initiates mechanical-chemical signal transduction to cause cardiac remodeling.^{186,187} Abnormal mechanical stretch upregulates piezo1 in cardiomyocytes, and the mechanical signal is converted into an intracellular oxidative stress signal.¹⁸⁸ Piezo1 is an important regulator of stretch-induced the expression of genes related to cardiac remodeling in cardiac fibroblasts.¹⁸⁹ After myocardial infarction, abnormal mechanical stress and mechanical load on the myocardium cause piezo1 upregulation, which induces pathological ventricular

remodeling by activating multiple signaling pathways, including those that mediate inflammatory cascades¹⁹⁰ and disrupt intracellular calcium cycling dynamics in cardiomyocytes.¹⁹¹ In pulmonary hypertension, enhanced cyclic stretch promotes endothelial–mesenchymal transition and stimulates SMC proliferation by upregulating piezo1,¹⁹² whereas high shear stress caused by high flow causes the upregulation of piezo1 in ECs and activates endothelial inflammation.¹⁹³ The pressure overload-induced activation of piezo1 causes platelet activation and thrombosis, promoting the occurrence of hypertensive complications.¹⁹⁴ In addition, under pressure overload, piezo1 upregulation disrupts cardiomyocyte calcium homeostasis causing myocardial hypertrophic remodeling.¹⁹⁵ The role of piezo1 in AS has been extensively studied, and many new discoveries concerning the role of piezo1 in hemodynamically related atherosclerosis have recently been reported. The upregulation of piezo1 under OSS causes the activation and nuclear translocation of YAP, thereby mediating endothelial dysfunction and promoting the formation of atherosclerotic lesions.^{196–198} In advanced atherosclerotic lesions, the activation of piezo1 is associated with increased plaque stability. The activation of piezo1 promotes foam cell apoptosis, reduces the expression of inflammatory factors, and increases the collagen content.¹⁹⁹ In addition, piezo1 has been found to be the target of many traditional Chinese medicines for the treatment of atherosclerosis which delay the progression of atherosclerotic lesions by inhibiting the expression of piezo1.^{198,200} Activated neutrophils release neutrophil extracellular traps (NETs) to mediate inflammation, which plays an important role in the formation of early atherosclerotic lesions. A recent study revealed that low shear stress downregulates piezo1 to promote the generation of NETs.²⁰¹ In the response of SMCs to increased ECM rigidity, piezo1 is among the key mechanotransduction molecules whose activation causes an increase in SMC volume.²⁰² The discovery of these new mechanisms has provided some new insights into the mechanotransduction function of piezo1, which is a continuous improvement in the biological function of piezo1 and further enhances its status in cardiovascular diseases related to mechanical regulation.

3.4.3. Cardiovascular homeostasis

In the past, researchers have focused mainly on the effects of mechanical stimulation on cardiovascular remodeling, but in recent years researchers have focused on the role of mechanical regulation in cardiovascular homeostasis. Mechanical homeostasis is the most important aspect of cardiovascular homeostasis and requires a combination of mechanical stimulation and mechanotransduction induced biological responses to maintain it.^{9,203} The regulation of homeostasis is characterized by negative feedback regulation. In the cardiovascular system, when mechanical stimulation deviates from the optimal value or set value, the corresponding negative feedback regulatory mechanism is activated to restore the system to the normal range.^{204,205} Blood pressure and flow homeostasis are essential for the physiological function of the cardiovascular system to ensure normal cardiovascular perfusion to cope with changing metabolic and physiological needs.²⁰⁶ When surgical operations or changes in tissue perfusion cause blood flow to increase or decrease, ECs sense that shear stress deviates from the set point. ECs respond quickly by regulating vasoconstriction or dilation through corresponding mechanisms, thereby changing the diameter of the blood vessels, maintaining blood flow shear stress within the physiological range, and maintaining cardiovascular homeostasis.²⁰⁶ When blood pressure increases acutely, the first to sense it in arterioles are SMCs, which maintain local hemodynamic homeostasis through myogenic contraction.^{179,207} The homeostasis of the ECM is also important for cardiovascular homeostasis, which is reflected in the balance of quantity, composition, assembly and mechanical properties.^{107,208} In different layers of the vascular wall, the components of the ECM also differ in terms of site specificity, which is a prerequisite for the normal physiological function of the cardiovascular system in the mechanical environment.²⁰⁹ Most ECM components are not static but rather dynamically changing, constantly produced, degraded and assembled. In this process,

the production and degradation of the ECM maintain a relatively balanced state,⁹⁵ and these relatively balanced states are the prerequisites for maintaining mechanical properties.¹⁰⁷ The state of cells is also important in the regulation of cardiovascular homeostasis, and this state is reflected in the phenotype of cells. Changes in the cell phenotype affect the perception of mechanical stimulation, thereby affecting the biological processes caused by subsequent mechanical transduction. ECs are the barrier cells of the cardiovascular system, and maintaining the normal phenotype of ECs is crucial for maintaining cardiovascular homeostasis.²¹⁰ Studies have shown that a more flexible ECM promotes longer elongation and tighter cell junctions in ECs under LSS, and promotes the production of nitric oxide, indicating that the quality of the ECM is related to the phenotype of ECs. Furthermore, suggests that the homeostasis of the interaction between the ECM and ECs is crucial for the maintenance of vascular homeostasis under hemodynamic conditions.²¹¹ Maintaining the contractile phenotype of VSMC is beneficial for resisting the pathological remodeling of the ECM caused by pressure overload and preventing the occurrence of aortic aneurysms.^{212,213} Mitochondria are the energy centers of cells, and their homeostasis is crucial for the maintaining of the contractility phenotype of SMCs.²¹⁴ The homeostasis of the interaction between SMCs and the ECM can maintain normal vascular compliance and prevent arteriosclerosis.¹¹⁸ Recent studies have shown that maintaining glutathione homeostasis in SMCs can prevent VSMC dysfunction and is an important means to improve pathological remodeling of the ECM in atherosclerosis and offset arterial stiffness.²¹⁵ In the future, the mechanical regulatory mechanisms of cardiovascular homeostasis will be revealed by an increasing number of studies, and the focus will gradually be refined from overall cardiovascular homeostasis to the homeostasis of the components of the cardiovascular system as well as the interactions among the components.

3.4.4. Mechanically regulated cardiac fibrosis

Cardiac fibrosis is the pathophysiological basis of HF.^{151,216} In recent years, many studies have investigated the role of mechanical regulation in cardiac fibrosis. Cardiac fibrosis is significantly related to hemodynamic changes. Pathological hemodynamics drives cardiac fibroblasts to differentiate into myofibroblasts through mechanotransduction,^{217–219} causing excessive accumulation of ECM components and secretion of pro-fibrotic factors.^{220–222} Myocardial mechanical overload has been confirmed to be the main driver of myocardial fibrosis in living myocardial slices that preserve structure and function.²²³ Abnormalities in cardiac mechanics can also promote cardiac fibrosis. In patients with mitral valve prolapse, myocardial fibrosis was found to be associated with impaired myocardial longitudinal strain caused by mitral valve prolapse.²²⁴ Another study revealed that decreased myocardial mechanical parameters, such as global longitudinal strain, circumferential strain and myocardial strain, can reflect the severity of myocardial fibrosis.²²⁵ Activated fibroblasts and myofibroblasts are the main cellular effectors of cardiac fibrosis.^{226,227} Studies have shown that ECs may also acquire a fibrotic phenotype through endothelial–mesenchymal transition under mechanical overload conditions, thereby promoting cardiac fibrosis.²²⁸ In pressure overload mediated cardiac fibrosis, macrophage-mediated inflammation also plays a role in the differentiation of fibroblasts into myofibroblasts.²²⁹ Many studies have confirmed that there is two-way communication between fibroblasts and the ECM. The mechanical properties of the ECM affect the behavior of fibroblasts, and the state of fibroblasts will also regulates the remodeling of the ECM. There is also a positive feedback loop between the two.^{230,231} A decrease in the viscoelasticity of the ECM and an increase in linear elasticity will activate the mechanically sensitive pathway and cause fibroblast activation.^{232,233} Activated fibroblasts are the main source of ECM proteins, causing increased ECM remodeling.^{167,234} Cardiac fibroblasts can also sense mechanical stress through mechanosensitive receptors, ion channels, and integrins, activating the intracellular fibrosis cascade and leading to fibrosis caused by pressure overload.^{235–239} The transformation of cardiac fibroblasts into myofibroblasts under pressure

overload is significantly associated with intracellular metabolic changes, manifested by increased aerobic glycolysis and lactate production.²⁴⁰ A recent study revealed that pressure overload activates reactive oxygen species production pathways to induce mitochondrial oxidative damage and promote myocardial fibrosis.²⁴¹ Exosomes also play important roles in cardiac fibrosis caused by pressure overload. Enhanced mechanical stretch induces cardiomyocytes to regulate the activation of cardiac fibroblasts through an exosome-mediated paracrine mechanism, thereby promoting cardiac fibrosis.²⁴² Crosstalk between fibroblasts and cardiomyocytes plays an important role in the process of cardiac fibrosis. A recent study revealed that during volume overload-induced cardiac fibrosis, integrin β -like1 is upregulated in fibroblasts, promoting cardiomyocyte hypertrophy.²⁴³ Mechanical stimulation is an important driving factor of cardiac fibrosis. Although the mechanical-related pathogenic mechanism has received much attention from researchers in recent years, there is still much room for research in the future, including mechanism regulating the crosstalk between cardiomyocytes and fibroblasts under mechanical stimulation, as well as the positive feedback regulation between fibroblast activation and ECM remodeling.

4. Summary and prospects

In this study, we used CiteSpace bibliometric analysis software to analyze the scientific literature concerning the mechanobiology of cardiovascular homeostasis and remodeling over the past 20 years and quantitatively and visually reviewed the research achievements and progress in this field. First, we conducted a quantitative analysis of basic information such as annual publication volume, authors, institutions, and countries. Second, we conducted a co-occurrence analysis of keywords, obtained hot keywords in the field, and summarized the research progress of hot keywords. Finally, we analyzed the citation burst keywords, obtained the emerging research directions in the field, and explained its current research status.

Cardiovascular disease is currently the leading cause of death worldwide, and most cardiovascular diseases are associated with pathological remodeling caused by mechanical stimulation.^{244,245} Over the past 20 years, owing to the continuous in-depth research on cardiovascular mechanobiology in various countries and the continuous development of technical means such as cell biology, molecular biology, in vitro mechanical stimulation models and mechanical simulation software, the number of published articles in this field has steadily increased. Among them, endothelial dysfunction induced by abnormal shear stress is the most concerning aspect for researchers, and many studies have been conducted on this topic. The focus of the research is this regulatory mechanism of abnormal stress on EC phenotypes. A large number of mechanosensitive signaling cascades have been discovered, and a deep understanding of these mechanosensitive signaling molecules will provide new potential targets for the treatment of cardiovascular pathological remodeling diseases. In the cardiovascular system, biological processes under mechanical stimulation are regulated by interactions between components within the system, including interactions between cardiovascular cells and the interaction between the ECM and cardiovascular cells. In addition, under mechanical stimulation, there are interactions between vascular cells that mediate vascular homeostasis. When blood flow increases and shear stress increases, ECs secrete vaso-dilator cytokines, inducing rapid relaxation of VSMCs and expansion of the arterial diameter, thereby restoring wall shear stress to the initial level.²⁴⁶ The ECM can also initiate mechanical signaling through matrix–cell interactions to affect the behavior and function of vascular cells, playing a vital role in cardiovascular homeostasis and remodeling.^{99,118,204} The quality and quantity of the ECM also affect the stiffness of blood vessels.¹⁰⁷ ECM–cell mechanotransduction interactions are mediated by focal adhesions and integrins. Focal adhesions connect the ECM and the cytoskeleton, and integrins transmit information about the chemical composition and mechanical state of the ECM to cells.^{247,248} The most critical aspect of the effects of abnormal mechanical

stimulation on cardiovascular cells, the ECM and the interactions between them is the process of cellular mechanotransduction.^{58,100} Mechanosensitive channels and mechanosensitive transcription factors play important roles in mechanotransduction and act as intermediate hubs for mechanical stimulation to elicit biological effects in cells.¹⁶¹

Cardiovascular remodeling diseases such as hypertension and HF usually occur under abnormal mechanical stimulation through positive feedback. Abnormal mechanical stimulation activates related signaling pathways, such as inflammation, oxidative stress, and immunity, causing pathological remodeling, such as vascular stiffness and myocardial hypertrophy, and this structural and functional remodeling further promotes the activation of pathogenic signaling pathways.²⁴⁹ Among these research hotspots and emerging research directions, mechanobiology has been continuously improved in cardiovascular homeostasis and remodeling, more mechanisms will be discovered in the future, and more potential therapeutic targets are expected to be identified in the clinic.

Although mechanobiology-related research on cardiovascular homeostasis and remodeling has made good progress in the past 20 years, there are still some challenges and difficulties in the future development process, which need to be solved and explored by researchers in the future. (1) More in depth, multimomics, and multidimensional research should be conducted on the mechanobiology of cardiovascular homeostasis and remodeling. Research progress on the role of mechanical stimulation in EC and VSMC dysfunction has revealed that its mechanism of action and effects, including mechanical sensing and mechanical transduction, have been continuously studied in depth. An increasing number of membrane-localized mechanical sensors and intracellular signaling molecule pathways have been discovered.^{250,251} Conducting more in-depth and comprehensive research will provide a theoretical basis for the development of new drugs and enable these findings to truly enter the clinic, which is highly important for improving the precision treatment of cardiovascular remodeling diseases. (2) The transmission of mechanical force within blood vessels requires physically interconnected structures, such as the ECM, focal adhesions, cytoskeleton and cell nucleus.^{99,252} The interaction mechanisms involved will require more research in the future, which will help to fully understand the mechanobiology of cardiovascular homeostasis and remodeling. (3) Currently, a complete quantitative system for evaluating the severity of lesions via mechanical stimulation is lacking. Many articles have reported the role of mechanical stimulation, such as pressure, circumferential wall stress, and shear stress, in diseases such as atherosclerosis, hypertension, and HF. However, it is still difficult to obtain accurate values at the lesion site, which limits its clinical application. Currently, most of them are simulated through computational fluid dynamics (CFD).^{253,254} The accuracy of the numerical values depends largely on the accuracy of CFD modeling, which limits the precise quantification of the relationship between mechanical stimulation and the severity of lesions. In the future, technology can be continuously improved to improve the accuracy of computational fluid dynamics simulations. In combination with medical imaging, advanced computational fluid dynamics and flow field testing technologies, cardiovascular system modeling and quantitative analysis can be carried out to explore new noninvasive detection technologies for cardiovascular diseases and personalized treatment and surgical design to provide mechanobiology solutions for the diagnosis, treatment and early warning of cardiovascular diseases.

CCRediT authorship contribution statement

Wei Liao: Visualization, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. **Yuxi Huang:** Visualization, Software, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. **Xiangxiu Wang:** Visualization, Software, Methodology, Investigation. **Ziqiu Hu:** Software, Investigation. **Chuanrong Zhao:** Writing – review & editing, Writing – original draft, Visualization, Resources, Project administration, Methodology, Investigation, Funding acquisition, Conceptualization. **Guixue Wang:** Writing – review &

editing, Writing – original draft, Resources, Project administration, Funding acquisition, Conceptualization.

Ethical approval

This study does not contain any studies with human or animal subjects performed by any of the authors.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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References

1. Fung Y-c. *Biomechanics: Motion, Flow, Stress, and Growth*. Springer Science & Business Media; 2013.
2. Jiang Z-L. Mechanobiology research in China. *Mechanobiology in Medicine*. 2023; 1(1):100002.
3. Wang Y, Shyy JY, Chien S. Fluorescence proteins, live-cell imaging, and mechanobiology: seeing is believing. *Annu Rev Biomed Eng*. 2008;10:1–38.
4. Zhao C, Wang X, Wang G. Hot topics and emerging trends in mechanobiology research. *Sichuan Da Xue Xue Bao Yi Xue Ban*. 2024;55(1):1–5 (in Chinese).
5. Zhang HZC, Wang G. Advances in vascular biomechanics and mechanobiology. *Medical Biomechanics*. 2024;39(1):17–23 (in Chinese).
6. Martinac B, Nikolaev YA, Silvani G, et al. Cell membrane mechanics and mechanosensory transduction. *Curr Top Membr*. 2020;86:83–141.
7. Weber KT, Anversa P, Armstrong PW, et al. Remodeling and repair of the cardiovascular system. *J Am Coll Cardiol*. 1992;20(1):3–16.
8. Baeyens N, Schwartz MA. Biomechanics of vascular mechanosensation and remodeling. *Mol Biol Cell*. 2016;27(1):7–11.
9. Humphrey JD, Schwartz MA. Vascular mechanobiology: homeostasis, adaptation, and disease. *Annu Rev Biomed Eng*. 2021;23:1–27.
10. Nakamura M, Sadoshima J. Mechanisms of physiological and pathological cardiac hypertrophy. *Nat Rev Cardiol*. 2018;15(7):387–407.
11. Wang X, Khalil RA. Matrix metalloproteinases, vascular remodeling, and vascular disease. *Adv Pharmacol*. 2018;81:241–330.
12. Klein LW. Atherosclerosis regression, vascular remodeling, and plaque stabilization. *J Am Coll Cardiol*. 2007;49(2):271–273.
13. Schiffrin EL. How structure, mechanics, and function of the vasculature contribute to blood pressure elevation in hypertension. *Can J Cardiol*. 2020;36(5):648–658.
14. Chen R, McVey DG, Shen D, Huang X, Ye S. Phenotypic switching of vascular smooth muscle cells in atherosclerosis. *J Am Heart Assoc*. 2023;12(20):e031121.
15. Mohindra R, Agrawal DK, Thankam FG. Altered vascular extracellular matrix in the pathogenesis of atherosclerosis. *J Cardiovasc Transl Res*. 2021;14(4):647–660.
16. Souilhol C, Harmsen MC, Evans PC, Krenning G. Endothelial-mesenchymal transition in atherosclerosis. *Cardiovasc Res*. 2018;114(4):565–577.
17. Chiu JJ, Chien S. Effects of disturbed flow on vascular endothelium: pathophysiological basis and clinical perspectives. *Physiol Rev*. 2011;91(1):327–387.
18. Jia M, Li Q, Guo J, et al. Deletion of BACH1 attenuates atherosclerosis by reducing endothelial inflammation. *Circ Res*. 2022;130(7):1038–1055.
19. Thompson AAR, Lawrie A. Targeting vascular remodeling to treat pulmonary arterial hypertension. *Trends Mol Med*. 2017;23(1):31–45.
20. Chen C, Hu Z, Liu S, Tseng H. Emerging trends in regenerative medicine: a scientometric analysis in CiteSpace. *Exp Opin Biol Ther*. 2012;12(5):593–608.
21. Liu X, Zhao S, Tan L, et al. Frontier and hot topics in electrochemiluminescence sensing technology based on CiteSpace bibliometric analysis. *Biosens Bioelectron*. 2022;201:113932.
22. Tan L, Wang X, Yuan K, et al. Structural and temporal dynamics analysis on drug-eluting stents: history, research hotspots and emerging trends. *Bioact Mater*. 2023; 23:170–186.

23. Yuan K, Deng C, Tan L, et al. Structural and temporal dynamics analysis of zinc-based biomaterials: history, research hotspots and emerging trends. *Bioact Mater*. 2024;35:306–329.
24. Chen X, Liu Y. Visualization analysis of high-speed railway research based on CiteSpace. Article. *Transp Policy*. 2020;85:1–17.
25. Chen C. Searching for intellectual turning points: progressive knowledge domain visualization. *Proc. Natl. Acad. Sci. U.S.A.* 2004;101(Suppl 1):5303–5310. Suppl 1.
26. Krawiec JT, Vorp DA. Adult stem cell-based tissue engineered blood vessels: a review. *Biomaterials*. 2012;33(12):3388–3400.
27. Cuspidi C, Giovannini R, Tadic M. Left atrial stiffness: a novel marker of hypertension-mediated organ damage on the horizon? *Hypertens Res*. 2021;44(3):365–367.
28. Tadic M, Cuspidi C, Grassi G, Ivanovic B. Gender-specific therapeutic approach in arterial hypertension - challenges ahead. *Pharmacol Res*. 2019;141:181–188.
29. Finol EA, Di Martino ES, Baek S. Cardiovascular biomechanics and biofluids: a special issue with a focus on modeling of cardiovascular structures. *Ann Biomed Eng*. 2013;41(7):1309–1310.
30. Fan J, Zhang X, Nie X, et al. Nuclear miR-665 aggravates heart failure via suppressing phosphatase and tensin homolog transcription. *Sci China Life Sci*. 2020;63(5):724–736.
31. Yao L, He J, Li B, et al. Regulation of YAP by mammalian target of rapamycin complex 1 in endothelial cells controls blood pressure through COX-2/mPGES-1/PGE(2) cascade. *Hypertension*. 2019;74(4):936–946.
32. Gong W, Yan M, Chen J, Chaugai S, Chen C, Wang D. Chronic inhibition of cyclic guanosine monophosphate-specific phosphodiesterase 5 prevented cardiac fibrosis through inhibition of transforming growth factor beta-induced Smad signaling. *Front Med*. 2014;8(4):445–455.
33. Davies PF. Flow-mediated endothelial mechanotransduction. *Physiol Rev*. 1995;75(3):519–560.
34. Wang C, Baker BM, Chen CS, Schwartz MA. Endothelial cell sensing of flow direction. *Arteriosclerosis, thrombosis, Vasc Biol*. 2013;33(9):2130–2136.
35. Blackman BR, Garcia-Cardena G, Gimbrone MA, et al. A new in vitro model to evaluate differential responses of endothelial cells to simulated arterial shear stress waveforms. *J Biomech Eng*. 2002;124(4):397–407.
36. Orr AW, Helmke BP, Blackman BR, Schwartz MA. Mechanisms of mechanotransduction. *Dev Cell*. 2006;10(1):11–20.
37. Hamrangsekachae M, Wen K, Bencherif SA, Ebong EE. Atherosclerosis and endothelial mechanotransduction: current knowledge and models for future research. *Am J Physiol Cell Physiol*. 2023;324(2):C488–C504.
38. Wang X, Shen Y, Shang M, Liu X, Munn LL. Endothelial mechanobiology in atherosclerosis. *Cardiovasc Res*. 2023;119(8):1656–1675.
39. Chatterjee S. Endothelial mechanotransduction, redox signaling and the regulation of vascular inflammatory pathways. *Front Physiol*. 2018;9:524.
40. Davies PF. Hemodynamic shear stress and the endothelium in cardiovascular pathophysiology. *Nat Clin Pract Cardiovasc Med*. 2009;6(1):16–26.
41. Karau KL, Krenz GS, Dawson CA. Branching exponent heterogeneity and wall shear stress distribution in vascular trees. *Am J Physiol Heart Circ Physiol*. 2001;280(3):H1256–H1263.
42. Chistiakov DA, Orekhov AN, Bobryshev YV. Effects of shear stress on endothelial cells: go with the flow. *Acta Physiol*. 2017;219(2):382–408.
43. Tarbell JM. Shear stress and the endothelial transport barrier. *Cardiovasc Res*. 2010;87(2):320–330.
44. Yang F, Zhang Y, Zhu J, et al. Laminar flow protects vascular endothelial tight junctions and barrier function via maintaining the expression of long non-coding RNA MALAT1. *Front Bioeng Biotechnol*. 2020;8:647.
45. Guo F, Li X, Peng J, et al. Autophagy regulates vascular endothelial cell eNOS and ET-1 expression induced by laminar shear stress in an ex vivo perfused system. *Ann Biomed Eng*. 2014;42(9):1978–1988.
46. Pan S. Molecular mechanisms responsible for the atheroprotective effects of laminar shear stress. *Antioxidants Redox Signal*. 2009;11(7):1669–1682.
47. Huang J, Pu Y, Zhang H, et al. KLF2 mediates the suppressive effect of laminar flow on vascular calcification by inhibiting endothelial BMP/SMAD1/5 signaling. *Circ Res*. 2021;129(4):e87–e100.
48. Psefteli PM, Kitscha P, Vizcay G, et al. Glycocalyx sialic acids regulate Nrf2-mediated signaling by fluid shear stress in human endothelial cells. *Redox Biol*. 2021;38:101816.
49. Ishii T, Warabi E, Mann GE. Mechanisms underlying unidirectional laminar shear stress-mediated Nrf2 activation in endothelial cells: Amplification of low shear stress signaling by primary cilia. *Redox Biol*. 2021;46:102103.
50. Wang Y, Sun HY, Kumar S, Puerta MDM, Jo H, Rezvan A. ZBTB46 is a shear-sensitive transcription factor inhibiting endothelial cell proliferation via gene expression regulation of cell cycle proteins. *Lab Invest*. 2019;99(3):305–318.
51. Bougaran P, Bats ML, Delobel V, et al. ROR2/PCP a new pathway controlling endothelial cell polarity under flow conditions. *Arteriosclerosis, thrombosis, Vasc Biol*. 2023;43(7):1199–1218.
52. Gimbrone Jr MA, Garcia-Cardena G. Endothelial cell dysfunction and the pathobiology of atherosclerosis. *Circ Res*. 2016;118(4):620–636.
53. Wang Y, Qiu J, Luo S, et al. High shear stress induces atherosclerotic vulnerable plaque formation through angiogenesis. *Regen Biomater*. 2016;3(4):257–267.
54. Qiu J, Lei D, Hu J, et al. Effect of intraplaque angiogenesis to atherosclerotic rupture-prone plaque induced by high shear stress in rabbit model. *Regen Biomater*. 2017;4(4):215–222.
55. J M, L N, A V, et al. Wall shear stress alteration: a local risk factor of atherosclerosis. *Curr Atheroscler Rep*. 2022;24(3):143–151.
56. Zhou M, Yu Y, Chen R, et al. Wall shear stress and its role in atherosclerosis. *Front Cardiovasc Med*. 2023;10:1083547.
57. Kim S, Woo CH. Laminar flow inhibits ER stress-induced endothelial apoptosis through PI3K/Akt-Dependent signaling pathway. *Mol Cells*. 2018;41(11):964–970.
58. Davies PF, Civelek M, Fang Y, Fleming I. The atherosusceptible endothelium: endothelial phenotypes in complex haemodynamic shear stress regions in vivo. *Cardiovasc Res*. 2013;99(2):315–327.
59. Babendreyer A, Molls L, Dreytmueller D, Uhlig S, Ludwig A. Shear stress counteracts endothelial CX3CL1 induction and monocytic cell adhesion. *Mediat Inflamm*. 2017;2017:1515389.
60. Qin X, Zhang K, Qiu J, et al. Uptake of oxidative stress-mediated extracellular vesicles by vascular endothelial cells under low magnitude shear stress. *Bioact Mater*. 2022;9:397–410.
61. Chen Z, Liu H, Zhao X, et al. Oridonin attenuates low shear stress-induced endothelial cell dysfunction and oxidative stress by activating the nuclear factor erythroid 2-related factor 2 pathway. *BMC Complement Med Ther*. 2022;22(1):180.
62. Wang Z, Wang F, Kong X, Gao X, Gu Y, Zhang J. Oscillatory shear stress induces oxidative stress via TLR4 activation in endothelial cells. *Mediat Inflamm*. 2019;2019:7162976.
63. Qu K, Wang C, Huang L, et al. Oscillatory shear stress-induced downregulation of TET1s injures vascular endothelial planar cell polarity by suppression of actin polymerization. *APL Bioeng*. 2023;7(3):036104.
64. Gulino-Debrac D. Mechanotransduction at the basis of endothelial barrier function. *Tissue Barriers*. 2013;1(2):e24180.
65. Cao R, Sun R, Ye Y, et al. Low shear stress-induced blockage of autophagic flux impairs endothelial barrier and facilitates atherosclerosis in mice. *Exp Cell Res*. 2024;439(1):114071.
66. Dayawansa NH, Baratchi S, Peter K. Uncoupling the vicious cycle of mechanical stress and inflammation in calcific aortic valve disease. *Front Cardiovasc Med*. 2022;9:783543.
67. Wang M, Lin BY, Sun S, Dai C, Long F, Butcher JT. Shear and hydrostatic stress regulate fetal heart valve remodeling through YAP-mediated mechanotransduction. *Elife*. 2023;12.
68. Fan L, Yao D, Fan Z, et al. Beyond VICs: shedding light on the overlooked VECs in calcific aortic valve disease. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*. 2024;178:117143.
69. Richards J, El-Hamamsy I, Chen S, et al. Side-specific endothelial-dependent regulation of aortic valve calcification: interplay of hemodynamics and nitric oxide signaling. *Am J Pathol*. 2013;182(5):1922–1931.
70. Weinberg EJ, Mack PJ, Schoen FJ, Garcia-Cardena G, Kaazempur Mofrad MR. Hemodynamic environments from opposing sides of human aortic valve leaflets evoke distinct endothelial phenotypes in vitro. *Cardiovasc Eng*. 2010;10(1):5–11.
71. Sun L, Rajamannan NM, Sucosky P. Defining the role of fluid shear stress in the expression of early signaling markers for calcific aortic valve disease. *PLoS one*. 2013;8(12):e84433.
72. Pang KT, Ghim M, Sarathchandra P, et al. Shear-mediated ALK5 expression regulates endothelial activation. *Biochem Biophys Res Commun*. 2023;642:90–96.
73. Parra-Izquierdo I, Sanchez-Bayuela T, Lopez J, et al. Interferons are pro-inflammatory cytokines in sheared-stressed human aortic valve endothelial cells. *Int J Mol Sci*. 2021;22(19).
74. Faure E, Bertrand E, Gaste A, Plaindoux E, Deplano V, Zaffran S. Side-dependent effect in the response of valve endothelial cells to bidirectional shear stress. *Int J Cardiol*. 2021;323:220–228.
75. Mutlu O, Salman HE, Al-Thani H, El-Menyar A, Qidwai UA, Yalcin HC. How does hemodynamics affect rupture tissue mechanics in abdominal aortic aneurysm: focus on wall shear stress derived parameters, time-averaged wall shear stress, oscillatory shear index, endothelial cell activation potential, and relative residence time. *Comput Biol Med*. 2023;154:106609.
76. Bappoo N, Syed MBJ, Khinsoe G, et al. Low shear stress at baseline predicts expansion and aneurysm-related events in patients with abdominal aortic aneurysm. *Circ Cardiovasc Imaging*. 2021;14(12):1112–1121.
77. Li STX, Qu C. The effect of wall shear stress on atherosclerosis and aneurysm. *Chin J Arteriosclerosis*. 2024;32(5):451–455 (in Chinese).
78. Hasan M, Al-Thani H, El-Menyar A, Zeidan A, Al-Thani A, Yalcin HC. Disturbed hemodynamics and oxidative stress interaction in endothelial dysfunction and AAA progression: focus on Nrf2 pathway. *Int J Cardiol*. 2023;389:131238.
79. Bjorck HM, Du L, Pulignani S, et al. Altered DNA methylation indicates an oscillatory flow mediated epithelial-to-mesenchymal transition signature in ascending aorta of patients with bicuspid aortic valve. *Sci Rep*. 2018;8(1):2777.
80. van de Pol V, Kurakula K, DeRuiter MC, Goumans MJ. Thoracic aortic aneurysm development in patients with bicuspid aortic valve: what is the role of endothelial cells? *Front Physiol*. 2017;8:938.
81. Filiberto AC, Spinoza MD, Elder CT, et al. Endothelial pannexin-1 channels modulate macrophage and smooth muscle cell activation in abdominal aortic aneurysm formation. *Nat Commun*. 2022;13(1):1521.
82. Ward MR, Tsao PS, Agrotis A, Dilley RJ, Jennings GL, Bobik A. Low blood flow after angioplasty augments mechanisms of restenosis: inward vessel remodeling, cell migration, and activity of genes regulating migration. *Arteri Throm Vasc Biol*. 2001;21(2):208–213.
83. Sanmartin M, Goicolea J, Garcia C, et al. Influencia de la tension de cizallamiento en la reestenosis intra-stent: estudio in vivo con reconstruccion 3D y dinamica de fluidos computacional. *Rev Esp Cardiol*. 2006;59(1):20–27.
84. Koskinas KC, Chatzizisis YS, Antoniadis AP, Giannoglou GD. Role of endothelial shear stress in stent restenosis and thrombosis: pathophysiologic mechanisms and implications for clinical translation. *J Am Coll Cardiol*. 2012;59(15):1337–1349.
85. Shishido K, Antoniadis AP, Takahashi S, et al. Effects of low endothelial shear stress after stent implantation on subsequent neointimal hyperplasia and clinical outcomes in humans. *J Am Heart Assoc*. 2016;5(9).

86. Liu CCF. High wall shear stress caused by arterio-venous fistula reduces neointimal hyperplasia induced by stent implantation. *Chin J Arteriosclerosis*. 2021;29(2): 129–134 (in Chinese).
87. Ward AO, Angelini GD, Caputo M, et al. NF-kappaB inhibition prevents acute shear stress-induced inflammation in the saphenous vein graft endothelium. *Sci Rep*. 2020;10(1):15133.
88. Ward AO, Caputo M, Angelini GD, George SJ, Zakkar M. Activation and inflammation of the venous endothelium in vein graft disease. *Atherosclerosis*. 2017; 265:266–274.
89. Jackson ML, Bond AR, Ascione R, Johnson JL, George SJ. FGL2/FcgammaRIIB signalling mediates arterial shear stress-mediated endothelial cell apoptosis: implications for coronary artery bypass vein graft pathogenesis. *Int J Mol Sci*. 2024; 25(14).
90. Lin PK, Davis GE. Extracellular matrix remodeling in vascular disease: Defining its regulators and pathological influence. *Arteriosclerosis, thrombosis, Vasc Biol*. 2023; 43(9):1599–1616.
91. Halper J. Basic components of vascular connective tissue and extracellular matrix. *Adv Pharmacol*. 2018;81:95–127.
92. Gonzalez BA, Harmeyer SW, Song T, Sadayappan S, Yutzey KE. Dynamic changes in mitral valve extracellular matrix, tissue mechanics and function in a mouse model of Marfan syndrome. *Matrix Biol*. 2024;126:1–13.
93. Tschumperlin DJ. Mechanotransduction. *Compr Physiol*. 2011;1(2):1057–1073.
94. Wang Y, Chatterjee E, Li G, Xu J, Xiao J. Force-sensing protein expression in response to cardiovascular mechanotransduction. *EBioMedicine*. 2024;110:105412.
95. Wang D, Brady T, Santhanam L, Gerecht S. The extracellular matrix mechanics in the vasculature. *Nat Cardiovasc Res*. 2023;2(8):718–732.
96. Mongkoldhumrongkul N, Latif N, Yacoub MH, Chester AH. Effect of side-specific valvular shear stress on the content of extracellular matrix in aortic valves. *Cardiovasc Eng Technol*. 2018;9(2):151–157.
97. Ma Z, Mao C, Jia Y, Fu Y, Kong W. Extracellular matrix dynamics in vascular remodeling. *Am J Physiol Cell Physiol*. 2020;319(3):C481–C499.
98. Yanagisawa H, Yokoyama U. Extracellular matrix-mediated remodeling and mechanotransduction in large vessels during development and disease. *Cell Signal*. 2021;86:110104.
99. Yamashiro Y, Thang BQ, Ramirez K, et al. Matrix mechanotransduction mediated by thrombospondin-1/integrin/YAP in the vascular remodeling. *Proc. Natl. Acad. Sci. U.S.A.* 2020;117(18):9896–9905.
100. Russo TA, Banuth AMM, Nader HB, Dreyfuss JL. Altered shear stress on endothelial cells leads to remodeling of extracellular matrix and induction of angiogenesis. *PLoS one*. 2020;15(11):e0241040.
101. Nakayama KH, Surya VN, Gole M, et al. Nanoscale patterning of extracellular matrix alters endothelial function under shear stress. *Nano Lett*. 2016;16(1):410–419.
102. Wagenseil JE, Mecham RP. Elastin in large artery stiffness and hypertension. *J Cardiovasc Transl Res*. 2012;5(3):264–273.
103. Kohn JC, Lampi MC, Reinhart-King CA. Age-related vascular stiffening: causes and consequences. *Front Genet*. 2015;6:112.
104. Xu Q, Chakravorty A, Bathgate RA, Dart AM, Du XJ. Relaxin therapy reverses large artery remodeling and improves arterial compliance in senescent spontaneously hypertensive rats. *Hypertension*. 2010;55(5):1260–1266.
105. Briones AM, Arribas SM, Salas M. Role of extracellular matrix in vascular remodeling of hypertension. *Curr Opin Nephrol Hypertens*. 2010;19(2):187–194.
106. Laurent S, Boutouyrie P. The structural factor of hypertension: large and small artery alterations. *Circ Res*. 2015;116(6):1007–1021.
107. Cai Z, Gong Z, Li Z, Li L, Kong W. Vascular extracellular matrix remodeling and hypertension. *Antioxidants Redox Signal*. 2021;34(10):765–783.
108. Garoffolo G, Pesce M. Vascular dysfunction and pathology: focus on mechanical forces. *Vasc Biol*. 2021;3(1):R69–R75.
109. You BA, Shen L, Li JF, Chen YG, Gu XH, Gao HQ. The correlation between carotid-femoral pulse wave velocity and composition of the aortic media in CAD patients with or without hypertension. *Swiss Med Wkly*. 2012;142:w13546.
110. Jiang WJ, Ren WH, Liu XJ, et al. Disruption of mechanical stress in extracellular matrix is related to Stanford type A aortic dissection through down-regulation of Yes-associated protein. *Aging (Albany NY)*. 2016;8(9):1923–1939.
111. Shen YH, LeMaire SA, Webb NR, Cassis LA, Daugherty A, Lu HS. Aortic aneurysms and dissections series. *Arteriosclerosis, thrombosis, Vasc Biol*. 2020;40(3):e37–e46.
112. Seim BE, Holt MF, Ratajska A, et al. Markers of extracellular matrix remodeling and systemic inflammation in patients with heritable thoracic aortic diseases. *Front Cardiovasc Med*. 2022;9:1073069.
113. Miller K, Mufty H, Catlin A, et al. Is there a relationship between stress in walls of abdominal aortic aneurysm and symptoms? *J Surg Res*. 2020;252:37–46.
114. Dahal S, Huang P, Murray BT, Mahler GJ. Endothelial to mesenchymal transformation is induced by altered extracellular matrix in aortic valve endothelial cells. *J Biomed Mater Res, Part A*. 2017;105(10):2729–2741.
115. Balachandran K, Suscosky P, Jo H, Yoganathan AP. Elevated cyclic stretch alters matrix remodeling in aortic valve cusps: implications for degenerative aortic valve disease. *Am J Physiol Heart Circ Physiol*. 2009;296(3):H756–H764.
116. Konduri S, Xing Y, Warnock JN, He Z, Yoganathan AP. Normal physiological conditions maintain the biological characteristics of porcine aortic heart valves: an ex vivo organ culture study. *Ann Biomed Eng*. 2005;33(9):1158–1166.
117. Lai A, Thurgood P, Cox CD, et al. Piezo1 response to shear stress is controlled by the components of the extracellular matrix. *ACS Appl Mater Interfaces*. 2022;14(36): 40559–40568.
118. Lacolley P, Regnault V, Segers P, Laurent S. Vascular smooth muscle cells and arterial stiffening: relevance in development, aging, and disease. *Physiol Rev*. 2017; 97(4):1555–1617.
119. Liu S, Lin Z. Vascular smooth muscle cells mechanosensitive regulators and vascular remodeling. *J Vasc Res*. 2022;59(2):90–113.
120. Elmarasi M, Elmakaty I, Elsayed B, et al. Phenotypic switching of vascular smooth muscle cells in atherosclerosis, hypertension, and aortic dissection. *J Cell Physiol*. 2024;239(4):e31200.
121. Miano JM, Fisher EA, Majesky MW. Fate and state of vascular smooth muscle cells in atherosclerosis. *Circulation*. 2021;143(21):2110–2116.
122. Frisantiene A, Philippova M, Erne P, Resink TJ. Smooth muscle cell-driven vascular diseases and molecular mechanisms of VSMC plasticity. *Cell Signal*. 2018; 52:48–64.
123. Owens GK, Kumar MS, Wamhoff BR. Molecular regulation of vascular smooth muscle cell differentiation in development and disease. *Physiol Rev*. 2004;84(3): 767–801.
124. Qiu J, Zheng Y, Hu J, et al. Biomechanical regulation of vascular smooth muscle cell functions: from in vitro to in vivo understanding. *J R Soc Interface*. 2014;11(90): 20130852.
125. Shinge SAU, Zhang D, Achu Muluh T, Nie Y, Yu F. Mechanosensitive Piezo1 channel evoked-mechanical signals in atherosclerosis. *J Inflamm Res*. 2021;14:3621–3636.
126. Scher C, Pfisterer L, Wagner AH, et al. Arterial wall stress controls NFAT5 activity in vascular smooth muscle cells. *J Am Heart Assoc*. 2014;3(2):e000626.
127. Touyz RM, Alves-Lopes R, Rios FJ, et al. Vascular smooth muscle contraction in hypertension. *Cardiovasc Res*. 2018;114(4):529–539.
128. Jiao X, Yu H, Du Z, et al. Vascular smooth muscle cells specific deletion of angiotensin-like protein 8 prevents angiotensin II-promoted hypertension and cardiovascular hypertrophy. *Cardiovasc Res*. 2023;119(9):1856–1868.
129. Liu G, Hitomi H, Hosomi N, et al. Mechanical stretch augments insulin-induced vascular smooth muscle cell proliferation by insulin-like growth factor-1 receptor. *Exp Cell Res*. 2011;317(17):2420–2428.
130. Brown IAM, Diederich L, Good ME, et al. Vascular smooth muscle remodeling in conductive and resistance arteries in hypertension. *Arteriosclerosis, thrombosis, Vasc Biol*. 2018;38(9):1969–1985.
131. McCormick ML, Gavrila D, Weintraub NL. Role of oxidative stress in the pathogenesis of abdominal aortic aneurysms. *Arteriosclerosis, thrombosis, Vasc Biol*. 2007;27(3):461–469.
132. Qian W, Hadi T, Silvestro M, et al. Microskeletal stiffness promotes aortic aneurysm by sustaining pathological vascular smooth muscle cell mechanosensation via Piezo1. *Nat Commun*. 2022;13(1):512.
133. Pedroza AJ, Shad R, Dalal AR, et al. Acute induced pressure overload rapidly incites thoracic aortic aneurysmal smooth muscle cell phenotype. *Hypertension*. 2022; 79(4):e86–e89.
134. Baker AB, Ettenson DS, Jonas M, Nugent MA, Iozzo RV, Edelman ER. Endothelial cells provide feedback control for vascular remodeling through a mechanosensitive autocrine TGF-beta signaling pathway. *Circ Res*. 2008;103(3):289–297.
135. Belo VA, Guimaraes DA, Castro MM. Matrix metalloproteinase 2 as a potential mediator of vascular smooth muscle cell migration and chronic vascular remodeling in hypertension. *J Vasc Res*. 2015;52(4):221–231.
136. Kim SA, Sung JY, Woo CH, Choi HC. Laminar shear stress suppresses vascular smooth muscle cell proliferation through nitric oxide-AMPK pathway. *Biochem Biophys Res Commun*. 2017;490(4):1369–1374.
137. Wang H, Yan S, Chai H, et al. Shear stress induces endothelial transdifferentiation from mouse smooth muscle cells. *Biochem Biophys Res Commun*. 2006;346(3): 860–865.
138. Duivenvoorden R, Vanbavel E, de Groot E, et al. Endothelial shear stress: a critical determinant of arterial remodeling and arterial stiffness in humans—a carotid 3.0-T MRI study. *Circ Cardiovasc Imaging*. 2010;3(5):578–585.
139. Goldman J, Zhong L, Liu SQ. Negative regulation of vascular smooth muscle cell migration by blood shear stress. *Am J Physiol Heart Circ Physiol*. 2007;292(2): H928–H938.
140. Yao G, Li H, Zuo X, et al. Oscillatory shear stress promotes vein graft intimal hyperplasia via NADPH oxidase-related pathways. *Front Surg*. 2023;10:1073557.
141. Fitzgerald TN, Shepherd BR, Asada H, et al. Laminar shear stress stimulates vascular smooth muscle cell apoptosis via the Akt pathway. *J Cell Physiol*. 2008;216(2): 389–395.
142. McMurray JJ, Pfeffer MA. Heart failure. *Lancet*. 2005;365(9474):1877–1889.
143. Boersma EM, Ter Maaten JM, Damman K, et al. Congestion in heart failure: a contemporary look at physiology, diagnosis and treatment. *Nat Rev Cardiol*. 2020; 17(10):641–655.
144. Wu J, You J, Wang S, Zhang L, Gong H, Zou Y. Insights into the activation and inhibition of angiotensin II type 1 receptor in the mechanically loaded heart. *Circ J*. 2014;78(6):1283–1289.
145. Uriel N, Sayer G, Annamalai S, Kapur NK, Burkhead D. Mechanical unloading in heart failure. *J Am Coll Cardiol*. 2018;72(5):569–580.
146. Barallobre-Barreiro J, Radovits T, Fava M, et al. Extracellular matrix in heart failure: role of ADAMTS5 in proteoglycan remodeling. *Circulation*. 2021;144(25): 2021–2034.
147. Travers JG, Wennersten SA, Pena B, et al. HDAC inhibition reverses preexisting diastolic dysfunction and blocks covert extracellular matrix remodeling. *Circulation*. 2021;143(19):1874–1890.
148. Hutchinson KR, Stewart Jr JA, Lucchesia PA. Extracellular matrix remodeling during the progression of volume overload-induced heart failure. *J Mol Cell Cardiol*. 2010; 48(3):564–569.
149. Korf-Klingebiel M, Reboll MR, Polten F, et al. Myeloid-Derived growth factor protects against pressure overload-induced heart failure by preserving Sarco/ Endoplasmic reticulum Ca(2+)-ATPase expression in cardiomyocytes. *Circulation*. 2021;144(15):1227–1240.

150. Li X, You J, Dai F, et al. TAK1 activation by NLRP3 deficiency confers cardioprotection against pressure overload-induced cardiomyocyte pyroptosis and hypertrophy. *JACC Basic Transl Sci.* 2023;8(12):1555–1573.
151. Bacmeister L, Schwarzl M, Warnke S, et al. Inflammation and fibrosis in murine models of heart failure. *Basic Res Cardiol.* 2019;114(3):19.
152. Wang G, Kong B, Shuai W, Fu H, Jiang X, Huang H. 3,3-Dimethyl-1-butanol attenuates cardiac remodeling in pressure-overload-induced heart failure mice. *J Nutr Biochem.* 2020;78:108341.
153. Shirakabe A, Zhai P, Ikeda Y, et al. Drp1-Dependent mitochondrial autophagy plays a protective role against pressure overload-induced mitochondrial dysfunction and heart failure. *Circulation.* 2016;133(13):1249–1263.
154. Zhang Y, Bauersachs J, Langer HF. Immune mechanisms in heart failure. *Eur J Heart Fail.* 2017;19(11):1379–1389.
155. Li Q, Li C, Elnwasany A, et al. PKM1 exerts critical roles in cardiac remodeling under pressure overload in the heart. *Circulation.* 2021;144(9):712–727.
156. Froese N, Szarozzyk M, Galuppo P, et al. Hypoxia attenuates pressure overload-induced heart failure. *J Am Heart Assoc.* 2024;13(3):e033553.
157. Havlenova T, Skaroupkova P, Miklovic M, et al. Right versus left ventricular remodeling in heart failure due to chronic volume overload. *Sci Rep.* 2021;11(1):17136.
158. Demirkiran A, van der Geest RJ, Hopman L, et al. Association of left ventricular flow energetics with remodeling after myocardial infarction: new hemodynamic insights for left ventricular remodeling. *Int J Cardiol.* 2022;367:105–114.
159. Leychenko A, Konorev E, Jijiwa M, Matter ML. Stretch-induced hypertrophy activates NfκB-mediated VEGF secretion in adult cardiomyocytes. *PLoS one.* 2011;6(12):e29055.
160. da Silva AR, Gunawan F, Boezio GLM, et al. egr3 is a mechanosensitive transcription factor gene required for cardiac valve morphogenesis. *Sci Adv.* 2024;10(20):ead10633.
161. Niu N, Xu S, Xu Y, Little PJ, Jin ZG. Targeting mechanosensitive transcription factors in atherosclerosis. *Trends Pharmacol Sci.* 2019;40(4):253–266.
162. McSweeney SR, Warabi E, Siow RC. Nrf2 as an endothelial mechanosensitive transcription factor: going with the flow. *Hypertension.* 2016;67(1):20–29.
163. Meli VS, Veerasubramanian PK, Downing TL, Wang W, Liu WF. Mechanosensation to inflammation: roles for YAP/TAZ in innate immune cells. *Sci Signal.* 2023;16(783):eadc9656.
164. Zheng Q, Zou Y, Teng P, et al. Mechanosensitive Channel PIEZO1 senses shear force to induce KLF2/4 expression via CaMKII/MEKK3/ERK5 Axis in endothelial cells. *Cells.* 2022;11(14).
165. Paoletti A, Fontana F, Pham VC, Rodel CJ, Abdelilah-Seyfried S. Mechanosensitive Notch-Dll4 and Klf2-Wnt9 signaling pathways intersect in guiding valvulogenesis in zebrafish. *Cell Rep.* 2021;37(1):109782.
166. Berecz T, Yiu A, Vittay O, et al. Transcriptional co-activators YAP1-TAZ of Hippo signalling in doxorubicin-induced cardiomyopathy. *ESC Heart Fail.* 2022;9(1):224–235.
167. Mia MM, Cibi DM, Ghani S, et al. Loss of Yap/Taz in cardiac fibroblasts attenuates adverse remodelling and improves cardiac function. *Cardiovasc Res.* 2022;118(7):1785–1804.
168. Ong YT, Andrade J, Armbruster M, et al. A YAP/TAZ-TEAD signalling module links endothelial nutrient acquisition to angiogenic growth. *Nat Metab.* 2022;4(6):672–682.
169. Li R, Shao J, Jin YJ, et al. Endothelial FAT1 inhibits angiogenesis by controlling YAP/TAZ protein degradation via E3 ligase MIB2. *Nat Commun.* 2023;14(1):1980.
170. Mannion AJ, Zhao H, Zhang Y, et al. Regulation of YAP promoter accessibility in endothelial mechanotransduction. *Arteriosclerosis, thrombosis, Vasc Biol.* 2024;44(3):666–689.
171. Liu J, Zhao C, Xiao X, et al. Endothelial discoidin domain receptor 1 senses flow to modulate YAP activation. *Nat Commun.* 2023;14(1):6457.
172. Dupont S, Morsut L, Aragona M, et al. Role of YAP/TAZ in mechanotransduction. *Nature.* 2011;474(7350):179–183.
173. Wang L, Luo JY, Li B, et al. Integrin-YAP/TAZ-JNK cascade mediates atheroprotective effect of unidirectional shear flow. *Nature.* 2016;540(7634):579–582.
174. Ritsvall O, Albinsson S. Emerging role of YAP/TAZ in vascular mechanotransduction and disease. *Microcirculation.* 2024;31(4):e12838.
175. Wang KC, Yeh YT, Nguyen P, et al. Flow-dependent YAP/TAZ activities regulate endothelial phenotypes and atherosclerosis. *Proc. Natl. Acad. Sci. U.S.A.* 2016;113(41):11525–11530.
176. Zhang Q, Cao Y, Liu Y, et al. Shear stress inhibits cardiac microvascular endothelial cells apoptosis to protect against myocardial ischemia reperfusion injury via YAP/miR-206/PDCD4 signaling pathway. *Biochem Pharmacol.* 2021;186:114466.
177. Pang ZD, Sun X, Bai RY, et al. YAP-galectin-3 signaling mediates endothelial dysfunction in angiotensin II-induced hypertension in mice. *Cell Mol Life Sci.* 2023;80(2):38.
178. Xu Q, Zhuo K, Zhang X, et al. The role of angiotensin II activation of yes-associated protein/PDZ-binding motif signaling in hypertensive cardiac and vascular remodeling. *Eur J Pharmacol.* 2024;962:176252.
179. Daoud F, Arevalo Martinez M, Ritsvall O, Bastrup JA, et al. YAP and TAZ in vascular smooth muscle confer protection against hypertensive Vasculopathy. *Arteriosclerosis, thrombosis, Vasc Biol.* 2022;42(4):428–443.
180. Arevalo Martinez M, Ritsvall O, Bastrup JA, et al. Vascular smooth muscle-specific YAP/TAZ deletion triggers aneurysm development in mouse aorta. *JCI Insight.* 2023;8(17).
181. Liu YN, Lv X, Chen X, et al. Specific overexpression of YAP in vascular smooth muscle attenuated abdominal aortic aneurysm formation by activating elastic fiber assembly via LTBP4. *J Cardiovasc Transl Res.* 2023;16(1):65–76.
182. Zhong HY, Yuan C, Liu XL, et al. Mechanical stretch aggravates vascular smooth muscle cell apoptosis and vascular remodeling by downregulating EHZ2. *Int J Biochem Cell Biol.* 2022;151:106278.
183. Douquet D, Patel A, Xu A, Vanhoutte PM, Honore E. Piezo ion channels in cardiovascular mechanobiology. *Trends Pharmacol Sci.* 2019;40(12):956–970.
184. Lai A, Cox CD, Chandra Sekar N, et al. Mechanosensing by Piezo1 and its implications for physiology and various pathologies. *Biol Rev Camb Phil Soc.* 2022;97(2):604–614.
185. Jin C, Su S, Yu S, et al. Essential roles of PIEZO1 in mammalian cardiovascular system: from development to diseases. *Cells.* 2024;13(17).
186. Yuan W, Zhang X, Fan X. The role of the Piezo1 mechanosensitive channel in heart failure. *Curr Issues Mol Biol.* 2023;45(7):5830–5848.
187. Huang J, Zhang K, Du R, et al. The Janus-faced role of Piezo1 in cardiovascular health under mechanical stimulation. *Genes Dis.* 2023;10(5):1956–1968.
188. Jiang F, Yin K, Wu K, et al. The mechanosensitive Piezo1 channel mediates heart mechano-chemo transduction. *Nat Commun.* 2021;12(1):869.
189. Ploeg MC, Munts C, Prinzen FW, Turner NA, van Bilsen M, van Nieuwenhoven FA. Piezo1 mechanosensitive ion channel mediates stretch-induced Nppb expression in adult rat cardiac fibroblasts. *Cells.* 2021;10(7).
190. Sun M, Mao S, Wu C, et al. Piezo1-Mediated neurogenic inflammatory cascade exacerbates ventricular remodeling after myocardial infarction. *Circulation.* 2024;149(19):1516–1533.
191. Su SA, Zhang Y, Li W, et al. Cardiac Piezo1 Exacerbates Lethal Ventricular Arrhythmogenesis by Linking Mechanical Stress with Ca(2+) Handling after Myocardial Infarction. vol. 6. Research (Wash D C). 2023:165.
192. Wang Z, Chen J, Babicheva A, et al. Endothelial upregulation of mechanosensitive channel Piezo1 in pulmonary hypertension. *Am J Physiol Cell Physiol.* 2021;321(6):C1010–C1027.
193. Chen J, Miao J, Zhou D, et al. Upregulation of mechanosensitive channel Piezo1 involved in high shear stress-induced pulmonary hypertension. *Thromb Res.* 2022;218:52–63.
194. Zhao W, Wei Z, Xin G, et al. Piezo1 initiates platelet hyperreactivity and accelerates thrombosis in hypertension. *J Thromb Haemostasis.* 2021;19(12):3113–3125.
195. Zhang Y, Su SA, Li W, et al. Piezo1-Mediated mechanotransduction promotes cardiac hypertrophy by impairing calcium homeostasis to activate Calpain/Calcineurin signaling. *Hypertension.* 2021;78(3):647–660.
196. Mao J, Yang R, Yuan P, et al. Different stimuli induce endothelial dysfunction and promote atherosclerosis through the Piezo1/YAP signaling axis. *Arch Biochem Biophys.* 2023;747:109755.
197. Lan Y, Lu J, Zhang S, et al. Piezo1-Mediated mechanotransduction contributes to disturbed flow-induced atherosclerotic endothelial inflammation. *J Am Heart Assoc.* 2024;13(21):e035558.
198. Wang YM, Chu TJ, Wan RT, Niu WP, Bian YF, Li J. Quercetin ameliorates atherosclerosis by inhibiting inflammation of vascular endothelial cells via Piezo1 channels. *Phytomedicine.* 2024;132:155865.
199. Pourtemour S, Fan J, Majhi RK, et al. PIEZO1 targeting in macrophages boosts phagocytic activity and foam cell apoptosis in atherosclerosis. *Cell Mol Life Sci.* 2024;81(1):331.
200. Pan X, Xu H, Ding Z, et al. Guizhitongluo Tablet inhibits atherosclerosis and foam cell formation through regulating Piezo1/NLRP3 mediated macrophage pyroptosis. *Phytomedicine.* 2024;132:155827.
201. Zhu Y, Wang T, Yang Y, et al. Low shear stress exacerbates atherosclerosis by inducing the generation of neutrophil extracellular traps via Piezo 1-mediated mechanosensation. *Atherosclerosis.* 2024;391:117473.
202. Johnson RT, Solanki R, Wostear F, et al. Piezo1-mediated regulation of smooth muscle cell volume in response to enhanced extracellular matrix rigidity. *Br J Pharmacol.* 2024;181(11):1576–1595.
203. Lv H, Ai D. Hippo/yes-associated protein signaling functions as a mechanotransducer in regulating vascular homeostasis. *J Mol Cell Cardiol.* 2022;162:158–165.
204. Yamashiro Y, Yanagisawa H. The molecular mechanism of mechanotransduction in vascular homeostasis and disease. *Clin Sci (Lond).* 2020;134(17):2399–2418.
205. Morales CR, Pedrozo Z, Lavandero S, Hill JA. Oxidative stress and autophagy in cardiovascular homeostasis. *Antioxidants Redox Signal.* 2014;20(3):507–518.
206. Baeyens N, Nicoli S, Coon BG, et al. Vascular remodeling is governed by a VEGFR3-dependent fluid shear stress set point. *Elife.* 2015;4.
207. Hill MA, Meininger GA. Arteriolar vascular smooth muscle cells: mechanotransducers in a complex environment. *Int J Biochem Cell Biol.* 2012;44(9):1505–1510.
208. Zhang L, Feng Q, Kong W. ECM microenvironment in vascular homeostasis: new targets for atherosclerosis. *Physiology.* 2024;39(5).
209. Fu Y, Zhou Y, Wang K, Li Z, Kong W. Extracellular matrix interactome in modulating vascular homeostasis and remodeling. *Circ Res.* 2024;134(7):931–949.
210. Pei J, Cai L, Wang F, et al. LPA(2) contributes to vascular endothelium homeostasis and cardiac remodeling after myocardial infarction. *Circ Res.* 2022;131(5):388–403.
211. Kohn JC, Zhou DW, Bordeleau F, et al. Cooperative effects of matrix stiffness and fluid shear stress on endothelial cell behavior. *Biophys J.* 2015;108(3):471–478.
212. Zhao G, Zhao Y, Lu H, et al. BAF60c prevents abdominal aortic aneurysm formation through epigenetic control of vascular smooth muscle cell homeostasis. *J Clin Invest.* 2022;132(21).
213. Du LJ, Sun JY, Zhang WC, et al. NCOR1 maintains the homeostasis of vascular smooth muscle cells and protects against aortic aneurysm. *Cell Death Differ.* 2023;30(3):618–631.
214. Xia Y, Zhang X, An P, Luo J, Luo Y. Mitochondrial homeostasis in VSMCs as a central hub in vascular remodeling. *Int J Mol Sci.* 2023;24(4).

215. Zhang J, Xie SA, Wang J, et al. Echinatin maintains glutathione homeostasis in vascular smooth muscle cells to protect against matrix remodeling and arterial stiffening. *Matrix Biol.* 2023;119:1–18.
216. Kong P, Christia P, Frangogiannis NG. The pathogenesis of cardiac fibrosis. *Cell Mol Life Sci.* 2014;71(4):549–574.
217. Gong L, Wang S, Shen L, et al. SLIT3 deficiency attenuates pressure overload-induced cardiac fibrosis and remodeling. *JCI Insight.* 2020;5(12).
218. Moore-Morris T, Guimaraes-Camboa N, Banerjee I, et al. Resident fibroblast lineages mediate pressure overload-induced cardiac fibrosis. *J Clin Invest.* 2014;124(7):2921–2934.
219. Adapala RK, Katari V, Kanugula AK, Ohanyan V, Paruchuri S, Thodeti CK. Deletion of endothelial TRPV4 protects heart from pressure overload-induced hypertrophy. *Hypertension.* 2023;80(11):2345–2356.
220. Xiang FL, Fang M, Yutzey KE. Loss of beta-catenin in resident cardiac fibroblasts attenuates fibrosis induced by pressure overload in mice. *Nat Commun.* 2017;8(1):712.
221. Li W, Zhang Z, Li X, et al. CGRP derived from cardiac fibroblasts is an endogenous suppressor of cardiac fibrosis. *Cardiovasc Res.* 2020;116(7):1335–1348.
222. Frangogiannis NG. Cardiac fibrosis. *Cardiovasc Res.* 2021;117(6):1450–1488.
223. Nunez-Toldra R, Kirwin T, Ferraro E, et al. Mechanosensitive molecular mechanisms of myocardial fibrosis in living myocardial slices. *ESC Heart Fail.* 2022;9(2):1400–1412.
224. Nagata Y, Bertrand PB, Baliyan V, et al. Abnormal mechanics relate to myocardial fibrosis and ventricular arrhythmias in patients with mitral valve prolapse. *Circ Cardiovasc Imaging.* 2023;16(4):e014963.
225. Hou X, Xiong X, Li X, et al. Predictive value of cardiac magnetic resonance mechanical parameters for myocardial fibrosis in hypertrophic cardiomyopathy with preserved left ventricular ejection fraction. *Front Cardiovasc Med.* 2022;9:1062258.
226. Pesce M, Duda GN, Forte G, et al. Cardiac fibroblasts and mechanosensation in heart development, health and disease. *Nat Rev Cardiol.* 2023;20(5):309–324.
227. Chalise U, Hale TM. Fibroblasts under pressure: cardiac fibroblast responses to hypertension and antihypertensive therapies. *Am J Physiol Heart Circ Physiol.* 2024;37(3):834–847.
228. Zhang L, Guo YN, Liu J, et al. Plantamajoside attenuates cardiac fibrosis via inhibiting AGEs activated-RAGE/autophagy/EndMT pathway. *Phytother Res.* 2023;37(3):834–847.
229. Zhang N, Ma Q, You Y, et al. CXCR4-dependent macrophage-to-fibroblast signaling contributes to cardiac diastolic dysfunction in heart failure with preserved ejection fraction. *Int J Biol Sci.* 2022;18(3):1271–1287.
230. Long Y, Niu Y, Liang K, Du Y. Mechanical communication in fibrosis progression. *Trends Cell Biol.* 2022;32(1):70–90.
231. Nguyen-Truong M, Wang Z. Biomechanical properties and mechanobiology of cardiac ECM. *Adv Exp Med Biol.* 2018;1098:1–19.
232. Niro F, Fernandes S, Cassani M, et al. Fibrotic extracellular matrix impacts cardiomyocyte phenotype and function in an iPSC-derived isogenic model of cardiac fibrosis. *Transl Res.* 2024;273:58–77.
233. Emig R, Knodt W, Krussig MJ, et al. Piezo1 channels contribute to the regulation of human atrial fibroblast mechanical properties and matrix stiffness sensing. *Cells.* 2021;10(3).
234. Garoffolo G, Casaburo M, Amadeo F, et al. Reduction of cardiac fibrosis by Interference with YAP-dependent transactivation. *Circ Res.* 2022;131(3):239–257.
235. Adapala RK, Katari V, Teegala LR, Thodeti S, Paruchuri S, Thodeti CK. TRPV4 mechanotransduction in fibrosis. *Cells.* 2021;10(11).
236. Bartoli F, Evans EL, Blythe NM, et al. Global PIEZO1 Gain-of-function Mutation causes cardiac hypertrophy and fibrosis in mice. *Cells.* 2022;11(7).
237. Zou Y, Pan L, Shen Y, et al. Cardiac Wnt5a and Wnt11 promote fibrosis by the crosstalk of FZD5 and EGFR signaling under pressure overload. *Cell Death Dis.* 2021;12(10):877.
238. Li R, Frangogiannis NG. Integrins in cardiac fibrosis. *J Mol Cell Cardiol.* 2022;172:1–13.
239. Meagher PB, Lee XA, Lee J, Visram A, Friedberg MK, Connelly KA. Cardiac fibrosis: key role of integrins in cardiac homeostasis and remodeling. *Cells.* 2021;10(4).
240. McCarty MF. Nutraceutical, Dietary, and Lifestyle Options for prevention and treatment of ventricular hypertrophy and heart failure. *Int J Mol Sci.* 2021;22(7).
241. Peng F, Liao M, Jin W, et al. 2-APQC, a small-molecule activator of Sirtuin-3 (SIRT3), alleviates myocardial hypertrophy and fibrosis by regulating mitochondrial homeostasis. *Signal Transduct Targeted Ther.* 2024;9(1):133.
242. Tang C, Hou YX, Shi PX, et al. Cardiomyocyte-specific Peli1 contributes to the pressure overload-induced cardiac fibrosis through miR-494-3p-dependent exosomal communication. *FASEB J official Publ Fed Am Soc Exp Biol.* 2023;37(1):e22699.
243. Chen X, Li X, Wu X, et al. Integrin beta-like 1 mediates fibroblast-cardiomyocyte crosstalk to promote cardiac fibrosis and hypertrophy. *Cardiovasc Res.* 2023;119(10):1928–1941.
244. Beech DJ, Kalli AC. Force sensing by piezo channels in cardiovascular health and disease. *Arteriosclerosis, thrombosis, Vasc Biol.* 2019;39(11):2228–2239.
245. Cecchi E, Giglioli C, Valente S, et al. Role of hemodynamic shear stress in cardiovascular disease. *Atherosclerosis.* 2011;214(2):249–256.
246. Hahn C, Schwartz MA. Mechanotransduction in vascular physiology and atherogenesis. *Nat Rev Mol Cell Biol.* 2009;10(1):53–62.
247. Humphrey JD, Dufresne ER, Schwartz MA. Mechanotransduction and extracellular matrix homeostasis. *Nat Rev Mol Cell Biol.* 2014;15(12):802–812.
248. Sun Z, Guo SS, Fässler R. Integrin-mediated mechanotransduction. *J cell Biol.* 2016;215(4):445–456.
249. Lai A, Zhou Y, Thurgood P, et al. Endothelial response to the combined biomechanics of vessel stiffness and shear stress is regulated via Piezo1. *ACS Appl Mater Interfaces.* 2023;15(51):59103–59116.
250. Li J, Zhu J, Gray O, et al. Mechanosensitive super-enhancers regulate genes linked to atherosclerosis in endothelial cells. *J cell Biol.* 2024;223(3).
251. Xu Y, Xu H, Yan J, Song G. Mechanical force induced activation of adhesion G protein-coupled receptor. *Mechanobiology in Medicine.* 2024;2(3):100078.
252. Zhu H, Miao R, Wang J, Lin M. Advances in modeling cellular mechanical perceptions and responses via the membrane-cytoskeleton-nucleus machinery. *Mechanobiology in Medicine.* 2024;2(1):100040.
253. Lee SN, Lin A, Dey D, Berman DS, Han D. Application of quantitative assessment of coronary atherosclerosis by coronary computed tomographic angiography. *Korean J Radiol.* 2024;25(6):518–539.
254. Ma YS, Gao J, Xie YH, Ren GS, Zhang ZK. The important role of wall shear stress in coronary atherosclerosis: finite element study of stenosis coronary artery. *Chin J Arteriosclerosis.* 2018;26(8):831–835 (in Chinese).