#### Check for updates

### **OPEN ACCESS**

EDITED BY Hetian Lei, Shenzhen Eye Hospital, China

REVIEWED BY Katrien Remaut, Ghent University, Belgium

\*CORRESPONDENCE Xinyi Su xysu@imcb.a-star.edu.sg

SPECIALTY SECTION This article was submitted to

Ophthalmology, a section of the journal Frontiers in Medicine

RECEIVED 21 May 2022 ACCEPTED 28 July 2022 PUBLISHED 18 August 2022

#### CITATION

Seah I, Ong C, Liu Z and Su X (2022) Polymeric biomaterials in the treatment of posterior segment diseases. *Front. Med.* 9:949543. doi: 10.3389/fmed.2022.949543

#### COPYRIGHT

© 2022 Seah, Ong, Liu and Su. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Polymeric biomaterials in the treatment of posterior segment diseases

### Ivan Seah<sup>1,2,3</sup>, Charles Ong<sup>4</sup>, Zengping Liu<sup>2,3,5</sup> and Xinyi Su<sup>1,2,3,5</sup>\*

<sup>1</sup>Department of Ophthalmology, National University Hospital, Singapore, Singapore, <sup>2</sup>Department of Ophthalmology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore, Singapore, <sup>3</sup>Institute of Molecular and Cell Biology (IMCB), Agency for Science, Technology and Research (A\*STAR), Singapore, Singapore, <sup>4</sup>Singapore National Eye Centre (SNEC), Singapore, Singapore, <sup>5</sup>Singapore Eye Research Institute (SERI), Singapore, Singapore

Polymeric biomaterials are biological or synthetic substances which can be engineered to interact with biological systems for the diagnosis or treatment of diseases. These biomaterials have immense potential for treating eyes diseases, particularly the retina—a site of many inherited and acquired diseases. Polymeric biomaterials can be engineered to function both as an endotamponade agent and to prevent intraocular scarring in retinal detachment repair surgeries. They can also be designed as a drug delivery platform for treatment of retinal diseases. Finally, they can be used as scaffolds for cellular products and provide non-viral gene delivery solutions to the retina. This perspective article explains the role of polymeric biomaterials in the treatment of retinal conditions by highlighting recent advances being translated to clinical practice. The article will also identify potential hurdles to clinical translation as future research directions in the field.

### KEYWORDS

retinal detachments, drug delivery system, macular degenerations, polymer, stem cell, retinal disease

### History of biomaterials in ophthalmology

Biomaterials are engineered substances which can interact with biological systems to diagnose or treat diseases. Early biomaterials were designed to perform mainly mechanical functions as implants. These implants can be found in all specialties of medicine. For example, metal and alloys like titanium are commonly used to produce heart valves, vessel stents and joint implants due to their high mechanical strength, immunity to corrosion and complete inertness to the body environment (1). Meanwhile bio-ceramics such as hydroxyapatite are used as bone and dental implants due to their excellent biocompatibility and wear resistance (2).

Unlike other body systems, the eye confers several key advantages for biomaterial applications. Firstly, the immune privilege status of the eye allows foreign biomaterials to be introduced with limited immunogenicity (3). Secondly, the existence of the blood-retinal-barrier limits the systemic penetration of most biomaterials. Furthermore, the small volume of biomaterial administered in the eye is unlikely to cause severe systemic effects. In addition, the implanted materials can be non-invasively monitored

by state-of-art multi-modal ophthalmic investigations. Thus, the eye is an extremely attractive organ for application of novel biomaterials.

The earliest applications of biomaterials in the eye were intraocular implants for cataract replacement (4). Cataract is the most common reversible cause of visual loss affecting approximately 16 million people globally (5). Cataract surgery involves removing the opacified lens and implanting an artificial intraocular lens (IOL). The earliest lenses developed were made of polymethylmethacrylate due to its inert nature and clarity (6). However, with the advent of smaller surgical incisions and phacoemulsification cataract surgery, silicone and acrylic IOLs are widely used today as they have mechanical flexibility, allowing them to be folded for insertion through small incisions (7).

Glaucoma drainage device, like the Ahmed tube, are examples of more recent implants used for the treatment of advanced glaucoma. To achieve optimal intraocular pressure, the tube-shaped glaucoma drainage device, acts as a conduit to shunt aqueous humor away from the anterior chamber of the eye (8). These devices are usually made of silicone and polypropylene due to their flexibility and inertness. Separately, silicone is also used to manufacture scleral buckle implants, used in the surgical treatment of retinal detachments. In orbital reconstructive surgery for fractures, hydroxyapatite and porous polyethylene are used as orbital implants post enucleation (9).

# Polymeric biomaterials and posterior segment applications

Advances in material science have enabled the development of next generation biomaterials through modifications at the molecular level. Polymeric biomaterials are one such example. They are made of repeated subunits of smaller molecules, and can be naturally-derived or synthetic. Naturally-derived polymers can be made of polysaccharides, polypeptides or polynucleotides. These can be further modified chemically to confer useful properties (10). Synthetic polymers are manmade and generated either through step-growth polymerization or chain-polymerization. Step-growth polymerization involves the addition of a single monomer per reaction while chainpolymerization allows the addition of another polymer chain to the original polymer. These processes have allowed the creation of novel polymeric biomaterials with unique properties for treating eye diseases. A key example is sodium hyaluronate, which has been widely adopted as viscoelastic agents for use during cataract surgery (11).

The posterior segment of the eye consists of key structures like the vitreous humor, retina, choroid and optic nerve. These structures are affected in diseases such as retinal detachment, proliferative vitreoretinopathy, retinal neovascular diseases and inherited retinal diseases. For the posterior segment of the eye, various polymeric biomaterials are currently being developed as vitreous substitutes, sustained and topical drug delivery systems, scaffolds for cellular therapeutics and non-viral gene delivery agents (Table 1). These polymers will be discussed in greater detail in the next segment.

### Polymeric biomaterials in the treatment of retinal detachments and proliferative vitreoretinopathy

Retinal detachment is characterized by the separation of the neuroretina from the underlying retinal pigment epithelium. In vitreo-retinal surgery, after repairing the detachment and closing the retinal break, the vitreous cavity is replaced with an endotamponade agent to keep the retina attached during post-operative recovery. Traditional endotamponade agents include intraocular gas agents, like sulfur hexafluoride (SF<sub>6</sub>) and perfluoropropane (C3F8), or non-biodegradable fluids such as silicone oil. Intraocular gases can result in corneal decompensation and cataract formation. Patients also have to adopt a prolonged face-down position. Most importantly, these patients cannot partake in air travel as low atmospheric pressure can result in gaseous expansion. Meanwhile, prolonged use of silicone oil can cause corneal decompensation, if migrated into the anterior chamber, and even silicone oil glaucoma (19). Thus, the use of silicone oil is usually accompanied by a second removal surgery, inadvertently predisposing a patient to further surgical risks.

Despite the shortfalls of both modalities, innovation in this field has largely been stagnant since the 1970s. Apart from biocompatibility, an endotamponade agent should ideally be: (1) optically clear to allow visual inspection of the retina during follow-ups, (2) easily administered during surgery, (3) exert sufficient surface tension across the retina to allow adequate tamponade of the repaired retina, and (4) biodegradable to avoid a removal surgery. Polymeric hydrogels have the potential to fulfill these criteria through careful polymer selection and various chemical modifications (20-25). A hydrogel is comprised of a three-dimensional network of hydrophilic polymers. Due to the cross-linkages between the individual polymer chains, the polymers can retain water molecules to maintain a gel-like property. In recent years, "smart" hydrogels which are capable of changing states from solution to gel in response to physical and chemical stimuli have also been developed (26). In particular, thermogels can change states based on the surrounding temperature (27, 28). Our group previously developed a urethane-based thermogel of a polyethylene glycol (PEG), poly(propylene glycol) (PPG) and poly(e-caprolactone) (PCL) polymer, termed EPC. The thermogel exists in liquid form at lower temperatures, and turns into gel status at body temperature. This unique feature

TABLE 1 Examples of biomaterials and applications in the posterior segment of the eye.

| Polymeric biomaterial  | Therapeutic area  | Application                        | Properties of biomaterial suitable for application  |
|--|---|------------------------------------|---|
| EPC Thermogel - Polyethylene glycol,<br>poly(propylene glycol), poly(e-caprolactone)<br>(12, 13) | Retinal Detachment  | • Retinal tamponade agent          | <ul> <li>Optical clarity</li> <li>Low swelling counter force</li> <li>Biodegradable</li> <li>Biocompatible</li> <li>Able to maintain prolonged<br/>endotamponade effect</li> <li>Able to suppress development of<br/>proliferative vitreoretinopathy</li> <li>Able to regenerate vitreous-like<br/>material over time</li> </ul>                |
| EPC Nanomicelles - Polyethylene glycol,<br>poly(propylene glycol), poly(e-caprolactone) (14)     | <ul> <li>Neovascular Retinal Diseases</li> <li>Age-related macular degeneration</li> <li>Diabetic retinopathy and<br/>macular oedema</li> </ul> | Drug delivery agent                | <ul> <li>Enables topical delivery of drugs<br/>to the posterior segment of the<br/>eye</li> <li>Enables delivery of biologics<br/>such as FDA-approved anti-<br/>vascular endothelial growth<br/>factor compounds</li> <li>Retains bioactivity of delivered<br/>drug to reduce area of<br/>neovascularisation</li> <li>Biocompatible</li> </ul> |
| Polyethylenimine (PEI)-based polymers (15, 16)   | Inherited retinal diseases  | Gene delivery agent                | <ul> <li>Able to form DNA-polymer complexes</li> <li>Enables delivery of genetic cargo into the cell of interest</li> <li>Able to undergo endosomal release to release genetic cargo intracellularly</li> <li>Biocompatible</li> </ul>  |
| Polyethylene terephthalate (PET) (17)<br>Poly(lactic-co-glycolic acid) (PLGA) (18)               | Advanced retinal degeneration <ul> <li>Inherited retinal diseases</li> <li>Age-related macular degeneration</li> </ul>                          | Scaffold for cellular therapeutics | <ul> <li>Biocompatible</li> <li>Biodegradable</li> <li>Able to promote and maintain<br/>a long-term retinal cell<br/>(RPE/photoreceptor) phenotype</li> <li>Have durability to tolerate<br/>implantation procedure</li> <li>Able to mimic physiological<br/>properties of retinal tissue</li> </ul>   |

allows easy administration by injecting *via* 25-gauge syringe into the vitreous cavity and enables the tamponade effect once heated to body temperature. While other polymeric hydrogel formulations have been researched as endotamponade agents (29), EPC's has distinct advantages of biodegradability and low swelling counter force. This allows patients to avoid a second removal surgery, with low risk of raised intraocular pressure. Interestingly, when we implanted EPC in vitrectomized rabbit eyes, we observed a vitreous-like substance re-formed after EPC biodegradation. Further characterization of the vitreous-like substance is currently underway to determine the possibility of vitreous regeneration associated with the polymer (12, 30).

5–10% of all retinal detachment surgeries fail due to proliferative vitreoretinopathy (PVR), a major complication characterized by the development of contractile cellular membranes, leading to tractional retinal detachments (31). Current treatment relies primarily on surgical removal of these membranes. We recently demonstrated that EPC hydrogel is able to function as a bio-functional polymer to prevent retinal scarring in an experimental rabbit model of proliferative vitreo-retinopathy *via* the nuclear factor erythroid 2-related factor (NRF2) signaling pathway. This is a first report, whereby a synthetic polymeric material alone can target intracellular signaling pathways to prevent retinal scarring. More importantly, it offers insight into how synthetic polymeric materials no longer function merely as inert drug carriers and challenges the conventional belief that a small molecule (drug) is always required to achieve a therapeutic effect at a cellular level. It lays the foundation for next generation nanomedicine, whereby polymers alone can be used to elicit specific biological responses (13).

# Polymeric biomaterials in the treatment of retinal neovascular diseases

Retinal diseases such as age-related macular degeneration (AMD), diabetic retinopathy (DR) and diabetic macular oedema (DMO) affect a significant amount of people globally. For instance, AMD is expected to affect approximately 288 million people by 2040 (32). Significant visual loss may occur in advanced disease. Due to the significant disease burden of retinal diseases, and dramatic reduction in quality of life due to vision loss, significant effort is being channeled into developing effective therapeutics. A notable breakthrough in the field was the development of intravitreal anti-vascular endothelial growth factor (anti-VEGF) treatment for neovascular retinal diseases in the early 2000s (33). Anti-VEGF therapies include bevacizumab (Avastin<sup>®</sup>) and FDA-approved, ranibizumab (Lucentis<sup>(R)</sup>) and aflibercept (Eylea<sup>(R)</sup>). These drugs have to be given by intravitreal injections at regular frequencies of between 1 to 3 months (34), which many patients find troublesome, resulting in reduced treatment compliance. Furthermore, intravitreal injections are associated with sight-threatening risks such as increased ocular pressure, retinal detachment, vitreous hemorrhage and endophthalmitis (35). The holy grail of drug delivery is to develop novel solutions for sustained delivery, thereby reducing the frequency of injections. One such example, is a poly lactide-co-glycolide (PLGA) implant that delivers dexamethasone (Ozurdex<sup>®</sup>) for up to 3-months. Other non-polymeric approaches have been utilized to reduce the frequency of intravitreal injections. These include FDA-approved Brolucizumab  $({\rm Brolu}^{(\rm I\!\!R)}),$  a humanized monoclonal single-chain variable fragment which has a longer duration of effect than current anti-VEGF compounds, and the Port Delivery System<sup>(R)</sup> (PDS), a surgically implanted drug reservoir which allows the slow administration of ranibizumab over time. However, Brolucizumab is potentially associated with higher

rates of intraocular inflammation, such as retinal vasculitis (36), while PDS requires an implantation surgical procedure, might be associated with increased risks of sight-threatening complications (37).

A key challenge in developing polymeric hydrogels suitable for sustained anti-VEGF delivery, is the preservation of antibody bioactivity during both hydrogel formulation and biodegradation. For instance, most hydrogels require chemical cross-linking agents for gelation. This can inadvertently result in drug inactivation. Thermogels, on the other hand, rely on small changes in temperature to induce physical cross-linking for gelation to occur, thus enabling preservation of anti-VEGF bioactivity. Indeed, we demonstrated in our EPC thermogel system, that sustained release of bevacizumab or aflibercept is achieved over 40 days, a duration greater than standard current clinical treatment interval. Furthermore, aflibercept released from biodegradable EPC gel retained its bioactivity and was capable of inhibiting vascular leakage in a rabbit choroidal neovascularisation model (38), suggesting the additional utility of the thermogel as a sustained drug release platform in the posterior segment of the eye.

Nano-micelles are globular structures which comprise of an internal hydrophobic fatty acyl chain and external hydrophilic polar head. The discovery of a topically administered nanomicellar system to deliver biologics to the retina-is the holy grail in ocular therapeutics-as it can overcome the multiple sight-threatening complications associated with invasive intravitreal injections. However, multiple static and dynamic ocular barriers between the cornea and the retina, prevent drugs from attaining a therapeutic concentration at the retina sufficient for disease control (39-42). A nanomicelle formulation of the EPC polymer is capable of topically delivering aflibercept to the retina of laser-induced choroidal neovascularisation (CNV) murine models. A single drop of the compound achieved a drug concentration in the murine vitreous that was above the clinically significant concentration required to inhibit VEGF activity. Most importantly, EPC micelle alone seems to have intrinsic anti-angiogenic properties, which works synergistically with aflibercept to result in CNV regression in murine models (14). Further characterization of the pharmacokinetics of EPC nano-micelles is currently underway. Topical compounds, if successful, have the potential to reduce the clinical burden of long-term clinical visits required for invasive intravitreal therapy.

### Future perspective: Biomaterials in retinal gene therapy for inherited retinal degenerations (IRDs)

Treatment options for IRDs are limited. Most of these patients eventually become visually impaired, relying on visual aids and rehabilitation for daily activities. To date, only 1

gene therapy exists for a specific IRD. Voretigene neparvovec (Luxturna<sup>(K)</sup>) is an adeno-associated serotype 2-delivered gene therapy for patients with biallelic RPE-65 mediated IRDs (43). RPE-65 mutations are usually associated with leber congenital amaurosis or retinitis pigmentosa. Since the FDA approval of this drug, many other novel gene therapeutic candidates have emerged. A key hurdle to developing a successful gene therapeutic is the method of gene delivery. Gene delivery platforms can be broadly classified into viral and non-viral methods (44). Viral methods are prevalently utilized as the adeno-associated viral (AAV) vector has been proven to be safe and effective through the approval of voretigene neparvovec. However, viral vectors have inherent limitations. The AAV vector has a packaging capacity of approximately 4.7kb (45). Hence, it is challenging to deliver larger genes such as ABCA4 and EYS which are implicated in common IRDs such as Stargardt disease and retinitis pigmentosa (46, 47). Dual and even triple AAV strategies have been proposed but have suffered from poor transfection efficiency (48). Other viruses with larger packaging capacities may not demonstrate the same tropism and safety profile as AAV.

Cationic polymers are effective non-viral methods of gene delivery. Polyethylenimine (PEI) has demonstrated effective transfection of various human cell lines through the protonsponge effect (49). However, the use of PEI is limited by its biocompatibility. Attempted strategies to improve biocompatibility include branching of PEI and conjugation with other polymers. For instance, Kurosaki et al. demonstrated that cationic complexes of DNA/PEI, when coated with ypolyglutamic acid or chondroitin sulfate, was capable of invivo gene delivery when administered intravitreally into eyes of mice (15). Natural polymers, which may have better biocompatibility profiles, have also been trialed as gene delivery vectors. Liposomes, which are vesicles of lipid with 1 or more phospholipid bilayers enclosing an aqueous core have also demonstrated the ability to deliver the RPE-65 gene into knock out mice (16). Lipid nanoparticles, which are also lipid nanoformulations, may not necessarily have phospholipid bilayers that liposomes have. Solid lipid nanoparticles, when conjugated with dextran have shown the ability to achieve a transfection efficiency of 50% in RPE cells (50). These successes highlight the potential of using polymeric biomaterials for non-viral gene delivery, especially since the use of viral vectors also incurs enormous manufacturing costs.

# Future perspective: Biomaterials in cellular therapy for retinal degenerations

Unfortunately, gene therapy is only viable for early stage IRDs. For patients who have undergone significant disease progression, retinal stem cell therapeutics hold greater promise. Since the proof-of-concept clinical trial by Schwartz et al. on embryonic stem cell (ES) derived retinal cell transplantation, research in this field has evolved tremendously (51). Many factors that contribute to transplantation success have been studied. These include cell sources, methods of delivery, degree of cellular maturation and the need for immunosuppression (52).

Currently, retinal cellular therapeutics can be classified into either RPE or photoreceptor transplantation. RPE is a cell monolayer sitting beneath the photoreceptors and above Bruch's membrane, thus there is a need to use either biostable or biodegradable scaffolds as a cell carrier to support long term function of the transplanted RPE cells. Two types of biostable scaffolds, polyethylene terephthalate (PET) (17) and parylene (53), have been successfully used for RPE cell monolayer transplants in clinical trials in AMD patients. A PLGA-based electrospun nanofibrous scaffolds with RPE patch were tested for safety and efficiency in Royal College of Surgeons (RCS) rats and pigs (18). One should be cautious in using biodegradable scaffolds in patients, as the by-products of biodegradation might be toxic to retinal cells. Moreover, bio-safety of scaffolds and its ability to support graft survival has been proven (54, 55), further studies are required to determine the scaffold's role in establishing visual recovery.

In photoreceptor transplantation, the goal is to prevent remaining healthy photoreceptors from undergoing further degeneration and eventual apoptosis (56). This can be achieved by transplanting photoreceptor cells of varying cellular maturity. Many studies have suggested that the orientation of transplanted photoreceptors and how they interface with host photoreceptor cells play a crucial role in post-transplant survival and integration (57). Thus, a polymeric substrate supporting a monolayer of transplanted photoreceptors can potentially achieve this, and promote inter-digitation between transplanted photoreceptor outer segments and endogenous RPE microvilli (58).

# **Translational hurdles**

Polymeric biomaterials hold great potential for treating many retinal diseases. However, significant hurdles have to be overcome to fully achieve clinical translation.

Firstly, better characterization of polymer-tissue relationship is required. Polymer development is largely based on an iterative process which requires significant resources, time and intuition. Increasingly, the field is developing machine-learning tools to predict properties of polymers in-silico (59). This data-driven approach may enable rational polymer design based on the required function in the tissue type the polymer interacts with.

Secondly, the ocular pharmacokinetics of novel polymers have to be explored. Important aspects such as biodistribution, metabolism and excretion mechanisms can affect the biocompatibility of the polymer significantly. While the systemic pharmacokinetics of polymers like PEG and PCL may have been established previously, their effects on local ocular tissue structure and function still needs to be studied in order to better understand the safety and clearance path of the byproducts while polymers are degrading (54).

Thirdly, manufacturing considerations have to be addressed for successful translation. A large part of the translation process is identifying a manufacturing process that is scalable, maintains the material's original physico-chemical properties and meets the sterility requirements of regulatory agencies. Most often, these aspects are inter-related. For instance, adjusting a polymer's structure to increase the mechanical strength can lead to the reduction of the material's clarity. Furthermore, common sterilization methods that are recognized by regulatory authorities are often physico-chemical methods involving heat, radiation and chemicals such as ethylene oxide, which may inadvertently alter the physico-chemical properties of polymers, rendering them unsuitable. Moreover, due to the complexity of the polymer synthesis processes, adequate monitoring and quality control is required to ensure functionality of final polymer product. As such, the final polymer synthesis process optimized for industry manufacturing, may be very different from the initial conceptualized process laid out in the research laboratory.

### Conclusion

Biomaterials have played significant roles in ophthalmology including intraocular lenses and glaucoma drainage devices. The applications of biomaterials are rapidly expanding, especially for polymeric biomaterials. Today, many polymeric biomaterials are being studied to address the challenges of treating retinal diseases. These applications range from endotamponade agents in retinal detachment surgery, drug delivery for retinal neovascular diseases and gene/cell delivery carriers. However, many translational hurdles still exist. Future work should

### References

1. Zuo W, Yu L, Lin J, Yang Y, Fei Q. Properties improvement of titanium alloys scaffolds in bone tissue engineering: a literature review. *Ann Transl Med.* (2021) 9:1259. doi: 10.21037/atm-20-8175

2. Cheah CW, Al-Namnam NM, Lau MN, Lim GS, Raman R, Fairbairn P, et al. Synthetic material for bone, periodontal, and dental tissue regeneration: where are we now, and where are we heading next? *Materials (Basel).* (2021) 14. doi: 10.3390/ma14206123

3. Keino H, Horie S, Sugita S. Immune privilege and eye-derived t-regulatory cells. J Immunol Res. (2018) 2018:1679197. doi: 10.1155/2018/1679197

4. Williams HP. Sir Harold Ridley's vision. Br J Ophthalmol. (2001) 85:1022–3. doi: 10.1136/bjo.85.9.1022

5. Asbell PA, Dualan I, Mindel J, Brocks D, Ahmad M, Epstein S. Age-related cataract. Lancet. (2005) 365:599–609. doi: 10.1016/S0140-6736(05)70803-5

focus on understanding the ocular pharmacokinetics of novel polymeric biomaterials and establishing synthesis methods that are suitable for large-scale manufacturing.

## Author contributions

Project conceptualization: IS and XS. Data collection: IS and CO. Manuscript writing: IS, CO, ZL, and XS. Project supervision: XS. All authors contributed to the article and approved the submitted version.

### Funding

This work was supported by the National Research Foundation (NRF), Singapore, under its Competitive Research Programme (CRP) (NRF-CRP21-2018-00103), Agency for Science, Technology and Research (A\*STAR), Singapore, and HMBS Domain under its Industry Alignment Fund Pre-Positioning (IAF-PP) programme (H20/H7/a0/034) awarded to XS.

### **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

### Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

6. Lloyd AW, Faragher RG, Denyer SP. Ocular biomaterials and implants. Biomaterials. (2001) 22:769–85. doi: 10.1016/S0142-9612(00)00237-4

7. Karayilan M, Clamen L, Becker ML. Polymeric materials for eye surface and intraocular applications. *Biomacromolecules*. (2021) 22:223–61. doi: 10.1021/acs.biomac.0c01525

8. Patel S, Pasquale LR. Glaucoma drainage devices: a review of the past, present, and future. *Semin Ophthalmol.* (2010) 25:265–70. doi: 10.3109/08820538.2010.518840

9. Baino F, Perero S, Ferraris S, Miola M, Balagna C, Verné E, et al. Biomaterials for orbital implants and ocular prostheses: overview and future prospects. *Acta Biomater.* (2014) 10:1064–87. doi: 10.1016/j.actbio.2013.12.014

10. Gopinath V, Kamath SM, Priyadarshini S, Chik Z, Alarfaj AA, Hirad AH. Multifunctional applications of natural polysaccharide starch and

cellulose: An update on recent advances. *Biomed Pharmacother*. (2021) 146:112492. doi: 10.1016/j.biopha.2021.112492

11. Higashide T, Sugiyama K. Use of viscoelastic substance in ophthalmic surgery - focus on sodium hyaluronate. *Clin Ophthalmol.* (2008) 2:21–30. doi: 10.2147/OPTH.S1439

12. Liu Z, Liow SS, Lai SL, Alli-Shaik A, Holder GE, Parikh BH, et al. Retinal-detachment repair and vitreous-like-body reformation via a thermogelling polymer endotamponade. *Nat Biomed Eng.* (2019) 3:598–610. doi: 10.1038/s41551-019-0382-7

13. Parikh BH, Liu Z, Blakeley P, Lin Q, Singh M, Ong JY, et al. A bio-functional polymer that prevents retinal scarring through modulation of NRF2 signalling pathway. *Nat Commun.* (2022) 13:2796. doi: 10.1038/s41467-022-30474-6

14. Zhao X, Seah I, Xue K, Wong W, Tan QSW, Ma X, et al. Antiangiogenic nanomicelles for the topical delivery of aflibercept to treat retinal neovascular disease. *Adv Mater*. (2021) 2021:e2108360. doi: 10.1002/adma.202108360

15. Kurosaki T, Uematsu M, Shimoda K, Suzuma K, Nakai M, Nakamura T, et al. Ocular gene delivery systems using ternary complexes of plasmid DNA, polyethylenimine, and anionic polymers. *Biol Pharm Bull.* (2013) 36:96–101. doi: 10.1248/bpb.b12-00728

16. Rajala A, Wang Y, Zhu Y, Ranjo-Bishop M, Ma JX, Mao C, et al. Nanoparticleassisted targeted delivery of eye-specific genes to eyes significantly improves the vision of blind mice in vivo. *Nano Lett.* (2014) 14:5257–63. doi: 10.1021/nl502275s

17. da Cruz L, Fynes K, Georgiadis O, Kerby J, Luo YH, Ahmado A, et al. Phase 1 clinical study of an embryonic stem cell-derived retinal pigment epithelium patch in age-related macular degeneration. *Nat Biotechnol.* (2018) 36:328–37. doi: 10.1038/nbt.4114

18. Sharma R, Khristov V, Rising A, Jha BS, Dejene R, Hotaling N, et al. Clinical-grade stem cell-derived retinal pigment epithelium patch rescues retinal degeneration in rodents and pigs. *Sci Transl Med.* (2019) 11. doi: 10.1126/scitranslmed.aat5580

19. Feng X, Li C, Zheng Q, Qian XG, Shao W, Li Y, et al. Risk of silicone oil as vitreous tamponade in pars plana vitrectomy: a systematic review and meta-analysis. *Retina*. (2017) 37:1989–2000. doi: 10.1097/IAE.000000000001553

20. Lin Q, Lim JYC, Xue K, Su X, Loh XJ. Polymeric hydrogels as a vitreous replacement strategy in the eye. *Biomaterials.* (2021) 268:120547. doi: 10.1016/j.biomaterials.2020.120547

21. Su X, Tan MJ Li Z, Wong M, Rajamani L, Lingam G, et al. Recent progress in using biomaterials as vitreous substitutes. *Biomacromolecules*. (2015) 16:3093– 102. doi: 10.1021/acs.biomac.5b01091

22. Xue K, Liu Z, Jiang L, Kai D, Li Z, Su X, et al. A new highly transparent injectable PHA-based thermogelling vitreous substitute. *Biomater Sci.* (2020) 8:926–36. doi: 10.1039/C9BM01603A

23. Schramm C, Spitzer MS, Henke-Fahle S, Steinmetz G, Januschowski K, Heiduschka P, et al. The cross-linked biopolymer hyaluronic acid as an artificial vitreous substitute. *Invest Ophthalmol Vis Sci.* (2012) 53:613–21. doi:10.1167/iovs.11-7322

24. Feng S, Chen H, Liu Y, Huang Z, Sun X, Zhou L, et al. A novel vitreous substitute of using a foldable capsular vitreous body injected with polyvinylalcohol hydrogel. *Sci Rep.* (2013) 3:1838. doi: 10.1038/srep01838

25. Hayashi K, Okamoto F, Hoshi S, Katashima T, Zujur DC Li X, et al. Fast-forming hydrogel with ultralow polymeric content as an artificial vitreous body. *Nat Biomed Eng.* (2017) 1:0044. doi: 10.1038/s41551-017-0044

26. El-Husseiny HM, Mady EA, Hamabe L, Abugomaa A, Shimada K, Yoshida T, et al. Smart/stimuli-responsive hydrogels: Cutting-edge platforms for tissue engineering and other biomedical applications. *Mater Today Bio.* (2022) 13:100186. doi: 10.1016/j.mtbio.2021.100186

27. Zhang K, Xue K, Loh XJ. Thermo-responsive hydrogels: from recent progress to biomedical applications. *Gels.* (2021) 7. doi: 10.3390/gels7030077

28. Santhanam S, Liang J, Struckhoff J, Hamilton PD, Ravi N. Biomimetic hydrogel with tunable mechanical properties for vitreous substitutes. *Acta Biomater.* (2016) 43:327–37. doi: 10.1016/j.actbio.2016.07.051

29. Laradji A, Shui YB, Karakocak BB, Evans L, Hamilton P, Ravi N. Bioinspired thermosensitive hydrogel as a vitreous substitute: synthesis, properties, and progress of animal studies. *Materials (Basel).* (2020) 13. doi: 10.3390/ma13061337

30. Xue K, Liu Z, Lin Q, Lim JYC, Tang KY, Wong SI, et al. PCL-based thermogelling polymer: molecular weight effects on its suitability as vitreous tamponade. ACS Appl Bio Mater. (2020) 3:9043–53. doi: 10.1021/acsabm.0c01266

31. Kwon OW, Song JH, Roh MI. Retinal detachment and proliferative vitreoretinopathy. *Dev Ophthalmol.* (2016) 55:154–62. doi: 10.1159/000438972

32. Wong WL, Su X, Li X, Cheung CM, Klein R, Cheng CY, et al. Global prevalence of age-related macular degeneration and disease burden projection for

2020 and 2040: a systematic review and meta-analysis. *Lancet Glob Health.* (2014) 2:e106-16. doi: 10.1016/S2214-109X(13)70145-1

33. Kim LA, D'Amore PA, A. brief history of anti-VEGF for the treatment of ocular angiogenesis. *Am J Pathol.* (2012) 181:376-9. doi: 10.1016/j.ajpath.2012.06.006

34. Luu KT, Seal J, Green M, Winskill C, Attar M. Effect of anti-VEGF therapy on the disease progression of neovascular age-related macular degeneration: a systematic review and model-based meta-analysis. *J Clin Pharmacol.* (2021). doi: 10.1002/jcph.2002

35. Sampat KM, Garg SJ. Complications of intravitreal injections. Curr Opin Ophthalmol. (2010) 21:178–83. doi: 10.1097/ICU.0b013e328338679a

36. Sharma A, Kumar N, Parachuri N, Sahyoun JY, Kuppermann BD, Bandello F. Brolucizumab – termination of 4 weekly trials – rebalancing the immunogenicity risk. *Expert Opin Biol Ther.* (2022). doi: 10.1080/14712598.2022.2028772

37. Sharma A, Parachuri N, Kumar N, Kuppermann BD, Bandello F. The Port Delivery System with ranibizumab-journey of mitigating vitreous hemorrhage. *Eye* (*Lond*). (2021). doi: 10.1038/s41433-021-01830-5

38. Xue K, Zhao X, Zhang Z, Qiu B, Tan QSW, Ong KH, et al. Sustained delivery of anti-VEGFs from thermogel depots inhibits angiogenesis without the need for multiple injections. *Biomater Sci.* (2019) 7:4603–14. doi: 10.1039/C9BM01049A

39. de Cogan F, Hill LJ, Lynch A, Morgan-Warren PJ, Lechner J, Berwick MR, et al. Topical delivery of anti-VEGF drugs to the ocular posterior segment using cell-penetrating peptides. *Invest Ophthalmol Vis Sci.* (2017) 58:2578–90. doi:10.1167/iovs.16-20072

40. Rodrigues GA, Lutz D, Shen J, Yuan X, Shen H, Cunningham J, et al. Topical drug delivery to the posterior segment of the eye: addressing the challenge of preclinical to clinical translation. *Pharm Res.* (2018) 35:245. doi: 10.1007/s11095-018-2519-x

41. Agrahari V, Mandal A, Trinh HM, Joseph M, Ray A, Hadji H, et al. A comprehensive insight on ocular pharmacokinetics. *Drug Deliv Transl Res.* (2016) 6:735–54. doi: 10.1007/s13346-016-0339-2

42. Mandal A, Pal D, Agrahari V, Trinh HM, Joseph M, Mitra AK. Ocular delivery of proteins and peptides: Challenges and novel formulation approaches. *Adv Drug Deliv Rev.* (2018) 126:67–95. doi: 10.1016/j.addr.2018.01.008

43. Aoun M, Passerini I, Chiurazzi P, Karali M, De Rienzo I, Sartor G, et al. Inherited retinal diseases due to rpe65 variants: from genetic diagnostic management to therapy. *Int J Mol Sci.* (2021) 22. doi: 10.3390/ijms22137207

44. Pan X, Veroniaina H, Su N, Sha K, Jiang F, Wu Z, et al. Applications and developments of gene therapy drug delivery systems for genetic diseases. *Asian J Pharm Sci.* (2021) 16:687–703. doi: 10.1016/j.ajps.2021.05.003

45. Wu Z, Yang H, Colosi P. Effect of genome size on AAV vector packaging. *Mol Ther.* (2010) 18:80–6. doi: 10.1038/mt.2009.255

46. Garcia-Delgado AB, Valdes-Sanchez L, Morillo-Sanchez MJ, Ponte-Zuñiga B, Diaz-Corrales FJ, de la Cerda B. Dissecting the role of EYS in retinal degeneration: clinical and molecular aspects and its implications for future therapy. *Orphanet J Rare Dis.* (2021) 16:222. doi: 10.1186/s13023-021-01843-z

47. Molday RS, Garces FA, Scortecci JF, Molday LL. Structure and function of ABCA4 and its role in the visual cycle and Stargardt macular degeneration. *Prog Retin Eye Res.* (2022) 89:101036. doi: 10.1016/j.preteyeres.2021.101036

48. Chamberlain K, Riyad JM, Weber T. Expressing transgenes that exceed the packaging capacity of adeno-associated virus capsids. *Hum Gene Ther Methods*. (2016) 27:1–12. doi: 10.1089/hgtb.2015.140

49. Sabin J, Alatorre-Meda M, Miñones J, Domínguez-Arca V, Prieto G New insights on the mechanism of polyethylenimine transfection and their implications on gene therapy and DNA vaccines. *Colloids Surf B Biointerfaces*. (2022) 210:112219. doi: 10.1016/j.colsurfb.2021.112219

50. Delgado D. del Pozo-Rodríguez A, Solinís M, Avilés-Triqueros M, Weber BH, Fernández E, et al. Dextran and protamine-based solid lipid nanoparticles as potential vectors for the treatment of X-linked juvenile retinoschisis. *Hum Gene Ther.* (2012) 23:345–55. doi: 10.1089/hum.2011.115

51. Schwartz SD, Hubschman JP, Heilwell G, Franco-Cardenas V, Pan CK, Ostrick RM, et al. Embryonic stem cell trials for macular degeneration: a preliminary report. *Lancet.* (2012) 379:713–20. doi: 10.1016/S0140-6736(12)60028-2

52. German OL, Vallese-Maurizi H, Soto TB, Rotstein NP, Politi LE. Retina stem cells, hopes and obstacles. *World J Stem Cells.* (2021) 13:1446-79. doi: 10.4252/wjsc.v13.i10.1446

53. Kashani AH, Uang J, Mert M, Rahhal F, Chan C, Avery RL, et al. Surgical method for implantation of a biosynthetic retinal pigment epithelium monolayer for geographic atrophy: experience from a phase 1/2a study. *Ophthalmol Retina*. (2020) 4:264–73. doi: 10.1016/j.oret.2019.09.017

54. Luo Z, Xian B, Li K, Li K, Yang R, Chen M, et al. Biodegradable scaffolds facilitate epiretinal transplantation of hiPSC-derived retinal neurons in nonhuman primates. *Acta Biomater.* (2021) 134:289–301. doi: 10.1016/j.actbio.2021.07.040

55. Becker S, Eastlake K, Jayaram H, Jones MF, Brown RA, McLellan GJ, et al. Allogeneic transplantation of müller-derived retinal ganglion cells improves retinal function in a feline model of ganglion cell depletion. *Stem Cells Transl Med.* (2016) 5:192–205. doi: 10.5966/sctm.2015-0125

56. Singh MS, Park SS, Albini TA, Canto-Soler MV, Klassen H, MacLaren RE, et al. Retinal stem cell transplantation: Balancing safety and potential. *Prog Retin Eye Res.* (2020) 75:100779. doi: 10.1016/j.preteyeres.2019.100779

57. Gasparini SJ, Llonch S, Borsch O, Ader M. Transplantation of photoreceptors into the degenerative retina: Current state and future perspectives. *Prog Retin Eye Res.* (2019) 69:1–37. doi: 10.1016/j.preteyeres.2018.11.001

58. Jung YH, Phillips MJ, Lee J, Xie R, Ludwig AL, Chen G, et al. 3D Microstructured scaffolds to support photoreceptor polarization and maturation. *Adv Mater.* (2018) 30:e1803550. doi: 10.1002/adma. 201803550

59. Suwardi A, Wang F, Xue K, Han MY, Teo P, Wang P, et al. Machine learning-driven biomaterials evolution. *Adv Mater.* (2022) 34:e2102703. doi: 10.1002/adma.202102703