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## Activin promotes oocyte development in ovine preantral follicles *in vitro*

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### Abstract

Activins have been implicated as important regulating factors for many reproductive processes. The aim of this study was to determine the effect of activin A on the development of ovine preantral follicles *in vitro*. Mechanically isolated preantral follicles ( $161 \pm 2$  microm) were cultured for 6 days in the presence of human recombinant activin A (0, 10 and 100 ng/ml). Half of the medium was replaced every second day and follicle diameters were measured. Conditioned medium was subsequently analysed for oestradiol content using a delayed enhancement lanthanide fluorometric immunoassay (DELFLIA). At the end of the culture period, follicles were fixed and processed for histology, after which oocyte diameter and granulosa cell death were measured. There was significant follicle growth over 6 days in all groups ( $p < 0.001$ ). Activin, at both concentrations, increased follicle growth over control levels by Day 6 ( $p < 0.05$ ). Oocyte diameters were also significantly increased by Day 6 of culture in all groups ( $p < 0.05$ ), with 100 ng/ml activin increasing oocyte diameter over control levels ( $p < 0.05$ ). Activin, at both concentrations, increased oestradiol production on Day 2 of culture, but this increase was not sustained during the culture period. Moreover, activin did not have any effect on antrum formation or follicle survival. In conclusion, activin promoted ovine preantral follicle and oocyte growth *in vitro*, but did not accelerate follicle differentiation over a six-day culture period. These results support a paracrine role for activin A during early oocyte and follicular development.

### Introduction

Over the last two decades, culture systems have been developed with the aim of growing oocytes from the earliest follicular stages through to maturation and fertilisation. The viability of rodent systems has been demonstrated by the production of live offspring from *in vitro* grown oocyte-granulosa cell complexes from murine preantral follicles [1] and from the culture of whole preantral follicles [2]. However, progress has been slow in developing these techniques for species with a long follicular growth period, such as humans and domestic species.

There are many technical reasons for the lack of progress in these species, but the major problem is that we have very little knowledge of how the oocyte acquires developmental competence during its growth within the follicle. At present the major benefits of culture systems that support the growth and development of immature oocytes are in advancing our knowledge of the co-ordination of oocyte development and granulosa cell differentiation, and understanding the developmental regulation of auto-crine/paracrine factors controlling these processes.

The TGF- $\beta$  superfamily is made up of a number of proteins with the potential to act as intraovarian regulators of ovarian function [3]. These are growth and differentiation factors such as growth differentiation factor-9 (GDF-9), mullerian inhibitory substance (MIS), activins and inhibins [4–6]. Activins are disulphide-linked dimeric glycoproteins, and the subunits inhibin  $\alpha$ , inhibin/activin  $\beta$ A and inhibin/activin  $\beta$ B, together with the activin receptors type I, IIA and IIB are expressed in ovarian cells during follicular development [7,8]. Activin A is composed of two  $\beta$ A subunits, and a role for this factor during early follicular development in sheep is supported by the presence of these peptides in oocytes and granulosa cells from the primordial stage onwards [9]. Activins are thought to play an autocrine/paracrine role in controlling early follicular development by promoting follicular growth and differentiation [5,10].

The role of activin during early follicular development has been reported in rodent species, where activin has been shown to both promote [11,12] and inhibit [13] preantral follicle growth and development. These ambiguous actions may be due to developmental regulation. In domestic ruminant species, the role(s) of activin during early follicle development has not been established. Therefore the aims of this study were to determine the effects of activin A on oocyte and granulosa cell development within ovine preantral follicles using a serum free culture system.

## Materials and Methods

### Preantral Follicle Isolation

Ovine ovaries from random stages of the oestrous cycle were obtained from an abattoir and transported at 25–30°C in M199 (HEPES buffered) media (GIBCO BRL, Life Technologies Ltd, Paisley, Renfrewshire, UK) supplemented with sodium pyruvate (2 mM), glutamine (2 mM), BSA (3 mg/ml), penicillin G (75  $\mu$ g/ml) and streptomycin (50  $\mu$ g/ml) (all chemicals from Sigma Chemicals, Poole, Dorset, UK unless otherwise stated). In a laminar flow hood, ovaries without large follicles or corpora lutea on the surface were rinsed with 70% ethanol, and fine slices of ovarian cortex were taken using a scalpel and placed in dissection medium [Leibovitz's medium (GIBCO BRL, Life Technologies Ltd.) supplemented with sodium pyruvate (2 mM), glutamine (2 mM), BSA (Fraction V, 3 mg/ml), penicillin G (75  $\mu$ g/ml) and streptomycin (50  $\mu$ g/ml)]. Preantral follicles ( $161 \pm 2$   $\mu$ m) were isolated from the cortical slices under a dissecting microscope using 25G needles. Follicles with an intact basement membrane and even distribution of granulosa and theca layers were selected for culture. The entire process from collection of material from the abattoir to the start of culture was always less than six hours.

### Preantral Follicle Culture

For the control group, preantral follicles ( $n = 34$ ) were cultured individually in 96 well plates (Bibby Sterilin Ltd, Stone, Staffs, UK) in 250  $\mu$ l of culture medium (McCoy's 5a medium with bicarbonate supplemented with HEPES (20 mM), BSA (0.1 %), L-glutamine (3 mM), penicillin (100 IU/ml), streptomycin (0.1 mg/ml), transferrin (2.5  $\mu$ g/ml), selenium (4 ng/ml), insulin (10 ng/ml) and L-ascorbic acid, sodium salt (50  $\mu$ g/ml), all obtained from Sigma Chemicals. Treatments added to the control medium were recombinant human activin A (R&D Systems) at a concentration of 10 ng/ml ( $n = 33$ ), or 100 ng/ml ( $n = 32$ ). Isolated preantral follicles were assigned randomly to the treatment groups. Plates were incubated for 6 days in a sterile humidified air atmosphere with 5 % CO<sub>2</sub> at 37°C. Each set of cultures ( $n = 5$ ) took place under identical conditions. Follicle diameters were measured using a crossed micrometer (basement membrane to basement membrane) under the dissection microscope on days 0, 2, 4 and 6. Half of the medium was replaced every second day, and this conditioned medium was stored at -70°C for subsequent analysis of oestradiol content. Any follicles that had lost basement membrane integrity during the first 24 hours of culture were likely to have been damaged during isolation, and were excluded from any further analysis.

### Detection of Oestradiol in Culture Medium using a Delayed Enhancement Lanthanide Fluorometric Immunoassay (DELFLIA)

Medium from preantral follicles cultured in control medium ( $n = 14$ ), or in the presence of activin A (10 ng/ml,  $n = 15$ ; 100 ng/ml,  $n = 15$ ) was analysed for oestradiol content. Oestradiol was biotinylated by standard procedures using 17 $\beta$ -oestradiol-3-(O-carboxy) methylether and EZ-Link biotin hydrazide (Pierce Warriner UK Ltd, 44 Upper Northgate Street Chester CH1 4EF). Nunc-Immuno Maxisorp 96 well plates were coated with donkey anti-sheep serum by incubating overnight at 4°C in the presence of donkey anti-sheep serum (250  $\mu$ g/ml) made up in carbonate buffer pH9.6 (100  $\mu$ l/well). The primary antibody was raised in sheep against 17 $\beta$ -estradiol 6-(O-carboxymethyl)-oxime: BSA [14].

Biotinylated oestradiol, follicle conditioned medium, oestradiol standards, and a 1:50,000 dilution of primary antibody, made up in 200  $\mu$ l of assay buffer, was added to the pre-coated wells of the microtitre plate. The assay buffer consisted of Tris buffer (50 mmol/l; pH 7.8) supplemented with sodium chloride (150 mmol/l), bovine gamma globulin (1%, w/v), Tween-20 (0.01%; v/v), thimerosal (0.0008%; w/v) and diethylenetriaminepenta-acetic acid (0.1 moles/l). After incubating the plates overnight at 4°C they were washed (4 $\times$ ) in a wash buffer consisting of Tris buffer (50 mmol/l; pH 7.8)

supplemented with sodium chloride (150 mmol/l), Tween-20 (0.01%; v/v) and thimerosal (0.0008%; w/v) before adding 100 µl of assay buffer containing 100 ng/ml europium labelled streptavidin (Perkin-Elmer Life Sciences address) followed by incubation at room temperature for 1 h with shaking. The plates were washed 4× in wash buffer before addition of 200 µl of DELFIA enhancement solution (Perkin-Elmer Life Sciences) to each well of the microtitre plate, and incubated for a further 5 mins with shaking at room temperature. The plates were analysed on a Victor 2 Multilabel Counter (Perkin-Elmer Life Sciences) by time resolved fluorimetry. The emission and excitation wavelengths were 615 nm and 340 nm respectively with a time delay of 400 µs. The inter- and intra-assay coefficients of variation were 13.2 and 9.6 % respectively. The minimum detectable level was 8.5 pg oestradiol per well.

#### **Histological Assessment**

At the end of the culture period, follicles were fixed overnight in Bouin's solution and dehydrated in ethanol (70%, 80%, 90%, 100%). Follicles were visualised during processing by addition of eosin to the 70% ethanol. Absolute ethanol was replaced with cedar wood oil for a minimum of 24 hours, then the oil was cleared from the follicles using toluene for 30 min. Follicles were embedded in paraffin wax (60°C), with changes every hour for 4 h to remove all traces of toluene. The samples were sectioned (6 µm) and mounted on gelatin-coated slides, and allowed to dry overnight at 37°C before staining with Haematoxylin and Eosin.

Histological measurements and observations were made under the light microscope with a crossed micrometer (Graticules Ltd.). The section containing the oocyte nucleolus, or if this was absent, the largest cross-section of the oocyte was used for observations and measurements. Oocyte diameters were measured, and granulosa cell death within a follicle was assessed by counting the number of pyknotic nuclei and expressing them as a percentage of the total number of granulosa cells. Only follicles that had maintained basement membrane integrity during culture were used for further analysis.

#### **Statistical Analyses**

Mean follicle diameter and oestradiol production on every second culture day were compared between experimental groups using a one-way ANOVA, with subsequent 2-sample t-tests to allow for individual comparisons between groups. The percentage of pyknotic granulosa cell nuclei were also compared between treatment groups using ANOVA and t-tests. A general linear model was applied to all data to take into account any differences between individual cultures. Oocyte diameters from freshly isolated and cultured preantral follicles were com-

pared using a one-way ANOVA, with subsequent 2-sample t-tests.

## **Results**

### **Follicle Growth**

Follicles were cultured for 6 days in control medium (n = 34) or in medium containing activin A (10 ng/ml, n = 33; 100 ng/ml, n = 32). As illustrated in Figure 1a, there was a significant increase in follicular diameter during culture in all treatment groups ( $p < 0.001$ ). At the end of the culture period, there was a significant difference between groups in terms of follicular diameter ( $p < 0.05$ ). Activin, at both concentrations, stimulated follicular growth over control levels ( $p < 0.05$ ).

### **Oestradiol Production**

Conditioned medium from control follicles (n = 14) and activin-treated follicles (10 ng/ml, n = 15; 100 ng/ml, n = 15) was analysed for oestradiol concentration on days 2, 4 and 6 of culture. Activin, at both 10 ng/ml and 100 ng/ml, increased oestradiol production on Day 2 of culture, compared with controls ( $p < 0.01$ ). However, by day 4 and 6 of culture, there was no significant difference in oestradiol production between activin-treated follicles and controls (Figure 1b).

### **Oocyte Growth**

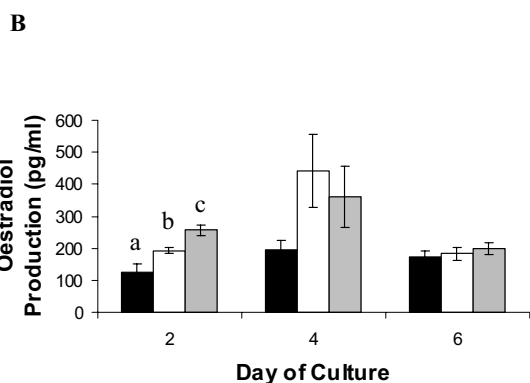
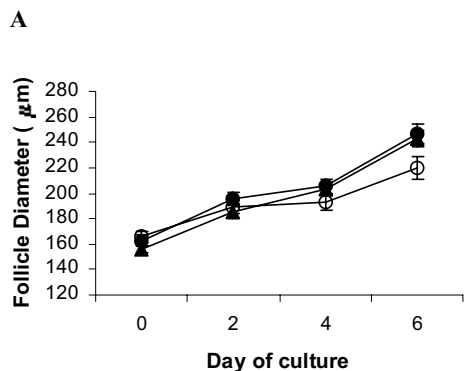
Histological observations were made using freshly isolated follicles (Day 0), and on Day 6 of culture using control follicles (n = 25) and follicles treated with activin A (10 ng/ml, n = 25; 100 ng/ml, n = 25). Oocyte diameter increased from Day 0 to Day 6 in all culture groups ( $p < 0.05$ ) (Figure 2). Moreover, activin, at a concentration of 100 ng/ml, increased oocyte diameter over control levels on Day 6 of culture ( $p < 0.05$ ) (Figure 2).

### **Antrum Formation and Granulosa Cell Death**

From histological observations, no significant differences in the incidence of antrum formation or the percentage of pyknotic granulosa cell nuclei were found between activin-treated follicles and controls on Day 6 of culture (Table 1). Preantral follicles fixed on Day 6 of culture, showing antrum formation (A) and pyknotic nuclei (B), are represented in Figure 3.

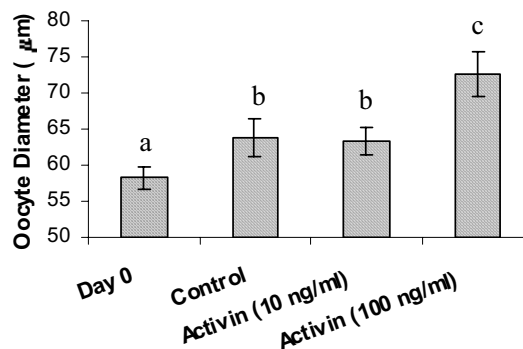
## **Discussion**

Previous *in vitro* studies have failed to produce competent oocytes from preantral follicles in species such as humans and domestic ruminants. Improved knowledge of the role of paracrine factors in regulating early oocyte and follicle development is important for optimisation of culture systems for the production of developmentally competent oocytes. In the current study, a serum-free culture system was used to investigate the effect of activin on early follicular growth, differentiation and survival *in vitro*.



**Figure 1**  
**(A)** Growth of ovine preantral follicles in control medium (open circles) or in the presence of activin A (10 ng/ml, closed circles; 100 ng/ml, triangles). Values are mean ± SEM. Growth is significant between Days 0 and 6 within treatment groups ( $p < 0.001$ ). Activin A, at both concentrations, stimulated follicular growth over control levels by Day 6 of culture ( $p < 0.05$ ). **(B)** Oestradiol production by ovine preantral follicles cultured in control medium (black bars), or in the presence of activin A [10 ng/ml (open bars), 100 ng/ml (striped bars)]. Values are mean ± SEM. Both concentrations of activin increased oestradiol concentration over control levels on Day 2 ( $p < 0.01$ ), but there were no differences between groups on Day 4 and 6 of culture.

In rodents, the precise role of activin during early follicular development is unclear. For example, activin increases the size of preantral follicles and promotes antrum formation in immature mice [12,15]. By contrast, activin does not stimulate growth of preantral follicles in adult mice, and blocks the effect of FSH on follicular growth [15]. Moreover, it has been reported that activin produced by secondary mouse follicles suppresses the growth of pri-



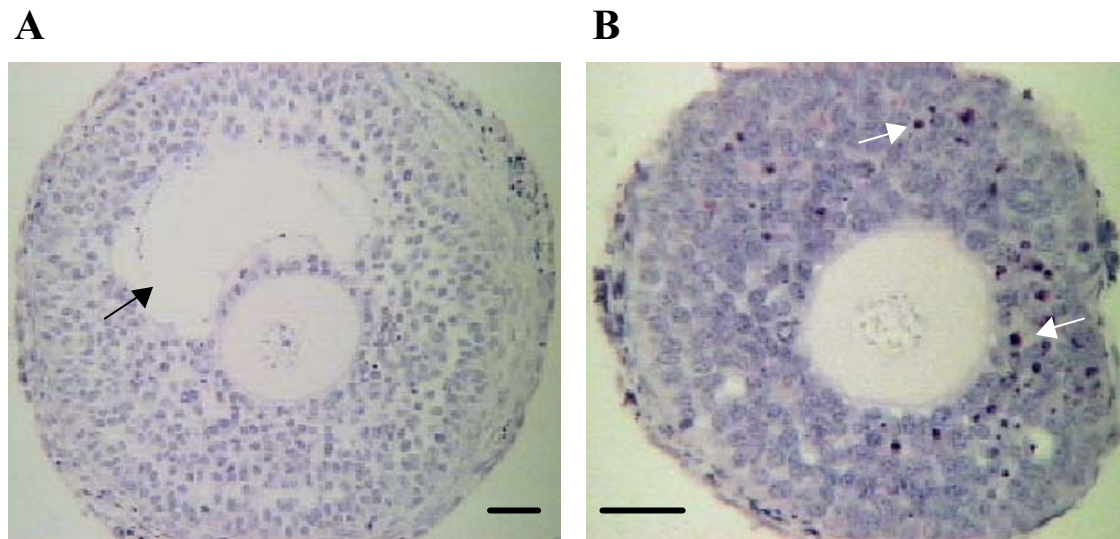
**Figure 2**  
 Oocyte diameters measured on Day 0, and in Day 6 cultured control follicles and follicles treated with activin A. Values are mean ± SEM. Oocyte diameter increased from Day 0 to Day 6 in all culture groups ( $p < 0.05$ ). Activin A, at a concentration of 100 ng/ml, increased oocyte diameter over control levels on Day 6 of culture ( $p < 0.05$ ).

**Table 1: Effect of activin on antrum formation and granulosa cell death on Day 6 of culture. Values are mean ± SEM.**

Treatment	% Follicles with Antrum	% Granulosa Cell Death
Control (n = 25)	25.0	1.4 ± 0.4
Activin A (10 ng/ml) (n = 25)	47.4	0.5 ± 0.1
Activin A (100 ng/ml) (n = 25)	33.3	2.5 ± 1.1

mary follicles *in vitro* [13], and it was hypothesised that a local decline in activin as a result of atresia of secondary follicles may be responsible for promotion of early folliculogenesis in response to circulating FSH [13]. Thus, according to the rodent culture models, there appears to be age- and stage-dependant effects of activin on follicular development. In the present study, medium-sized and large preantral follicles (< 200 µm diameter) were isolated from ovaries obtained from animals of unknown age; therefore investigation into any similar age- or stage-dependent effects of activin in sheep follicles was out-with the scope of this investigation.

TGF-β superfamily members are likely to play a significant role in folliculogenesis in ruminants, as mutations in the TGF-β signalling system have been implicated in the increased ovulation rate observed in Booroola and Inverdale sheep [16,17]. Although activin A has been shown to promote oestradiol synthesis whilst suppressing prog-



**Figure 3**  
 Histological sections on Day 6 of culture representing **(A)** a follicle cultured with activin showing early antrum formation (arrow), and **(B)** a degenerating follicle containing pyknotic nuclei in granulosa cells (arrows). Bar = 20  $\mu\text{m}$ .

terone in ovine granulosa cell culture [18], there is little information on the role of this factor during the early stages of ovine follicle development.

As a member of the TGF- $\beta$  family, activin executes its actions through a group of intracellular signal transducers, the Smad protein family [19]. There is evidence of an activin signal transduction pathway in oocytes, as activin receptors and Smads 2, 4 and 8 are present [20]. Granulosa cells also express activin receptors, as well as Smads 2 and 4 [20]. Thus activin is capable of directly influencing early oocyte and follicle development, as supported by the present study. The presence of receptors on both the oocyte and somatic cells may provide an explanation for the dose-dependent effect of activin on oocyte growth presented here. However, further investigations will be necessary to ascertain activin receptor levels during ovine follicle development *in vitro*.

Ovine preantral follicles have previously been developed to the antral stage in culture medium containing serum and high levels of FSH [21]. In serum-free conditions, ovine oocyte-granulosa complexes from fresh and frozen-thawed tissue have been grown to the antral stage during

long term culture, and these have produced increasing levels of oestradiol [22]. However, these studies have proved to be unsatisfactory in terms of producing oocytes capable of maturation and further development. Due to the protracted nature of preantral follicle development in domestic ruminants, the ideal culture system should accelerate oocyte development, without inducing inappropriate follicular differentiation. Androgens have been shown to increase murine preantral follicle growth and oestradiol secretion [23,24]. However, we have recently shown that androstenedione increases follicular differentiation but decreases oocyte survival within ovine preantral follicles [25]. A recent study using rat preantral follicles found that activin increased FSH-induced follicular inhibin- $\alpha$  expression (a marker of differentiation) during a 72 hour culture period [6]. In contrast, another TGF- $\beta$  family member, MIS, was found to promote follicle growth and proliferation, without potentiating FSH-induced follicular differentiation [6]. In the current investigation, although activin promoted oestradiol production by Day 2 of culture, this increase was not sustained during the subsequent 4 days of culture. It should be noted that growth of ovine preantral and early antral follicles is thought to be independent of gonadotrophins,

and expression of steroidogenic enzymes such as cytochrome P450 aromatase increase during antral follicle development [26]. This would provide an explanation for the low oestradiol levels produced by preantral follicles in the present study. Further investigations into the effect of activin in combination with FSH, using an extended culture system, will be useful to determine the role of activin in steroidogenesis in ovine follicles after antrum formation. As sheep follicles have a much longer growth period compared with rodent follicles, there are likely to be species differences in the role of paracrine factors, including activin, during preantral follicle growth and differentiation.

In mice, a combination of FSH and insulin, which act synergistically to promote differentiation and function of granulosa cells [27–29], causes precocious differentiation of immature murine oocyte-associated granulosa cells in culture [30]. Interestingly, this differentiation was found to be associated with reduced competence of oocytes to undergo fertilisation and pre-implantation development [30]. In the present study, activin was found to have a positive effect on both follicle and oocyte growth after six days. Moreover, activin did not promote inappropriate differentiation, as oestradiol concentrations were not increased over control levels for the majority of the culture period. In addition, activin did not significantly increase the incidence of antrum formation. Future studies employing an extended culture period will be important to assess the developmental potential of oocytes after treatment with activin.

The culture system reported here has previously been shown to maintain bovine preantral follicle survival and basement membrane integrity over six days [31]. Here, using ovine follicles, we have shown that the incidence of granulosa cell death was low across all culture groups. Thus the increased follicular growth observed in the presence of activin was likely to be due to granulosa cell proliferation, rather than a result of increased atresia of control follicles.

In summary, this study has shown that activin promotes ovine preantral follicle and oocyte development over a six day culture period, without accelerating follicular differentiation. Improved knowledge of the species- and stage-specific roles of paracrine factors in the co-ordination of oocyte development and somatic cell differentiation is essential for the development of culture systems capable of supporting oocyte development and acquisition of developmental competence. A technique for obtaining a source of homogeneous mature oocytes from ovine ovaries would provide a model for *in vitro* maturation and fertilisation systems for human oocytes, as well as permitting

investigations into postovulatory and embryonic development.

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