



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Exosomes and the kidney: prospects for diagnosis and therapy of renal diseases

Bas W.M. van Balkom¹, Trairak Pisitkun², Marianne C. Verhaar¹ and Mark A. Knepper²

¹Department of Nephrology and Hypertension, University Medical Center Utrecht, Utrecht, The Netherlands and ²Epithelial Systems Biology Laboratory, National Heart, Lung and Blood Institute, Bethesda, Maryland, USA

Exosomes are 40–100 nm membrane vesicles secreted into the extracellular space by numerous cell types. These structures can be isolated from body fluids including urine and plasma. Exosomes contain proteins, mRNAs, miRNAs, and signaling molecules that reflect the physiological state of their cells of origin and consequently provide a rich source of potential biomarker molecules. Aside from diagnostic uses, exosome-mediated transfer of proteins, mRNAs, miRNAs, and signaling molecules offer the promise that they may be used for therapeutic purposes. In this review, we integrate new knowledge about exosomes from outside the field of nephrology with recent progress by renal researchers in order to provide a basis for speculation about how the study of exosomes may affect the fields of nephrology and renal physiology in the next few years.

Kidney International (2011) **80**, 1138–1145; doi:10.1038/ki.2011.292; published online 31 August 2011

KEYWORDS: disease biomarkers; mass spectrometry; microvesicles; miRNA; proteomics

Over the past several years, there has been increasing interest in the nephrology community in a newly recognized biological entity, namely, the exosome. Exosomes are small (40–100 nm) membrane-bound vesicles, secreted upon fusion of the limiting membrane of multivesicular bodies with the plasma membrane.¹ Besides the vesicular exosome, there is an entirely different structure in eukaryotic cells that is called an ‘exosome’, that is, the RNA–exosome, a multi-protein complex that degrades various types of RNA molecules.² Vesicular exosomes are also referred to as microvesicles in some works. As discussed in this article, vesicular exosomes may contain RNA exosomes. For simplicity, we use the term ‘exosome’ to refer to ‘vesicular exosomes’ throughout. They are present not only in urine³ but also in a variety of other body fluids including blood plasma.¹ On the basis of protein mass spectrometry (MS) results, urinary exosomes appear to derive from each of the epithelial cell types facing the renal tubule lumen.³ Similarly, exosomes in plasma most likely derive from the many cell types that face the vascular lumen, including various types of blood cells and endothelial cells. We have previously published reviews on the topic of urinary exosomes, focusing mainly on the isolation of urinary exosomes as the starting material for protein biomarker discovery experiments.^{4–7} Here we take a broader view, with an attempt to integrate progress outside of the field of nephrology with recent progress by renal researchers to provide a basis for speculation about what impact the study of exosomes will have on the fields of nephrology and renal physiology in the next few years.

BIOLOGY OF EXOSOMES

In 1981, Trams *et al.*⁸ proposed the term ‘exosomes’ for exfoliated membrane vesicles, appearing as large (500–1000 nm) and small (approximately 40 nm) vesicles, which they identified to be secreted by a variety of cell types. A few years later, Johnstone *et al.*⁹ discovered that during reticulocyte maturation, specific proteins, including the transferrin receptor, are shed via secretion of <100 nm vesicles that they termed ‘exosomes’. In the current literature, exosomes are defined as 40–100 nm vesicles that are secreted upon fusion of multivesicular bodies (late endosomes) with the plasma membrane. This fusion event results in the release of the intraluminal vesicles of multivesicular bodies, after

Correspondence: Mark A. Knepper, Epithelial Systems Biology Laboratory, National Heart, Lung and Blood Institute, National Institutes of Health, 10 Center Drive MSC-1603, Building 10, Room 6N260, Bethesda, Maryland 20892-1603, USA. E-mail: knep@helix.nih.gov

Received 26 April 2011; revised 9 June 2011; accepted 14 June 2011; published online 31 August 2011

which they are termed ‘exosomes’, into the extracellular space.¹ Exosomes are known to be produced by many different cell types, including dendritic cells, B-lymphocytes, various stem cells, epithelial cells, and endothelial cells,^{3,10–15} and can be isolated from cell culture supernatant, as well as from a variety of biological fluids, such as blood, urine, semen (prostasomes), amniotic fluid, and pleural fluid.^{3,14,16–19} Multivesicular bodies are late endosomes that are populated with intraluminal vesicles by fusion of small cytoplasmic vesicles derived from early endosomes with the outer membranes of multivesicular bodies, followed by invagination of the recruited membrane, inward budding, and scission (Figure 1). These events are mediated through the concerted action of the so-called ESCRT complexes (endosomal complexes required for transport).^{20,21} As vesicles bud inward, the lumina of these future exosomes capture a small portion of the cytosol, taking along a set of soluble proteins, mRNAs, microRNAs (miRNAs), and other cytosolic molecules. The orientation of the lipid membranes of exosomes is identical to that of cells; that is, integral membrane proteins are oriented such that the amino acid sequences facing the outside of the plasma membrane of cells also face to the outside of exosomes.¹ It has been proposed that in addition to random selection of a portion of the cytoplasm, proteins and RNA molecules may be selectively incorporated into exosomes.^{22–24}

Besides exosomes, other types of microvesicles can also be isolated from cell culture supernatants and body fluids (reviewed by Camussi *et al.*²⁵). These microvesicles are not derived from multivesicular bodies, but appear to be shed by the plasma membrane. Usually, these microvesicles tend to be larger in size (up to 1 µm), although smaller microvesicles, which fall in the range of exosomes, have been described.²⁶ In addition, it has been shown that there are microvesicles in urine that are derived from microvilli of podocytes.²⁷ Because of the overlap in size, microvesicles may be included among exosomes when they are isolated from urine.

Proteomic analyses show that many of the proteins detectable in exosomes are common to exosomes from all cell types.^{3,13,28} These include ribosomal components, cytoskeletal proteins, small and heterotrimeric GTPases, tetraspanin proteins, and the components of the ESCRT complexes involved in forming multivesicular bodies. Furthermore, exosomes contain many cell-specific proteins. The incorporation of certain proteins into internal vesicles of multivesicular bodies is not a random selection of proteins expressed in a given cell type. For example, proteomic profiling of proteins in urinary exosomes revealed an abundance of integral membrane proteins targeted to the apical plasma membranes of epithelial cells, but a dearth of proteins associated with the basolateral domain.³ Further evidence for selective protein sorting to exosomes comes from the observations in nonpolarized cells showing that particular proteins are enriched in exosomes compared with the whole cell. Such proteins include the transmembrane proteins CD55, CD59, CD63, CD81, CD82, the transferrin

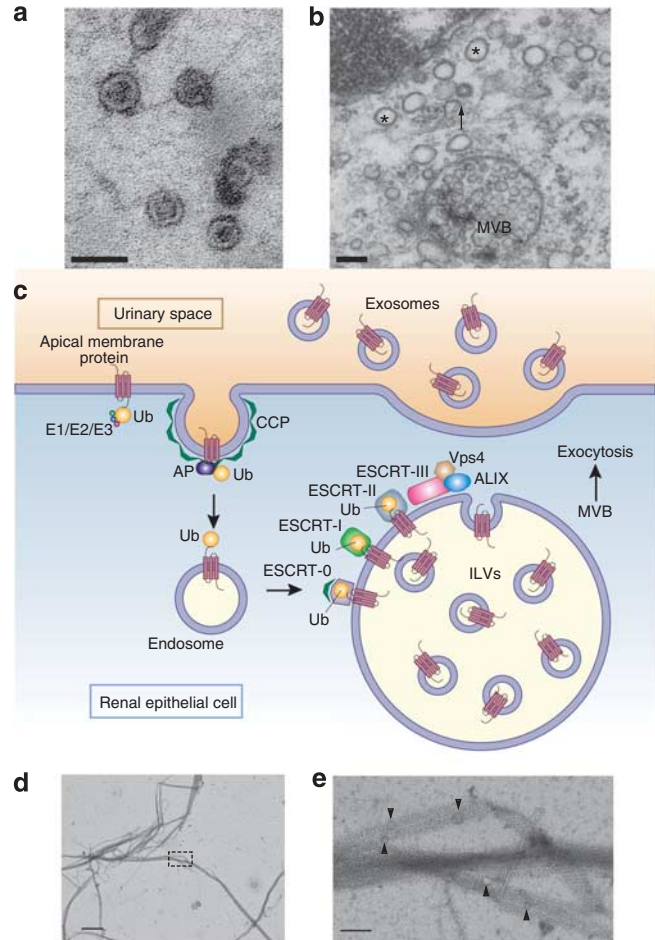


Figure 1 | Exosomes in urine. (a) Electron micrograph of negatively stained urinary exosomes (scale bar, 50 nm). (b) Electron micrograph of a renal inner medullary collecting duct cell (scale bar, 100 nm). Uncoated vesicles (asterisks) and coated vesicles (arrow) are indicated. MVB, multivesicular body. (c) Schematic of urinary exosome formation and release into the urine. AP, adaptor protein; ALIX, ALG-2 interacting protein X; CCP, clathrin-coated pit (clathrin molecules are shown in green); E1, ubiquitin-activating enzyme; E2, ubiquitin-conjugating enzyme; E3, ubiquitin-protein ligase; ESCRT, endosomal sorting complex required for transport; ILVs, intraluminal vesicles; Ub, ubiquitin; Vps4, vacuolar protein sorting 4. (d and e) Electron microscope images of the 17,000 g pellets from pooled normal human urine. Tamm-Horsfall protein (THP) forms long polymeric filaments that are associated laterally to form rope-like structures (d, scale bar, 800 nm and e, depicting the dashed box in d; scale bar, 100 nm). The THP network depicted contains small (40–100 nm) vesicles compatible with exosomes (e, arrowheads).

receptor, and phospholipase D2, as well as many soluble proteins such as certain heatshock proteins.^{14,22,23,29,30} Ubiquitin- and lipid raft-associated protein sorting have been reported to be involved in this selective incorporation of proteins into exosomes.^{28,31}

PHYSIOLOGICAL ROLES OF EXOSOMES

Besides a likely role in elimination of excess or senescent proteins and lipids, there is considerable evidence that

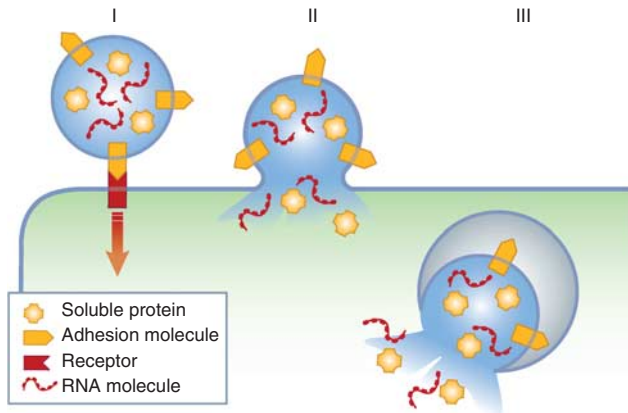


Figure 2 | Mechanisms for exosome-mediated signaling to target cells. Interaction of exosome surface proteins with adhesion molecules or receptors on target cells can initiate a downstream signaling cascade (I). Direct fusion of the exosome membrane with the target cell plasma membrane results in the release of content (RNA, proteins) into the cytoplasm of the target cell (II). Transfer of exosome content to the cytoplasm may also occur after endocytosis and subsequent fusion of the exosome membrane with the endosomal membrane (III).

exosomes can have roles in intercellular signaling in a cell-selective manner.^{32–36} This role has been reviewed recently by Camussi *et al.*²⁵ in this journal. Exosomes may elicit effects on target cells by at least three possible mechanisms (Figure 2): (I) They can adhere with high specificity to the target cell surface (without membrane fusion) through adhesion molecules and receptors present on their surfaces, leading to receptor activation and downstream signaling in the target cell.^{33,37} (II) Exosomes could hypothetically fuse directly with target cells, resulting in transfer of the contents of exosomes (mRNAs, miRNAs, proteins, and signaling molecules). (III) Another possibility that is better supported in the literature is that the contents of exosomes may incorporate into target cells after endocytosis of exosomes and processing in the endosomal pathway.^{34–36,38}

These mechanisms have been demonstrated in exosomes secreted into the blood and extracellular fluid. If urinary exosomes can carry out intercellular signaling in the same way as exosomes secreted into blood and extracellular fluid, they could have important roles in renal physiology. For example, exosomes may mediate downstream information transfer in the process of renal tubule hypertrophy seen with nephron loss. In this process, increases in single-nephron glomerular filtration rate are matched by increases in the transport capacity for salt and water in downstream nephron segments, in association with marked enlargement of renal tubule cells and widening of the tubule lumen.³⁹ It is possible that such downstream information transfer is occurring in normal physiological states. Indeed, proteins that are nominally proximal tubule proteins have been detected in the renal-collecting duct, including the water channel aquaporin-1 (ref. 40) and the ammonium-generating enzyme glutaminase.^{41,42} Such downstream information transfer may

not always be beneficial. We speculate that Tamm–Horsfall protein (uromodulin), an abundant polymeric protein in normal urine, has a role to limit exosomal fusion in downstream nephron segments. Urinary exosomes are typically shrouded by large polymeric fibers formed from Tamm–Horsfall protein, which would prevent them from coming into contact with cell surfaces unless the polymeric network is locally dissolved.⁴³ If these speculations are true, they may provide a basis for understanding how mutations or deficiency of Tamm–Horsfall protein could cause renal disease^{44,45} and could therefore warrant further investigation.

An additional way that urinary exosomes could have roles in kidney physiology is through actions of exosome-resident proteins in the renal tubule lumen. An example of this may be the demonstrated presence of abundant angiotensin-converting enzyme in urinary exosomes,^{3,46} which could have a role in the well-known intraluminal renin–angiotensin system described by Navar *et al.*⁴⁷

It is certainly possible that the main physiological role for urinary exosomes is the disposal of senescent proteins from cells, which may be a more efficient means of protein elimination than proteasomal degradation and lysosomal degradation.

Although the major focus of this article is on urinary exosomes, it is likely that exosomes secreted into the blood and extracellular fluid have roles in renal physiology and pathophysiology, especially among cell types with their plasma membranes in direct contact with the vascular compartment such as cells of the immune system and endothelial cells. Prime examples are the roles of exosomes and microvesicles in cell–cell communication in immune cell and stem cell signaling. For example, exosome-mediated communication is involved in the priming of CD8+ and CD4+ cells by antigen-loaded major histocompatibility complex class I and II molecules on exosomes, showing target cell-specific effects.^{48,49} Receptor–ligand interaction between exosomes and cells are also essential to identify specific target cells, as demonstrated by the specific binding of dendritic cell-derived exosomes to activated, and not resting, T cells. The selection of the target cell is mediated by the interaction between inter-cellular adhesion molecule 1 on exosomes and its ligand, lymphocyte function-associated antigen 1, on activated T cells,³² suggesting a mechanism by which, on a broader scale, exosomes may be targeted to specific cell types. In addition, it has been demonstrated that mesenchymal stem cells contain specific miRNA signatures, which are selectively incorporated and subsequently transferred to target cells.⁵⁰ Transferred miRNAs affect gene expression in target cells⁵¹, demonstrating that besides transfer of proteins, exosomes can modulate the physiology of the target cell by transfer of RNA. Further examples for the role of microvesicle-mediated transfer of RNA include the modulation of stem cells and the stem cell niche, which could be a crucial stem in stem cell-mediated tissue repair (reviewed by Deregibus *et al.*⁵² and Quesenberry *et al.*⁵³), and potentially represent a system that is efficiently hijacked by tumors for

the stimulation of angiogenesis. Blood-borne exosomes may also be involved in angiogenesis, at least in tumors. Specifically, tumors promote their vascularization not only through the secretion of known angiogenic cytokines and growth factors, but also via exosomes.^{38,54,55} On the basis of these observations and others, one could well imagine that blood-borne exosomes could have a role in various glomerulopathies in graft rejection, in hypertension, and in other kidney-related diseases.

EXOSOMES AS A SOURCE OF PROTEIN BIOMARKERS

Urinary proteomics studies have identified potential urinary biomarkers for several pathological entities, for example, acute kidney transplant rejection⁵⁶ and diabetic nephropathy.⁵⁷ Despite these and other successes, the number of kidney-derived proteins and peptides detectable in whole urine (or 'minimally processed' urine) by MS has been limited in part by the presence of filtered plasma proteins and very abundant kidney-derived proteins, especially Tamm–Horsfall protein or uromodulin. Abundant proteins compete with less abundant proteins for identification in the mass spectrometer. Consequently, we may be missing the biomarker candidates that would provide the best sensitivity and specificity for diagnosis of a given disease. One approach to enrichment of kidney-derived proteins has been the isolation of exosomes from urine.³ Normal urine contains exosomes that derive from every epithelial cell type facing the urinary space (Figure 1), offering the potential to monitor physiological and pathophysiological changes throughout the nephron through the expedient of urine collection and analysis.

The advent of detailed protein sequence data from the human genome project and marked technological improvements in MS of proteins and peptides may lead to the discovery of even more protein biomarkers. It has become possible to identify and quantify literally thousands of proteins from a single sample using shotgun proteomics based on MS systems that combine liquid chromatography and tandem mass spectrometry (MS/MS). We have used liquid chromatography–MS/MS-based protein MS to carry out large-scale profiling of proteins present in urinary exosomes from normal humans⁴⁶ and have made the data available on a publicly accessible database (<http://dir.nhlbi.nih.gov/papers/lkem/exosome/>). This database provides a listing of 1160 proteins present in urinary exosomes and contains potential biomarker proteins that can be the basis of hypotheses regarding the mechanism of the disease.

A general analysis of urinary proteins by Adachi *et al.*⁵⁸ also detected large numbers of membrane proteins, presumably because of the presence of exosomes in the samples. About 3% of total urinary protein in samples from normal subjects is derived from exosomes.⁵⁹ Thus, isolating exosomes from urine provides a more than 30-fold enrichment of exosomal proteins, allowing proteins that are minor components of whole urine to be readily detectable immunochemically or by protein MS.

As noted above, exosomes are not uniquely found in urine and in fact have been identified in multiple body fluids including blood plasma, where they derive from reticulocytes, leukocytes, endothelial cells, and presumably other cell types that contact the intravascular space. Their presence in blood therefore offers an advantage for biomarker discovery in plasma, which is analogous to their advantage for biomarker discovery in urine. Specifically, the isolation of exosomes allows marked enrichment of biomarkers that may not be readily detectable in whole plasma or even plasma that has been stripped of its most abundant soluble proteins. Major efforts have been undertaken to define the proteomes of plasma-derived exosomes and exosomes from other body fluids, aiming especially at the discovery of novel biomarkers for prevalent diseases such as cancer and atherosclerosis.^{16,18,60–62} A general database of exosomal proteins called 'ExoCarta' can be found at exocarta.ludwig.edu.au/.⁶³

What disease processes in the kidney would be the best targets for exosome-based biomarker discovery? The answer, we believe, is 'those diseases that require clinical decision making, that is, currently non-optimal or too slow with current diagnostic methodologies'. Therefore, there may be renal diseases that are prevalent and have large negative impacts on length or quality of life, but are not good targets for urinary exosome-based biomarker discovery because the addition of a new biomarker would not significantly influence clinical decision making. On the basis of these considerations, one example of a good target for biomarker discovery in urinary exosomes may be the decision-making process encountered in renal allograft patients who experience an increase in serum creatinine levels. The discrimination between rejection and kidney injury, as well as the discrimination between different mechanisms of rejection, is generally addressed through renal biopsy for which a full battery of analyses generally requires many hours or days. Here, a rapid immunological test could speed the initiation of appropriate therapy. Another prime target is early diagnosis of acute kidney injury in surgical and intensive care settings. In studies reported thus far, several potential markers have been identified including KIM1,⁶⁴ HSP72,⁶⁵ Klotho,⁶⁶ IL-6,⁶⁷ NGAL,⁶⁸ L-FABP,⁶⁹ netrin-1,⁷⁰ or fetuin-A.⁷¹ Among these, only the study identifying fetuin-A as a potential acute kidney injury biomarker was conducted using exosomes as starting materials. Exosome analysis may also be useful for classification of other disease processes involving the renal tubule, such as polycystic kidney disease,⁷² lysosomal storage diseases (for example, Nieman–Pick disease and cystinosis), and transporter mutations (such as Gitelman and Bartter syndromes⁴⁶). Urinary exosome analysis may also be useful in the detection and classification of liver damage, which can secondarily affect the kidney.⁷³ In addition, it has been proposed that analysis of urinary exosomes could be performed in patients with hypertension,⁷⁴ possibly to find biomarkers to predict which drugs will be the most effective in lowering the blood pressure in a given patient (personalized medicine). Multiple transcription factors have been

found in urinary exosomes and their analysis has been proposed as a means of noninvasively detecting and monitoring various glomerular diseases including focal segmental glomerulosclerosis.⁷⁵ Furthermore, it has been proposed that exosome analysis of urine may provide better ways to monitor responses to the treatment of prostatic cancer.⁷⁶ The above list of biomarker targets is not exhaustive, and other prime clinical decision-making processes that are amenable to urinary exosome-based biomarker discovery may be readily apparent to the reader.

Genetic diseases may also be diagnosable through urinary exosome analysis. Looking towards the future, the continual improvement in mass spectrometers is making it more and more feasible to use MS in *de novo* sequencing mode to screen for mutations and polymorphisms that affect the primary sequence of proteins. Thus, although significant strides are currently being made with regard to DNA sequencing using so-called 'deep-sequencing technologies', MS may provide an alternative way to discover sequence variations in proteins that appear in the urinary exosomal proteome.

EXOSOMES AS A SOURCE OF RNA BIOMARKERS

Besides proteins and peptides, exosomes contain mRNA and miRNAs.^{1,28,34,77–79} Such RNAs are potentially useful as disease biomarkers. Although efficient exosome isolation protocols have been introduced for urinary RNA analysis,⁷⁹ most studies of urinary RNAs thus far have bypassed exosome isolation, opting for direct analysis of mRNA levels using RT-PCR in sediments from whole urine, which undoubtedly contains RNA from both exosomes and whole cells. An example is a recent study showing increased glycoprotein B7-1 to nephrin mRNA ratios in urinary sediments from patients with minimal change disease compared with focal segmental glomerulosclerosis.⁸⁰ Another recent example is the finding that urinary granzyme A mRNA levels can potentially distinguish patients with cellular rejection from those with acute kidney injury.⁸¹ Exosome isolation can potentially increase the sensitivity and precision of urinary mRNA analysis.

MicroRNA profiling can also be used to identify potential biomarkers. Initially, exosomes from tumors were investigated for the presence of biomarkers, and in 2008, Skog *et al.*³⁸ discovered that mRNA encoding a specific variant of the VEGF-receptor (VEGFvIII) predicts a better treatment response in the treatment of glioblastoma. Furthermore, miRNA signatures of circulating exosomes may serve as a useful tool for the diagnosis of lung cancer and ovarian cancer,^{82,83} and recently a method for the isolation of mRNA and miRNA for diagnostic purposes from urine exosomes were developed.⁷⁹ Even without enrichment by exosome isolation, the abundances of several miRNAs (miR-200a, miR-200b, and miR-429) were found to be decreased in urinary sediments from patients with immunoglobulin A nephropathy, and the degree of reduction correlated with the severity of the disease.⁸⁴

It may be possible to increase both the sensitivity and the specificity of RNA biomarker approaches through the enrichment of exosomes specific to the given cell type. Flow cytometry approaches for the latter task are under development.^{85,86}

EXOSOMES AS POTENTIAL THERAPEUTIC AGENTS

The finding of mRNAs and miRNAs in exosomes and evidence for a role for exosomes in cell-cell communication (reviewed above) foreshadows an important new direction, that is, the use of exosomes as delivery vehicles for therapeutics. The concept is that RNA-bearing exosomes can potentially deliver their contents to specific target cells in order to transiently correct dysregulated processes.

Already, several researchers have preliminarily explored the possibility of using exosomes as therapeutic delivery vehicles. In 1998, Zitvogel *et al.*⁸⁷ proposed the use of exosomes in the immunotherapy of cancer, showing that exosomes derived from tumor peptide-pulsed dendritic cells injected into tumor-bearing mice resulted in eradication or reduced growth of the tumor. More recently, others have pioneered the application of exosomes in cancer treatment.^{88,89} Two phase I clinical trials studied injection of antigen-loaded exosomes from autologous dendritic cells into patients with melanoma or lung cancer and demonstrated feasibility and safety of exosome-based therapy, although the effects on reduction of disease progression were only minor.^{90,91} Similar approaches have the potential for treatment of renal cancers.⁹²

In 2007, Valadi *et al.*³⁴ demonstrated that exosomes are able to transfer miRNAs from their cell of origin to target cells. Besides miRNAs, pre-miRNA could be identified in mesenchymal stem cell-derived exosomes.⁹³ Functionally, this offers cells the possibility to increase (mRNA) or reduce (miRNA, pre-miRNA) protein expression levels in specific target cells. Transfer of mRNA and miRNA molecules to target cells can influence their function, which may be the mechanism by which endothelial progenitor cell-derived exosomes stimulate angiogenesis in endothelial cells.⁷⁷

Another potential use of exosomes is as vehicles for the delivery of specific antigens. This approach has been applied for vaccination against severe acute respiratory syndrome, using exosomes containing the severe acute respiratory syndrome S protein⁹⁴ and against *Toxoplasma gondii*, using antigen-containing exosomes.⁹⁵ Both vaccines showed positive results, displayed as higher levels of neutralizing antibodies and, in the *T. gondii* study, there was a reduction of disease severity in mice.

Exosomes have been reported to be the active component in the conditioned medium of mesenchymal stem cells that display cardioprotective effects by reducing cardiac infarct size after experimental ischemia-reperfusion.⁹⁶ Cardiomyocyte progenitor cell-derived exosomes may also have this potential.¹² A role for exosomes may be found in the paracrine effects that have been observed in experimental stem cell therapy. For example, in experimental stem cell

therapy of acute kidney injury, mesenchymal stem cells have been shown to improve recovery in part through paracrine factors derived from secreted exosomes.^{97,98} In experimental stem cell therapy of experimental glomerulonephritis in rats (anti-Thy1.1 glomerulonephritis), Kunter *et al.*⁹⁹ found a benefit that they attributed to paracrine factors from the injected mesenchymal stem cells rather than from the cells themselves. Conceivably, exosome secretion is involved in these observed paracrine effects.

For many kidney-related diseases, a prime target for potential exosome-based therapy are endothelial cells, which have essential roles in regulation of blood pressure, local regulation of blood flow, regulation of blood clotting, and clearance of plasma lipids. Failure of these processes is responsible for a large fraction of common chronic diseases that affect the kidney, including atherosclerosis and hypertension. Because the endothelial cells face the blood compartment, they might be considered 'low-hanging fruit' for potential exosome-based therapies, as the problem of targeting is largely obviated.

On the basis of the above observations and additional ongoing research, we conclude that exosomes have considerable promise for treatment of a variety of renal diseases. To succeed, however, there is a need to develop methods for efficient isolation of exosomes of appropriate composition, allowing targeting to specific cell types and allowing transfer of selected biomolecule cargos. This can only be achieved through further basic research based on the following questions: (1) How do multivesicular bodies select biomolecules for inclusion in their intraluminal vesicles? (2) How do exosomes interact with cell surfaces of some cells and not others? (3) What is the mechanism of fusion of the limiting membrane of exosomes with plasma membranes?, and (4) How are exosomes in plasma normally cleared and how can this process be selectively delayed for therapeutic exosomes?

CONCLUSION

Our objective in this short review has been to provide a brief synopsis of knowledge about exosomes with a view toward future exploitation in the diagnosis and therapy of kidney diseases and kidney-related diseases. Nephrologists are in a continual search for new tools to improve their ability to rapidly and accurately diagnose renal disease via noninvasive methodologies. The advent of sensitive and accurate MS and genomics techniques has facilitated this quest, and it is likely that in the near future several exosomal biomarkers will come into play in clinical practice. Current efforts to replace ultracentrifugation with more efficient exosome isolation methods, such as filtration,^{100,101} size-exclusion chromatography,¹⁰² and affinity methods,¹⁰³ can be expected to lead to more practical protocols for the profiling of exosomal proteins and RNAs. The potential use of exosomes as therapeutic vehicles is based on the fact that exosome secretion and reuptake can move molecules (and information) between cells, processes that can be interrupted or modified by design. Possibly, in the coming years, we can

extend this frontier beyond its current main focus in the areas of oncology and immune diseases to achieve new ways to treat kidney diseases and kidney-related diseases.

DISCLOSURE

All the authors declared no competing interests.

ACKNOWLEDGMENTS

MAK and TP are supported by the intramural budget of the National Heart, Lung and Blood Institute (project Z01-HL-001285). BWMvB is supported by the Netherlands Organisation for Scientific Research (NWO; NGL/ZonMw Horizon Grant 93519028); MCV is supported by the Netherlands Organisation for Scientific Research (NWO; VIDI grant 016.096.359).

REFERENCES

1. Stoorvogel W, Kleijmeer MJ, Geuze HJ *et al.* The biogenesis and functions of exosomes. *Traffic* 2002; **3**: 321–330.
2. Chen CY, Gherzi R, Ong SE *et al.* AU binding proteins recruit the exosome to degrade ARE-containing mRNAs. *Cell* 2001; **107**: 451–464.
3. Pisitkun T, Shen RF, Knepper MA. Identification and proteomic profiling of exosomes in human urine. *Proc Natl Acad Sci USA* 2004; **101**: 13368–13373.
4. Pisitkun T, Johnstone R, Knepper MA. Discovery of urinary biomarkers. *Mol Cell Proteomics* 2006; **5**: 1760–1771.
5. Gonzales P, Pisitkun T, Knepper MA. Urinary exosomes: is there a future? *Nephrol Dial Transplant* 2008; **23**: 1799–1801.
6. Knepper MA, Pisitkun T. Exosomes in urine: who would have thought? *Kidney Int* 2007; **72**: 1043–1045.
7. Gonzales PA, Zhou H, Pisitkun T *et al.* Isolation and purification of exosomes in urine. *Methods Mol Biol* 2010; **641**: 89–99.
8. Trams EG, Lauter CJ, Salem Jr N *et al.* Exfoliation of membrane ectoenzymes in the form of micro-vesicles. *Biochim Biophys Acta* 1981; **645**: 63–70.
9. Johnstone RM, Adam M, Hammond JR *et al.* Vesicle formation during reticulocyte maturation. Association of plasma membrane activities with released vesicles (exosomes). *J Biol Chem* 1987; **262**: 9412–9420.
10. van Niel G, Raposo G, Candalh C *et al.* Intestinal epithelial cells secrete exosome-like vesicles. *Gastroenterology* 2001; **121**: 337–349.
11. van Niel G, Mallego J, Bevilacqua C *et al.* Intestinal epithelial exosomes carry MHC class II/peptides able to inform the immune system in mice. *Gut* 2003; **52**: 1690–1697.
12. Vrijse J, Sluijter JP, Schuchardt MW *et al.* Cardiomyocyte progenitor cell-derived exosomes stimulate migration of endothelial cells. *J Cell Mol Med* 2010; **14**: 1064–1070.
13. Wubbolts R, Leckie RS, Veenhuizen PT *et al.* Proteomic and biochemical analyses of human B cell-derived exosomes. Potential implications for their function and multivesicular body formation. *J Biol Chem* 2003; **278**: 10963–10972.
14. Thery C, Regnault A, Garin J *et al.* Molecular characterization of dendritic cell-derived exosomes. Selective accumulation of the heat shock protein hsc73. *J Cell Biol* 1999; **147**: 599–610.
15. Zhan R, Leng X, Liu X *et al.* Heat shock protein 70 is secreted from endothelial cells by a non-classical pathway involving exosomes. *Biochem Biophys Res Commun* 2009; **387**: 229–233.
16. Bard MP, Hegmans JP, Hemmes A *et al.* Proteomic analysis of exosomes isolated from human malignant pleural effusions. *Am J Respir Cell Mol Biol* 2004; **31**: 114–121.
17. Caby MP, Lankar D, Vincendeau-Scherrer C *et al.* Exosomal-like vesicles are present in human blood plasma. *Int Immunol* 2005; **17**: 879–887.
18. Keller S, Rupp C, Stoeck A *et al.* CD24 is a marker of exosomes secreted into urine and amniotic fluid. *Kidney Int* 2007; **72**: 1095–1102.
19. Utleug AG, Yi EC, Xie T *et al.* Proteomic analysis of human prostatesomes. *Prostate* 2003; **56**: 150–161.
20. Babst M, Katzmans DJ, Estepa-Sabal EJ *et al.* Escrt-III: an endosome-associated heterooligomeric protein complex required for mvb sorting. *Dev Cell* 2002; **3**: 271–282.
21. Wollert T, Hurley JH. Molecular mechanism of multivesicular body biogenesis by ESCRT complexes. *Nature* 2010; **464**: 864–869.
22. Escola JM, Kleijmeer MJ, Stoorvogel W *et al.* Selective enrichment of tetraspan proteins on the internal vesicles of multivesicular endosomes and on exosomes secreted by human B-lymphocytes. *J Biol Chem* 1998; **273**: 20121–20127.

23. Laulagnier K, Motta C, Hamdi S *et al.* Mast cell- and dendritic cell-derived exosomes display a specific lipid composition and an unusual membrane organization. *Biochem J* 2004; **380**: 161–171.
24. Ohshima K, Inoue K, Fujiwara A *et al.* Let-7 microRNA family is selectively secreted into the extracellular environment via exosomes in a metastatic gastric cancer cell line. *PLoS One* 2010; **5**: e13247.
25. Camussi G, Deregibus MC, Bruno S *et al.* Exosomes/microvesicles as a mechanism of cell-to-cell communication. *Kidney Int* 2010; **78**: 838–848.
26. McConnell RE, Higginbotham JN, Shifrin Jr DA *et al.* The enterocyte microvillus is a vesicle-generating organelle. *J Cell Biol* 2009; **185**: 1285–1298.
27. Hara M, Yanagihara T, Hirayama Y *et al.* Podocyte membrane vesicles in urine originate from tip vesiculation of podocyte microvilli. *Hum Pathol* 2010; **41**: 1265–1275.
28. Buschow SI, van Balkom BW, Aalberts M *et al.* MHC class II-associated proteins in B-cell exosomes and potential functional implications for exosome biogenesis. *Immunol Cell Biol* 2010; **88**: 851–856.
29. Rabesandratana H, Toutant JP, Reggio H *et al.* Decay-accelerating factor (CD55) and membrane inhibitor of reactive lysis (CD59) are released within exosomes during *In vitro* maturation of reticulocytes. *Blood* 1998; **91**: 2573–2580.
30. Johnstone RM, Bianchini A, Teng K. Reticulocyte maturation and exosome release: transferrin receptor containing exosomes shows multiple plasma membrane functions. *Blood* 1989; **74**: 1844–1851.
31. de Gassart A, Geminard C, Fevrier B *et al.* Lipid raft-associated protein sorting in exosomes. *Blood* 2003; **102**: 4336–4344.
32. Nolte-t Hoen EN, Buschow SI, Anderton SM *et al.* Activated T cells recruit exosomes secreted by dendritic cells via LFA-1. *Blood* 2009; **113**: 1977–1981.
33. Denzer K, Kleijmeijer MJ, Heijnen HF *et al.* Exosome: from internal vesicle of the multivesicular body to intercellular signaling device. *J Cell Sci* 2000; **113**(Pt 19): 3365–3374.
34. Valadi H, Ekstrom K, Bossios A *et al.* Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat Cell Biol* 2007; **9**: 654–659.
35. Fevrier B, Raposo G. Exosomes: endosomal-derived vesicles shipping extracellular messages. *Curr Opin Cell Biol* 2004; **16**: 415–421.
36. Smalheiser NR. Exosomal transfer of proteins and RNAs at synapses in the nervous system. *Biol Direct* 2007; **2**: 35.
37. Clayton A, Turkes A, Dewitt S *et al.* Adhesion and signaling by B cell-derived exosomes: the role of integrins. *FASEB J* 2004; **18**: 977–979.
38. Skog J, Wurdinger T, van RS *et al.* Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. *Nat Cell Biol* 2008; **10**: 1470–1476.
39. Fine LG, Schlondorff D, Trizna W *et al.* Functional profile of the isolated uremic nephron. *J Clin Invest* 1978; **78**: 1519–1527.
40. Sabolic I, Valenti G, Verbatatz JM *et al.* Localization of the CHIP28 water channel in rat kidney. *Am J Physiol* 1992; **263**: C1225–C1233.
41. Wright PA, Burg MB, Knepper MA. Microdissection of kidney tubule segments. In: Fleischer S (ed). *Methods in Enzymology*, 191 edn. Academic Press: San Diego, 1990, pp 191–231.
42. Wright PA, Packer RK, Garcia-Perez A *et al.* Time course of renal glutamate dehydrogenase induction during NH₄Cl loading in rats. *Am J Physiol* 1992; **262**: F999–F1006.
43. Fernandez-Llama P, Khositseth S, Gonzales PA *et al.* Tamm-Horsfall protein and urinary exosome isolation. *Kidney Int* 2010; **77**: 736–742.
44. Wolf MT, Beck BB, Zaucke F *et al.* The uromodulin C744G mutation causes MCKD2 and FJHN in children and adults and may be due to a possible founder effect. *Kidney Int* 2007; **71**: 574–581.
45. Bachmann S, Mutig K, Bates J *et al.* Renal effects of Tamm-Horsfall protein (uromodulin) deficiency in mice. *Am J Physiol Renal Physiol* 2005; **288**: F559–F567.
46. Gonzales PA, Pisitkun T, Hoffert JD *et al.* Large-scale proteomics and phosphoproteomics of urinary exosomes. *J Am Soc Nephrol* 2009; **20**: 363–379.
47. Navar LG, Harrison-Bernard LM, Nishiyama A *et al.* Regulation of intrarenal angiotensin II in hypertension. *Hypertension* 2002; **39**: 316–322.
48. Thery C, Zitvogel L, Amigorena S. Exosomes: composition, biogenesis and function. *Nat Rev Immunol* 2002; **2**: 569–579.
49. Utsugi-Kobukai S, Fujimaki H, Hotta C *et al.* MHC class I-mediated exogenous antigen presentation by exosomes secreted from immature and mature bone marrow derived dendritic cells. *Immunol Lett* 2003; **89**: 125–131.
50. Collino F, Deregibus MC, Bruno S *et al.* Microvesicles derived from adult human bone marrow and tissue specific mesenchymal stem cells shuttle selected pattern of miRNAs. *PLoS One* 2010; **5**: e11803.
51. Pegtel DM, Cosmopoulos K, Thorley-Lawson DA *et al.* Functional delivery of viral miRNAs via exosomes. *Proc Natl Acad Sci USA* 2010; **107**: 6328–6333.
52. Deregibus MC, Tetta C, Camussi G. The dynamic stem cell microenvironment is orchestrated by microvesicle-mediated transfer of genetic information. *Histol Histopathol* 2010; **25**: 397–404.
53. Quesenberry PJ, Aliotta JM. Cellular phenotype switching and microvesicles. *Adv Drug Deliv Rev* 2010; **62**: 1141–1148.
54. Iero M, Valenti R, Huber V *et al.* Tumour-released exosomes and their implications in cancer immunity. *Cell Death Differ* 2008; **15**: 80–88.
55. Janowska-Wieczorek A, Wysoczynski M, Kijowski J *et al.* Microvesicles derived from activated platelets induce metastasis and angiogenesis in lung cancer. *Int J Cancer* 2005; **113**: 752–760.
56. Ling XB, Sigdel TK, Lau K *et al.* Integrative urinary peptidomics in renal transplantation identifies biomarkers for acute rejection. *J Am Soc Nephrol* 2010; **21**: 646–653.
57. Jiang H, Guan G, Zhang R *et al.* Identification of urinary soluble E-cadherin as a novel biomarker for diabetic nephropathy. *Diabetes Metab Res Rev* 2009; **25**: 232–241.
58. Adachi J, Kumar C, Zhang Y *et al.* The human urinary proteome contains more than 1500 proteins, including a large proportion of membrane proteins. *Genome Biol* 2006; **7**: R80.
59. Zhou H, Yuen PS, Pisitkun T *et al.* Collection, storage, preservation, and normalization of human urinary exosomes for biomarker discovery. *Kidney Int* 2006; **69**: 1471–1476.
60. Simpson RJ, Jensen SS, Lim JW. Proteomic profiling of exosomes: current perspectives. *Proteomics* 2008; **8**: 4083–4099.
61. varez-Llamas G, De la CF, Barderas ME *et al.* Recent advances in atherosclerosis-based proteomics: new biomarkers and a future perspective. *Expert Rev Proteomics* 2008; **5**: 679–691.
62. Mears R, Craven RA, Hanrahan S *et al.* Proteomic analysis of melanoma-derived exosomes by two-dimensional polyacrylamide gel electrophoresis and mass spectrometry. *Proteomics* 2004; **4**: 4019–4031.
63. Mathivanan S, Simpson RJ. ExoCarta: a compendium of exosomal proteins and RNA. *Proteomics* 2009; **9**: 4997–5000.
64. Han WK, Bailly V, Abichandani R *et al.* Kidney Injury Molecule-1 (KIM-1): a novel biomarker for human renal proximal tubule injury. *Kidney Int* 2002; **62**: 237–244.
65. Barrera-Chimal J, Perez-Villalva R, Cortes-Gonzalez C *et al.* Hsp72 is an early and sensitive biomarker to detect acute kidney injury. *EMBO Mol Med* 2011; **3**: 5–20.
66. Aiello S, Noris M. Klotho in acute kidney injury: biomarker, therapy, or a bit of both? *Kidney Int* 2010; **78**: 1208–1210.
67. Dennen P, Altmann C, Kaufman J *et al.* Urine interleukin-6 is an early biomarker of acute kidney injury in children undergoing cardiac surgery. *Crit Care* 2010; **14**: R181.
68. Bolignano D, Donato V, Coppolino G *et al.* Neutrophil gelatinase-associated lipocalin (NGAL) as a marker of kidney damage. *Am J Kidney Dis* 2008; **52**: 595–605.
69. Matsui K, Kamijo-Ikemori A, Hara M *et al.* Clinical significance of tubular and podocyte biomarkers in acute kidney injury. *Clin Exp Nephrol* 2010; **15**: 220–225.
70. Ramesh G, Krawczeski CD, Woo JG *et al.* Urinary netrin-1 is an early predictive biomarker of acute kidney injury after cardiac surgery. *Clin J Am Soc Nephrol* 2010; **5**: 395–401.
71. Zhou H, Pisitkun T, Aponte A *et al.* Exosomal Fetuin-A identified by proteomics: a novel urinary biomarker for detecting acute kidney injury. *Kidney Int* 2006; **70**: 1847–1857.
72. Hogan MC, Manganeli L, Woollard JR *et al.* Characterization of PKD protein-positive exosome-like vesicles. *J Am Soc Nephrol* 2009; **20**: 278–288.
73. Conde-Vancells J, Rodriguez-Suarez E, Gonzalez E *et al.* Candidate biomarkers in exosome-like vesicles purified from rat and mouse urine samples. *Proteomics Clin Appl* 2010; **4**: 416–425.
74. Esteva-Font C, Wang X, Ars E *et al.* Are sodium transporters in urinary exosomes reliable markers of tubular sodium reabsorption in hypertensive patients? *Nephron Physiol* 2010; **114**: 25–34.
75. Zhou H, Cheruvanky A, Hu X *et al.* Urinary exosomal transcription factors, a new class of biomarkers for renal disease. *Kidney Int* 2008; **74**: 613–621.
76. Mitchell PJ, Welton J, Staffurth J *et al.* Can urinary exosomes act as treatment response markers in prostate cancer? *J Transl Med* 2009; **7**: 4.

77. Deregibus MC, Cantaluppi V, Calogero R *et al.* Endothelial progenitor cell derived microvesicles activate an angiogenic program in endothelial cells by a horizontal transfer of mRNA. *Blood* 2007; **110**: 2440–2448.
78. Michael A, Bajracharya SD, Yuen PS *et al.* Exosomes from human saliva as a source of microRNA biomarkers. *Oral Dis* 2010; **16**: 34–38.
79. Miranda KC, Bond DT, McKee M *et al.* Nucleic acids within urinary exosomes/microvesicles are potential biomarkers for renal disease. *Kidney Int* 2010; **78**: 191–199.
80. Navarro-Munoz M, Ibernorn M, Perez V *et al.* Messenger RNA expression of B7-1 and NPHS1 in urinary sediment could be useful to differentiate between minimal change disease and focal segmental glomerulosclerosis in adult patients. *Nephrol Dial Transplant* 2011; doi:10.1093/ndt/gfr128.
81. van Ham SM, Heutink KM, Jorritsma T *et al.* Urinary granzyme A mRNA is a biomarker to diagnose subclinical and acute cellular rejection in kidney transplant recipients. *Kidney Int* 2010; **78**: 1033–1040.
82. Taylor DD, Gercel-Taylor C. MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. *Gynecol Oncol* 2008; **110**: 13–21.
83. Rabinowits G, Gercel-Taylor C, Day JM *et al.* Exosomal microRNA: a diagnostic marker for lung cancer. *Clin Lung Cancer* 2009; **10**: 42–46.
84. Wang G, Kwan BC, Lai FM *et al.* Expression of microRNAs in the urinary sediment of patients with IgA nephropathy. *Dis Markers* 2010; **28**: 79–86.
85. Gelderman MP, Simak J. Flow cytometric analysis of cell membrane microparticles. *Methods Mol Biol* 2008; **484**: 79–93.
86. Orozco AF, Lewis DE. Flow cytometric analysis of circulating microparticles in plasma. *Cytometry A* 2010; **77**: 502–514.
87. Zitvogel L, Regnault A, Lozier A *et al.* Eradication of established murine tumors using a novel cell-free vaccine: dendritic cell-derived exosomes. *Nat Med* 1998; **4**: 594–600.
88. Chaput N, Taieb J, Andre F *et al.* The potential of exosomes in immunotherapy. *Expert Opin Biol Ther* 2005; **5**: 737–747.
89. Viaud S, Thery C, Ploix S *et al.* Dendritic cell-derived exosomes for cancer immunotherapy: what's next? *Cancer Res* 2010; **70**: 1281–1285.
90. Escudier B, Dorval T, Chaput N *et al.* Vaccination of metastatic melanoma patients with autologous dendritic cell (DC) derived-exosomes: results of the first phase I clinical trial. *J Transl Med* 2005; **3**: 10.
91. Morse MA, Garst J, Osada T *et al.* A phase I study of dexosome immunotherapy in patients with advanced non-small cell lung cancer. *J Transl Med* 2005; **3**: 9.
92. Zhang Y, Luo CL, He BC *et al.* Exosomes derived from IL-12-anchored renal cancer cells increase induction of specific antitumor response *in vitro*: a novel vaccine for renal cell carcinoma. *Int J Oncol* 2010; **36**: 133–140.
93. Chen TS, Lai RC, Lee MM *et al.* Mesenchymal stem cell secretes microparticles enriched in pre-microRNAs. *Nucleic Acids Res* 2010; **38**: 215–224.
94. Kuate S, Cinatl J, Doerr HW *et al.* Exosomal vaccines containing the S protein of the SARS coronavirus induce high levels of neutralizing antibodies. *Virology* 2007; **362**: 26–37.
95. Beauvillain C, Juste MO, Dion S *et al.* Exosomes are an effective vaccine against congenital toxoplasmosis in mice. *Vaccine* 2009; **27**: 1750–1757.
96. Lai RC, Arslan F, Lee MM *et al.* Exosome secreted by MSC reduces myocardial ischemia/reperfusion injury. *Stem Cell Res* 2010; **4**: 214–222.
97. Gatti S, Bruno S, Deregibus MC *et al.* Microvesicles derived from human adult mesenchymal stem cells protect against ischaemia-reperfusion-induced acute and chronic kidney injury. *Nephrol Dial Transplant* 2011; **26**: 1474–1483.
98. Bruno S, Grange C, Deregibus MC *et al.* Mesenchymal stem cell-derived microvesicles protect against acute tubular injury. *J Am Soc Nephrol* 2009; **20**: 1053–1067.
99. Kunter U, Rong S, Djuric Z *et al.* Transplanted mesenchymal stem cells accelerate glomerular healing in experimental glomerulonephritis. *J Am Soc Nephrol* 2006; **17**: 2202–2212.
100. Merchant ML, Powell DW, Wilkey DW *et al.* Microfiltration isolation of human urinary exosomes for characterization by MS. *Proteomics Clin Appl* 2010; **4**: 84–96.
101. Cheruvanky A, Zhou H, Pisitkun T *et al.* Rapid isolation of urinary exosomal biomarkers using a nanomembrane ultrafiltration concentrator. *Am J Physiol Renal Physiol* 2007; **292**: F1657–F1661.
102. Rood IM, Deegens JK, Merchant ML *et al.* Comparison of three methods for isolation of urinary microvesicles to identify biomarkers of nephrotic syndrome. *Kidney Int* 2010; **78**: 810–816.
103. Chen C, Skog J, Hsu CH *et al.* Microfluidic isolation and transcriptome analysis of serum microvesicles. *Lab Chip* 2010; **10**: 505–511.